

doi: 10.1093/femsle/fnaa010 Advance Access Publication Date: 16 January 2020 Minireview

# MINIREVIEW – Environmental Microbiology

# Understanding the challenges of non-food industrial product contamination

Edward Cunningham-Oakes<sup>1,†</sup>, Rebecca Weiser<sup>1,†</sup>, Tom Pointon<sup>2</sup> and Eshwar Mahenthiralingam<sup>1,\*,‡</sup>

<sup>1</sup>Microbiomes, Microbes and Informatics Group, Organisms and Environment Division, Cardiff School of Biosciences, Cardiff University, Park Place, Cardiff, UK, Wales CF10 3AX, UK and <sup>2</sup>Unilever Research and Development, Port Sunlight, Wirral, CH62 4ZD, UK

\*Corresponding author: Organisms and Environment Division, School of Biosciences, Cardiff University, Main Building, Room 0.23, Museum Avenue, Cardiff CF10 3AX, United Kingdom. Tel: +44 0 29 20875875; Fax: +44 0 29 20874305; E-mail: mahenthiralingame@cardiff.ac.uk

**One sentence summary:** Microbial contamination in non-food industrial products was reviewed and it is shown that while antimicrobial resistant Gram-negative species are the most common cause, nearly 50% of recorded recall incidents report the microorganisms present as unidentified. <sup>†</sup>Equal contribution.

Editor: Laura Bowater

<sup>‡</sup>Eshwar Mahenthiralingam, http://orcid.org/0000-0001-9014-3790

#### ABSTRACT

Preventing microbial contamination of non-food products is a major area of industrial microbiology where preservatives are used to stop microbial growth. However, microorganisms occasionally overcome product preservation, causing recalls and the implementation of multiple procedures to prevent further contamination. Correct reporting of microbial contamination in non-food industrial products is vital, especially if spoilage organisms are antimicrobial resistant and pose a health threat. Gram-negative bacteria such as *Pseudomonas, Burkholderia* and *Enterobacteriaceae* are frequently reported as non-food product contaminants, including species that overlap current antimicrobial resistance priorities. Historical analysis of recall databases highlighted that for greater than 15% of contamination incidents, the causative microbial agents are reported as unidentified. Here we review the current antimicrobial resistant bacterial species associated with non-food product contaminants are reported as unidentified despite frequent detection of antimicrobial resistant pathogens; in contrast, 98% of food-related microbial contaminants are classified. Recommendations to fill this microbial identification gap in non-food product recalls are made. Overall, reporting standards for microbial contamination in non-food products must be improved to enable surveillance and for understanding the risks associated with antimicrobial resistant microorganisms

Keywords: contamination; non-food products; antimicrobial resistance; classification; identification

Preventing microbial spoilage is a key issue for several industrial sectors including the manufacture and sale of non-food products such as home care products (cleaning products), personal care products (cosmetics and toiletries) and toys. Preservatives and antimicrobial formulations are used to prevent microbial growth in non-food products, but since manufacture of these products is not a sterile process, growth of certain resilient and intrinsically antimicrobial resistant (AMR) microorganisms may occur. Non-food product microbial contamination is a costly issue for industry and reporting of incidents

© FEMS 2020. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Received: 30 May 2019; Accepted: 14 January 2020

associated with publicly recalled products is required. We review bacterial groups most commonly associated with non-food product contamination, focussing on *Pseudomonas aeruginosa*, *Burkholderia cepacia* complex and members of *Enterobacteriaceae*. Evaluation of current recall data for Europe as part of this exercise worryingly identified that half of all non-food microbiological contamination incidents reported the causative agent as unidentified. The issues behind the lack of microbial contaminant identification in the non-food product sector are discussed and recommendations to improve reporting are made.

The emergence of treatment refractory infections such as the notorious ESKAPE pathogens (Enterococcus faecium, Staphylococcus aureus, Klebsiella pneumoniae, Acinetobacter baumannii, P. aeruginosa and Enterobacter spp.) has been extensively highlighted, and these organisms contribute greatly to patient morbidity and mortality (Pendleton, Gorman and Gilmore 2013). Intrinsically resistant microorganisms are also problematic in manufacturing settings and can overcome antimicrobial preservatives to contaminate industrial products. Pseudomonas and multiple Enterobacteriaceae (eg. Enterobacter and Klebsiella) are globally recognised clinical AMR threats (Tacconelli et al. 2018a), but they and other intrinsically resistant bacteria such as Burkholderia may also be recovered as microbial contaminants of non-food products (Jimenez 2007; Sutton and Jimenez 2012; Vincze, Al Dahouk and Dieckmann 2019). Multiple Gram-negative bacteria have intrinsic resistance and the potential to acquire AMR elements (Pendleton, Gorman and Gilmore 2013), but what is their role in non-sterile product contamination and do they constitute potential public health risks? The identification and tracking of antimicrobial resistant contaminants in non-food consumer products is far less stringent than in clinical settings and the food product industry, making a thorough risk assessment difficult. In this review we also expand on key non-food product contaminating AMR bacteria and issues in relation to their identification and reporting.

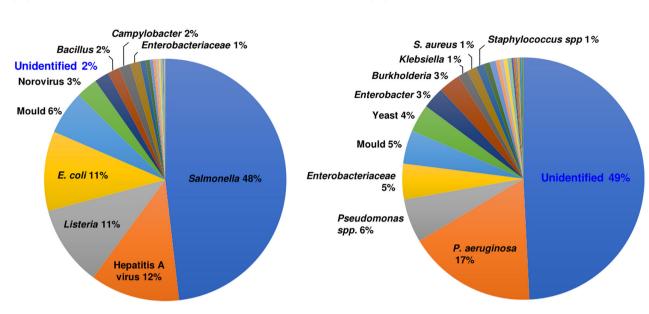
### MICROBIAL ISSUES ASSOCIATED WITH THE MANUFACTURE OF NON-STERILE INDUSTRIAL PRODUCTS

