












DATA NOTE

The genome sequence of the Eurasian river otter, *Lutra lutra* Linnaeus 1758 [version 1; peer review: awaiting peer review]

Dan Mead¹, Frank Hailer ², Elisabeth Chadwick², Roberto Portela Miguez ³, Michelle Smith¹, Craig Corton¹, Karen Oliver¹, Jason Skelton¹, Emma Betteridge¹, Jale Doulcan Doulcan ¹, Olga Dudchenko⁴, Arina Omer⁴, David Weisz⁴, Erez Lieberman Aiden⁴, Shane McCarthy ¹, Kerstin Howe ¹, Ying Sims¹, James Torrance¹, Alan Tracey ¹, Richard Challis ¹, Richard Durbin ¹, Mark Blaxter ¹

¹Wellcome Genome Campus, Wellcome Sanger Institute, Hinxton, CB10 1SA, UK

²Cardiff Otter Project, Cardiff University School of Biosciences, Cardiff, CF10 3AX, UK

³Department of Life Sciences, Natural History Museum, London, SW7 5BD, UK

⁴Baylor College of Medicine, Houston, Texas, TX 77030 USA, USA

v1 **First published:** 19 Feb 2020, 5:33 (<https://doi.org/10.12688/wellcomeopenres.15722.1>)

Latest published: 19 Feb 2020, 5:33 (<https://doi.org/10.12688/wellcomeopenres.15722.1>)

Abstract

We present a genome assembly from an individual male *Lutra lutra* (the Eurasian river otter; Vertebrata; Mammalia; Eutheria; Carnivora; Mustelidae). The genome sequence is 2.44 gigabases in span. The majority of the assembly is scaffolded into 20 chromosomal pseudomolecules, with both X and Y sex chromosomes assembled.

Keywords

Lutra lutra river otter genome sequence chromosomal

Open Peer Review

Reviewer Status AWAITING PEER REVIEW

Any reports and responses or comments on the article can be found at the end of the article.

Corresponding author: Mark Blaxter (mark.blaxter@sanger.ac.uk)

Author roles: **Mead D:** Conceptualization, Investigation, Writing – Review & Editing; **Hailer F:** Formal Analysis, Investigation, Resources, Writing – Review & Editing; **Chadwick E:** Writing – Review & Editing; **Portela Miguez R:** Data Curation, Formal Analysis, Investigation, Resources, Writing – Review & Editing; **Smith M:** Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Corton C:** Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Oliver K:** Formal Analysis, Investigation, Methodology, Supervision, Writing – Review & Editing; **Skelton J:** Writing – Review & Editing; **Betteridge E:** Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Doullan JD:** Formal Analysis, Investigation, Methodology, Writing – Review & Editing; **Dudchenko O:** Formal Analysis, Investigation, Methodology, Visualization, Writing – Review & Editing; **Omer A:** Formal Analysis, Investigation, Methodology, Software, Writing – Review & Editing; **Weisz D:** Writing – Review & Editing; **Lieberman Aiden E:** Software, Supervision, Writing – Review & Editing; **McCarthy S:** Data Curation, Formal Analysis, Investigation, Software, Writing – Review & Editing; **Howe K:** Data Curation, Formal Analysis, Investigation, Software, Supervision, Validation, Writing – Review & Editing; **Sims Y:** Data Curation, Formal Analysis, Investigation, Software, Writing – Review & Editing; **Torrance J:** Data Curation, Formal Analysis, Investigation, Software, Supervision, Validation, Writing – Review & Editing; **Tracey A:** Data Curation, Formal Analysis, Investigation, Software, Validation, Writing – Review & Editing; **Challis R:** Data Curation, Formal Analysis, Investigation, Software, Validation, Writing – Review & Editing; **Durbin R:** Conceptualization, Data Curation, Formal Analysis, Investigation, Software, Validation, Visualization, Writing – Review & Editing; **Blaxter M:** Conceptualization, Data Curation, Formal Analysis, Investigation, Software, Validation, Visualization, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This work was supported by the Wellcome Trust through core funding to the Wellcome Sanger Institute (WT206194). SMcC and RD were supported by Wellcome grant WT207492. MB was supported by Wellcome grant WT218328. ELA was supported by an NSF Physics Frontiers Center Award (PHY1427654), the Welch Foundation (Q-1866), a USDA Agriculture and Food Research Initiative Grant (2017-05741), and an NIH Encyclopedia of DNA Elements Mapping Center Award (UM1HG009375).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2020 Mead D *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Mead D, Hailer F, Chadwick E *et al.* **The genome sequence of the Eurasian river otter, *Lutra lutra* Linnaeus 1758 [version 1; peer review: awaiting peer review]** Wellcome Open Research 2020, 5:33 (<https://doi.org/10.12688/wellcomeopenres.15722.1>)

