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The ProTide Prodrug Technology: From the Concept to the Clinic

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Abstract

The ProTide technology is a prodrug approach developed for the efficient intracellular delivery of nucleoside analogue monophosphates and monophosphonates. In this approach, the hydroxyls of the monophosphate or monophosphonate groups are masked by an aromatic group and an amino acid ester moiety, which are enzymatically cleaved-off inside cells to release the free nucleoside monophosphate and monophosphonate species. Structurally, this represents the current end-point of an extensive medicinal chemistry endeavour that spans almost three decades. It started from the masking of nucleoside monophosphate and monophosphonate groups by simple alkyl groups and evolved into the sophisticated ProTide system as known today. This technology has been extensively employed in drug discovery and it has already led to the discovery of two FDA-approved (antiviral) ProTides. In this work, we will review the development of the ProTide technology, its application in drug discovery and its role in the improvement of drug delivery and efficacy.

Keywords: ProTide, Prodrug, Nucleoside, Antiviral, Cancer.

1. Introduction

Nucleoside analogues were first introduced a number of decades ago as an effective way of treating various diseases, such as cancer and viral infections like herpes simplex virus (HSV), human cytomegalovirus (HCMV), human immunodeficiency virus (HIV), hepatitis B virus (HBV) and recently also hepatitis C virus (HCV).¹⁻⁶ Currently, there are more than twenty different nucleoside analogues available in the clinic to treat the viral infections and cancer.^{7, 8} These molecules enter the cell with the aid of transporters, such as concentrative nucleoside transporters as well as peptide transporters, or via passive diffusion.^{9, 10} Once inside the cell, the nucleoside analogues are activated by a number of nucleoside and nucleotide kinases which phosphorylate them in a stepwise manner resulting in the formation of the mono-, di- and tri-phosphorylated nucleoside analogue metabolites (Figure 1).^{6, 11}

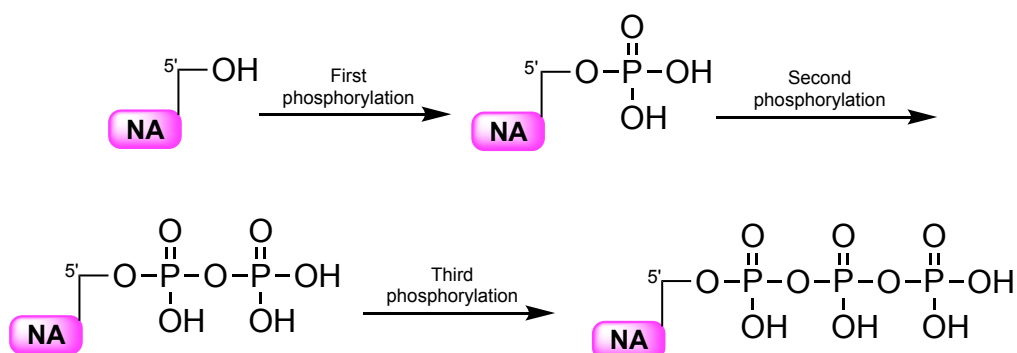


Figure 1. Intracellular activation of therapeutic and experimental nucleoside analogues. NA: nucleoside analogue.

The activated (phosphorylated) antiviral nucleoside analogues exert their therapeutic effects by targeting and inhibiting intracellular enzymes (usually the virus-encoded DNA or RNA polymerases) and/or by being incorporated into the viral nucleic acid chains leading to the termination of the elongation process.^{7, 8, 12} However, there are a number of limitations associated with the administration of nucleoside analogue drugs. Nucleoside analogues are structurally different to natural nucleosides making their phosphorylation by cellular or viral

kinases often inefficient.^{13, 14} This ultimately limits the formation of the active triphosphate metabolites, often the nucleoside analogue active metabolite.¹³ They also often demonstrate poor oral bioavailability due to their low intestinal permeability, as they are usually polar molecules that hinder their transport through the cell border *via* the paracellular route.¹⁵ Additionally, the therapeutic use of many antiviral and anticancer nucleoside analogues is further limited by resistance, which occurs via a number of mechanisms⁸ that include 1) down regulation of the nucleoside kinases responsible for activating nucleoside analogues,¹⁶ 2) depletion of transporters that affect their efficient trafficking into the target cells,¹⁷⁻¹⁹ 3) activation of phosphatases (i.e. 5'-nucleotidases) that dephosphorylate the active metabolites of nucleoside analogues,^{20, 21} and 4) increased catabolic bioconversion of the nucleoside analogues, e.g. (deoxy)cytidine deamination, which may yield inactive versions of some of the nucleoside analogues^{22, 23}.

To overcome these limitations, it was initially suggested that delivering phosphorylated metabolites of the nucleoside analogue would address some of these shortages. This notion was further supported by numerous studies that reported the poor phosphorylation of nucleoside analogues into their nucleoside monophosphates as the rate-limiting step in their *in situ* activation.²⁴⁻²⁷ However, the idea of using nucleoside analogue monophosphates as therapeutics faced two main challenges. First, these compounds would have poor *in vivo* stability due to de-phosphorylation in the blood stream, and second, their transport into cells will be inefficient because of the charged nature of the phosphorylated nucleoside. To address the P-O bond instability, Holý and De Clercq switched the P-O- bonds in the monophosphate group of nucleoside analogues into monophosphonate P-CH₂- bonds.²⁸ This led to improved *in situ* stability, which translated into longer half-lives of the nucleoside monophosphonates as compared to their monophosphates.²⁸ This advance in the design of stable nucleotide therapeutics inspired the discovery of many nucleoside analogue monophosphonates some of which were eventually approved for clinical use as antivirals against HCMV [i.e. cidofovir], HBV [i.e. adefovir and tenofovir], and HIV [i.e. tenofovir] (Figure 2).⁷

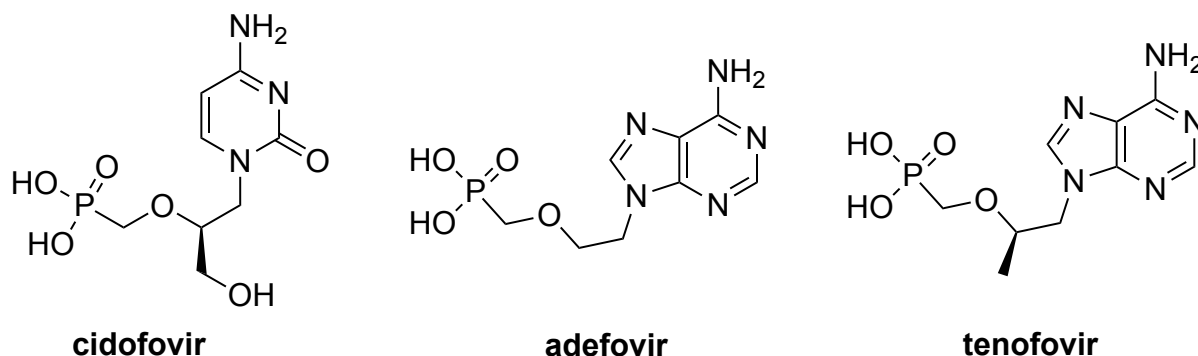
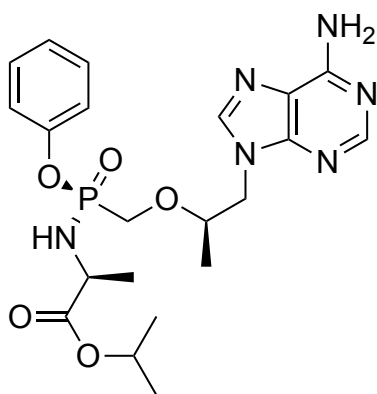
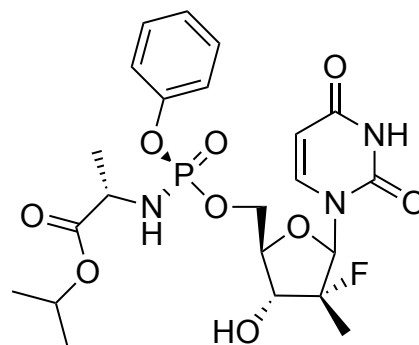


Figure 2. Chemical structures of therapeutic nucleoside analogue monophosphonates, cidofovir, adefovir and tenofovir.

Although the use of monophosphonates efficiently addressed the *in situ* and *in vivo* instability issue associated with monophosphate (P-O-) bonds, these compounds still had poor cellular uptake due to the negative charges on the monophosphonate group at physiological pH (< 7.4). This limitation acted as a challenge in the medicinal chemistry field to develop prodrugs of nucleoside analogue monophosphates and monophosphonates that can deliver them into cells more efficiently. Numerous prodrug technologies that mask either one or two of the oxygens of the monophosphate and monophosphonate groups making the molecule more lipophilic and hence improve transport into the cells have been developed.^{13, 29-31} Among these prodrug approaches is the ProTide prodrug technology that was pioneered by Prof. Chris McGuigan (Cardiff, UK).³² This prodrug technology has proved to be a powerful tool in the discovery of efficacious nucleoside monophosphate and monophosphonate prodrug therapeutics.³³ To date, it has delivered at least ten clinical candidates that reached, or successfully passed, phase I clinical trials.³³ Although most of these nucleotide prodrugs were developed as antiviral agents against HIV, HBV, HCV and Ebola virus, the most recent ones are also being pursued as potential anticancer agents. Moreover, the ProTide prodrug technology has so far led to the discovery of two FDA-approved (antiviral) drugs, tenofovir alafenamide (TAF)³⁴ and sofosbuvir³⁵ (Figure 3).¹³



Tenofovir Alafenamide



Sofosbuvir

Figure 3. Chemical structures of the two FDA-approved antiviral ProTides, tenofovir alafenamide (TAF) and sofosbuvir.

Due to the effectiveness of the ProTide technology in the *in vivo* intracellular delivery of nucleoside analogue monophosphates and monophosphonates, there has been a growing interest in the application of this technology in drug discovery. The last comprehensive review on the development of the ProTide technology appeared in 2009 (Mehellou, Balzarini and McGuigan)³². However, at that time, the ProTides had just entered clinical trials and significant advances in their synthesis, metabolism and clinical studies have been achieved since then. Thus, we felt that there is a need for an updated and comprehensive report on the ProTide technology, which this review aims to address and therefore provide a much needed updated resource for the scientific community.

2. Inception and evolution of the ProTides

Initial work on the ProTides can be tracked to the late 1980s.³⁶⁻³⁸ The mission was to mask the oxygen atoms of phosphate groups in nucleoside monophosphate analogues so that they are neutral at physiological pH and hence have a better uptake into cells. The process started from the simple masking of the phosphate groups by alkyl groups but eventually evolved into the intelligent masking of the phosphate with aryl groups and amino acid esters, which can be cleaved-off inside the intact cells. Such development process, which took at least two decades, could be divided into six phases as discussed below.

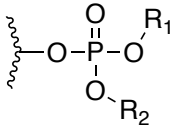
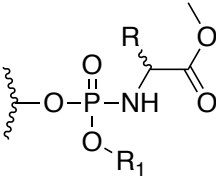
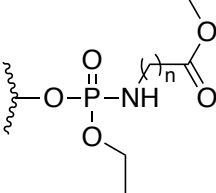
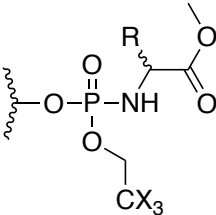
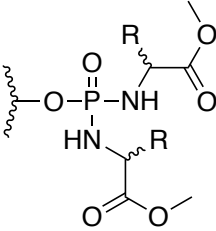
Stage One: Alkyl and haloalkyl phosphate esters

The first attempt of making nucleoside analogue monophosphate prodrugs was to mask the phosphate group of 9- β -D-arabinofuranosyladenine (AraA)³⁹ monophosphate, which exerts antiviral activity, by simple dialkyl triesters (*Stage 1*, Table 1).³⁶ Although these prodrugs led to increased stability towards adenosine/adenylate deaminases, they demonstrated a moderate biological activity.³⁶ Similar results were found when the same principle was applied to the anti-cancer agent β -D-arabinofuranosylcytosine (AraC)^{40,37} However, when the dialkyl phosphate approach was used to mask the monophosphate group of the anti-HIV agent 3'-azidothymidine (AZT)⁴¹, no significant activity was observed *in vitro*.⁴² This was attributed to several parameters, but the most likely explanation for the lack of activity was due to the high stability of the dialkyl triesters, which hindered the eventual conversion to the biologically active 5'-O-triphosphate form of AZT.⁴³ Masking the monophosphate groups of AZT and 2',3'-dideoxycytidine (ddC)⁴⁴ with haloalkyl derivatives also yielded agents with poor biological activity.⁴⁵ In order to enhance the intracellular cleavage of the haloalkyl masking groups, more haloalkyl phosphate masking groups with halogen atoms incorporated either on one or in both masking groups were used for making AZT monophosphate prodrugs. However, these prodrugs also displayed moderate antiviral activities.⁴⁶ However, the use of haloalkyl groups in AraA and AraC monophosphates displayed significant increases in biological activity (*Stage 1*, Table 1).⁴⁷ This increase in activity was mainly attributed to increased lipophilicity, resulting in better membrane permeability of the prodrug.⁴⁷ The lack of increased AZT activity through administration of haloalkyl prodrugs of AZT could now be explained by the fact that the first phosphorylation step of AZT is not the rate-limiting step in its activation pathway while it certainly is for AraA and AraC.⁴⁸ Nevertheless, the alkyl and haloalkyl phosphate masking groups, especially for their application to AraA and AraC, indicated that these masking groups showed good activity due to better membrane-crossing ability, since the biological activity correlated with lipophilicity, rather than increased intracellular levels of the phosphate species.^{36, 37, 47} Such conclusion shifted the thinking towards phosphate prodrugs to a different strategy.

Stage Two: Alkyloxy and haloalkyloxy phosphoramidates

Subsequently, the design of nucleoside phosphate prodrugs focused on alkyloxy and haloalkyl phosphoramidate prodrugs (*Stage 2*, Table 1). The move towards phosphoramidate prodrugs was provoked by the findings that the HIV protease can cleave an oligopeptide from the phosphate group of blocked nucleotide phosphoramidates.⁴⁹ Initially, a selection of AZT alkyloxy phosphoramidates with methyl-esterified amino acids were synthesised.⁴² These compounds showed better anti-HIV activity than the dialkyl AZT prodrugs.⁴² Although the anti-HIV activity did not correlate with the alkyl phosphate chain, it did correlate with the amino acid side chain with L-alanine showing the better activity.^{42, 50} Retrospectively, this observation proved to be critical for the success of the ProTide technology in drug discovery since all of the ProTides that entered clinical trials and those that have been approved for clinical use bear an L-alanine moiety.³³ Further studies into different, i.e. α , β and γ , types of amino acids, indicated the superiority of α -amino acids.⁵¹ Encouraged by the influence of the amino acid moiety on the anti-HIV activity of AZT alkyloxy phosphoramidates, the masking of phosphate groups by the methyl-esterified amino acids and haloalkyl groups, which increased the membrane-crossing ability, were explored next. Haloalkyl phosphoramidates of AZT were synthesised with glycine, alanine and valine amino acids as well as ethyl, trifluoroethyl or trichloroethyl as the haloalkyl motifs.⁵² These, surprisingly, did not show good anti-HIV activity with the exception of the trichloroethyl alanine prodrug. Since no clear structure-activity relationship was apparent from the haloalkyl phosphoramidates, they were not pursued any further. However, the key finding of the enhancement of activity with nucleoside phosphoramidate prodrugs, especially with L-alanine, proved to be important for the future evolution of this nucleoside monophosphate prodrug strategy.

Stages of development	Phosphate masking group(s)
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<p>1. Alkyl and haloalkyl phosphate triesters</p>	 <p>R_1 and R_2 = Me, Et, Pr, $-\text{CH}_2\text{CCl}_3$ and $-\text{CH}_2\text{CF}_3$</p>
<p>2. Alkyloxy phosphoramidates</p>	 <p>R = H, Me, $-\text{CH}(\text{CH}_3)_2$, $-\text{CH}_2\text{CH}(\text{CH}_3)_2$, $-\text{CH}(\text{CH}_3)\text{Et}$ or Bn R_1 = Me, Et, Pr, Bu or Hex</p>  <p>n = 1-6</p>  <p>R = H, Me or iPr X = H, F or Cl</p>
<p>3. Phosphorodiamidates</p>	 <p>R = H, Me, iPr, $-\text{CH}_2\text{Ph}$ or</p>

	-CH ₂ CO ₂ CH ₃ .
4. Lactyl-derived systems	<p>R = CH₃ or Et, R₁ = H or Me, R₂ = Me, nPr or nC₁₂H₂₅.</p>
5. Diaryl phosphates	<p>Ar = Ph or p-X-Ph (x = various)</p>
6. Aryloxy phosphoramidates	<p>Ar = Ph or substituted Ph R = H, Me, -CH₂iPr, -CH₂Ph...etc. R₁ = Me, Et, iPr, tBu, Bn...etc.</p>

Table 1. Stages of the development of the ProTide prodrug technology.