Non-food consumer products encompass an extensive range of items including cosmetics, personal hygiene and home care products, chemical products such as tattoo ink and children's toys such as soap bubbles, plasticine and paints (European Commission 2018). These products are manufactured, sold and used world-wide and are not required to be sterile, but must be safe for use (Orus and Leranoz 2005). Antimicrobial preservatives are incorporated into products to maintain their quality, extend shelf life and protect the consumer (Neza and Centini 2016). If a product is inadequately preserved, microbial contamination may occur which can lead to product spoilage and recall, financial or reputational damage to the manufacturer, and more seriously, poses a risk to human health (Orth et al. 2006). Alarmingly, the presence of opportunistic pathogens in non-food products has been linked with infections in vulnerable consumers (Lundov et al. 2009). Whilst incidents are most likely to be reported in hospital settings, contaminated products do reach the market place, as has been shown by product recall data and surveys of off-the-shelf products (Lundov et al. 2009). Despite being a major area of routine global microbiology, the risks stemming from contaminated non-food products are poorly characterised and literature in this area is limited.

#### RECALL INFORMATION IS USEFUL FOR SURVEILLANCE OF MICROBIAL CONTAMINATION

As there is no requirement for industry to publish information concerning products which have failed quality control or contamination incidents, systematic literature in this area is limited. Product recall data, however, is collated by the US Food and Drug Administration (FDA) (Food and Drug Administration 2017) and the European Commission (European Commission 2018), and made publicly available. Reports based on FDA data for personal care and non-sterile pharmaceutical products (Jimenez 2007; Sutton and Jimenez 2012) and Safety Gate (previously known as RAPEX: the rapid alert system for non-food consumer products) (European Commission 2018) in the EU (Wong et al. 2000; Lundov et al. 2009), indicate the range of microorganisms found as contaminants. A report published in 2007 (Jimenez 2007) surveying the FDA database from 1995 to 2006, documented that Gram-negative bacteria accounted for 60% of product recalls, with Grampositive bacteria and fungal species accounting for approximately 4% and 23%, respectively. Microorganisms recorded as unidentified comprised 22% of the recall cases evaluated at the time (Jimenez 2007: 383-99). A subsequent update examined FDA recalls from 2004 to 2011 (Sutton and Jimenez 2012), and identified a similar proportion of Gram-negative bacteria (50%), Gram-positive bacteria (10%) and fungal (19%) contaminants, but showed limited improvement in the number of recall incidents reported due to unidentified microorganisms (15%). Overall, these pioneering surveillance reports clearly showed that Gram-negative bacterial species caused over half of the incidents which resulted in recall of non-sterile industrial products, and also demonstrated that a significant proportion of the non-food product contamination microorganisms were left unidentified.

Gram-negative bacterial species stand out as the most common contaminants of non-sterile industrial products. Historically, P. aeruginosa and B. cepacia complex bacteria, have been recognised as problematic, objectionable industrial contaminants, identified in a high proportion of recall incidents. P. aeruginosa accounted for 14% and 6% of recall incidents respectively for 1995 to 2006 and 2004 to 2011 FDA database surveys (Jimenez 2007; Sutton and Jimenez 2012); over the same time periods B. cepacia complex bacteria were identified in 22% and 25% of incident surveys (Jimenez 2007; Sutton and Jimenez 2012). P. aeruginosa was the most common contaminant reported in recalled cosmetic products in a short survey (2005 to 2008) of the RAPEX Safety Gate database (Lundov and Zachariae 2008). Enterobacteriaceae are also prominent contaminants with significant numbers of Enterobacter species identified within recalled products in the United States (Jimenez 2007; Sutton and Jimenez 2012). A recent update examining contaminant notifications in the Safety Gate database between 2005 and 2017 also found that P. aeruginosa, other Pseudomonas species and Enterobacteriaceae ranked as the top three identified microorganisms encountered in contaminated toys, cosmetics and chemical products (Vincze, Al Dahouk and Dieckmann 2019). However, Vincze et al. (Vincze, Al Dahouk and Dieckmann 2019) also determined that the main microbial hazard was an unacceptably high total count of unidentified aerobic microorganisms; 218 out of 240 products were recalled for this reason, only half of which had additional microbial identification information.



# (A) Food

# (B) Non-food

Figure 1. Lack of microbial species identification in non-food product recalls compared to food products. The diversity of microorganisms responsible for product recalls in the European databases was analysed for food and non-food products between 2005 and 2018. The proportion of organism types detected for the 7577 food product and 378 non-food product isolations made in recall incidents is shown in panels A and B, respectively. Each labelled segment highlights where organisms represented  $\geq 1\%$  of the total number of recall reports in the period. Unidentified organisms in each database are highlighted in the blue font.

# THE IDENTIFICATION GAP: A LACK OF ORGANISM CLASSIFICATION FOR NON-FOOD PRODUCT RECALL INCIDENTS

The previously mentioned meta-analyses of both FDA and EU product recall databases have determined that unidentified microorganisms are associated with a large proportion of product recalls. To update these past analyses (Jimenez 2007; Lundov and Zachariae 2008; Sutton and Jimenez 2012), investigate recall reporting in different industrial sectors, and determine the extent of AMR microorganisms within contamination incidents, we conducted a meta-analysis of both food and non-food product recalls between 2005 and 2018 in the EU. To do this we used two publicly available EU databases, (i) the European Commission Rapid Alert System for Food and Feed (RASFF) (European Commission 2019) to search food product recalls, and (ii) Safety Gate (European Commission 2018) to search non-food product recalls. These databases record a variety of parameters for each recall including the nature and severity of risk, product type, country of origin, notifying country and overall outcomes. In relation to contaminated foods, searches of RASFF database were performed for: (i) product type 'Food', and (ii) hazard type 'Pathogenic microorganisms' or 'Non-pathogenic microorganisms' and each incident recorded within a spreadsheet. An analogous search of the Safety Gate database for all product categories and the risk type 'Microbiological' was also performed. The classification of each reported microorganism was recorded to the family, genus or species level. Organisms not classified to at least family level or grouped together under collective descriptions such as 'coliforms', 'moulds' or 'fungus', were designated 'unidentified'. If multiple organisms were found in a product recall, each organism was tallied separately.