First published: 19 Feb 2020, 5:33 (<https://doi.org/10.12688/wellcomeopenres.15722.1>)

Species taxonomy

Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Laurasiatheria; Carnivora; Caniformia; Mustelidae; Lutrinae; Lutra; *Lutra lutra* Linnaeus 1758 (NCBI txid 9657).

Background

The Eurasian river otter, *Lutra lutra*, is found along the coasts and inland waters of Europe, Asia, China, Japan, Java, Sri Lanka, the Middle East and North Africa. Eurasia. Throughout Europe, populations of *L. lutra* declined precipitously through the latter half of the 20th century, and the species is of active conservation concern. In Ireland, *L. lutra* populations have remained relatively stable¹, and in Britain river restoration and active intervention have resulted in increased populations, and recolonisation of watersheds from which otters had been eliminated². There is active research of the continuing impacts

of pollutants on otters (Pountney *et al.*, 2015), and on the population genetic patterns that have resulted from their near-extinction and subsequent recovery in Britain (Stanton *et al.*, 2014). Here we present a chromosomally assembled genome sequence for *L. lutra*, based on a male specimen from Britain.

Genome sequence report

The genome was sequenced from a naturally deceased single male *L. lutra* collected by the Cardiff Otter Project from Wincanton, Somerset. A total of 63-fold coverage in Pacific Biosciences single-molecule long reads (N50 24 kb) and 58-fold coverage in 10X Genomics read clouds (from molecules with an estimated N50 of 57 kb) were generated. Primary assembly contigs were scaffolded with chromosome conformation HiC data (17-fold coverage). The final assembly has a total length of 2.44 Gb in 43 sequence scaffolds with a scaffold N50 of 149.0 Mb (Table 1). The majority, 92.7%, of the assembly sequence was assigned to 20 chromosomal-level scaffolds representing 18 autosomes (numbered by sequence length), and the X and Y sex chromosomes (Figure 1–Figure 4; Table 2). The assembly has a BUSCO (Simão *et al.*, 2015) completeness

¹ Vincent Wildlife Trust <https://www.vincentwildlife.ie/species/otter>

² National Biodiversity network Atlas <https://species.nbnatlas.org/species/NBNSYS0000005133#overview>

Table 1. Genome data for *Lutra lutra* mLutLut1.

Project accession data	
Assembly identifier	mLutLut1
Species	<i>Lutra lutra</i>
Specimen	NHMKUK ZD 2019.215
NCBI taxonomy ID	9657
BioProject	PRJEB35340
Biosample ID	SAMEA994731
Isolate information	Wild casualty; male
Raw data accessions	
PacificBiosciences SEQUEL I	ERR3313238, ERR3313239-ERR3313241, ERR3313246, ERR3313327, ERR3313330, ERR3313333-ERR3313341
10X Genomics Illumina	ERR3316145-ERR3316148, ERR3316169-ERR3316171
Hi-C Illumina	SRR10119468
Genome assembly	
Assembly accession	GCA_902655055.1
Accession of alternate haplotype	GCA_902653095.1
Span (Mb)	2,438.00
Number of contigs	228
Contig N50 length (Mb)	30.40
Number of scaffolds	43
Scaffold N50 length (Mb)	149.00
Longest scaffold (Mb)	223.45
BUSCO* genome score	C:95.8%[S:94.3%,D:1.5%],F:1.9%,M:2.3%,n:4104

*BUSCO scores based on the mammalia_odb9 BUSCO set using v3.0.2. C= complete [S= single copy, D=duplicated], F=fragmented, M=missing, n=number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/mLutLut1_1/dataset/mLutLut1_1/busco.