Stage Three: *Phosphoramidates*

Given that the introduction of amino acids to generate phosphoramidates led to enhanced anti-HIV activity, it was logical that the next step would be to mask both oxygen groups of the phosphate group by amino acid esters. Indeed, a series of amino acid methyl esters with different side-chains were used to mask both oxygens of the phosphate group of AZT monophosphate (Stage 3, Table 1).⁵³ Unlike previous studies where L-alanine appeared to give the better biological activity, in these phosphoramidate systems amino acids with aromatic side-chains, e.g. phenylalanine, showed better improvement of biological activity.⁵³ Although this was surprising in the context of the ProTides' development, such finding was consistent with other studies where a preference was found for amino acids with aromatic side-chains in the development of unrelated phosphoramidate diesters.^{29, 31} Although no

further studies with these phosphorodiamidates were conducted since the early nineties and the development of the ProTides moved on, phosphorodiamidates as phosphate prodrugs recently (in 2011) made a comeback.⁵⁴

Stage Four: *Lactyl-derived systems*

Continuing with the probing of the phosphoramidates and particularly studies on the importance of the P-N bond, isosteric O-linked nucleoside analogues derived from lactic and glycolic acids of AZT were prepared and evaluated for their biological activity (Stage 4, Table 1).⁵⁵ Although no significant improvement in the anti-HIV activity was noted for the lactyl-derived AZT monophosphate prodrugs, the observed activity was found to correlate with the lengthening of the alkyl phosphate chain (R_2) since longer chains resulted in poorer anti-HIV activity. As for the glycol systems ($R_1 = H$), the prodrugs were generally more active than the lactyl systems. The poor anti-HIV activity of these phosphate prodrug systems was not very encouraging and they were therefore not optimised or explored any further.

Stage Five: *Diaryl phosphates*

The next masking groups studied in the design of a novel phosphate prodrug strategy were aryl motifs. Indeed, the masking of AZT monophosphate by diaryl motifs was pursued (Stage 5, Table 1).⁵⁶ These prodrugs showed good anti-HIV activity and, remarkably, some of them showed much better anti-HIV activity than the parent compound AZT.⁵⁶ In the journey towards the ProTides, this was the first time a phosphate prodrug was more active than the parent compound. Interestingly, these phosphate prodrugs retained their anti-HIV activity in cell lines that were considered to be resistant to AZT.⁵⁶ However, when these cells (T lymphocyte cell line, JM) were used for this study back in 1992, they were considered as AZT insensitive due to poor phosphorylation of AZT.⁵⁷ But, it was later shown that such AZT resistance was a result of an active AZT-efflux pump rather than inefficient intracellular phosphorylation.⁵⁸ Regardless of the exact AZT-insensitivity mechanism in this cell line, the AZT diaryl phosphate prodrugs were more active than their parent compound providing a clear indication of an increased delivery of AZT monophosphate into the cells. This notion was further supported by the fact that *para*-nitrophenyl phosphate masking groups had ca.

100-fold better anti-HIV activity than their unsubstituted phenyl masking groups most probably due to the electron-withdrawing effect of the *para*-nitro group that speeds-up the de-masking of the prodrugs in cells.⁵⁶ A further study on the effect of the nature of the phenyl substituents on the antiviral activity of diaryl phosphate prodrugs of AZT was then conducted.^{59, 60} This study confirmed that stronger *para*-electron-withdrawing groups on the phenyl phosphate masking groups gave better anti-HIV activity.^{59, 60} Intriguingly, these AZT diaryl phosphate prodrugs as well as AZT did not exhibit any anti-HIV activity in a thymidine kinase (TK)-deficient human T-lymphocyte cell line (CEM/TK⁻).⁶⁰ McGuigan and colleagues concluded that this might have been due to poor intracellular release of AZT monophosphate. The development of diaryl monophosphate prodrugs was not progressed any further and the focus turned back to phosphoramidates, which showed good biological activity.⁵⁶

Stage Six: Aryloxy phosphoramidates (generally referred to as ProTides)

Inspired by the improvement of the biological activities of the earlier examples of nucleoside monophosphate prodrugs particularly the *para*-substituted aryls and the amino acid esters, McGuigan and co-workers took the next logical step, which was to combine these two masking groups in the design of new monophosphate prodrugs.⁶¹ Indeed, the masking of the AZT monophosphate group with *para*-substituted aryls and an amino acid ester was conducted and these efforts yielded prodrugs that showed improved anti-HIV activity as compared to the parent nucleoside AZT. In particular, the combination of a phenyl and L-alanine methyl ester showed the most potent anti-HIV activity in this study, $EC_{50} = 0.8 \mu\text{M}$ (AZT, IC_{50} (toxicity) ca. $100 \mu\text{M}$).⁶¹ Interesting structure-activity relationships on these aryloxy phosphoramidate prodrugs emerged from this study. Notably, there was a preference for the L-alanine amino acid as compared to L-leucine (> 10-fold) and glycine (> 100-fold) while electron-withdrawing *para*-substituted aryls were less potent than their unsubstituted moieties.⁶¹ Further studies into these substitutions concluded that *para*-fluoro aryls did not largely affect the biological activity while *para*-nitro-substitutions had detrimental effects on the biological activity, namely a 100-fold loss in activity.⁶² In a separate study it

was shown that *para*-nitrophenyl-containing nucleoside monophosphate prodrugs were toxic even to uninfected cells.⁶³ Remarkably, AZT aryloxy phosphoramidates retained their anti-HIV activities in thymidine kinase deficient (TK⁻) cells.⁶² These observations provided a direct proof of the ability of aryloxy phosphoramidates to deliver intact nucleoside analogue monophosphates into intact cells. Hence, the masking of nucleoside analogue monophosphates and monophosphonates with an aryl motif and an amino acid ester, which is nowadays known as the ProTide technology, was extensively pursued in the future discovery of novel antiviral and anticancer nucleotide therapeutics.

1. Synthesis of the ProTides

Although the synthesis of the aryloxy triester phosphoramidate ProTides has recently been excellently reviewed by Pradere *et. al.*,²⁹ for the completion of this Review, we will briefly discuss the different strategies used in the synthesis of the ProTides. We will also include recent reports on the development and use of new catalysts that improve the synthesis of the ProTides, which have been published after the comprehensive review by Pradere *et. al.*²⁹. There are three different strategies for synthesising ProTides: 1) the coupling of a nucleoside with a diarylphosphite followed by subsequent oxidative amination 2), the coupling of the nucleoside with a phosphorochloridate reagent, and 3) coupling of an amino acid to a nucleoside aryl phosphate (Figure 4).

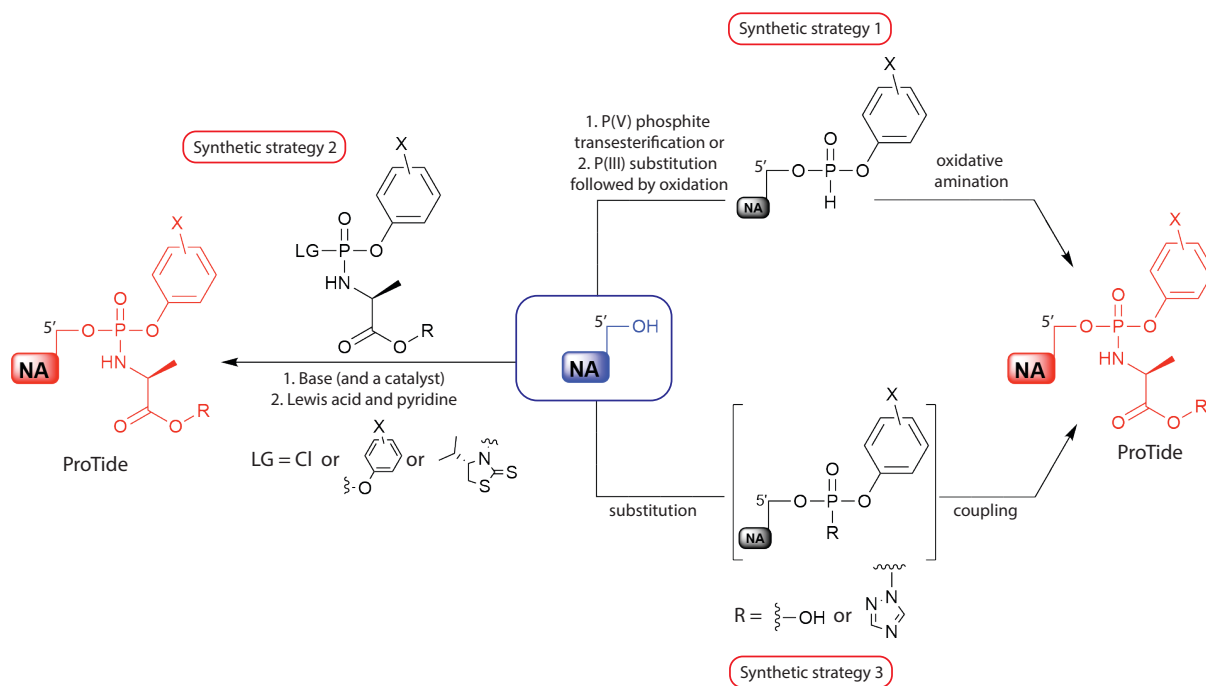


Figure 4. A representation of the three main strategies for synthesising ProTides. LG: leaving group. NA: nucleoside analogue. R: any ester. X: any aromatic substitution.

Among these three strategies, the second synthetic strategy, which involves the coupling of the nucleoside to a phosphorochloridate is the most common method and has been the method of choice of the initial synthesis of the ProTides. The coupling reaction takes place in the presence of either the Grignard reagent *tert*-butyl magnesium chloride (*t*BuMgCl) or *N*-methylimidazole (NMI), which act as a base. The choice of the base is often influenced by the nucleoside substrate, most precisely the number of hydroxyl groups that could couple to the phosphorochloridate. For instance, in ribonucleosides, the base used is often NMI and this generates mostly the 5'-*O*- phosphoramidates due to steric hindrance and the fact that it is a weaker base than *t*BuMgCl. However, in cases where the nucleoside has only one hydroxyl group, the base of choice is *t*BuMgCl, which often gives higher yields of the product as compared to NMI. Notably, the approach of coupling the nucleoside to a phosphorochloridate yields a mixture of diastereoisomers. In some cases, the two ProTide diastereoisomers exhibit similar potency and their development therefore is pursued as a mixture of diastereoisomers (see examples in Figure 10).⁶⁴ In other cases, the two

diastereoisomers may have different rates of metabolism and hence varying potency. For instance, the *Sp*-diastereoisomer of the clinically used ProTide sofosbuvir exhibits a 10-fold increase in antiviral activity than its *Rp*-diastereoisomer.³⁵ In these instances, the separation of the two diastereoisomers becomes necessary. The separation of these by column chromatography has proved to be a stern challenge.

To address this, Ross et al⁶⁵ developed a system that allows for the facile generation and crystallisation of single isomer phosphoramidating reagents that could be coupled to nucleosides to yield pure single ProTide isomers. From this study, phosphorochloridates with a pentafluorophenol were found to be optimal since the desired *Sp*-isomer phosphorochloridate was readily crystallised, stable and allowed for the formation of the *Sp*-isomer ProTides, which is often the more active isomer.⁶⁵

Continuing on the theme of synthetic procedures with stereocontrol of the phosphorous chirality of the ProTides, researchers from Merck recently reported the discovery of a small molecule catalyst that, upon the addition to the reaction of a nucleoside with a phosphorochloridate, leads to the generation of a single isomer with significant preference for the 5'-hydroxyl compared to the 3'-one.⁶⁶ However, the reported catalyst generates the *Rp*-isomer and not the *Sp*-one, which often shows better pharmacological activity. This represents a challenge for the wide use of this catalyst. Impressively, more work from Merck researchers led to the identification of various Lewis acids as being able to improve the yield, stereoselectivity and regioselectivity of the reaction of phosphorochlorides with nucleosides to generate ProTides.⁶⁷ Indeed, a selection of Lewis acids were studied and dimethyl aluminium chloride emerged as the optimal catalyst for this reaction.

In the synthesis of nucleoside monophosphonate ProTides three different synthetic strategies are employed (Figure 5). The first strategy involves the *bis*-chlorination of a phosphate group to form a monophosphonate dichloridate. This is then followed by the addition of 1:1 molar equivalents of the phenol and amino acid ester yielding the desired ProTides. Alternatively, in the second synthetic strategy, the monophosphonate group is

initially coupled to one phenol group using *N,N*-dicyclohexylcarbodiimide (DCC) and the product is then chlorinated. Finally the chloride group is substituted by the amino acid ester.

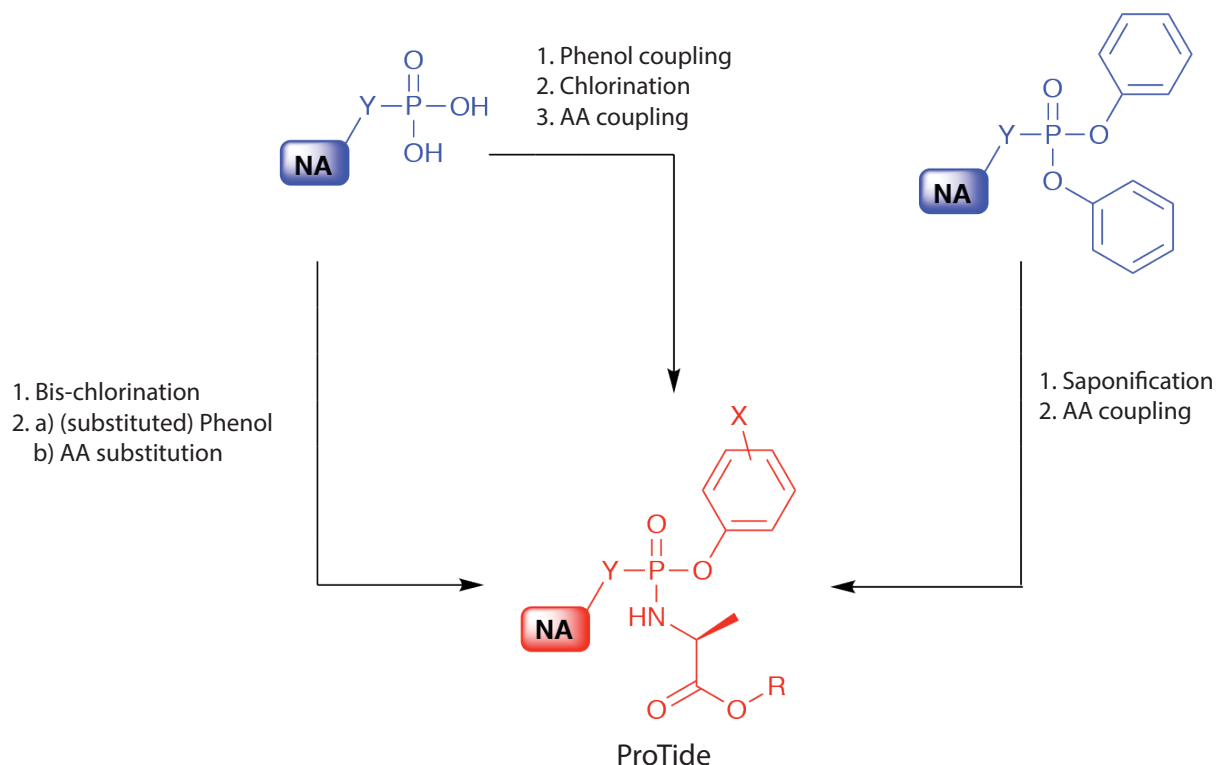


Figure 5. Synthetic strategies for accessing monophosphate ProTides. AA: amino acid. NA: nucleoside analogue. R: any ester. X: any aromatic substitution. Y: O or CH₂.

For the third synthetic strategy, the starting material employed is the diphenyl-masked nucleoside monophosphate, which first undergoes selective saponification that results in the removal of one of the phenyl masking groups. The product is subsequently coupled to the amino acid ester to generate the desired nucleoside analogue monophosphate ProTide. Further details on the synthesis of the ProTides can be found in Pradere *et al.*²⁹ and Serpi *et al.*⁶⁸.