For food products over the 2005 to 2018 period, a total of 7577 microorganisms were reported in 1016 recalls, while in contrast a total of 378 microorganisms were reported for 254 non-food product incidents over the same time period. To probe the extent of useful epidemiological information within the RASFF and Safety Gate databases, information related to the identity of the contaminating organism for each product recall was examined. Proportional analysis of the microbial groups encountered in food and non-food product contamination revealed major differences in the type of microorganisms reported in recalls (Fig. 1). The top five microorganisms encountered in food product recalls in rank order were Salmonella, Hepatitis A virus, Listeria, Escherichia coli and moulds (Fig. 1A). Given the major importance of Salmonella as a food-bourne pathogen it was not surprising that this Gram-negative genus was identified in 48% of food product recalls. An additional key feature of the recall information in the food product database was the accuracy of identification, with 98% of organisms being identified and only 2% being reported as unidentified (Fig. 1A).

In striking contrast to the food related recalls, 49% of microorganisms associated with non-food product recalls were reported as unidentified (Fig. 1B). After this, the top ten most common microorganisms in non-food product recalls, from highest to lowest prevalence, were P. aeruginosa, Pseudomonas species, Enterobacteriaceae, Moulds, Yeasts, Enterobacter, Burkholderia, Klebsiella, S. aureus and Staphylococcus species (Fig. 1B). Overall, the intrinsically antimicrobial resistant Gram-negative bacteria Pseudomonas, Enterobacteriaceae, Enterobacter, Burkholderia, Klebsiella and Achromobacter, were associated with 30% of non-food product recalls (Fig. 1B). In addition, the ESKAPE pathogen group of bacteria (Pendleton, Gorman and Gilmore 2013) accounted for 23% of all non-food recalls (data not shown). If the same proportions of antimicrobial resistant ESKAPE species are mirrored in the 49% of recall microorganisms recorded as unidentified, then the non-food product manufacturing industry is considerably under-reporting potential hazards in terms of global health risk and AMR. Given the recent significant improvements in methods for accurate microbial identification (Maiden et al. 2013; van

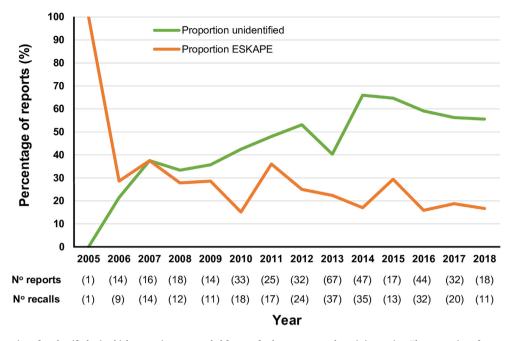


Figure 2. The proportion of unclassified microbial contaminants recorded for non-food consumer products is increasing. The proportion of reports of microbial contaminants in EU Safety Gate non-food product recalls from 2005 to 2018 associated with unidentified microorganisms (green line) and ESKAPE pathogens (*E. faecium*, *S. aureus*, *K. pneumoniae*, *A. baumannii*, *P. aeruginosa* and *Enterobacter* spp.; orange line). The total number of reports of microorganisms and corresponding number of product recalls is given below the year in brackets.

Belkum et al. 2013) it would be expected that more contamination incidents would be accurately reported over time. Between 2005 to 2018, the number of non-food product recalls varied between 1 and 37 incidents per year, with the number of reports of microorganisms varying between 1 and 67 (Fig. 2). Proportionally, the lack of identification in reporting for non-food product recall microorganisms increased from 2010 onwards to a high in 2014 where 66% of incidents were reported as unidentified (Fig. 2). Overall, to the current time there remains a high proportion of unidentified contaminants (>55%) in non-food product recalls (Fig. 2).

#### NON-FOOD PRODUCT TYPES MOST AFFECTED BY MICROBIAL CONTAMINATION

The top 10 identified microorganisms in product recalls between 2005 and 2018 collectively represented 172 reports of microbial contamination, which were linked to 138 product recalls belonging to nine different product categories (Fig. 3; soap bubble products, face/hand/feet/body care, toys, hair care, make-up, baby care, tattoo ink, tooth care and chemical products). Soap bubbles (33% of reports) and face/hand/feet/body care (skin care) products (30% of reports) were the most commonly reported contaminated products (Fig. 3). In rank order below this, contamination of toys was linked to 13% of the microbial reports, while hair care, make-up, baby-care, tattoo ink, tooth care and undefined chemical products each accounted 6% or less (Fig. 3). P. aeruginosa showed the most widespread distribution being identified in recalls of eight of the nine product types (it was not found in chemical products), as well as being most commonly found in soap bubble and skin care products (Fig. 3). Other Pseudomonas species were encountered in four of the nine products types, and in correlation with P. aeruginosa, they affected soap bubble and skin care products most commonly. Enterobacteriaceae also showed a propensity to contaminate soap bubble and skin care products, and 10 of the 11 'Enterobacter' species-linked reports occurred specifically in skin care products (Fig. 3). Burkholderia, Klebsiella and S. aureus were most commonly encountered in recalled skin care products (Fig. 3). Overall, P. aeruginosa represented the most substantive risk of contamination across multiple non-food product types (Fig. 3).

#### FREQUENT CONTAMINANTS OF NON-FOOD PRODUCTS: P. AERUGINOSA

P. aeruginosa is an extremely versatile microorganism with the ability to survive in diverse habitats including soil, water, plant and animal tissues, community and hospital environments. In particular, its presence within water via natural, tap and potable sources is a recognised problem for manufacturing industries reliant on water as a major raw material (Jimenez 2007). P. aeruginosa is a rapidly growing microorganism and is straightforward to culture with a range of positive or negative selective media (Weiser et al. 2014). Standard industrial practices for microbiology and quality control are effective for recovering it as a contaminant from a range of products and industrial sources (White et al. 2011; Weiser et al. 2014). Accurate identification of P. aeruginosa can be achieved using multiple methods including biochemical profiling or matrix-assisted laser desorptionionisation time of flight mass spectroscopy (MALDI TOF MS) (van Belkum et al. 2013), 16S rRNA or oprL gene (De Vos et al. 1997) sequence analysis and international databases such as the public multilocus sequencing typing (MLST) resource (https://pubm lst.org/paeruginosa/) (Curran et al. 2004) can enable placement of strains in the context of their wider epidemiological prevalence.