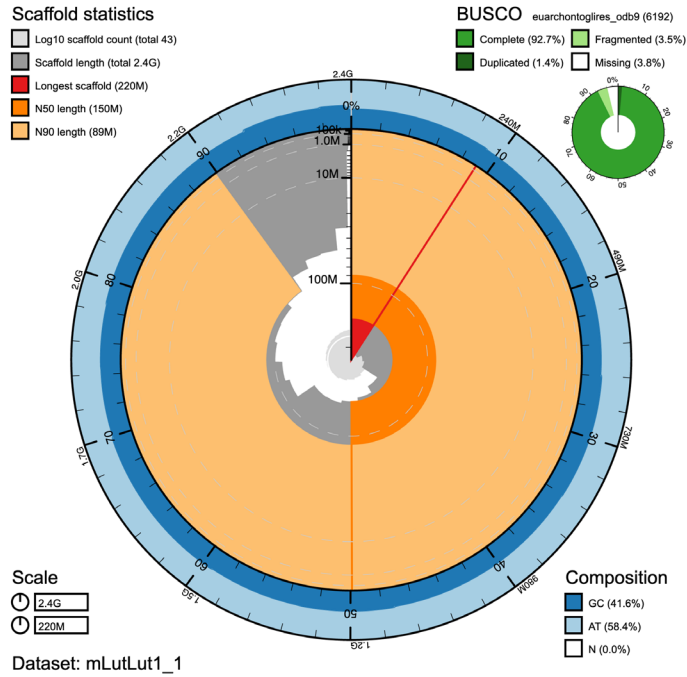


Figure 1. Genome assembly of *Lutra lutra* mLutLut1: BlobToolKit Snailplot. The plot shows N50 metrics for *L. lutra* assembly mLutLut1 and BUSCO scores for the Euarchontoglires set of orthologues. The interactive version of this figure is hosted [here](#).

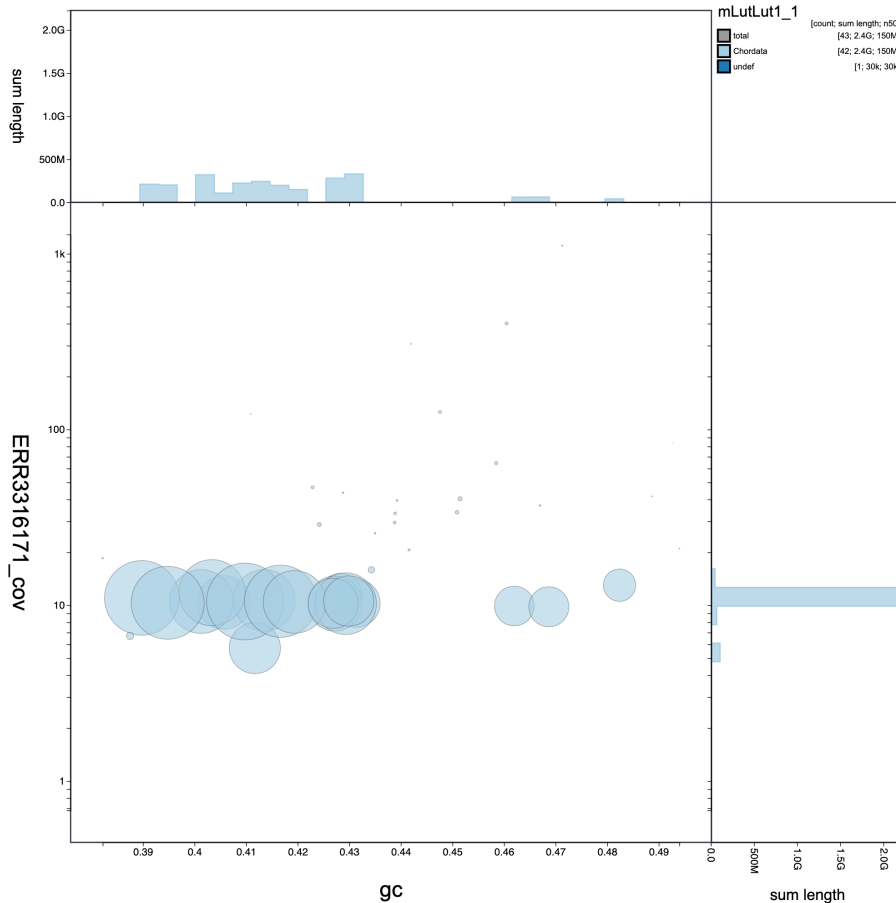


Figure 2. Genome assembly of *Lutra lutra* mLutLut1: BlobToolKit GC-coverage plot. The interactive version of this figure is hosted [here](#).