2. Metabolism of the ProTides

Once inside the cell, the ProTides require metabolism to release the nucleoside monophosphate. The metabolic conversion of the 2',3'-didehydro-2',3'-dideoxythymidine (d4T)⁶⁹⁻⁷¹ monophosphate (d4T-MP) ProTide to the eventual active metabolite d4T-TP has been extensively studied in several cell systems, including human lymphocytes (Figure 6).

Beside efficient formation of d4T-MP, d4T-diphosphate (d4T-DP) and d4T-triphosphate (d4T-TP), an additional metabolite markedly accumulated upon exposure of the cell culture to the d4T-MP ProTide, which was identified as being the alaninyl d4T-MP. This metabolite may act as an intracellular depot form of d4T-MP, explaining not only the superior antiviral activity of the ProTide *versus* the parental d4T, but also the marked retainment of antiviral activity in thymidine kinase-deficient cells in which the parental d4T markedly loses antiviral efficacy.^{72, 73} Also, the activation pathway of the ProTides of d4T-MP and AZT-MP containing different amino acids and/or ester moieties has been investigated. It was found that overall, there was a close correlation between the anti-HIV activity of these prodrugs and their conversion rate to the amino acyl nucleotide metabolite (metabolite **C**, Figure 6).^{74, 75} The metabolic conversion process is believed to be mediated by a two-step enzymatic process, which eventually releases the active monophosphate (or monophosphonate) form of the parent nucleoside as summarised in Figure 6.

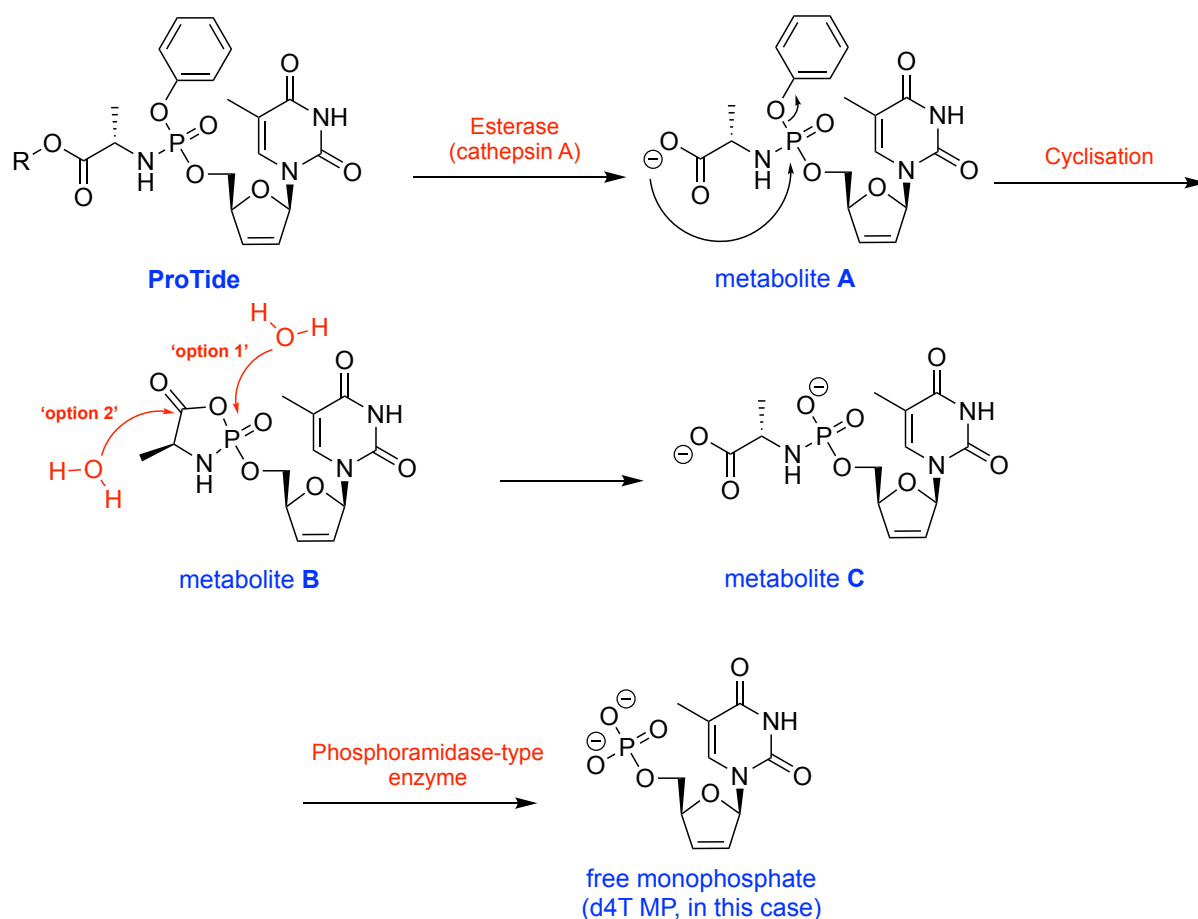


Figure 6. Postulated mechanism of ProTide metabolism illustrated using the ProTide of the anti-HIV agent d4T as an example.

The first step involves the cleavage of the amino acid ester by intracellular esterases. Researchers at Gilead Sciences identified cathepsin A as a major esterase that mediates the hydrolysis of ester motifs of ProTides.⁷⁶ Such finding has made cathepsin A widely used in the field in probing the hydrolysis of ProTides *in vitro*.^{64, 77-79} Notably, a second enzyme that was able to hydrolyse the ester group of the two antiviral ProTides studied, TAF and 9-(*R*)-4-(*R*)-[[[(*S*)-1-[(ethoxycarbonyl)ethyl]amino]phenoxy-phosphonyl]methoxy]-2-fluoro-1-furanyl-adenine (GS-9131)⁸⁰ (structure not disclosed), was also reported in this study but its identity was not determined.⁷⁶ In 2011, however, this identity of this second enzyme was established and was shown to be carboxylesterase 1 (CES1).⁸¹ Although both cathepsin A and CES1 were found to be expressed in primary hepatocytes, CES1 expression was

undetectable in clone A replicon cells.⁸¹ This indicated that in primary human hepatocytes, both cathepsin A and CES1 are involved in the hydrolysing of the carboxyl ester motif of the ProTides, whereas in clone A cells, cathepsin A is the major enzyme that mediates such process. Beyond cathepsin A and CES1, a series of cysteine and (lysosomal) serine proteases have also been identified as being able to hydrolyse the carboxyl ester motif of the ProTides in quiescent cells and peripheral blood mononuclear cells (PBMC).⁸²

Earlier work by McGuigan and colleagues⁸³ into the efficiency of the ester group hydrolysis in various ProTides indicated that this is dependent on the structure of the ester moiety. In particular, it was noted that ProTides with a *tert*-butyl ester group were slower hydrolysed than those bearing other esters, e.g. methyl, isopropyl and benzyl, which correlated with the biological activity.⁸³ Although the lipophilicity of the ester motif seemed to give an increase in biological activity most likely due to increased cellular uptake, the rate of ester hydrolysis is another influencing factor in the ultimate biological activity of the ProTides.⁸³ For instance, ProTides with naphthyl esters yield more active ProTides than their phenyl ester counterparts due to higher lipophilicity and faster hydrolysis (Figure 7). However, propylbenzene esters, which are more lipophilic, are not as active as phenyl esters probably due to poor *in vivo* hydrolysis.

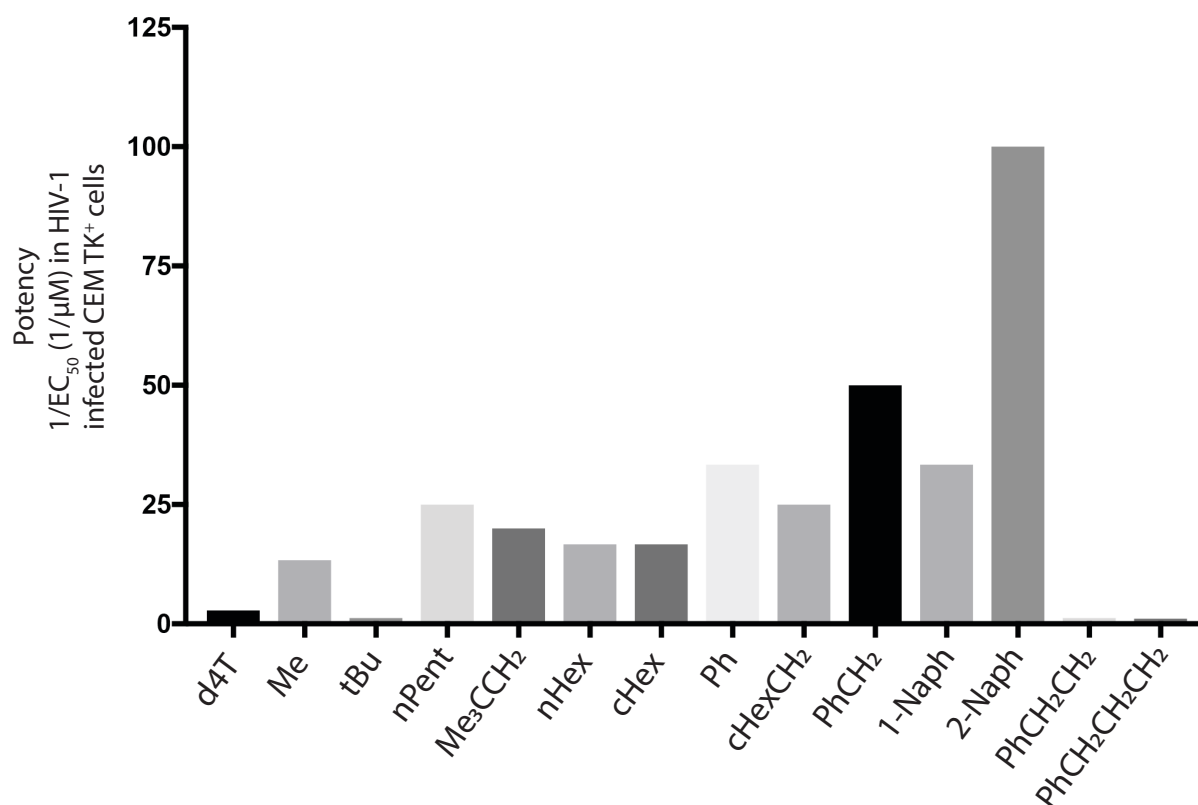


Figure 7. Effect of the ester motif on the ProTides' biological activity. The data presented regard d4T, ProTides and were obtained from McGuigan et al.⁸³

In addition to the ester group's influence on the overall lipophilicity of the ProTides and its effect on the biological activity, the aryl motif has significant impact on the lipophilicity. In McGuigan's work, the aryl group mostly used was the phenyl group though the naphthyl group has also been explored. The biological activity between these two aryl motifs often correlated with lipophilicity and hence the naphthyl-containing ProTides exhibit a better biological activity than their phenyl counterparts. Notably, substitutions on these aryl motifs exerts a significant impact on the biological activity (Figure 8). Electron-withdrawing substitutions were found to be associated with increased biological activity as compared to the electron-donating groups. The position on the aryl group that allows maximum electron-withdrawing effects, i.e. ortho and para, had more pronounced effects than the meta position.

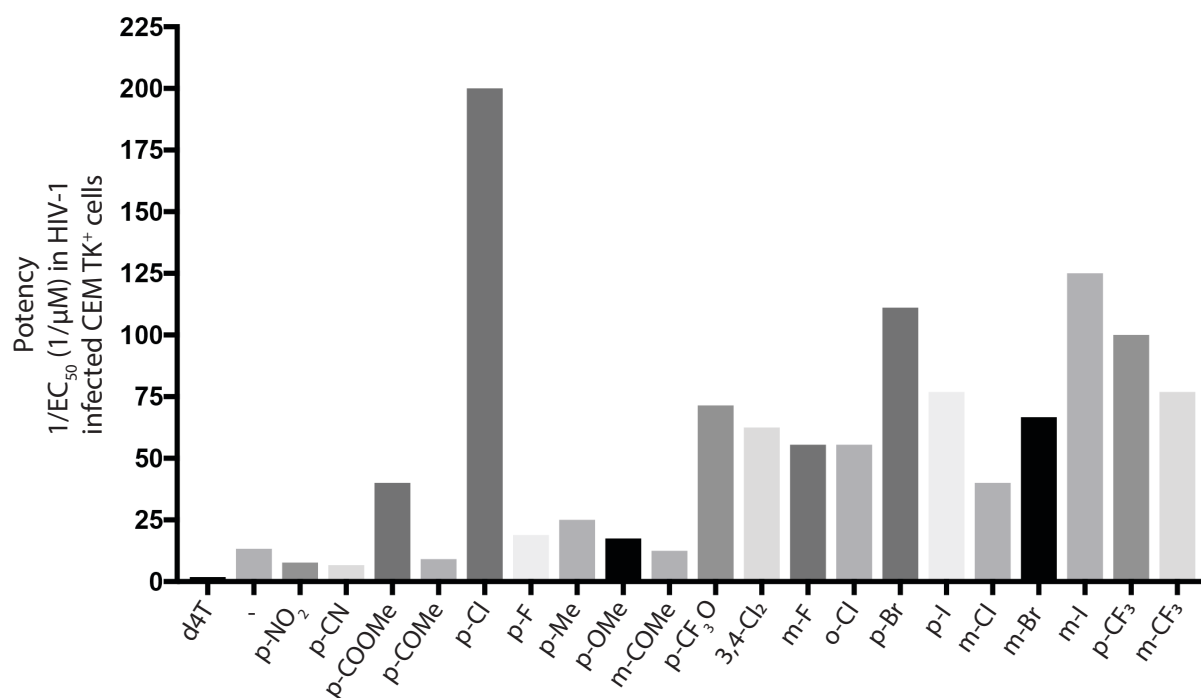


Figure 8. Effect of the aryl group substitutions on the ProTides' biological activity. The data presented regard d4T, ProTides and were obtained from Siddiqui et al.⁸⁴

Beyond the nature of the ester group, it has been shown that the nature of the amino acid of the ProTides also affects the rate of cathepsin A-mediated hydrolysis of the ester group (Figure 9).⁷⁶ Indeed, cathepsin A was unable to hydrolyse ProTides with branched amino acids such as valine and isoleucine. Subsequent studies showed that the nature of the amino acid side chain, primarily its size (bulkiness) and charge, has great influence on what enzyme hydrolyzes the ester motif.⁸² Indeed, an interesting structure-activity relationship emerged from the influence of the amino acid on the rate of the metabolism of ProTides. For instance, it was found that the ester motif of ProTides with alanine were efficiently processed by esterases (e.g. leukocyte elastase and proteinase 3) and serine proteases, but not cysteine, aspartic, or metalloproteases.⁸² Amino acids with bulkier side chains, e.g. phenylalanine and leucine, were more efficiently hydrolysed by chymotrypsin-related serine proteases such as cathepsin G and chymase.⁸²

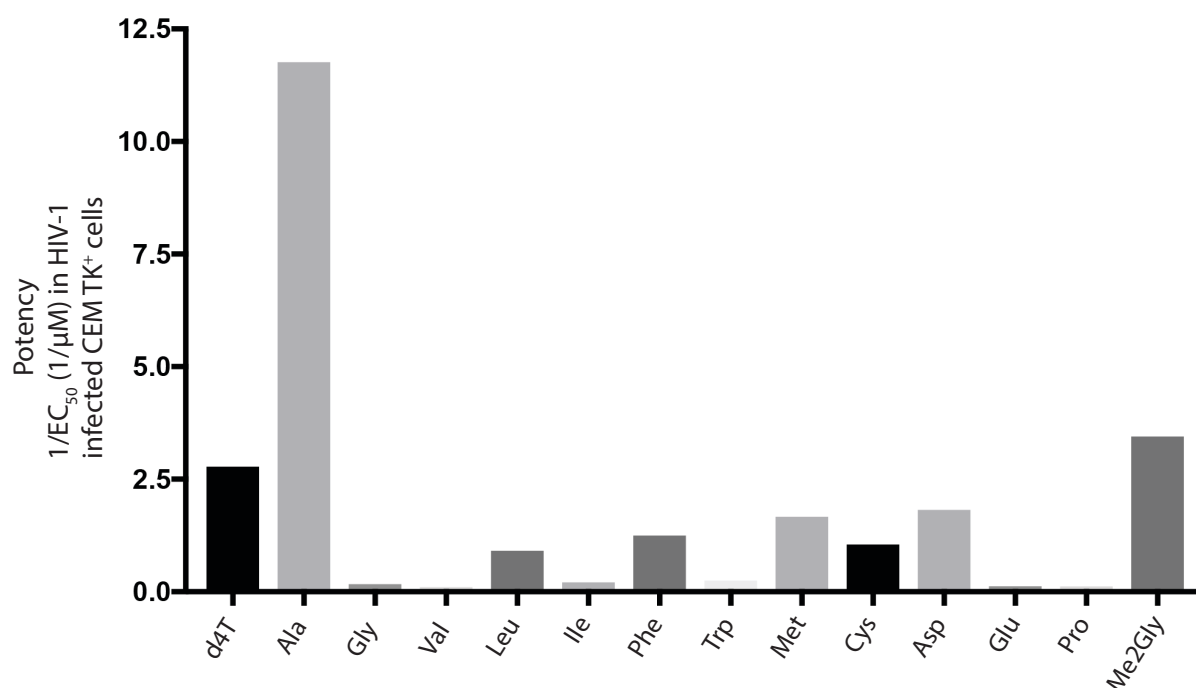


Figure 9. Effect of the amino acid on the ProTides' biological activity. The data presented regard d4T, ProTides and were obtained from McGuigan et al.⁸⁵