Contamination of taps, surfaces, instruments and patients has been associated with multiple incidents of nosocomially acquired *P. aeruginosa* infections. The presence of the pathogen in home and personal care products has also initiated infections in hospital settings. A historical case associated with a

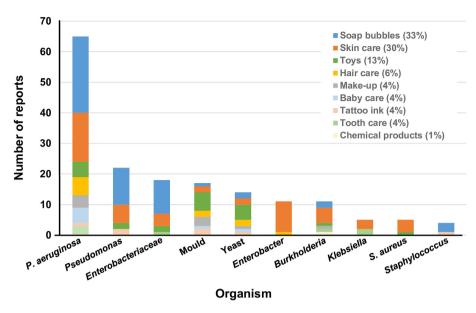


Figure 3. The microbial groups and species most commonly reported in non-food product types. The top 10 microorganisms identified from recalled non-food product types within the Safety Gate database are shown. The data derives from 172 reports of microbial contamination from 138 product recalls between 2005 to 2018 where an organism identification was provided for the non-food product type. The percentage of reports belonging to each product category is shown in the colour key at the top right of the figure.

contaminated hand lotion used by healthcare workers in a neonatal intensive care unit inadvertently led to vulnerable infants being infected (Becks and Lorenzoni 1995). Interestingly, the Enterobacteriaceae species Enterobacter agglomerans was also cultured from the contaminated hand lotion bottles, but did not cause infection in the infants (Becks and Lorenzoni 1995). More recently an outbreak with extensively drug-resistant P. aeruginosa was associated with sharing of an aromatherapy oil among hospitalised patients in an intensive care unit (Mayr et al. 2017). Such localised incidents of contamination leading to infection can be normally be controlled by infection control, but outbreaks in wider community settings are problematic to resolve. A national P. aeruginosa outbreak in the UK occurred among individuals undergoing ear piercing, which continued for 3 months in 2016 before it was stopped by a source tracing enabled product recall (Evans et al. 2018). Using variable Number Tandem Repeat (VNTR) strain typing and cohort analysis, the outbreak was linked to the use of a cosmetic after-care solution which had become contaminated during production (Evans et al. 2018). Within the same industrial sector of beauty and cosmetic products, contaminated tattoo ink has also been implicated in multiple cases of P. aeruginosa infection (Hogsberg et al. 2013). Overall, P. aeruginosa in addition to being frequently described as ubiquitous bacterial species found in multiple niches, is consistently encountered in relation to incidents of non-food product contamination (Jimenez 2007; Sutton and Jimenez 2012).

#### **B. CEPACIA COMPLEX BACTERIA**

*B. cepacia* complex bacteria are also historically linked with the ability to cause contamination in both clinical and non-food industrial settings (Torbeck *et al.* 2011). This group of Gramnegative bacteria has undergone multiple taxonomic revisions over the last 20 years with over 20 species defined within the *B. cepacia* complex (Depoorter *et al.* 2016). The historical prevalence of individual *B. cepacia* species in contamination is more difficult to assess since it was reported as '*Pseudomonas cepacia*' up to 1997 prior to taxonomic reclassification (Vandamme

et al. 1997), and even post 2010, it is still occasionally referred to as 'Burkholderia cepacia' in the published literature (Torbeck et al. 2011). Unlike P. aeruginosa, B. cepacia complex bacteria are not straightforward to identify, requiring selective media for their enrichment (Henry et al. 1999), and molecular tests such as recA gene sequencing (Mahenthiralingam et al. 2000) and MLST (Baldwin et al. 2005) for accurate identification. Surveillance and tracking of infections caused by B. cepacia complex bacteria has been extensive because of the problematic lung disease they cause in people with cystic fibrosis, and MLST resources for strain identification are comprehensive (Baldwin et al. 2005) (https://pubmlst.org/bcc/).

In the last 5 years, analysis of non-food product contamination caused by B. cepacia complex bacteria has been more detailed in relation to reporting taxonomic identity. Species associated with a variety of non-food product sources were accurately identified within a collection of 60 industrial isolates, and Burkholderia lata, Burkholderia cenocepacia and Burkholderia vietnamiensis were the top three species accounting for 25%, 18% and 13%, respectively, of the collection assessed (Rushton et al. 2013). In 2016, a strain of Burkholderia stabilis was shown to cause an outbreak associated with contaminated washing gloves in Switzerland using a comprehensive range of molecular and genomic tests (Sommerstein et al. 2017). Whole genome sequencing including whole genome MLST (Maiden et al. 2013) was carried out to identify that the B. stabilis isolates causing infections in multiple patients at different treatment centers were identical to those recovered from the contaminated gloves (Sommerstein et al. 2017). In the United States, a widespread outbreak associated with the laxative docusate occurred in 2016, and isolates causing the resulting infections in patients shown to be identical using a PCR genotyping method (Marquez et al. 2017). However further molecular analysis demonstrated that the docusate outbreak strain belonged to a potentially novel taxonomic group within the B. cepacia complex.