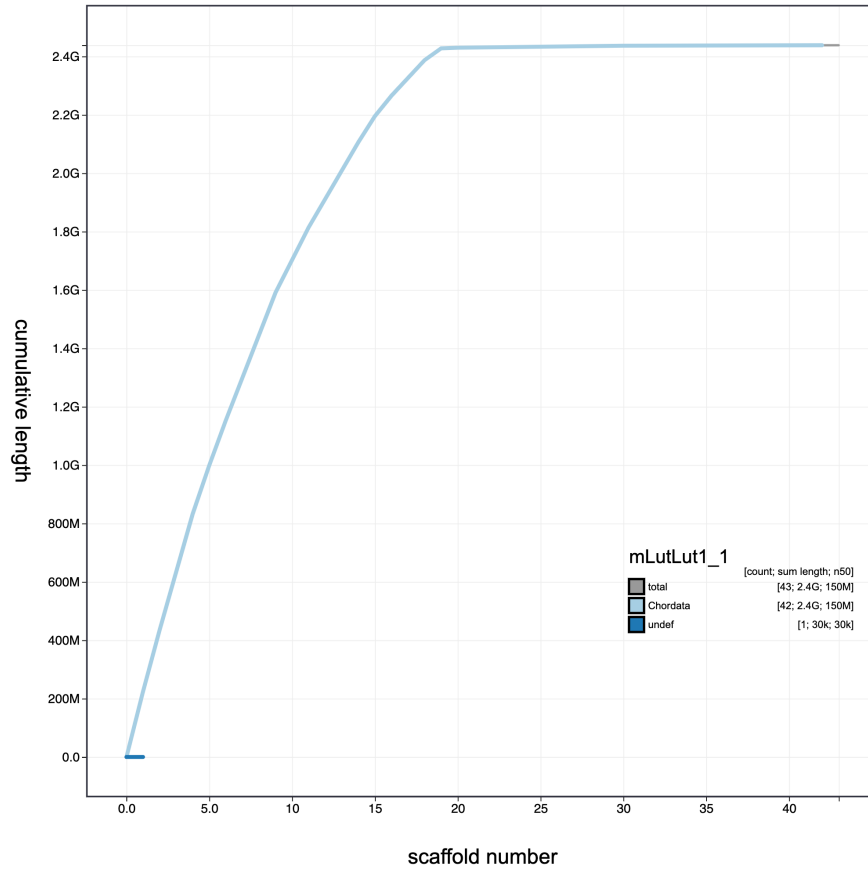


Figure 3. Genome assembly of *Lutra lutra* mLutLut1: BlobToolKit Cumulative sequence plot. The interactive version of this figure is hosted [here](#).

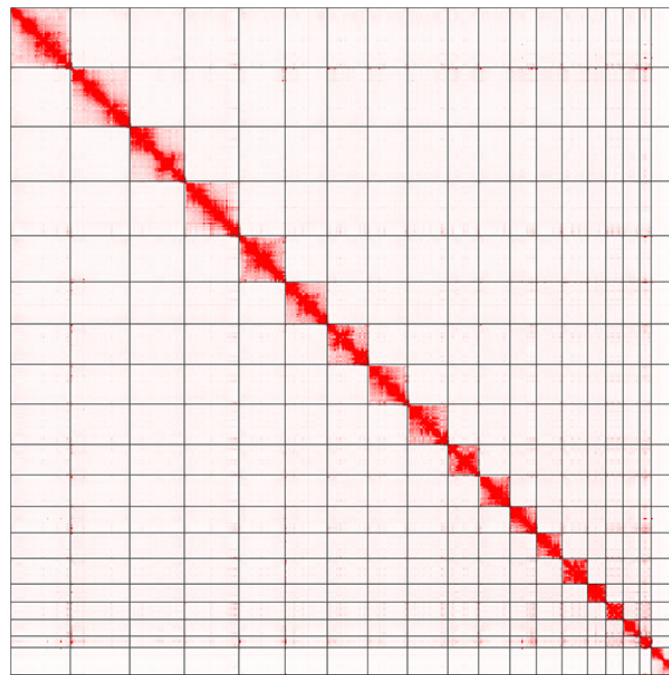


Figure 4. Genome assembly of *Lutra lutra* mLutLut1: Hi-C contact map. Hi-C contact map of the *L. lutra* mLutLut1 assembly, visualized in Juicebox ([Durand et al., 2016](#)). An interactive version of the map hosted [here](#), powered by Juicebox.js ([Robinson et al., 2018](#)).

of 95.8% using the *mammalia_odb9* reference set. While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited.