After the ester motif at the amino acid moiety is cleaved-off, a free carboxylate, which under physiological pH carries a negative charge, is generated. Subsequently, a nucleophilic attack by the carboxylate functionality on the phosphate or phosphonate group causes the loss of the aryl-leaving group yielding a highly unstable five-membered anhydride ring. The speed of the nucleophilic attack from the carboxylate on the phosphate or phosphonate group varies depending on the nucleoside. For instance, AZT metabolite **A** formation (Figure 6) was slower than that of d4T.⁷⁵ Interestingly, the hydrolysis of ProTides with β -alanine did not release the nucleoside monophosphate, since the hydrolysis did not proceed beyond the hydrolysis of the ester motif.⁷⁵ Metabolite **B** formation is followed by a rapid nucleophilic attack by a water molecule on the phosphate (option 1, Figure 6) or carbon of the carbonyl group of the amino acid (option 2, Figure 6) to open-up the ring and to generate the phosphoramidate metabolite (metabolite **C**, Figure 6). Although no compelling evidence exists to whether the water molecule attacks the phosphate or the carbon of the carbonyl

group of the amino acid, which both yield the same product, the cleavage of the P-N bond at this stage has never been observed.⁸⁶

The final step in the metabolism of the ProTides is the cleavage of the P-N bond by a second enzyme known as a phosphoramidase-type enzyme (histidine triad nucleotide binding protein 1,⁸⁷ HINT1).^{81, 88, 89} This eventually leads to the release of the nucleoside monophosphate/phosphonate. Notably, the cleavage of the P-N bond can be resistant to HINT1 as is the case with TAF. It is hypothesised that the hydrolysis of the P-N bond in this case proceeds because of the chemical instability of the P-N bond in the acidic environment of endosomes/lysosomes.⁹⁰

3. ProTides in Drug Discovery

Since the invention of the ProTide technology by the McGuigan laboratory in the early 1990s, this modality has been widely employed in the synthesis of many nucleoside monophosphate or monophosphonate ProTides. Although initially the structure activity relationship (SAR) studies were performed on pyrimidine nucleosides such as AZT and d4T, a wide variety of other ProTide derivatives have also been synthesized and antivirally evaluated. Notably, application of the phosphoramidate ProTide technology on purine nucleoside analogues such as 2',3'-dideoxyadenosine (ddA) and 2',3'-dideoxyadenosine(D4A),⁹¹ carbocyclic adenosine derivatives,⁹² and carbovir/abacavir^{93, 94} often led to markedly enhanced antiviral potencies of such ProTides. More recently, it was even shown that ProTides of the antiherpetic acyclovir resulted in both anti-herpetic and anti-HIV activity,^{95, 96} and thus, enabled broadening of the antiviral spectrum of such drug. In this respect, it should be mentioned that the ProTide technology not only enables significant optimization of existing antiviral drugs, but it also allows conversion of apparently looking inactive molecules to active antivirals by the improved uptake and direct delivery of their activated (monophosphate) species. A series of ProTides have reached clinical trials³³ with two of them already approved for clinical use.¹³ To highlight the usefulness of this prodrug technology in drug discovery, key ProTides that are currently reported to be undergoing clinical trials and those that have received FDA-approval will be discussed.

5.1. ProTides as Clinical Candidates

5.1.1. Benzyl CCCCC1=CN(C(=O)N1C2=CC=CC=C2)COP(=O)(OC3=CC=CC=C3)C4=CC=CC=C4 (NUC-1031)

NUC-1031 (**1**, Figure 10)⁶⁴ is a phenyl L-alanine benzyl ester ProTide of the FDA-approved anticancer nucleoside analogue gemcitabine (Gemzar™)⁹⁷ and developed for clinical use by NuCana Biomed Ltd. Gemcitabine emerged as an ideal candidate for the application of the ProTide technology because of the emerging resistance against this anticancer therapeutic agent. Gemcitabine resistance mechanisms include inadequate conversion of gemcitabine into its pharmacologically active di- and tri-phosphate forms due to downregulation of deoxycytidine kinase, deamination of gemcitabine to its much less active uridine derivative and mutations in the membrane transporter that mediates its active uptake into cells.⁹⁸

A series of gemcitabine ProTides were synthesised and probed for their cytostatic activities.⁶⁴ The lead structure from these studies was **1** as it exhibited significant antiproliferative activity, superior *in vivo* stability and an excellent PK profile.⁶⁴

A notable result that emerged from the ProTide was that it was able to effectively overcome gemcitabine resistance. This was attributed to three main reasons: firstly, cytidine deaminase is an enzyme found in both serum and cells and is known to catabolize gemcitabine. In order for deamination to occur, unprotected hydroxyl groups on the 3'- and 5'-position of the nucleoside analogue must be present.⁹⁹ ProTide **1** demonstrated resistance to deamination because of the presence of the phosphoramidate moiety on the 5'-O-position of the ribose moiety. A second advantage of being resistant to deaminase breakdown is an increased safety profile, as lesser toxic metabolite (2',2'-difluorodeoxyuridine) would be produced.⁶⁴ Secondly, it was found that the cellular uptake of **1** was not dependent upon nucleoside transporters and that the ProTide was lipophilic enough to enter the cells via passive diffusion. This was deduced following an experiment involving dipyridamole –a known inhibitor of the human equilibrative nucleoside transporter 1 (hENT1)– to mimic the downregulation of nucleoside transport in cancer cells. Gemcitabine was incubated with pancreatic PANC1 cancer cells in the presence of dipyridamole, and it

was found that its cytotoxic activity was significantly reduced, with the cytostatic activity decreasing from 611 nM to > 2000 nM (CC₅₀). Contrastingly, even with blockage of hENT1, the activity of **1** remained fairly constant (CC₅₀ = 162 μM), thus confirming that the cellular uptake of **1** is independent of nucleoside transporters.⁶⁴ Thirdly, the structure of the ProTide is such that it is able to overcome the rate-limiting first phosphorylation-mediated by deoxycytidine kinase which is downregulated in certain cancers. This was confirmed by little change in the activity of **1** when incubated with RT112 cancer cells in the presence of 2'-deoxycytidine being the competing natural substrate for gemcitabine activation to its 5'-monophosphate by 2'-deoxycytidine kinase (EC₅₀ = 0.7 μM in the presence of 2'-deoxycytidine compared to 0.2 μM in the dCyd absence). Conversely, the activity of gemcitabine decreased when 2'-deoxycytidine was co-administered as the two compounds competed for the same active site on the deoxycytidine kinase.⁶⁴

In vivo animal studies of compound **1** were conducted, focusing in particular on gemcitabine (partially) resistant pancreatic cancers. The initial results obtained were very promising. In mouse models with human tumour xenografts, **1** reduced the tumour size faster than gemcitabine and significant reductions in tumour size were noticeable on day 7 post administration of the ProTide. The body weight of the animals was also recorded to assess their tolerance to the compound. Overall, ProTide **1** was associated with less than 4% reduction in body weight and it was found that over the course of the treatment, the mean body weight change was less pronounced in mice that were treated with **1** compared to those treated with gemcitabine. This indicated that the gemcitabine ProTide was better tolerated than the parent nucleoside analogue.⁶⁴ The pharmacokinetics of the ProTide were also considerably better than those of gemcitabine: the half-life of **1** was 7.9 h compared to 1.5 h for gemcitabine. The ProTide also led to intracellular concentrations of the active gemcitabine triphosphate metabolite that was 13 times higher than achieved by gemcitabine.¹⁰⁰

Phase I/II clinical trials were conducted involving 68 patients and showed that **1** was effective in a variety of different cancers but most notably in patients with ovarian, pancreatic

and biliary cancers. Phase III studies will be conducted focusing on these cancers in particular. The ProTide was found to be stable *in vivo*, exhibiting a plasma half-life of 8.3 h. Maximum intracellular concentrations of the active metabolite were quickly achieved, peaked at 475 μM and were maintained for 24 h. In several patients, the tumour had shrunk by more than 30%, whereas in 33 patients, the disease had stabilised.¹⁰¹

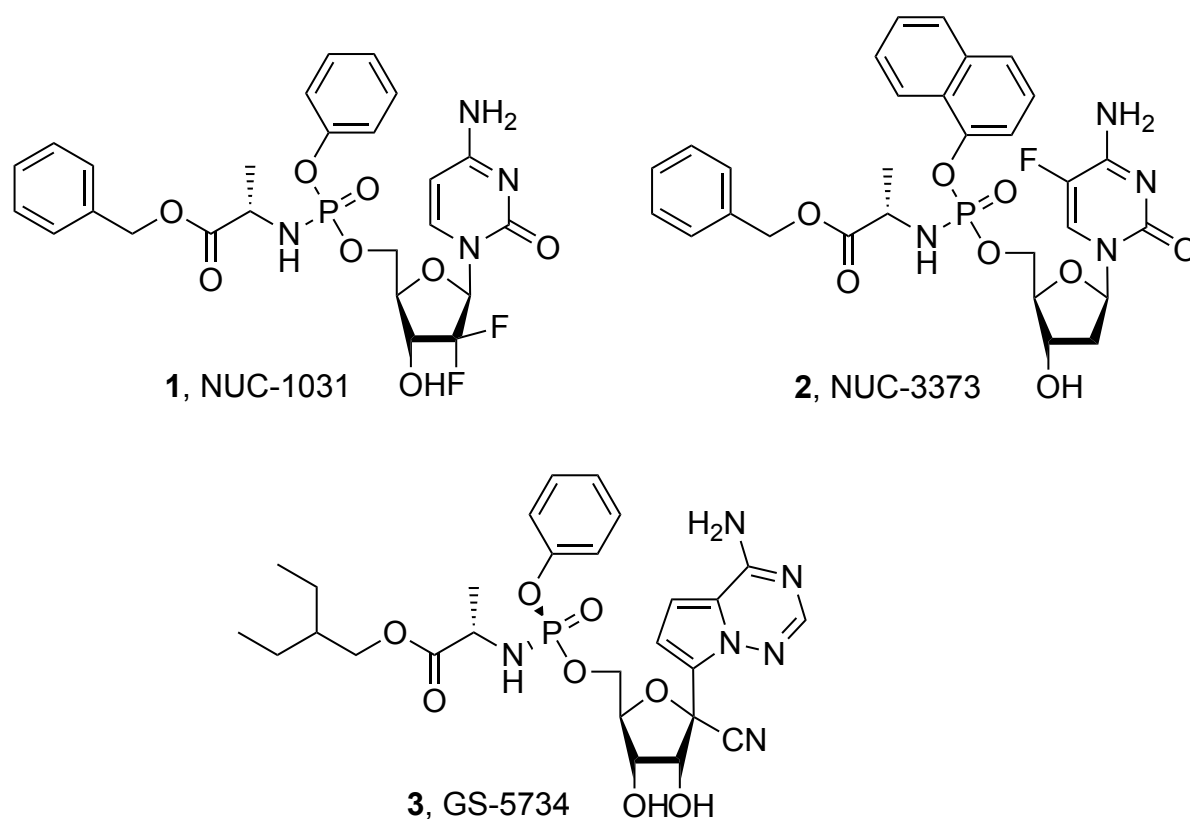


Figure 10. Chemical structures of key ProTides undergoing clinical trials.

5.1.2. Benzyl CCCCOC(=O)C[C@@H](N)OP(=O)(OC1=CC=CC=C1)OC2=CC=CC=C2 (((((2*R*,3*S*,5*R*)-5-(4-amino-5-fluoro-2-oxypyrimidin-1(2*H*)-yl)-3-hydroxytetrahydrofuran-2-yl)methoxy)(naphthalen-1-yloxy)phosphoryl)-*L*-alaninate (NUC-3373)

Another ProTide currently being developed for clinical use by NuCana Biomed Ltd is NUC-3373 (**2**, Figure 10),^{79, 102} a phosphoramidate prodrug of 5-fluoro-2'-deoxyuridine (FdU). FdU is a metabolite which gets formed *in vivo* from the pharmacologically active anti-cancer agent 5-fluorouracil (5-FU). Initially discovered by McGuigan and his colleagues in 2011 after extensive SAR studies,⁷⁹ ProTide **2** displayed promising preclinical results and initial

studies highlighted its superiority over its parent nucleoside, 5-FU. The path towards the final structure of **2** began with McGuigan and his co-workers by synthesising and comparing 39 different ProTides, with variations in the aryl, ester and amino acid regions.⁷⁹ There was little difference observed in the potency of the ProTides upon varying the amino acid ester region, and also upon changing the nature of the amino acid. L-alanine has long been the amino acid of choice when formulating ProTides, but there was little to discern between L-alanine and the other amino acids in these experiments. Generally, the ProTides yielding the 1-naphthyl group displayed more potency than phenyl-containing ProTides, thus leading to the deduction that the 1-naphthyl, benzyl ester-containing ProTide was the most effective.⁷⁹ In order to determine the cytotoxic activity of **2**, *in vitro* studies were conducted involving human tumour cell lines (colorectal, lung, ovarian, acute lymphoblastic leukaemia and cervix). Attention to thymidine kinase and the nucleoside transporter hENT1 was also given to investigate the characteristics of cancer resistance. Xenograft mouse models were also used to further analyse the anticancer activity of the ProTide.¹⁰³

It was found that compound **2** was, on the whole, significantly more cytotoxic than 5-FU, displaying IC₅₀ values 2- to 333-times lower in the majority of the tumor cell lines tested. The ProTide also produced 363-times more of the active metabolite (FdU) than 5-FU, thus further displaying its superiority in efficacy. Thymidine kinase inhibition only slightly affected ProTide **2** (~ 4-fold) compared to FdU, that showed reductions in cytotoxicity of up to 136-fold, respectively, indicating the independence of the ProTide for the FdU-activating enzyme. Significant results were obtained with the colorectal tumour xenografts: ProTide **2** inhibited tumour growth by 47%, compared to 25% tumour growth inhibition demonstrated by 5-FU.¹⁰³ Compound **2** was found to overcome the main mechanisms of resistance that is associated with 5-FU and other drugs within this family, such as decreased levels of the necessary phosphorylating (activating) enzyme thymidine kinase, which is required to produce the active metabolite; an overexpression of thymidylate synthase (an enzyme responsible for DNA synthesis and a target for FdUMP (the active metabolite of 5-FU), altered levels of thymidine phosphorylase, and reduced entry into the cells via nucleoside transporters.^{79, 102}

ProTide **2** overcomes these pathways to resistance by preventing degradation by phosphorolytic enzymes, bypassing the need to be activated by thymidine kinase and being able to diffuse into cells, independent of nucleoside transporters such as hENT1.¹⁰⁴ Given the encouraging results obtained in the pre-clinical studies, ProTide **2** entered phase I clinical trials in the first quarter of 2016 in pursuance of determining the pharmacokinetic profile of the ProTide, the safety of the compound in human participants and clinical activity of **2** in tumour-bearing patients.¹⁰³ The trial itself consists of two parts: the first part assessing the optimum dose that will maximise the risk/benefit reward to the participants when given a weekly dose of the ProTide, administered as a single intravenous infusion. The second part will assess the same as part one, except the dosing interval which will be increased to fortnightly (www.clinicaltrials.gov, trial identifier: NCT02723240).