Although improvements in the level of resolution being applied to the investigation of *B. cepacia* complex contamination are clearly being made, studies may still fail to follow up on

the resources obtained during the investigation of outbreak incidents. For example, while whole genome sequencing was applied to demonstrate that a clonal B. cepacia complex strain present in a contaminated octenidine mouthwash had caused an outbreak of clinical infections, further follow-up to identify the strain to a species level was not performed (Becker et al. 2018). The availability of whole genome sequence data offers multiple bioinformatic approaches to accurately assign species status (Bull et al. 2012) and excellent comparative resources for sequence data are available at the B. cepacia complex MLST site (https://pubmlst.org/bcc/). Using the deposited whole genome sequence of the octenidine contamination isolates (Becker et al. 2018), an MLST sequence type was derived, ST-881. This showed that the mouth wash and outbreak strain (Becker et al. 2018) was a member of the B. cepacia complex species Burkholderia arboris. Although, this retrospective analysis of recently deposited genome sequence data for a B. cepacia complex contaminant (Becker et al. 2018) was carried out using command-line bioinformatics, the same identification result of B. arboris was also obtained when the genome sequence data was uploaded on the web to the MLST database and a search for matching sequence loci initiated.

#### **ENTEROBACTERIACEAE**

Multiple Enterobacteriaceae including Klebsiella, Enterobacter (European Commission 2018) and Serratia (Polilli et al. 2011) have been encountered as non-food product contaminants, and recent reports show certain species within this highly related group of Gram-negative bacteria have become quite problematic. In particular, Enterobacter gergoviae is a well-known contaminant of food and cosmetic products with high intrinsic preservative resistance (Periame, Pages and Davin-Regli 2014). Recent taxonomic reclassification of several Enterobacter species placed E. gergoviae isolated within a new genus Pluralibacter using multilocus sequence-based methods (Brady et al. 2013). A recall of 15 000 tubes of a best-selling skincare product because of high levels of Pluralibacter (Enterobacter) gergoviae was reported in the media in 2016, bringing public attention to the issue of nonfood product contamination by this species. In addition, P. gergoviae, has been responsible for seven other major European recall incidents involving skin care products in the past ten years, as documented by Safety Gate (European Commission 2018).

The close taxonomic relationships between Enterobacteriaceae species and their propensity to cause infection and product contamination underpin the need for accurate and high resolution identification methods. Over the past two decades, such methods have been developed and applied predominantly in clinical and research settings, but also have application in non-food product contamination. Many genera within Enterobacteriaceae are polyphyletic by 16S rRNA gene sequence analysis alone and therefore expanded genotyping analyses have been required for taxonomic assignments (Brady et al. 2013). As previously mentioned, multilocus sequencebased methods were used to re-classify Enterobacter species as different genera (Brady et al. 2013) and a limited number of studies have applied similar techniques to contamination scenarios. An investigation of a contaminated batch of liquid hand soap identified a single strain of Klebsiella oxytoca was present in the product bottles and clearly illustrates that accurate species identification can be achieved in the non-food product sector (Dieckmann et al. 2016). In this study, a combination of rapid phenotypic methods (MALDI TOF MS and infrared spectroscopy) and genotypic methods (pulsed-field gel electrophoresis, MLST and whole genome sequencing) were used to accurately identify the contaminant to both the species and strain level. Whilst there are multiple media for culture and putative phenotypic identification of *Enterobacteriaceae* (Perry 2017) genotyping and whole genome sequencing have vastly increased accuracy and resolution. Resources such as the extensive genomic databases within Enterobase (https://enterobase.warwick.ac.uk) are available to facilitate detailed characterisation of strains from certain *Enterobacteriaceae* (Alikhan et al. 2018).

### THE ISSUES BEHIND A LACK OF IDENTIFICATION OF NON-FOOD PRODUCT CONTAMINANTS

The large discrepancy in the proportion of unidentified organisms reported in food and non-food product recall information (Figs 1 and 2) may reflect a number of issues with how contaminants are identified and validated for the hygienic integrity of products. The RASFF (European Commission 2019) and Safety Gate (European Commission 2018) databases do not publish microbial identification methods and there is likely to be considerable variability between reporting sites and resource limitations when routine testing only detects certain microorganisms. The perceived risk may also influence the extent of reporting, with more stringent quality control being applied to foods that will be ingested, compared to non-food products having more limited direct contact with consumers. The non-food global ISO 11 930 and European Pharmacopeia challenge testing methods require the absence of E. coli, P. aeruginosa, S. aureus, Candida albicans and Aspergillus brasiliensis (Vincze, Al Dahouk and Dieckmann 2019); but do not require evaluation against opportunistic pathogens Enterobacter, Burkholderia or Klebsiella species. From our meta-analysis (Fig. 1B) it is clear that Enterobacter, Burkholderia and Klebsiella are causing a growing number of non-food product recalls and should be integrated into challenge test methodologies to ensure products are adequately preserved for their intended purpose.

Current regulations require a mandatory absence from a representative sample (1 g or 1 mL) of the following microorganisms: for cosmetics, P. aeruginosa, S. aureus and C. albicans must be absent; and for toys, the latter organisms in addition to E. coli and Salmonella species must be absent (Vincze, Al Dahouk and Dieckmann 2019). Hence a significant number of incidents reported as containing unidentified microorganisms could be attributable to genera such as Enterobacter, Burkholderia and Klebsiella that are not listed as mandatory for reporting. Overall, in relation to the clinical and environmental research areas, industrial microbiology receives limited attention despite the global scale and daily usage of non-food products. It is apparent from the information recorded in the recall database that a range of antimicrobial resistant microorganisms and specific ESKAPE pathogens (Pendleton, Gorman and Gilmore 2013) may be encountered as non-food product contaminants (Fig. 1B). Since these organisms must survive in the presence of antimicrobials to contaminate preserved non-food products, they represent an additional ecological niche for investigation of AMR. Whilst biocide and preservative resistance is mediated by complex, multifactorial mechanisms and is often transient (Maillard 2007), stable adaptive changes resulting in preservative tolerance and cross-resistance to antibiotics have been observed for the opportunistic pathogens such as P. aeruginosa (Abdel Malek and Badran 2010: 588–92) and B. cepacia complex bacteria (Rushton et al. 2013). Isolation and characterisation of strains of industrial origin with increased AMR is not just important to understand resistance mechanisms but can also impact industrial practices directly. In a case study from 2003 (Ferrarese, Paglia and Ghirardini 2003), P. aeruginosa and P. gergoviae strains originating from an industrial plant were found to have increased resistance to cosmetic preservatives, in particular formaldehyde releasing agents. This led to the evaluation of disinfection procedures in the plant and change to a more efficacious decontamination regime.