Table 2. Chromosomal pseudomolecules in the genome assembly of *Lutra lutra* mLutLut1.

ENA accession	Chromosome	Size (Mb)	GC%
LR738403.1	1	223.45	41
LR738404.1	2	210.65	39
LR738405.1	3	201.32	39.5
LR738406.1	4	197.71	41.7
LR738407.1	5	165.81	40.3
LR738408.1	6	154.43	40.1
LR738409.1	7	149.01	41.9
LR738410.1	8	144.75	41.3
LR738411.1	9	144.09	42.9
LR738412.1	10	114.66	42.7
LR738413.1	11	108.79	40.6
LR738414.1	12	96.45	43
LR738415.1	13	95.73	42.7
LR738416.1	14	89.08	43.1
LR738417.1	15	69.99	42.8
LR738418.1	16	61.48	46.9
LR738419.1	17	60.35	46.2
LR738420.1	18	40.43	48.2
LR738421.1	X	99.69	41.2
LR738422.1	Y	2.25	38.8

Methods

The river otter specimen was collected from Wincanton, Somerset by the Cardiff Otter Project. A full tissue dissection and preservation in 80% ethanol was undertaken and the specimen accessioned by the Natural History Museum, London.

DNA was extracted using an agarose plug extraction from spleen tissue following the Bionano Prep Animal Tissue DNA Isolation Soft Tissue Protocol. Pacific Biosciences CLR long read and 10X Genomics read cloud sequencing libraries were constructed according to the manufacturers' instructions. Sequencing was performed by the Scientific Operations core at the Wellcome Sanger Institute on Pacific Biosciences SEQUEL I and Illumina HiSeq X instruments. Hi-C data were generated by the Aiden lab using an optimised version of their protocols (Dudchenko *et al.*, 2017).

Assembly was carried out using Falcon-unzip (Chin *et al.*, 2016), haplotypic duplication was identified and removed with purge_dups (Guan *et al.*, 2020) and a first round of scaffolding carried out with 10X Genomics read clouds using scaff10x (<https://github.com/wtsi-hpag/Scaff10X>). Scaffolding with Hi-C data (Rao *et al.*, 2014) was carried out with 3D-DNA (Dudchenko *et al.*, 2017), followed by manual curation with Juicebox Assembly Tools (Dudchenko *et al.*, 2018; Durand *et al.*, 2016; Robinson *et al.*, 2018) and visualisation in HiGlass (Kerpedjiev *et al.*, 2018). The Hi-C scaffolded assembly was polished with arrow using the PacBio data, then polished with the 10X Genomics Illumina data by aligning to the assembly with longranger align, calling variants with freebayes (Garrison & Marth, 2012) and applying homozygous non-reference edits using bcftools consensus (<https://github.com/VGP/vgp-assembly/tree/master/pipeline/freebayes-polish>). Two rounds of the Illumina polishing were applied. The assembly was checked for contamination and corrected using the gEVAL system (Chow *et al.*, 2016). We removed two

Table 3. Software tools used.

Software tool	Version	Source
Falcon-unzip	falcon-kit 1.2.2	(Chin <i>et al.</i> , 2016)
purge_dups	1.0.0	(Guan <i>et al.</i> , 2020)
3D-DNA	180419	(Dudchenko <i>et al.</i> , 2018)
scaff10x	4.2	https://github.com/wtsi-hpag/Scaff10X
arrow	GenomicConsensus 2.3.3	https://github.com/PacificBiosciences/GenomicConsensus
longranger align	2.2.2	https://support.10xgenomics.com/genome-exome/software/pipelines/latest/advanced/other-pipelines
freebayes	v1.1.0-3-g961e5f3	(Garrison & Marth, 2012)
bcftools consensus	1.9	http://samtools.github.io/bcftools/bcftools.html
gEVAL	2016	(Chow <i>et al.</i> , 2016)
BlobToolKit	1	(Challis <i>et al.</i> , 2019)

low-coverage scaffolds that were likely to have derived from the ribosomal DNA cistron of a *Sarcocystis* species (most similar to *Sarcocystis lutrae*). The genome was analysed within the BlobToolKit environment (Challis *et al.*, 2019).