5.1.3. 2-Ethylbutyl ((S)-(((2*R*,3*S*,4*R*,5*R*)-5-(4-aminopyrrolo[2,1-*f*][1,2,4]triazin-7-yl)-5-cyano-3,4-dihydroxytetrahydrofuran-2-yl)methoxy)(phenoxy)phosphoryl)-*L*-alaninate (GS-5734)

In 2014, there was a mass outbreak of the Ebola virus, most prevalent in Western Africa, although there were a number of reported cases in the USA and the United Kingdom. In West-Africa alone, there were a total of 28,616 confirmed cases in Guinea, Liberia and Sierra Leone, leading to 11,310 deaths (WHO 2016). To this date there is no drug treatment available on the market to treat this disease. However, GS-5734 (**3**, Figure 10)¹⁰⁵, developed by Gilead Sciences, has shown promising results as a potential treatment for the Ebola virus. Although most of the interest in this compound has been due to its activity against Ebola, recently, **3** was reported to be a broad-spectrum antiviral agent since it exhibited potent antiviral activity against a variety of viruses including filoviruses, coronaviruses, and paramyxoviruses.¹⁰⁶

Compound **3** is a ProTide of an adenosine analogue which, following intracellular delivery, gets metabolised into the pharmacologically active nucleoside triphosphate form, which then inhibits viral RNA dependent RNA polymerase.¹⁰⁵

The activity of **3** was initially tested using cell-based assays, followed by the administration of the ProTide to rhesus monkeys that were infected with the Ebola virus. Rodent studies

were deemed not suitable to determine the pharmacokinetics of **3** due to the high serum concentrations of esterases present in many rodent species, which would prematurely degrade the ProTide and impact on the findings. Thus, rhesus monkeys were chosen as they closely resemble humans and they do not express a high serum esterase concentration.¹⁰⁵ 10 μM of **3** was incubated in a 2 h-pulse or 72 h-continuous incubation of Ebola virus-infected human primary macrophages, endothelial cells and liver cells to deduce its activity. 10 μM of the nucleoside analogue was also incubated for 72 h in human monocyte-derived macrophages for comparison. ProTide **3** was found to be rapidly metabolised in the primary macrophages, with maximum intracellular concentrations of nucleoside triphosphate being achieved after 4 h post incubation, with a half-life of 16 h. It was found that **3** displayed an EC_{50} value against Ebola virus of 0.086 μM in the human primary macrophages, 0.14 μM in the endothelial cells and 0.07 μM in the Huh-7 liver cells. In comparison, the nucleoside analogue displayed an EC_{50} value of greater than 20 μM in both the human macrophages and endothelial cells, and 1.5 μM in the liver cells. The cytotoxic concentration (CC_{50}) of the ProTide in all cell lines was above 20 μM . The different cell lines were also infected with different variants of the Ebola virus (Makona and Kikwit variants) and other filoviruses. The cell-based assays gave strong indication that compound **3** would be effective in treating the other viruses as well.¹⁰⁵

The pharmacokinetics of **3** were examined in rhesus monkeys. After being administered intravenously with 10 mg/kg of the ProTide, **3** displayed a very short plasma half-life of just 0.39 h. There was rapid distribution into the peripheral compartments within 2 h post administration, and a half-life in the peripheral blood mononuclear cells of 14 h was observed. Levels required to achieve a 50% inhibition of the Ebola virus were maintained for 24 h.¹⁰⁵ Compiling the pharmacokinetics results and tissue distribution data led to the conclusion that once daily dosing would be effective to achieve a clinical effect.

In order to assess the efficacy of the ProTide, a blinded placebo-controlled study was conducted involving 12 Ebola-infected rhesus monkeys (6 per treatment group) in two different modalities.¹⁰⁵ The first part involved intramuscular administration of **3** either on day

0 (same day as being infected with Ebola), or day 2. Part two of the study administered a loading dose of 10 mg/kg followed by once daily doses of 3 mg/kg, commencing at various days after virus exposure or 10 mg/kg administered on day 3 post virus exposure. All of the monkeys given **3** survived until the end of the trial; 50% survival was noted with the treated group of animals given 3 mg/kg compared to no survival in the placebo group. It was noted that monkeys given the 10 mg/kg dose after day 3 had 100% survival along with reduced signs associated with the Ebola virus (thrombocytopenia, coagulopathy).¹⁰⁵

Given the promising results obtained in the rhesus monkey study, a phase II clinical trial is due to be conducted in Liberia. Sponsored by the National Institute of Allergy & Infectious Diseases, 60 to 120 men infected with Ebola virus will be recruited and will be given a five-day course of **3**, administered once daily, followed by monitoring over a six-month period to determine the clearance of virus from a persistent genital reservoir (www.clinicaltrials.gov, trial identifier: NCT02818582).

5.2. ProTides as Drugs

There have been two ProTide drugs approved by the FDA for clinical use. Sofosbuvir (Sovaldi™), Figure 3, was approved by the FDA in December 2013 for the treatment of HCV.¹⁰⁷ Sofosbuvir is a phosphoramidate prodrug of 2'-deoxy-2'-fluoro-2'-C-methyluridine, the parent nucleoside. Having emerged as the superior ProTide out of 50 other ProTides developed,³⁵ sofosbuvir was acquired by Gilead Sciences, where it adopted the identity sofosbuvir and underwent further extensive clinical evaluation. Sofosbuvir was found to act by inhibiting the HCV NS5B polymerase, an enzyme responsible for the replication of the viral genome. Inhibition of this enzyme would result in RNA chain termination and ultimately stop the replication of HCV.³⁵ Following cellular uptake by hepatocytes, sofosbuvir undergoes sequential hydrolysis by cathepsin A and CES1 to form the active nucleoside monophosphate. Further hydrolysis by HINT1 and subsequent phosphorylations yielded the pharmacologically active triphosphate metabolite, that exerts its action on NS5B polymerase.¹⁰⁸ Initial animal studies of sofosbuvir were encouraging. The ProTide was well absorbed following oral administration in portal vein cannulated dogs, bioavailability was

9.9%, which equated to a fraction absorbed of 36%.¹⁰⁹ In human subjects, the absolute bioavailability was not determined, although over 80% of the administered dose was absorbed. The absorption was rapid, with maximum concentrations (C_{max}) of sofosbuvir achieved 0.5-2 h post oral administration and C_{max} of the inactive parent nucleoside metabolite was achieved 2-4 h post administration and a half-life of 0.48-0.75 h. *Ex vivo* studies of the distribution of the anti-HCV agent showed a plasma protein binding of around 82% in healthy subjects and 85% in subjects with end stage renal disease.¹⁰⁸

Studies using the diastereomeric mixture of sofosbuvir in humans showed that it was rapidly absorbed and metabolised in the liver according to the typical *in vivo* degradation of the ProTides (see Figure 6). Briefly, the isopropyl ester motif was hydrolysed into the naked carboxylate group and then the P-N bond of the phosphoramidate group was cleaved-off to afford parent nucleoside monophosphate.¹¹⁰ Notably, the parent nucleoside, an inactive metabolite, was detectable in the plasma and remained detectable for at least 24 h following the 100 and 800 mg dosing of the diastereomeric mixture of sofosbuvir.¹¹⁰ Although sofosbuvir showed little indication of being excreted renally (3.5% recovered in urine; renal clearance = 0.238 L/min), the inactive parent nucleoside was the predominant compound present in the urine (77.7%), indicating excretion via the kidneys. This was further confirmed when studies were conducted observing the pharmacokinetics of sofosbuvir in patients with impaired renal function – the renal clearance of the parent nucleoside was around 2-fold higher than the glomerular filtration rate, signifying active renal excretion of the metabolites of sofosbuvir.¹⁰⁹ Phase II studies of sofosbuvir in combination with pegylated interferon and ribavirin established the doses required to provide an optimal effect, whilst minimising any potential adverse effects.¹¹¹ Initial studies postulated that 200 mg or 400 mg doses of sofosbuvir with pegylated interferon and ribavirin were most effective, as 100 mg dosing was found to be sub-optimal due to low viral load decline than observed with the 200 mg and 400 mg doses. At these doses, the incidence of adverse effects was low, with the most frequent adverse effects being fatigue and nausea.¹¹¹ Coupled with the short half-life of sofosbuvir, once daily dosing was recommended as there was little accumulation of the drug.¹⁰⁹

The safety of sofosbuvir was later confirmed in phase II clinical trials. It was postulated that as sofosbuvir is not metabolised via the cytochrome P450-3A4 enzymes, drug-drug interactions would be minimal. However, because it seems to induce the P-glycoprotein efflux pump, caution was advised with co-administration of carbamazepine and rifabutin as the concentrations of sofosbuvir can decrease significantly.^{112, 113}

Single-group, open label phase III study of treatment-naïve patients with HCV genotypes 1, 4, 5 and 6 treated with sofosbuvir plus pegylated interferon and ribavirin for 12 weeks indicated high potency of sofosbuvir.¹⁰⁷ Indeed, 80% of cirrhotic patients versus 92% of noncirrhotic patients achieved a viral load below the lower limit of quantifications 12 weeks after completion of HCV treatment (SVR12). This indicated the efficacy of sofosbuvir in treating different HCV genotypes confirming similar earlier preclinical observations.¹¹⁴ In a placebo-controlled phase III study comparing sofosbuvir and ribavirin for 12 weeks with matched interferon ineligible controls in genotype 2 and 3 patients, sustained viral response was achieved in 78% of treated patients with 93% response rate in genotype 2 patients and 61% in genotype 3 patients.¹¹⁵ In a study involving patients who had previously been treated with interferon who were treated with either sofosbuvir and ribavirin for 12 weeks followed by placebo for 4 weeks or sofosbuvir and ribavirin for 16 weeks, response rates showed a 73% SVR12 in the 16-week arm compared to 50% of the 12-week arm. Impressively, 94% of genotype 2 patients achieved SVR12 within 12 weeks of treatment with sofosbuvir and ribavirin, while 87% achieved it within 16 weeks. As for genotype 3 patients, 30% of patients achieved SVR12 within 12 weeks compared to 62% within 16 weeks. Notably, 46% of the patients treated with sofosbuvir and ribavirin for 12 weeks had viral relapse compared to 27% of the 16-week treatment arm. In another study, the efficacy of extended treatment of 24 weeks of sofosbuvir plus ribavirin in genotype 3 patients was evaluated.¹¹⁶ The results showed that 93% of the 73 patients with genotype 2 who took part achieved SVR12 within 12 weeks of sofosbuvir and ribavirin treatment, whereas 85% of the 250 patients with genotype 3 treated for 24 weeks with sofosbuvir and ribavirin achieved SVR12. This was the first large scale study showing the efficacy of prolonged interferon-free treatments in

genotype 3 patients. Given the excellent safety profile and pan-genotypic efficacy of sofosbuvir, it has been licenced for clinical use in combination with pegylated interferon, ribavirin and the NS5A inhibitors ledipasvir and velpatasvir depending on the HCV genotype to be treated.¹¹⁷ Collectively, this has made sofosbuvir a cornerstone in HCV therapy.

TAF, Figure 3, is the latest ProTide-based drug approved by the FDA for the treatment of HIV.¹¹⁸ Developed by Gilead Sciences Inc., TAF is a ProTide of the acyclic nucleoside phosphonate drug tenofovir. Another prodrug of tenofovir, tenofovir disoproxil fumarate (TDF), had long been an FDA-approved treatment for HIV, however, the ProTide TAF demonstrated increased anti-HIV activity, higher *in vivo* stability and was endowed with lesser side-effects than both tenofovir and TDF.¹¹⁸ TAF is specifically accumulating in lymphatic tissue, and in the liver, and thus, holds also great potential for the treatment of HBV infections. Markedly lower plasma levels but significantly higher intracellular tenofovir concentrations are observed with TAF as compared to TDF, explaining the lower incidence of tenofovir-associated side-effects with respect of bone mineral density losses and kidney nephrotoxicity.¹¹⁹ The drug itself becomes phosphorylated following cellular uptake into the pharmacologically active diphosphate form and is a potent inhibitor of reverse transcriptase. Inhibition of this enzyme results in DNA chain termination and viral replication ceases to progress.¹²⁰

Early studies of TAF were conducted in dogs in order to characterise its pharmacokinetic profile and understand its permeability and efficacy as a potential treatment for HIV. Initial *in vitro* studies using hepatic and intestinal S9 fractions from both humans and dogs yielded similar results, highlighting the suitability of using dogs for the pre-clinical studies. *In vitro* experiments showed TAF to have half-lives of 31 minutes in both dog intestinal and hepatic S9 fractions, a plasma half-life of 92 minutes and a half-life of 28.3 minutes in an MT-2 cell extract.⁸⁰ 5 mg/kg of TAF were administered to the dogs; plasma concentrations peaked at 1.58 μM , achieved at 0.14 h post administration. The levels of TAF decreased rapidly (undetectable after 2 h post administration), which was accompanied by an increase in tenofovir levels. Rapid accumulation of the active metabolite tenofovir diphosphate in the

peripheral blood mononuclear cells followed, with peak concentrations of 18 μ M achieved at 1 h post administration. The bioavailability of TAF was found to be dependent on the administered dose, with bioavailabilities of 1.7% and 24.7% at 2 mg/kg and 20 mg/kg, respectively.¹²¹ However, other studies found that oral administration of 10 mg/kg of TAF yielded a bioavailability of 17% when compared to the intravenous dose,⁸⁰ with the majority of the drug metabolised in the kidneys and excreted in the urine.¹²²

An earlier study conducted by Eisenberg et al. in 2001¹²³ compared the activity of the ProTide with tenofovir and its prodrug TDF. Following incubation with peripheral blood mononuclear cells, it was found that the levels of tenofovir and its metabolites were 10-30 times higher compared to TDF and tenofovir, respectively.¹²³ The *in vitro* anti-HIV activity of TAF was determined to be 1000-fold and 10-fold greater than tenofovir and TDF.⁸⁰

Following successful *in vitro* assessments of TAF, the ProTide progressed into a number of phase I and II trials, where the *in vivo* pharmacokinetics/dynamics of the ProTide was evaluated and compared to its predecessors, along with establishing the optimum dose of TAF. An initial study was conducted by Markowitz et al.,¹²⁴ where either 40 mg or 120 mg of TAF or 300 mg of TDF were administered once daily for 14 days and the activity of both prodrugs as monotherapy for the HIV-1 infection was compared. The study showed that both 40 mg and 120 mg of TAF resulted in higher intracellular concentrations of tenofovir diphosphate in PBMC than tenofovir. TAF also produced a greater net change in HIV-1 RNA compared to TDF, thus illustrating its superiority over the existing prodrug in terms of potency.¹²⁴

Phase II/III studies of TAF focused on its efficacy in different patient groups including, but not limited to, adults and adolescents. As tenofovir is eliminated via the kidneys, it was imperative to understand how renal function would affect both the accumulation of tenofovir in the body and also its excretion. TDF has been associated with effects on the bone and renal toxicity, leading to decreased glomerular filtration rates, increases in serum creatinine and increased risk of adverse renal events. This toxicity was attributed to its effect in the proximal tubule, with efficient uptake from plasma but less efficient efflux into urine resulting

in accumulation of tenofovir in the body.¹²⁵ Therefore, a Phase 3 double-blinded study was conducted in which TAF was administered to patients with impaired renal function. Patients were given either 10 mg of TAF or 300 mg of TDF, each co-formulated with elvitegravir, cobicistat and emtricitabine. The study found that participants on the TAF treatment regime displayed a smaller increase in creatinine, less proteinuria, and significant decreases in bone mineral density.¹²⁶

TAF was demonstrated to be superior not only in its antiviral activity but also in the fact that therapeutic effects could be achieved at much lower doses than TDF, thus reducing the incidence of adverse effects. This led to the FDA approval of the ProTide in the treatment of HIV-1 in a combination therapy with other anti-HIV agents.¹¹⁸ Moreover, in November 2016, the FDA granted approval for a once-daily treatment of 25 mg TAF in patients suffering of chronic HBV infection (Gilead Sciences 2016). This followed extensive studies which highlighted the ProTides superiority over current therapies used to treat not only HIV, but also chronic HBV infections.¹²⁷

6. Conclusions and Outlook

The ProTide technology has facilitated the discovery and application of a variety of nucleoside monophosphate and monophosphonate analogues in the clinic. It took many years of trial and error, mainly by the McGuigan group, to optimise the masking groups of the phosphate to yield the successful ProTide technology as we know it today. Although this technology has largely been used in the discovery of nucleoside-based ProTides as antiviral and anti-cancer agents, recently it was applied in the early discovery of nucleoside-based ProTides for Parkinson's disease.⁷⁸ Additionally, the ProTide prodrug approach is receiving increasing attention as a prodrug strategy for the intracellular delivery on monophosphorylated non-nucleoside compounds such as glucosamine,^{128, 129} Sphingosine 1-phosphate (S1P),¹³⁰ 4-phospho-D-erythronhydroxamic acid¹³¹ and 5-phospho erythronhydroxamic acid¹³². Together, these examples show the broad usefulness of this prodrug technology and highlight its potential in the discovery of non-nucleoside monophosphate and monophosphonate prodrugs as potential therapeutics as well.