Correct classification of bacterial infections is essential for accurate local and global microbiological surveillance. It has been deemed vital to improve surveillance to understand the health threat of AMR and ensure antibiotic stewardship programmes are successful (Tacconelli et al. 2018b). Multiple strategies are currently available to accurately identify microorganisms to genus or species level, including phenotypic (biochemical analysis or MALDI TOF MS) (van Belkum et al. 2013) and genotypic methods (16S rRNA gene sequencing, MLST or whole genome sequencing) (Maiden et al. 2013). Whole genome sequencing has revolutionised our understanding of bacteria, facilitating detailed epidemiological understanding of both hospital acquired infection and food-contamination incidents (Maiden et al. 2013). Currently, routine microbial identification in the non-food product industry is based on culture-dependent methods (European Commission 2016; Food and Drug Administration 2017) which have numerous limitations. The expense of molecular diagnostics and DNA sequencing is no longer prohibitive due to the rapid improvement in technology and curtailment in costs. It is therefore the opportune time for wider industrial sectors to take advantage of the accurate microbial identification afforded by DNA sequence-based and gold standard genomic methods (Maiden et al. 2013). All of these techniques, including genome sequencing are now employed regularly in clinical microbiology and food microbiology to identify and track priority pathogens.

### **RECOMMENDATIONS TO FILL THE NON-FOOD PRODUCT MICROBIAL IDENTIFICATION GAP**

If the trend towards reporting microorganisms as unidentified in the Safety Gate database is not corrected, researchers, manufacturers and the public will not have an accurate means to understand the risks posed by non-food product contaminating microorganisms. Methods to accurately identify microorganisms have improved considerably (van Belkum et al. 2013) and multiple commercial companies offer both MALDI TOF MS and gene/genome sequence-based analysis for relatively limited costs. The costs of microbial identification in relation to recall incidents is also small in relation to the overall costs of manufacturers having to deal with removing products, cleaning manufacturing facilities, or develop new formulations to prevent non-food product contamination. Further to the methods to identify P. aeruginosa, B. cepacia complex and Enterobacteriaceae discussed above, Stapyloccocus and fungal species were also among the top five non-food product contaminants (Fig. 1B). Staphylococcus species can be selectively cultured and identified on a range of growth media (Perry 2017). DNA sequence-based identification using the 16S rRNA and rpoB genes is also effective (Mellmann et al. 2006) and MLST databases are available for multiple pathogenic species (https://pubmlst.org/databases). Systematic identification of fungi is less advanced than bacteria but sequence analysis of the nuclear ribosomal DNA internal transcribed (ITS) is an accurate means to validate species groups (Raja et al. 2017).

In terms of improving recall and surveillance information, a primary recommendation would be that non-food product recall reports require microbial identification to be carried out to a minimum of the genus level for both bacteria and fungi. A secondary recommendation would be that for bacterial species which pose a significant health or AMR risk as opportunistic pathogens (e.g. P. aeruginosa, the Enterobacteriaceae, B. cepacia complex and Staphylococcus spp.), identification to the species level should also be reported. In addition, identification should be checked against the current taxonomy of each species, for example by comparison to the List of Prokaryotic names with Standing in Nomenclature (www.bacterio.net) (Parte 2018), to enable fine-grain surveillance and accurate comparative analysis of microbial contamination incidents. Harmonisation of detection methods and minimum identification criteria across EU laboratories is necessary to improve the consistency of reporting, bringing it closer to the level of reporting seen in food products (Vincze, Al Dahouk and Dieckmann 2019).

If manufacturers and regulatory bodies apply whole genome sequencing to the characterization of contaminant and outbreak strains, then they have all the information required for accurate species classification (Bull et al. 2012). Such analysis can also future-proof our understanding by archiving sequence data within nucleotide sequence databases to enable retrospective and comparative analysis of contamination incidents. In addition, since the non-food product and cosmetics industry landscape is changing, methods for surveillance and recall recording need to be updated to facilitate the identification of contamination trends. The rise of new market trends such as natural preservative ingredients in non-food products such as cosmetics (Papageorgiou et al. 2010) may result in a shift in the diversity of microorganisms encountered as contaminants. Improved vigilance of non-food microbial contamination is also key to ensure that testing methodologies remain relevant to the products they govern. For example, the non-food global ISO 11 930 and European Pharmacopeia testing methods do not currently include the prevalent contaminating bacteria Enterobacter, Burkholderia or Klebsiella (Fig. 1), and only recommend using E. coli, P. aeruginosa, S. aureus, C. albicans and A. brasiliensis (Siegert 2013). The inclusion of Burkholderia as mandatory for reporting has already been discussed (Torbeck et al. 2011), and as other important opportunistic pathogens, Enterobacter and Klebsiella should also be considered.

#### **CONCLUSIONS AND FUTURE PERSPECTIVES**

Overall, we have highlighted that there is a significant gap in the accurate identification and reporting of microorganisms associated with recall of contaminated non-food consumer products. We must change the accuracy of recall reporting information to avoid significantly under-reporting the presence of potentially pathogenic and known AMR organisms in non-food products. In this respect, the non-food consumer product industry and the microbiological analyses required of them, lag behind those of the food and clinical microbiology sectors. Going forward, it is paramount that microbial contaminants in non-food products are accurately identified and reported to isolate problematic organisms and sources of contamination, improve quality control and preservation system design, identify and characterise AMR organisms and ultimately protect consumers.

#### **AUTHOR INFORMATION**

RW conceived and initiated the metadata analysis of nonsterile food products. EC-O updated the analysis and conceived the comparison to food product recalls. EM and TP secured funding support. EC-O and EM drafted the initial manuscript, and all authors contributed to further review, analysis and writing.

#### **ACKNOWLEDGEMENTS**

Barry Murphy, Thomas Connor, Denise Donoghue, Peter Jay and Andrew Weightman are acknowledged for general advice.