Data availability

European Nucleotide Archive: Lutra lutra (Eurasian otter) genome assembly, mLutLut1. BioProject accession number PRJEB35340; <https://www.ebi.ac.uk/ena/data/view/PRJEB35340>.

The genome sequence is released openly for reuse. The *L. lutra* genome sequencing initiative is part of the Wellcome Sanger Institute's "25 genomes for 25 years" project³. It is also part of the Vertebrate Genome Project (VGP)⁴ ordinal references

³ <https://www.sanger.ac.uk/science/collaboration/25-genomes-25-years>

⁴ <https://vertebrategenomesproject.org/>

programme, the DNA Zoo Project⁵ and the Darwin Tree of Life (DToL) project⁶. The specimen has been preserved in ethanol and deposited with the Natural History Museum, London under registration number NHMUK ZD 2019.215 where it will remain accessible to the research community for posterity. All raw data and the assembly have been deposited in the ENA. The genome will be annotated and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1.

Acknowledgements

We thank Mike Stratton and Julia Wilson for their continuing support for the 25 genomes for 25 years project.

⁵ <https://www.dnazoo.org/>

⁶ <https://www.darwintreeoflife.org/>

References

- Challis R, Richards E, Rajan J, *et al.*: **BlobToolKit – Interactive Quality Assessment of Genome Assemblies.** *bioRxiv.* 2019.
[Publisher Full Text](#)
- Chin CS, Peluso P, Sedlazeck FJ, *et al.*: **Phased diploid genome assembly with single-molecule real-time sequencing.** *Nat Methods.* 2016; **13**(12): 1050–54.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Chow W, Brugger K, Caccamo M, *et al.*: **gEVAL - a web-based browser for evaluating genome assemblies.** *Bioinformatics.* 2016; **32**(16): 2508–10.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Dudchenko O, Batra SS, Omer AD, *et al.*: **De novo assembly of the *Aedes aegypti* genome using Hi-C yields chromosome-length scaffolds.** *Science.* 2017; **356**(6333): 92–95.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Dudchenko O, Shamim MS, Batra SS, *et al.*: **The Juicebox Assembly Tools Module Facilitates de Novo Assembly of Mammalian Genomes with Chromosome-Length Scaffolds for under \$1000.** *bioRxiv.* 2018.
[Publisher Full Text](#)
- Durand NC, Robinson JT, Shamim MS, *et al.*: **Juicebox Provides a Visualization System for Hi-C Contact Maps with Unlimited Zoom.** *Cell Syst.* 2016; **3**(1): 99–101.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Garrison E, Marth G: **Haplotype-Based Variant Detection from Short-Read Sequencing.** *arXiv [q-bio.GN].* arXiv. 2012.
[Reference Source](#)
- Guan D, McCarthy SA, Wood J, *et al.*: **Identifying and removing haplotypic duplication in primary genome assemblies.** *Bioinformatics.* 2020; pii: btaa025.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Kerpedjiev P, Abdennur N, Lekschas F, *et al.*: **HiGlass: web-based visual exploration and analysis of genome interaction maps.** *Genome Biol.* 2018; **19**(1): 125.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Pountney A, Filby AL, Thomas GO, *et al.*: **High liver content of polybrominated diphenyl ether (PBDE) in otters (*Lutra lutra*) from England and Wales.** *Chemosphere.* 2015; **118**: 81–86.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Rao SS, Huntley MH, Durand NC, *et al.*: **A 3D map of the human genome at kilobase resolution reveals principles of chromatin looping.** *Cell.* 2014; **159**(7): 1665–80.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Robinson JT, Turner D, Durand NC, *et al.*: **Juicebox.js Provides a Cloud-Based Visualization System for Hi-C Data.** *Cell Syst.* 2018; **6**(2): 256–58.e1.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
- Simão FA, Waterhouse RM, Ioannidis P, *et al.*: **BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs.** *Bioinformatics.* 2015; **31**(19): 3210–12.
[PubMed Abstract](#) | [Publisher Full Text](#)
- Stanton DWG, Hobbs GI, McCafferty DJ, *et al.*: **Contrasting Genetic Structure of the Eurasian Otter (*Lutra lutra*) across a Latitudinal Divide.** *J Mammal.* 2014; **95**(4): 814–823.
[Publisher Full Text](#)