Regardless of the substrates to which the ProTide technology is applied to either nucleosides or non-nucleosides, it has already proven to be a powerful technology in drug discovery and efficient intracellular delivery of the (activated) parental drug. Given its expanding applications, there is a real possibility that this technology will deliver new (nucleoside and non-nucleoside) ProTides for the treatment of a variety of other diseases beyond microbial infections and cancer.

Abbreviations

5-FU, 5-fluorouracil; AraA, 9- β -D-arabinofuranosyladenine; AraC, β -D-arabinofuranosylcytosine; AZT, 3'-azidothymidine; d4A, 2',3'-didehydro-2',3'-dideoxyadenosine; CES1, carboxylesterase 1; d4T, 2',3'-didehydro-2',3'-dideoxythymidine; ddC, 2',3'-dideoxyadenosine; ddC, 2',3'-dideoxycytidine; DCC, N,N'-Dicyclohexylcarbodiimide; FDA, Food and Drug Administration; FdU, 5-fluoro-2'-deoxyuridine; FdUMP, 5-fluoro-2'-deoxyuridine-5'-monophosphate; HBV, hepatitis B virus; HCMV, human cytomegalovirus; HCV, hepatitis C virus; hENT1, human equilibrative nucleoside transporter 1; HINT1, Histidine triad nucleotide-binding protein 1; HIV, human immunodeficiency virus; HSV, herpes simplex virus; NA, nucleoside analogue; NMI, N-methylimidazole; PK, Pharmacokinetics, SVR12, the lower limit of quantifications 12 weeks after completion of HCV treatment; TAF, tenofovir alafenamide; TDF, tenofovir disoproxil fumarate; SAR, structure-activity relationship; tBuMgCl, *tert*-butyl magnesium chloride; TK, thymidine kinase.

Dedication

This review was written in memory of Prof. Christopher McGuigan (1958-2016), who provided the basis for the ProTide technology.

Biographies

Youcef Mehellou obtained his M.Pharm. (Pharmacy) degree from King's College London in 2005. This was followed by a Ph.D. on 'ProTide' chemistry under the supervision of Prof. Chris McGuigan in Cardiff University, U.K. Following postdoctoral work initially with Prof. Sidney M. Hecht in Arizona State University (2009/2010) and then with Prof. Dario R. Alessi FRS in the University of Dundee, U.K., (2010/2013) Youcef moved to the University of Birmingham, U.K., in early 2013 as a Lecturer in Medicinal Chemistry. Since January 2017, Youcef has been a Lecturer in Medicinal Chemistry in Cardiff University, U.K., where his group is focusing on the discovery of small molecule modulators of signal transduction and the development of novel phosphate and phosphonate prodrug therapeutics.

Hardeep S. Rattan is currently a fourth and final year Pharmacy student in the University of Birmingham, U.K. He has a keen interest in medicinal chemistry particularly the discovery and development of nucleoside and nucleotide therapeutics. Hardeep's final year Masters research project focused on the ProTide prodrug technology and was carried out under the supervision of Dr. Mehellou.

Jan Balzarini graduated at the KU Leuven (Belgium) as Master in Biology (1975), Master in Bioengineering (1977) and PhD in Bioengineering (1984). He is affiliated to the Rega Institute for Medical Research at the KU Leuven. In 1984, he became tenured, performed a postdoctoral stay at the National Cancer Institute of the NIH (1985/1986) with Sam Broder and David G. Johns. In 2001, he was awarded the prestigious René Descartes Research Prize of the European Commission (in collaboration with several European Research Teams) and he received several other (inter)national awards. In 2001, he became full professor at the KU Leuven; in 2006, he was appointed as Laboratory Head of Virology and Chemotherapy at the Rega Institute; and in 2014, he retired as Emeritus Professor.

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References

1. Perkins, E. S.; Wood, R. M.; Sears, M. L.; Prusoff, W. H.; Welch, A. D. Anti-viral activities of several iodinated pyrimidine deoxyribonucleosides. *Nature* **1962**, *194*, 985-986.
2. Fyfe, J. A.; Keller, P. M.; Furman, P. A.; Miller, R. L.; Elion, G. B. Thymidine kinase from herpes simplex virus phosphorylates the new antiviral compound, 9-(2-hydroxyethoxymethyl)guanine. *J. Biol. Chem.* **1978**, *253*, 8721-8727.
3. De Clercq, E.; Descamps, J.; De Somer, P.; Barr, P. J.; Jones, A. S.; Walker, R. T. (E)-5-(2-Bromovinyl)-2'-deoxyuridine: a potent and selective anti-herpes agent. *Proc. Natl. Acad. Sci. U. S. A.* **1979**, *76*, 2947-2951.
4. De Clercq, E. Nucleoside analogues as antiviral agents. *Acta. Microbiol. Acad. Sci. Hung.* **1981**, *28*, 289-306.
5. Robins, R. K. The potential of nucleotide analogs as inhibitors of retroviruses and tumors. *Pharm. Res.* **1984**, *1*, 11-18.
6. Jordheim, L. P.; Durantel, D.; Zoulim, F.; Dumontet, C. Advances in the development of nucleoside and nucleotide analogues for cancer and viral diseases. *Nat. Rev. Drug Discovery* **2013**, *12*, 447-464.
7. De Clercq, E.; Li, G. Approved antiviral drugs over the past 50 years. *Clin. Microbiol. Rev.* **2016**, *29*, 695-747.
8. Shelton, J.; Lu, X.; Hollenbaugh, J. A.; Cho, J. H.; Amblard, F.; Schinazi, R. F. Metabolism, biochemical actions, and chemical synthesis of anticancer nucleosides, nucleotides, and base analogs. *Chem. Rev.* **2016**, *116*, 14379-14455.
9. Loffler, M.; Morote-Garcia, J. C.; Eltzschig, S. A.; Coe, I. R.; Eltzschig, H. K. Physiological roles of vascular nucleoside transporters. *Arterioscler., Thromb., Vasc. Biol.* **2007**, *27*, 1004-1013.
10. Molina-Arcas, M.; Casado, F. J.; Pastor-Anglada, M. Nucleoside transporter proteins. *Curr. Vasc. Pharmacol.* **2009**, *7*, 426-434.

11. Anderson, P. L.; Kakuda, T. N.; Lichtenstein, K. A. The cellular pharmacology of nucleoside- and nucleotide-analogue reverse-transcriptase inhibitors and its relationship to clinical toxicities. *Clin. Infect. Dis.* **2004**, *38*, 743-753.
12. Mehellou, Y.; De Clercq, E. Twenty-six years of anti-HIV drug discovery: where do we stand and where do we go? *J. Med. Chem.* **2010**, *53*, 521-538.
13. Thornton, P. J.; Kadri, H.; Miccoli, A.; Mehellou, Y. Nucleoside phosphate and phosphonate prodrug clinical candidates. *J. Med. Chem.* **2016**, *59*, 10400–10410.
14. Smal, C.; Vertommen, D.; Bertrand, L.; Ntamashimikiro, S.; Rider, M. H.; Van Den Neste, E.; Bontemps, F. Identification of in vivo phosphorylation sites on human deoxycytidine kinase. Role of Ser-74 in the control of enzyme activity. *J. Biol. Chem.* **2006**, *281*, 4887-4893.
15. Li, F.; Maag, H.; Alfredson, T. Prodrugs of nucleoside analogues for improved oral absorption and tissue targeting. *J. Pharm. Sci.* **2008**, *97*, 1109-1134.
16. Saiki, Y.; Yoshino, Y.; Fujimura, H.; Manabe, T.; Kudo, Y.; Shimada, M.; Mano, N.; Nakano, T.; Lee, Y.; Shimizu, S.; Oba, S.; Fujiwara, S.; Shimizu, H.; Chen, N.; Nezhad, Z. K.; Jin, G.; Fukushige, S.; Sunamura, M.; Ishida, M.; Motoi, F.; Egawa, S.; Unno, M.; Horii, A. DCK is frequently inactivated in acquired gemcitabine-resistant human cancer cells. *Biochem. Biophys. Res. Commun.* **2012**, *421*, 98-104.
17. Jamieson, G. P.; Brocklebank, A. M.; Snook, M. B.; Sawyer, W. H.; Buolamwini, J. K.; Paterson, A. R.; Wiley, J. S. Flow cytometric quantitation of nucleoside transporter sites on human leukemic cells. *Cytometry* **1993**, *14*, 32-38.
18. Wiley, J. S.; Jones, S. P.; Sawyer, W. H.; Paterson, A. R. Cytosine arabinoside influx and nucleoside transport sites in acute leukemia. *J. Clin. Invest.* **1982**, *69*, 479-489.
19. Damaraju, V. L.; Damaraju, S.; Young, J. D.; Baldwin, S. A.; Mackey, J.; Sawyer, M. B.; Cass, C. E. Nucleoside anticancer drugs: the role of nucleoside transporters in resistance to cancer chemotherapy. *Oncogene* **2003**, *22*, 7524-7536.

20. Eriksson, S. Is the expression of deoxynucleoside kinases and 5'-nucleotidases in animal tissues related to the biological effects of nucleoside analogs? *Curr. Med. Chem.* **2013**, *20*, 4241-4248.
21. Hunsucker, S. A.; Mitchell, B. S.; Spychala, J. The 5'-nucleotidases as regulators of nucleotide and drug metabolism. *Pharmacol. Ther.* **2005**, *107*, 1-30.
22. Abraham, A.; Varatharajan, S.; Abbas, S.; Zhang, W.; Shaji, R. V.; Ahmed, R.; Abraham, A.; George, B.; Srivastava, A.; Chandy, M.; Mathews, V.; Balasubramanian, P. Cytidine deaminase genetic variants influence RNA expression and cytarabine cytotoxicity in acute myeloid leukemia. *Pharmacogenomics* **2012**, *13*, 269-282.
23. Steuart, C. D.; Burke, P. J. Cytidine deaminase and the development of resistance to arabinosyl cytosine. *Nat. New Biol.* **1971**, *233*, 109-110.
24. Balzarini, J.; Herdewijn, P.; De Clercq, E. Differential patterns of intracellular metabolism of 2',3'-didehydro-2',3'-dideoxythymidine and 3'-azido-2',3'-dideoxythymidine, two potent anti-human immunodeficiency virus compounds. *J. Biol. Chem.* **1989**, *264*, 6127-6133.
25. Herdewijn, P.; Balzarini, J.; De Clercq, E.; Pauwels, R.; Baba, M.; Broder, S.; Vanderhaeghe, H. 3'-substituted 2',3'-dideoxynucleoside analogues as potential anti-HIV (HTLV-III/LAV) agents. *J. Med. Chem.* **1987**, *30*, 1270-1278.
26. Balzarini, J.; Kang, G. J.; Dalal, M.; Herdewijn, P.; De Clercq, E.; Broder, S.; Johns, D. G. The anti-HTLV-III (anti-HIV) and cytotoxic activity of 2',3'-didehydro-2',3'-dideoxyribonucleosides: a comparison with their parental 2',3'-dideoxyribonucleosides. *Mol. Pharmacol.* **1987**, *32*, 162-167.
27. Hao, Z.; Cooney, D. A.; Farquhar, D.; Perno, C. F.; Zhang, K.; Masood, R.; Wilson, Y.; Hartman, N. R.; Balzarini, J.; Johns, D. G. Potent DNA chain termination activity and selective inhibition of human immunodeficiency virus reverse transcriptase by 2',3'-dideoxyuridine-5'-triphosphate. *Mol. Pharmacol.* **1990**, *37*, 157-163.
28. De Clercq, E.; Holy, A. Acyclic nucleoside phosphonates: a key class of antiviral drugs. *Nat. Rev. Drug Discovery* **2005**, *4*, 928-940.

29. Pradere, U.; Garnier-Amblard, E. C.; Coats, S. J.; Amblard, F.; Schinazi, R. F. Synthesis of nucleoside phosphate and phosphonate prodrugs. *Chem. Rev.* **2014**, *114*, 9154-9218.
30. Hecker, S. J.; Erion, M. D. Prodrugs of phosphates and phosphonates. *J. Med. Chem.* **2008**, *51*, 2328-2345.
31. Wagner, C. R.; Iyer, V. V.; McIntee, E. J. Pronucleotides: toward the in vivo delivery of antiviral and anticancer nucleotides. *Med. Res. Rev.* **2000**, *20*, 417-451.
32. Mehellou, Y.; Balzarini, J.; McGuigan, C. Aryloxy phosphoramidate triesters: a technology for delivering monophosphorylated nucleosides and sugars into cells. *ChemMedChem* **2009**, *4*, 1779-17791.
33. Mehellou, Y. The ProTides boom. *ChemMedChem* **2016**, *11*, 1114-1116.
34. Chapman, H.; Kernan, M.; Prisbe, E.; Rohloff, J.; Sparacino, M.; Terhorst, T.; Yu, R. Practical synthesis, separation, and stereochemical assignment of the PMPA pro-drug GS-7340. *Nucleosides, Nucleotides Nucleic Acids* **2001**, *20*, 621-628.
35. Sofia, M. J.; Bao, D.; Chang, W.; Du, J.; Nagarathnam, D.; Rachakonda, S.; Reddy, P. G.; Ross, B. S.; Wang, P.; Zhang, H. R.; Bansal, S.; Espiritu, C.; Keilman, M.; Lam, A. M.; Steuer, H. M.; Niu, C.; Otto, M. J.; Furman, P. A. Discovery of a beta-d-2'-deoxy-2'-alpha-fluoro-2'-beta-C-methyluridine nucleotide prodrug (PSI-7977) for the treatment of hepatitis C virus. *J. Med. Chem.* **2010**, *53*, 7202-7218.
36. McGuigan, C.; Tollerfield, S. M.; Riley, P. A. Synthesis and biological evaluation of some phosphate triester derivatives of the anti-viral drug AraA. *Nucleic Acids Res.* **1989**, *17*, 6065-6075.
37. Jones, B. C.; McGuigan, C.; Riley, P. A. Synthesis and biological evaluation of some phosphate triester derivatives of the anti-cancer drug araC. *Nucleic Acids Res.* **1989**, *17*, 7195-7201.
38. McGuigan, C.; Shackleton, J. M.; Tollerfield, S. M.; Riley, P. A. Synthesis and evaluation of some novel phosphate and phosphinate derivatives of araA. Studies on the mechanism of action of phosphate triesters. *Nucleic Acids Res.* **1989**, *17*, 10171-10177.

39. Schabel, F. M., Jr. The antiviral activity of 9-beta-D-arabinofuranosyladenine (Ara-A). *Chemotherapia* **1968**, *13*, 321-338.
40. Talley, R. W.; Vaitkevicius, V. K. Megaloblastosis produced by a cytosine antagonist, 1-beta-D-arabinofuranosylcytosine. *Blood* **1963**, *21*, 352-362.
41. Mitsuya, H.; Weinhold, K. J.; Furman, P. A.; St Clair, M. H.; Lehrman, S. N.; Gallo, R. C.; Bolognesi, D.; Barry, D. W.; Broder, S. 3'-Azido-3'-deoxythymidine (BW A509U): an antiviral agent that inhibits the infectivity and cytopathic effect of human T-lymphotropic virus type III/lymphadenopathy-associated virus in vitro. *Proc. Natl. Acad. Sci. U. S. A.* **1985**, *82*, 7096-7100.
42. Devine, K. G.; McGuigan, C.; O'Connor, T. J.; Nicholls, S. R.; Kinchington, D. Novel phosphate derivatives of zidovudine as anti-HIV compounds. *AIDS* **1990**, *4*, 371-373.
43. McGuigan, C.; Nicholls, S. R.; O'Connor, T. J.; Kinchington, D. Synthesis of some novel dialkyl phosphate derivatives of 3'-modified nucleosides as potential anti-AIDS drugs. *Antiviral Chem. Chemother.* **1990**, *1*, 25-33.
44. Mitsuya, H.; Broder, S. Inhibition of the in vitro infectivity and cytopathic effect of human T-lymphotropic virus type III/lymphadenopathy-associated virus (HTLV-III/LAV) by 2',3'-dideoxynucleosides. *Proc. Natl. Acad. Sci. U. S. A.* **1986**, *83*, 1911-1915.
45. McGuigan, C.; O'Connor, T. J.; Nicholls, S. R.; Nickson, C.; Kinchington, D. Synthesis and anti-HIV activity of some novel substituted dialkyl phosphate derivatives of AZT and ddCyd. *Antiviral Chem. Chemother.* **1990**, *1*, 355-360.
46. McGuigan, C.; Turner, S.; Nicholls, S. R.; O'Connor, T. J.; Kinchington, D. Haloalkyl phosphate derivatives of AZT as inhibitors of HIV- studies in the phosphate region. *Antiviral Chem. Chemother.* **1994**, *5*, 162-168.
47. McGuigan, C.; Jones, B. C. N. M.; Tollerfield, S. M.; Riley, P. A. Synthesis and biological evaluation of haloalkyl phosphate triester derivatives of AraA and AraC. *Antiviral Chem. Chemother.* **1992**, *3*, 79-94.
48. Stein, D. S.; Moore, K. H. Phosphorylation of nucleoside analog antiretrovirals: a review for clinicians. *Pharmacotherapy* **2001**, *21*, 11-34.