#### **FUNDING**

This work was supported by Biotechnology and Biological Sciences Research Council (BBSRC) and Unilever R&D industrial CASE studentship awards (RW, BB/F016557/1; EC-O, BB/M009122/1). RW acknowledges current funding from a Wellcome Trust Institutional Strategic Support Fund award held at Cardiff University.

**Competing interests.** TP is an employee of Unilever R&D, the company that part funded the studentships to RW and EC-O. The remaining authors have no conflicts of interest.

#### REFERENCES

- Abdel Malek SM, Badran YR. Pseudomonas aeruginosa PAO1 adapted to 2-phenoxyethanol shows cross-resistance to dissimilar biocides and increased susceptibility to antibiotics. Folia Microbiol (Praha) 2010;55:588–92.
- Alikhan NF, Zhou Z, Sergeant MJ et al. A genomic overview of the population structure of Salmonella. PLos Genet 2018;14:e1007261.
- Baldwin A, Mahenthiralingam E, Thickett KM et al. Multilocus sequence typing scheme that provides both species and strain differentiation for the Burkholderia cepacia complex. J Clin Microbiol 2005;43: 4665–73.
- Becker SL, Berger FK, Feldner SK et al. Outbreak of Burkholderia cepacia complex infections associated with contaminated octenidine mouthwash solution, Germany, August to September 2018. Euro Surveill 2018;23: doi: 10.2807/1560-7917.ES.2018.23.42.1800540.
- Becks VE, Lorenzoni NM. *Pseudomonas aeruginosa* outbreak in a neonatal intensive care unit: a possible link to contaminated hand lotion. *Am J Infect Control* 1995;23:396–8.
- Brady C, Cleenwerck I, Venter S et al. Taxonomic evaluation of the genus Enterobacter based on multilocus sequence analysis (MLSA): proposal to reclassify E. nimipressuralis and E. amnigenus into Lelliottia gen. nov. as Lelliottia nimipressuralis comb. nov. and Lelliottia amnigena comb. nov., respectively, E. gergoviae and E. pyrinus into Pluralibacter gen. nov. as Pluralibacter gergoviae comb. n ov. and Pluralibacter pyrinus comb. nov., respectively, E. cowanii, E. radicincitans, E. oryzae and E. arachidis into Kosakonia gen. nov. as Kosakonia cowanii comb. nov., Kosakonia radicincitans comb. nov., Kosakonia oryzae comb. nov. and Kosakonia arachidis comb. nov., respectively, and E. turicensis, E. helveticus and E. pulveris into Cronobacter as Cronobacter zurichensis nom. nov ., Cronobacter helveticus comb. nov. and Cronobacter pulveris comb. nov., respectively, and emended description of the genera Enterobacter and Cronobacter. Syst Appl Microbiol 2013;36: 309-19.

- Bull MJ, Marchesi JR, Vandamme P et al. Minimum taxonomic criteria for bacterial genome sequence depositions and announcements. J Microbiol Methods 2012;89: 18–21.
- Curran B, Jonas D, Grundmann H et al. Development of a multilocus sequence typing scheme for the opportunistic pathogen *Pseudomonas aeruginosa. J Clin Microbiol* 2004;**42**: 5644–9.
- De Vos D, Lim A, Jr., Pirnay JP et al. Direct detection and identification of *Pseudomonas aeruginosa* in clinical samples such as skin biopsy specimens and expectorations by multiplex PCR based on two outer membrane lipoprotein genes, *oprI* and *oprL. J Clin Microbiol* 1997;**35**: 1295–9.
- Depoorter E, Bull MJ, Peeters C et al. Burkholderia: an update on taxonomy and biotechnological potential as antibiotic producers. Appl Microbiol Biotechnol 2016;**100**: 5215–29.
- Dieckmann R, Hammerl JA, Hahmann H et al. Rapid characterisation of *Klebsiella* oxytoca isolates from contaminated liquid hand soap using mass spectrometry, FTIR and Raman spectroscopy. *Faraday Discuss* 2016;**187**: 353–75.
- European Commission. The Scientific Committee on Consumer Safety Notes of Guidance for the Testing of Cosmetic Ingredients and their Safety Evaluation. 2016: http://ec.europa.eu/health/ scientific\_committees/consumer\_safety/docs/sccs\_o\_190.pdf (8 January 2020, date last accessed).
- European Commission. Safety Gate: The Rapid Alert System for Dangerous Non-food Products. 2018: https://ec.europa.eu/con sumers/consumers\_safety/safety\_products/rapex/alerts/rep ository/content/pages/rapex/index\_en.htm (8 January 2020, date last accessed).
- European Commission. Rapid Alert System for Food and Feed (RASFF). 2019, https://ec.europa.eu/food/safety/rasff\_en (8 January 2020, date last accessed).
- Evans H, Bolt H, Heinsbroek E et al. National outbreak of Pseudomonas aeruginosa associated with an aftercare solution following piercings, July to September 2016, England. Euro Surveill 2018;23: doi: 10.2807/1560-7917.ES.2018.23.37.1700795.
- Ferrarese L, Paglia R, Ghirardini A. Bacterial resistance in cosmetics industrial plant: connected problems and their solution. *Ann Microbiol* 2003;**53**: 477–90.
- Food and Drug Administration. Bacteriological Analytical Manual, 8th Edition, Revision A. 2017: https://www.fda.gov/food/f oodscienceresearch/laboratorymethods/ucm2006949.htm (8 January 2020, date last accessed).
- Henry D, Campbell M, McGimpsey C et al. Comparison of isolation media for recovery of Burkholderia cepacia complex from respiratory secretions of patients with cystic fibrosis. J Clin Microbiol 1999;**37**:1004–7.
- Hogsberg T, Saunte DM, Frimodt-Moller N et al. Microbial status and product labelling of 58 original tattoo inks. *J Eur Acad Dermatol Venereol* 2013;**27**: 73–80.
- Jimenez L. Microbial diversity in pharmaceutical product recalls and environments. PDA J Pharm Sci Technol 2007;61: 383–99.
- Lundov MD, Moesby L, Zachariae C et al. Contamination versus preservation of cosmetics: a review on legislation, usage, infections, and contact allergy. *Contact Dermatitis* 2009;**60**: 70– 8.
- Lundov MD, Zachariae C. Recalls of microbiologically contaminated cosmetics in EU from 2005 to May 2008. Int J Cosmet Sci 2008;**30**: 471–4.
- Mahenthiralingam E, Bischof J, Byrne SK et al. DNA-Based diagnostic approaches for identification of Burkholderia cepacia complex, Burkholderia vietnamiensis, Burkholderia multivorans, Burkholderia stabilis, and Burkholderia cepacia genomovars I and III. J Clin Microbiol 2000;**38**:3165–73.

- Maiden MCJ, Jansen van Rensburg MJ, Bray JE et al. MLST revisited: the gene-by-gene approach to bacterial genomics. Nat *Rev Microbiol* 2013;11:728–36.
- Maillard JY. Bacterial resistance to biocides in the healthcare environment: should it be of genuine concern? J Hosp Infect 2007;65(Suppl 2):60–72.
- Marquez L, Jones KN, Whaley EM et al. An outbreak of Burkholderia cepacia complex infections associated with contaminated liquid docusate. Infect Control Hosp Epidemiol 2017;**38**: 567–73.
- Mayr A, Hinterberger G, Lorenz IH et al. Nosocomial outbreak of extensively drug-resistant *Pseudomonas aeruginosa* associated with aromatherapy. Am J Infect Control 2017;**45**: 453–5.
- Mellmann A, Becker K, von Eiff C et al. Sequencing and staphylococci identification. Emerg Infect Dis 2006;12: 333–6.
- Neza E, Centini M. Microbiologically contaminated and overpreserved cosmetic products according RAPEX 2008–2014. *Cosmetics* 2016;**3**:3.
- Orth DS, Kabara JJ, Denyer SP et al. Cosmetic and Drug Microbiology. New York: Informa Healthcare, 2006.
- Orus P, Leranoz S. Current trends in cosmetic microbiology. International Microbiology: The Official Journal of the Spanish Society for Microbiology 2005;**8**:77–9.
- Papageorgiou S, Varvaresou A, Tsirivas E et al. New alternatives to cosmetics preservation. J Cosmet Sci 2010;61: 107–23.
- Parte AC. LPSN List of Prokaryotic names with Standing in Nomenclature (bacterio.net), 20 years on. Int J Syst Evol Microbiol 2018;68:1825–9.
- Pendleton JN, Gorman SP, Gilmore BF. Clinical relevance of the ESKAPE pathogens. *Expert Rev Anti Infect Ther* 2013;11:297–308.
- Periame M, Pages JM, Davin-Regli A. Enterobacter gergoviae adaptation to preservatives commonly used in cosmetic industry. Int J Cosmetic Sci 2014;**36**:386–95.
- Perry JD. A decade of development of chromogenic culture media for clinical microbiology in an era of molecular diagnostics. *Clin Microbiol Rev* 2017;**30**:449–79.
- Polilli E, Parruti G, Fazii P et al. Rapidly controlled outbreak of Serratia marcescens infection/colonisations in a neonatal intensive care unit, Pescara General Hospital, Pescara, Italy, April 2011. Euro Surveill 2011;16:19892.
- Raja HA, Miller AN, Pearce CJ et al. Fungal identification using molecular tools: a primer for the natural products research community. J Nat Prod 2017;**80**: 756–70.

- Rushton L, Sass A, Baldwin A et al. Key role for efflux in the preservative susceptibility and adaptive resistance of Burkholderia cepacia complex bacteria. Antimicrob Agents Chemother 2013;57:2972–80.
- Siegert W. Comparison of microbial challenge testing methods for cosmetics. Household and Personal Care Today 2013;8:32–9.
- Sommerstein R, Fuhrer U, Lo Priore E et al. Burkholderia stabilis outbreak associated with contaminated commerciallyavailable washing gloves, Switzerland, May 2015 to August 2016. Euro Surveill 2017;22: doi: 10.2807/1560-7917.ES.2017.22.49.17-00213.
- Sutton S, Jimenez L. A review of reported recalls involving microbiological control 2004–2011 with emphasis on FDA considerations of 'objectionable organisms'. Am Pharmaceut Rev 2012;15:42–57.
- Tacconelli E, Carrara E, Savoldi A et al. Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect* Dis 2018a;**18**:318–27.
- Tacconelli E, Sifakis F, Harbarth S et al. Surveillance for control of antimicrobial resistance. Lancet Infect Dis 2018b;18:e99–e106.
- Torbeck L, Raccasi D, Guilfoyle DE et al. Burkholderia cepacia: This decision is overdue. PDA J Pharm Sci Tech PDA 2011;65: 535–43.
- van Belkum A, Durand G, Peyret M et al. Rapid clinical bacteriology and its future impact. Ann Lab Med 2013;33: 14–27.
- Vandamme P, Holmes B, Vancanneyt M et al. Occurrence of multiple genomovars of Burkholderia cepacia in cystic fibrosis patients and proposal of Burkholderia multivorans sp. nov. Int J Syst Bacteriol 1997;47: 1188–200.
- Vincze S, Al Dahouk S, Dieckmann R. Microbiological safety of non-food products: what can we learn from the RAPEX database? Int J Environ Res Public Health 2019;16: doi: 10.3390/ijerph16091599.
- Weiser R, Donoghue D, Weightman A et al. Evaluation of five selective media for the detection of *Pseudomonas aeruginosa* using a strain panel from clinical, environmental and industrial sources. J Microbiol Methods 2014;**99**: 8–14.
- White J, Gilbert J, Hill G et al. Culture-independent analysis of bacterial fuel contamination provides insight into the level of concordance with the standard industry practice of aerobic cultivation. Appl Environ Microbiol 2011;77: 4527–38.
- Wong S, Street D, Delgado SI et al. Recalls of foods and cosmetics due to microbial contamination reported to the U.S. Food and Drug Administration. J Food Protect 2000;**63**:1113–6.