49. Navia, M. A.; Fitzgerald, P. M.; McKeever, B. M.; Leu, C. T.; Heimbach, J. C.; Herber, W. K.; Sigal, I. S.; Darke, P. L.; Springer, J. P. Three-dimensional structure of aspartyl protease from human immunodeficiency virus HIV-1. *Nature* **1989**, *337*, 615-620.
50. McGuigan, C.; Devine, K. G.; O'Connor, T. J.; Galpin, S. A.; Jeffries, D. J.; Kinchington, D. Synthesis and evaluation of some novel phosphoramidate derivatives of 3'-azido-3'-deoxythymidine (AZT) as anti-HIV compounds. *Antiviral Chem. Chemother.* **1990**, *1*, 107-113.
51. Curley, D.; McGuigan, C.; Devine, K. G.; O'Connor, T. J.; Jeffries, D. J.; Kinchington, D. Synthesis and anti-HIV evaluation of some phosphoramidate derivatives of AZT- studies on the effect of chain elongation on biological-activity. *Antiviral Res.* **1990**, *14*, 345-356.
52. McGuigan, C.; Devine, K. G.; O'Connor, T. J.; Kinchington, D. Synthesis and anti-HIV activity of some haloalkyl phosphoramidate derivatives of 3'-azido-3'-deoxythymidine (AZT)- potent activity of the trichloroethyl methoxyalaninyl compound. *Antiviral Res.* **1991**, *15*, 255-263.
53. Jones, B. C.; McGuigan, C.; O'Connor, J. T.; Jeffries, D. J.; Kinchington, D. Synthesis and anti-HIV activity of some novel phosphorodiamidate derivatives of 3'-azido-3'-deoxythymidine (AZT). *Antiviral Chem. Chemother.* **1991**, *2*, 35-39.
54. McGuigan, C.; Madela, K.; Aljarah, M.; Bourdin, C.; Arrica, M.; Barrett, E.; Jones, S.; Kolykhalov, A.; Bleiman, B.; Bryant, K. D.; Ganguly, B.; Gorovits, E.; Henson, G.; Hunley, D.; Hutchins, J.; Muhammad, J.; Obikhod, A.; Patti, J.; Walters, C. R.; Wang, J.; Vernachio, J.; Ramamurty, C. V.; Battina, S. K.; Chamberlain, S. Phosphorodiamidates as a promising new phosphate prodrug motif for antiviral drug discovery: application to anti-HCV agents. *J. Med. Chem.* **2011**, *54*, 8632-8645.
55. McGuigan, C.; Nickson, C.; O'Connor, T. J.; Kinchington, D. Synthesis and anti-HIV activity of some novel lactyl and glycolyl phosphate derivatives. *Antiviral Res.* **1992**, *17*, 197-212.

56. McGuigan, C.; Pathirana, R. N.; Mahmood, N.; Devine, K. G.; Hay, A. J. Aryl phosphate derivatives of AZT retain activity against HIV1 in cell lines which are resistant to the action of AZT. *Antiviral Res.* **1992**, *17*, 311-321.
57. Roberts, N. A.; Martin, J. A.; Kinchington, D.; Broadhurst, A. V.; Craig, J. C.; Duncan, I. B.; Galpin, S. A.; Handa, B. K.; Kay, J.; Krohn, A.; Lambert, R. W.; Merrett, J. H.; Mills, J. S.; Parkes, K. E. B.; Redshaw, S.; Ritchie, A. J.; Taylor, D. L.; Thomas, G. J.; Machin, P. J. Rational design of peptide-based HIV proteinase inhibitors. *Science* **1990**, *248*, 358-361.
58. Snyder, R. D.; Brennan, T.; Taylor, D. L.; Tyms, A. S. Basis of relative insensitivity of HIV-infected JM cells to AZT. *Antiviral Chem. Chemother.* **1995**, *6*, 307-311.
59. McGuigan, C.; Davies, M.; Pathirana, R.; Mahmood, N.; Hay, A. J. Synthesis and anti-HIV activity of some novel diaryl phosphate derivatives of AZT. *Antiviral Res.* **1994**, *24*, 69-77.
60. McGuigan, C.; Pathirana, R. N.; Davies, M. P. H.; Balzarini, J.; Declercq, E. Diaryl phosphate derivatives act as pro-drugs of AZT with reduced cytotoxicity compared to the parent nucleoside. *Bioorg. Med. Chem. Lett.* **1994**, *4*, 427-430.
61. McGuigan, C.; Pathirana, R. N.; Mahmood, N.; Hay, A. J. Aryl phosphate derivatives of AZT inhibit HIV replication in cells where the nucleoside is poorly active. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 701-704.
62. McGuigan, C.; Pathirana, R. N.; Balzarini, J.; De Clercq, E. Intracellular delivery of bioactive AZT nucleotides by aryl phosphate derivatives of AZT. *J. Med. Chem.* **1993**, *36*, 1048-1052.
63. McGuigan, C.; Pathirana, R. N.; Mahmood, N.; Devine, K. G.; Hay, A. J. Aryl phosphate derivatives of AZT retain activity against HIV-1 in cell-lines which are resistant to the action of AZT. *Antiviral Res.* **1992**, *17*, 311-321.
64. Slusarczyk, M.; Lopez, M. H.; Balzarini, J.; Mason, M.; Jiang, W. G.; Blagden, S.; Thompson, E.; Ghazaly, E.; McGuigan, C. Application of ProTide technology to gemcitabine: a successful approach to overcome the key cancer resistance mechanisms

- leads to a new agent (NUC-1031) in clinical development. *J. Med. Chem.* **2014**, *57*, 1531-1542.
65. Ross, R. S.; Reddy, P. G.; Zhang, H. R.; Rachakonda, S.; Sofia, M. J. Synthesis of diastereomerically pure nucleotide phosphoramidates. *J. Org. Chem.* **2011**, *76*, 8311-8319.
66. DiRocco, D. A.; Ji, Y.; Sherer, E. C.; Klapars, A.; Reibarkh, M.; Dropinski, J.; Mathew, R.; Maligres, P.; Hyde, A. M.; Limanto, J.; Brunskill, A.; Ruck, R. T.; Campeau, L. C.; Davies, I. W. A multifunctional catalyst that stereoselectively assembles prodrugs. *Science* **2017**, *356*, 426-430.
67. Simmons, B.; Liu, Z.; Klapars, A.; Bellomo, A.; Silverman, S. M. Mechanism-based solution to the ProTide synthesis problem: selective access to Sofosbuvir, Acelarin, and INX-08189. *Org. Lett.* **2017**, *19*, 2218-2221.
68. Serpi, M.; Madela, K.; Pertusati, F.; Slusarczyk, M. Synthesis of phosphoramidate prodrugs: ProTide approach. *Curr. Protoc. Nucleic Acid Chem.* **2013**, Chapter 15, Unit 15.5.
69. Hamamoto, Y.; Nakashima, H.; Matsui, T.; Matsuda, A.; Ueda, T.; Yamamoto, N. Inhibitory effect of 2',3'-didehydro-2',3'-dideoxynucleosides on infectivity, cytopathic effects, and replication of human immunodeficiency virus. *Antimicrob. Agents Chemother.* **1987**, *31*, 907-910.
70. Baba, M.; Pauwels, R.; Herdewijn, P.; De Clercq, E.; Desmyter, J.; Vandeputte, M. Both 2',3'-dideoxythymidine and its 2',3'-unsaturated derivative (2',3'-dideoxythymidinene) are potent and selective inhibitors of human immunodeficiency virus replication in vitro. *Biochem. Biophys. Res. Commun.* **1987**, *142*, 128-134.
71. Lin, T. S.; Schinazi, R. F.; Prusoff, W. H. Potent and selective in vitro activity of 3'-deoxythymidin-2'-ene (3'-deoxy-2',3'-didehydrothymidine) against human immunodeficiency virus. *Biochem. Pharmacol.* **1987**, *36*, 2713-2718.

72. Balzarini, J.; Karlsson, A.; Aquaro, S.; Perno, C. F.; Cahard, D.; Naesens, L.; De Clercq, E.; McGuigan, C. Mechanism of anti-HIV action of masked alaninyl d4T-MP derivatives. *Proc. Natl. Acad. Sci. U. S. A.* **1996**, *93*, 7295-7299.
73. Balzarini, J.; Egberink, H.; Hartmann, K.; Cahard, D.; Vahlenkamp, T.; Thormar, H.; De Clercq, E.; McGuigan, C. Antiretrovirus specificity and intracellular metabolism of 2',3' - didehydro-2',3'-dideoxythymidine (stavudine) and its 5'-monophosphate triester prodrug So324. *Mol. Pharmacol.* **1996**, *50*, 1207-1213.
74. Naesens, L.; Cahard, D.; Salgado, A.; Bidois, L.; De Clercq, E.; McGuigan, C.; Balzarini, J. Metabolism and anti-HIV activity of phosphoramidate derivatives of d4T-MP with variations in the amino acid moiety. *Adv. Exp. Med. Biol.* **1998**, *431*, 753-757.
75. Saboulard, D.; Naesens, L.; Cahard, D.; Salgado, A.; Pathirana, R.; Velazquez, S.; McGuigan, C.; De Clercq, E.; Balzarini, J. Characterization of the activation pathway of phosphoramidate triester prodrugs of stavudine and zidovudine. *Mol. Pharmacol.* **1999**, *56*, 693-704.
76. Birkus, G.; Wang, R.; Liu, X.; Kutty, N.; MacArthur, H.; Cihlar, T.; Gibbs, C.; Swaminathan, S.; Lee, W.; McDermott, M. Cathepsin A is the major hydrolase catalyzing the intracellular hydrolysis of the antiretroviral nucleotide phosphonoamidate prodrugs GS-7340 and GS-9131. *Antimicrob. Agents Chemother.* **2007**, *51*, 543-550.
77. Mehellou, Y.; Valente, R.; Mottram, H.; Walsby, E.; Mills, K. I.; Balzarini, J.; McGuigan, C. Phosphoramidates of 2'-beta-D-arabinouridine (AraU) as phosphate prodrugs; design, synthesis, in vitro activity and metabolism. *Bioorg. Med. Chem.* **2010**, *18*, 2439-2446.
78. Osgerby, L.; Lai, Y. C.; Thornton, P. J.; Amalfitano, J.; Le Duff, C. S.; Jabeen, I.; Kadri, H.; Miccoli, A.; Tucker, J. H.; Muqit, M. M.; Mehellou, Y. Kinetin riboside and its ProTides activate the Parkinson's disease associated PTEN-induced putative kinase 1 (PINK1) independent of mitochondrial depolarization. *J. Med. Chem.* **2017**, *60*, 3518-3524.
79. McGuigan, C.; Murziani, P.; Slusarczyk, M.; Gonczy, B.; Vande Voorde, J.; Liekens, S.; Balzarini, J. Phosphoramidate ProTides of the anticancer agent FUDR successfully

- deliver the preformed bioactive monophosphate in cells and confer advantage over the parent nucleoside. *J. Med. Chem.* **2011**, *54*, 7247-7258.
80. Lee, W. A.; He, G. X.; Eisenberg, E.; Cihlar, T.; Swaminathan, S.; Mulato, A.; Cundy, K. C. Selective intracellular activation of a novel prodrug of the human immunodeficiency virus reverse transcriptase inhibitor tenofovir leads to preferential distribution and accumulation in lymphatic tissue. *Antimicrob. Agents Chemother.* **2005**, *49*, 1898-1906.
81. Murakami, E.; Tolstykh, T.; Bao, H.; Niu, C.; Steuer, H. M.; Bao, D.; Chang, W.; Espiritu, C.; Bansal, S.; Lam, A. M.; Otto, M. J.; Sofia, M. J.; Furman, P. A. Mechanism of activation of PSI-7851 and its diastereoisomer PSI-7977. *J. Biol. Chem.* **2010**, *285*, 34337-34347.
82. Birkus, G.; Kutty, N.; He, G. X.; Mulato, A.; Lee, W.; McDermott, M.; Cihlar, T. Activation of 9-[(R)-2-[[[(S)-[[[(S)-1-(Isopropoxycarbonyl)ethyl]amino] phenoxyphosphinyl]-methoxy]propyl]adenine (GS-7340) and other tenofovir phosphonoamidate prodrugs by human proteases. *Mol. Pharmacol.* **2008**, *74*, 92-100.
83. McGuigan, C.; Sutton, P. W.; Cahard, D.; Turner, K.; O'Leary, G.; Wang, Y.; Gumbleton, M.; De Clercq, E.; Balzarini, J. Synthesis, anti-human immunodeficiency virus activity and esterase lability of some novel carboxylic ester-modified phosphoramidate derivatives of stavudine (d4T). *Antiviral Chem. Chemother.* **1998**, *9*, 473-479.
84. Siddiqui, A. Q.; McGuigan, C.; Ballatore, C.; Zuccotto, F.; Gilbert, I. H.; De Clercq, E.; Balzarini, J. Design and synthesis of lipophilic phosphoramidate d4T-MP prodrugs expressing high potency against HIV in cell culture: structural determinants for in vitro activity and QSAR. *J. Med. Chem.* **1999**, *42*, 4122-4128.
85. McGuigan, C.; Tsang, H. W.; Cahard, D.; Turner, K.; Velazquez, S.; Salgado, A.; Bidois, L.; Naesens, L.; De Clercq, E.; Balzarini, J. Phosphoramidate derivatives of d4T as inhibitors of HIV: the effect of amino acid variation. *Antiviral Res.* **1997**, *35*, 195-204.
86. Anderson, B. M.; Cordes, E. H.; Jencks, W. P. Reactivity and catalysis in reactions of the serine hydroxyl group and of O-acyl serines. *J. Biol. Chem.* **1961**, *236*, 455-463.

87. Brenner, C. Hint, Fhit, and GalT: function, structure, evolution, and mechanism of three branches of the histidine triad superfamily of nucleotide hydrolases and transferases. *Biochemistry* **2002**, *41*, 9003-9014.
88. McIntee, E. J.; Remmel, R. P.; Schinazi, R. F.; Abraham, T. W.; Wagner, C. R. Probing the mechanism of action and decomposition of amino acid phosphomonoester amidates of antiviral nucleoside prodrugs. *J. Med. Chem.* **1997**, *40*, 3323-3331.
89. Cheng, J.; Zhou, X.; Chou, T. F.; Ghosh, B.; Liu, B.; Wagner, C. R. Identification of the amino acid-AZT-phosphoramidase by affinity T7 phage display selection. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 6379-6381.
90. Birkus, G.; Kutty, N.; Frey, C. R.; Shribata, R.; Chou, T.; Wagner, C.; McDermott, M.; Cihlar, T. Role of cathepsin A and lysosomes in the intracellular activation of novel antipapillomavirus agent GS-9191. *Antimicrob. Agents Chemother.* **2011**, *55*, 2166-2173.
91. Balzarini, J.; Kruining, J.; Wedgwood, O.; Pannecouque, C.; Aquaro, S.; Perno, C. F.; Naesens, L.; Witvrouw, M.; Heijtkink, R.; De Clercq, E.; McGuigan, C. Conversion of 2',3'-dideoxyadenosine (ddA) and 2',3'-didehydro-2',3'-dideoxyadenosine (d4A) to their corresponding aryloxyphosphoramidate derivatives markedly potentiates their activity against human immunodeficiency virus and hepatitis B virus. *FEBS Lett.* **1997**, *410*, 324-328.
92. McGuigan, C.; Hassan-Abdallah, A.; Srinivasan, S.; Wang, Y.; Siddiqui, A.; Daluge, S. M.; Gudmundsson, K. S.; Zhou, H.; McLean, E. W.; Peckham, J. P.; Burnette, T. C.; Marr, H.; Hazen, R.; Condreay, L. D.; Johnson, L.; Balzarini, J. Application of phosphoramidate ProTide technology significantly improves antiviral potency of carbocyclic adenosine derivatives. *J. Med. Chem.* **2006**, *49*, 7215-7226.
93. Balzarini, J.; Aquaro, S.; Hassan-Abdallah, A.; Daluge, S. M.; Perno, C. F.; McGuigan, C. Improved antiviral activity of the aryloxymethoxyalaninyl phosphoramidate (APA) prodrug of abacavir (ABC) is due to the formation of markedly increased carbovir 5'-triphosphate metabolite levels. *FEBS Lett.* **2004**, *573*, 38-44.

94. McGuigan, C.; Harris, S. A.; Daluge, S. M.; Gudmundsson, K. S.; McLean, E. W.; Burnette, T. C.; Marr, H.; Hazen, R.; Condreay, L. D.; Johnson, L.; De Clercq, E.; Balzarini, J. Application of phosphoramidate pronucleotide technology to abacavir leads to a significant enhancement of antiviral potency. *J. Med. Chem.* **2005**, *48*, 3504-3515.
95. Lisco, A.; Vanpouille, C.; Tchesnokov, E. P.; Grivel, J. C.; Biancotto, A.; Brichacek, B.; Elliott, J.; Fromentin, E.; Shattock, R.; Anton, P.; Gorelick, R.; Balzarini, J.; McGuigan, C.; Derudas, M.; Gotte, M.; Schinazi, R. F.; Margolis, L. Acyclovir is activated into a HIV-1 reverse transcriptase inhibitor in herpesvirus-infected human tissues. *Cell Host Microbe* **2008**, *4*, 260-270.
96. Vanpouille, C.; Lisco, A.; Derudas, M.; Saba, E.; Grivel, J. C.; Brichacek, B.; Scrimieri, F.; Schinazi, R.; Schols, D.; McGuigan, C.; Balzarini, J.; Margolis, L. A new class of dual-targeted antivirals: monophosphorylated acyclovir prodrug derivatives suppress both human immunodeficiency virus type 1 and herpes simplex virus type 2. *J. Infect. Dis.* **2010**, *201*, 635-643.
97. Hertel, L. W.; Boder, G. B.; Kroin, J. S.; Rinzel, S. M.; Poore, G. A.; Todd, G. C.; Grindey, G. B. Evaluation of the antitumor activity of gemcitabine (2',2'-difluoro-2'-deoxycytidine). *Cancer Res.* **1990**, *50*, 4417-4422.
98. Saif, M. W.; Lee, Y.; Kim, R. Harnessing gemcitabine metabolism: a step towards personalized medicine for pancreatic cancer. *Ther. Adv. Med. Oncol.* **2012**, *4*, 341-346.
99. Camiener, G. W.; Smith, C. G. Studies of enzymatic deamination of cytosine arabinoside .1. Enzyme distribution and species specificity. *Biochem. Pharmacol.* **1965**, *14*, 1405-1416.
100. Ghazaly, E. A.; Slusarczyk, M.; Mason, M.; Gribben, J.; McGuigan, C.; Blagden, S. NUC-1031: a novel ProTide that overcomes the key cancer resistance mechanisms associated with poor survival. *Cancer Res.* **2014**, *74*, CT401.
101. Blagden, S. P.; Rizzuto, I.; Stavrika, C.; O'Shea, D.; Suppiah, P.; Patel, M. K.; Sukumaran, A.; Loyse, N.; Bharwani, N.; Rockall, A.; Gabra, H.; El-Bahrawy, M.; Wasan, H. S.; Leonard, R. C.; Habib, N. A.; Gribben, J. G.; Ghazaly, E. A.; McGuigan, C. Final

- results of ProGem1, the first in-human phase I/II study of NUC-1031 in patients with solid malignancies. *J. Clin. Oncol.* **2015**, *33*, 2514.
102. Vande Voorde, J.; Liekens, S.; McGuigan, C.; Murziani, P. G.; Slusarczyk, M.; Balzarini, J. The cytostatic activity of NUC-3073, a phosphoramidate prodrug of 5-fluoro-2'-deoxyuridine, is independent of activation by thymidine kinase and insensitive to degradation by phosphorolytic enzymes. *Biochem. Pharmacol.* **2011**, *82*, 441-452.
103. Ghazaly, E. A.; Slusarczyk, M.; McGuigan, C.; Harrison, D.; Blagden, S. P. NUC-3373: A novel pyrimidine nucleotide analogue that overcomes key cancer drug resistance limiting patient survival. *Mol. Cancer Ther.* **2016**, *14*, B46.
104. Alexander, P.; Kucera, G.; Pardee, T. S. Improving nucleoside analogs via lipid conjugation: Is fatter any better? *Crit. Rev. Oncol. Hematol.* **2016**, *100*, 46-56.
105. Warren, T. K.; Jordan, R.; Lo, M. K.; Ray, A. S.; Mackman, R. L.; Soloveva, V.; Siegel, D.; Perron, M.; Bannister, R.; Hui, H. C.; Larson, N.; Strickley, R.; Wells, J.; Stuthman, K. S.; Van Tongeren, S. A.; Garza, N. L.; Donnelly, G.; Shurtleff, A. C.; Retterer, C. J.; Gharaibeh, D.; Zamani, R.; Kenny, T.; Eaton, B. P.; Grimes, E.; Welch, L. S.; Gomba, L.; Wilhelmsen, C. L.; Nichols, D. K.; Nuss, J. E.; Nagle, E. R.; Kugelman, J. R.; Palacios, G.; Doerffler, E.; Neville, S.; Carra, E.; Clarke, M. O.; Zhang, L.; Lew, W.; Ross, B.; Wang, Q.; Chun, K.; Wolfe, L.; Babusis, D.; Park, Y.; Stray, K. M.; Trancheva, I.; Feng, J. Y.; Barauskas, O.; Xu, Y.; Wong, P.; Braun, M. R.; Flint, M.; McMullan, L. K.; Chen, S. S.; Fearn, R.; Swaminathan, S.; Mayers, D. L.; Spiropoulou, C. F.; Lee, W. A.; Nichol, S. T.; Cihlar, T.; Bavari, S. Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys. *Nature* **2016**, *531*, 381-385.
106. Lo, M. K.; Jordan, R.; Arvey, A.; Sudhamsu, J.; Shrivastava-Ranjan, P.; Hotard, A. L.; Flint, M.; McMullan, L. K.; Siegel, D.; Clarke, M. O.; Mackman, R. L.; Hui, H. C.; Perron, M.; Ray, A. S.; Cihlar, T.; Nichol, S. T.; Spiropoulou, C. F. GS-5734 and its parent nucleoside analog inhibit Filo-, Pneumo-, and Paramyxoviruses. *Sci. Rep.* **2017**, *7*, 43395.

107. Lawitz, E.; Mangia, A.; Wyles, D.; Rodriguez-Torres, M.; Hassanein, T.; Gordon, S. C.; Schultz, M.; Davis, M. N.; Kayali, Z.; Reddy, K. R.; Jacobson, I. M.; Kowdley, K. V.; Nyberg, L.; Subramanian, G. M.; Hyland, R. H.; Arterburn, S.; Jiang, D.; McNally, J.; Brainard, D.; Symonds, W. T.; McHutchison, J. G.; Sheikh, A. M.; Younossi, Z.; Gane, E. J. Sofosbuvir for previously untreated chronic hepatitis C infection. *N. Engl. J. Med.* **2013**, *368*, 1878-1887.
108. Kattakuzhy, S.; Levy, R.; Kottlil, S. Sofosbuvir for treatment of chronic hepatitis C. *Hepatol. Int.* **2015**, *9*, 161-173.
109. Kirby, B. J.; Symonds, W. T.; Kearney, B. P.; Mathias, A. A. Pharmacokinetic, pharmacodynamic, and drug-interaction profile of the hepatitis C virus NS5B polymerase inhibitor Sofosbuvir. *Clin. Pharmacokinet.* **2015**, *54*, 677-690.
110. Denning, J.; Cornpropst, M.; Flach, S. D.; Berrey, M. M.; Symonds, W. T. Pharmacokinetics, safety, and tolerability of GS-9851, a nucleotide analog polymerase inhibitor for hepatitis C virus, following single ascending doses in healthy subjects. *Antimicrob. Agents Chemother.* **2013**, *57*, 1201-1208.
111. Rodriguez-Torres, M.; Lawitz, E.; Kowdley, K. V.; Nelson, D. R.; Dejesus, E.; McHutchison, J. G.; Cornpropst, M. T.; Mader, M.; Albanis, E.; Jiang, D.; Hebner, C. M.; Symonds, W. T.; Berrey, M. M.; Lalezari, J. Sofosbuvir (GS-7977) plus peginterferon/ribavirin in treatment-naive patients with HCV genotype 1: a randomized, 28-day, dose-ranging trial. *J. Hepatol.* **2013**, *58*, 663-668.
112. Cholongitas, E.; Papatheodoridis, G. V. Sofosbuvir: a novel oral agent for chronic hepatitis C. *Ann. Gastroenterol.* **2014**, *27*, 331-337.
113. Papatheodoridis, G. V.; Dimou, E.; Papadimitropoulos, V. Nucleoside analogues for chronic hepatitis B: antiviral efficacy and viral resistance. *Am. J. Gastroenterol.* **2002**, *97*, 1618-1628.
114. Lam, A. M.; Espiritu, C.; Bansal, S.; Micolochick Steuer, H. M.; Niu, C.; Zennou, V.; Keilman, M.; Zhu, Y.; Lan, S.; Otto, M. J.; Furman, P. A. Genotype and subtype profiling

- of PSI-7977 as a nucleotide inhibitor of hepatitis C virus. *Antimicrob. Agents Chemother.* **2012**, *56*, 3359-3368.
115. Jacobson, I. M.; Gordon, S. C.; Kowdley, K. V.; Yoshida, E. M.; Rodriguez-Torres, M.; Sulkowski, M. S.; Shiffman, M. L.; Lawitz, E.; Everson, G.; Bennett, M.; Schiff, E.; Al-Assi, M. T.; Subramanian, G. M.; An, D.; Lin, M.; McNally, J.; Brainard, D.; Symonds, W. T.; McHutchison, J. G.; Patel, K.; Feld, J.; Pianko, S.; Nelson, D. R. Sofosbuvir for hepatitis C genotype 2 or 3 in patients without treatment options. *N. Engl. J. Med.* **2013**, *368*, 1867-1877.
116. Zeuzem, S.; Dusheiko, G. M.; Salupere, R.; Mangia, A.; Flisiak, R.; Hyland, R. H.; Illeperuma, A.; Svarovskaia, E.; Brainard, D. M.; Symonds, W. T.; Subramanian, G. M.; McHutchison, J. G.; Weiland, O.; Reesink, H. W.; Ferenci, P.; Hezode, C.; Esteban, R. Sofosbuvir and ribavirin in HCV genotypes 2 and 3. *N. Engl. J. Med.* **2014**, *370*, 1993-2001.
117. Lynch, S. M.; Wu, G. Y. Hepatitis C Virus: A Review of treatment guidelines, cost-effectiveness, and access to therapy. *J. Clin. Transl. Hepatol.* **2016**, *4*, 310-319.
118. De Clercq, E. Tenofovir alafenamide (TAF) as the successor of tenofovir disoproxil fumarate (TDF). *Biochem. Pharmacol.* **2016**, *119*, 1-7.
119. Gibson, A. K.; Shah, B. M.; Nambiar, P. H.; Schafer, J. J. Tenofovir alafenamide. *Ann. Pharmacother.* **2016**, *50*, 942-952.
120. Menendez-Arias, L.; Alvarez, M.; Pacheco, B. Nucleoside/nucleotide analog inhibitors of hepatitis B virus polymerase: mechanism of action and resistance. *Curr. Opin. Virol.* **2014**, *8*, 1-9.
121. Babusis, D.; Phan, T. K.; Lee, W. A.; Watkins, W. J.; Ray, A. S. Mechanism for effective lymphoid cell and tissue loading following oral administration of nucleotide prodrug GS-7340. *Mol. Pharm.* **2013**, *10*, 459-466.
122. Ray, A. S.; Fordyce, M. W.; Hitchcock, M. J. Tenofovir alafenamide: a novel prodrug of tenofovir for the treatment of human immunodeficiency virus. *Antiviral Res.* **2016**, *125*, 63-70.

123. Eisenberg, E. J.; He, G. X.; Lee, W. A. Metabolism of GS-7340, a novel phenyl monophosphoramidate intracellular prodrug of PMPA, in blood. *Nucleosides, Nucleotides Nucleic Acids* **2001**, *20*, 1091-1098.
124. Markowitz, M.; Zolopa, A.; Squires, K.; Ruane, P.; Coakley, D.; Kearney, B.; Zhong, L.; Wulfsohn, M.; Miller, M. D.; Lee, W. A. Phase I/II study of the pharmacokinetics, safety and antiretroviral activity of tenofovir alafenamide, a new prodrug of the HIV reverse transcriptase inhibitor tenofovir, in HIV-infected adults. *J. Antimicrob. Chemother.* **2014**, *69*, 1362-1369.
125. Ray, A. S.; Fordyce, M. W.; Hitchcock, M. J. Tenofovir alafenamide: a novel prodrug of tenofovir for the treatment of human immunodeficiency virus. *Antiviral Res.* **2016**, *125*, 63-70.
126. Sax, P. E.; Wohl, D.; Yin, M. T.; Post, F.; DeJesus, E.; Saag, M.; Pozniak, A.; Thompson, M.; Podzamczar, D.; Molina, J. M.; Oka, S.; Koenig, E.; Trottier, B.; Andrade-Villanueva, J.; Crofoot, G.; Custodio, J. M.; Plummer, A.; Zhong, L.; Cao, H.; Martin, H.; Callebaut, C.; Cheng, A. K.; Fordyce, M. W.; McCallister, S.; Team, G.-U.-S. Tenofovir alafenamide versus tenofovir disoproxil fumarate, coformulated with elvitegravir, cobicistat, and emtricitabine, for initial treatment of HIV-1 infection: two randomised, double-blind, phase 3, non-inferiority trials. *Lancet* **2015**, *385*, 2606-2615.
127. Agarwal, K.; Fung, S. K.; Nguyen, T. T.; Cheng, W.; Sicard, E.; Ryder, S. D.; Flaherty, J. F.; Lawson, E.; Zhao, S.; Subramanian, G. M.; McHutchison, J. G.; Gane, E. J.; Foster, G. R. Twenty-eight day safety, antiviral activity, and pharmacokinetics of tenofovir alafenamide for treatment of chronic hepatitis B infection. *J. Hepatol.* **2015**, *62*, 533-540.
128. McGuigan, C.; Serpi, M.; Bibbo, R.; Roberts, H.; Hughes, C.; Caterson, B.; Gibert, A. T.; Verson, C. R. Phosphate prodrugs derived from N-acetylglucosamine have enhanced chondroprotective activity in explant cultures and represent a new lead in antiosteoarthritis drug discovery. *J. Med. Chem.* **2008**, *51*, 5807-5812.

129. Serpi, M.; Bibbo, R.; Rat, S.; Roberts, H.; Hughes, C.; Caterson, B.; Alcaraz, M. J.; Gibert, A. T.; Verson, C. R.; McGuigan, C. Novel phosphoramidate prodrugs of N-acetyl-(D)-glucosamine with antidegenerative activity on bovine and human cartilage explants. *J. Med. Chem.* **2012**, *55*, 4629-4639.
130. James, E.; Pertusati, F.; Brancale, A.; McGuigan, C. Kinase-independent phosphoramidate S1P1 receptor agonist benzyl ether derivatives. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 1371-1378.
131. Ruda, G. F.; Alibu, V. P.; Mitsos, C.; Bidet, O.; Kaiser, M.; Brun, R.; Barrett, M. P.; Gilbert, I. H. Synthesis and biological evaluation of phosphate prodrugs of 4-phospho-D-erythronhydroxamic acid, an inhibitor of 6-phosphogluconate dehydrogenase. *ChemMedChem* **2007**, *2*, 1169-1180.
132. Ruda, G. F.; Wong, P. E.; Alibu, V. P.; Norval, S.; Read, K. D.; Barrett, M. P.; Gilbert, I. H. Aryl phosphoramidates of 5-phospho erythronhydroxamic acid, a new class of potent trypanocidal compounds. *J. Med. Chem.* **2010**, *53*, 6071-6078.

Table of Contents Graphic

