



A molecular basis for the T cell response in HLA-DQ2.2 mediated celiac disease

Yi Tian Ting^{a,b,1}, Shiva Dahal-Koirala^{c,d,1}, Hui Shi Keshia Kim^{a,b}, Shuo-Wang Qiao^{c,d}, Ralf S. Neumann^{c,d}, Knut E. A. Lundin^{c,d,e}, Jan Petersen^{a,b,f}, Hugh H. Reid^{a,b,f}, Ludvig M. Sollid^{c,d,2,3}, and Jamie Rossjohn^{a,b,f,g,2,3}

^aInfection and Immunity Program, Biomedicine Discovery Institute, Monash University, Clayton, VIC 3800, Australia; ^bThe Department of Biochemistry and Molecular Biology, Monash University, Clayton, VIC 3800, Australia; ^cDepartment of Immunology, University of Oslo and Oslo University Hospital-Rikshospitalet, 0372 Oslo, Norway; ^dK. G. Jebsen Centre for Coeliac Disease Research, University of Oslo, 0372 Oslo, Norway; ^eDepartment of Gastroenterology, Oslo University Hospital-Rikshospitalet, 0372 Oslo, Norway; ^fAustralian Research Council Centre of Excellence in Advanced Molecular Imaging, Monash University, Clayton, VIC 3800, Australia; and ^gInstitute of Infection and Immunity, Cardiff University School of Medicine, Heath Park, CF14 4XN Cardiff, United Kingdom

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The highly homologous human leukocyte antigen (HLA)-DQ2 molecules, HLA-DQ2.5 and HLA-DQ2.2, are implicated in the pathogenesis of celiac disease (CeD) by presenting gluten peptides to CD4⁺ T cells. However, while HLA-DQ2.5 is strongly associated with disease, HLA-DQ2.2 is not, and the molecular basis underpinning this differential disease association is unresolved. We here provide structural evidence for how the single polymorphic residue (HLA-DQ2.5-Tyr22 α and HLA-DQ2.2-Phe22 α) accounts for HLA-DQ2.2 additionally requiring gluten epitopes possessing a serine at the P3 position of the peptide. In marked contrast to the biased T cell receptor (TCR) usage associated with HLA-DQ2.5-mediated CeD, we demonstrate with extensive single-cell sequencing that a diverse TCR repertoire enables recognition of the immunodominant HLA-DQ2.2-glut-L1 epitope. The crystal structure of two CeD patient-derived TCR in complex with HLA-DQ2.2 and DQ2.2-glut-L1 (PFSEQEQPV) revealed a docking strategy, and associated interatomic contacts, which was notably distinct from the structures of the TCR:HLA-DQ2.5:gliadin epitope complexes. Accordingly, while the molecular surfaces of the antigen-binding clefts of HLA-DQ2.5 and HLA-DQ2.2 are very similar, differences in the nature of the peptides presented translates to differences in responding T cell repertoires and the nature of engagement of the respective antigen-presenting molecules, which ultimately is associated with differing disease penetrance.

celiac disease | human leukocyte antigen | X-ray crystallography | T cell receptor

Celiac disease (CeD) is a heritable and chronic inflammatory disorder that is caused by a maladaptive immune response to cereal gluten proteins (1). While exhibiting features of food intolerance, the disease also has many autoimmune characteristics including highly disease-specific autoantibodies targeting the enzyme transglutaminase 2 (TG2). CeD patients, but not healthy subjects, have gluten-specific CD4⁺ T cells (2). These T cells, persisting for decades in patients (3), are considered drivers of the CeD pathophysiology that includes the formation of an inflammatory lesion in the proximal small intestine and the generation of autoantibodies (4). The gluten-specific T cells are uniquely restricted by disease-associated HLA-DQ allotypes, namely HLA-DQ2.5 (*HLA-DQA1*05/HLA-DQB1*02:01*), HLA-DQ2.2 (*HLA-DQA1*02:01/HLA-DQB1*02:02*), and HLA-DQ8 (*HLA-DQA1*03:01/HLA-DQB1*03:02*) (5–7). About 90% of patients affected by CeD are HLA-DQ2.5 positive, and those who do not carry HLA-DQ2.5 are equally often either HLA-DQ2.2 or HLA-DQ8. Interestingly, while the risk of the three disease-associated HLA-DQ allotypes are very different, the clinical presentations of patients with the different HLA-DQ allotypes are the same (7). This suggests that the differential risks relate to likelihood of generating an anti-gluten T cell response, and once the gluten-specific T cells are established and clonally expanded, exposure to gluten leads to the same pathophysiological reactions.

The gluten peptides recognized by T cells of CeD patients are posttranslationally modified. TG2 is essential in this process as the enzyme can convert glutamine residues in polypeptides to glutamate, mostly following a QXP recognition motif (8, 9). Gluten proteins are exceptionally rich in such sequence motifs but naturally sparse in negatively charged residues that is required for a high-affinity binding to peptide-binding pockets within the antigen-binding cleft of the disease-associated HLA-DQ allotypes (7). The TG2-mediated deamidation of gluten peptides leads to an increase in binding affinity and the formation of kinetically stable peptide–HLA-DQ complexes (10). Hence, this post-translational modification is essential for the immunogenicity of these antigens (11–13), and the immune reactions inflicting damage that ensue T cell recognition of HLA-DQ gluten peptide complexes is likely the mechanism behind the absolute association of certain HLA-DQ allomorphs with CeD (7). Generally, T cell clones restricted by the three CeD-associated HLA allotypes recognize distinct and separate sets of gluten epitopes (4, 14, 15). This selectivity is based on the capacity of peptides to form kinetically stable peptide–HLA-DQ complexes as only peptides that

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Significance

Celiac disease (CeD) is an autoimmune-like disorder that is driven by the ingestion of dietary gluten. There is a strong human leukocyte antigen (HLA) association with the disease, with the majority of CeD patients being HLA-DQ2.5 positive. While HLA-DQ2.2 is closely related to HLA-DQ2.5, it is less of a risk factor in CeD. Our understanding behind this differential disease association remained unclear. We give the molecular basis for the differential risks of the two HLA-DQ variants, and show that small differences in the peptide–HLA landscape cause profound differences in the responding T cell repertoires.

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The authors declare no competing interest.

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Data deposition: The atomic coordinates and structure factors have been deposited in the Protein Data Bank, <http://www.rcsb.org/> (PDB ID codes 6PX6 and 6PY2). Single-cell TCR sequencing raw data generated in this study are available in the European Genome-phenome Archive ([EGAS00001003673](https://www.ebi.ac.uk/ena/browser/view/EGAS00001003673)).

¹Y.T.T. and S.D.-K. contributed equally to this work.

²L.M.S. and J.R. contributed equally to this work.

³To whom correspondence may be addressed. Email: l.m.sollid@medisin.uio.no or jamie.rossjohn@monash.edu.

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form stable peptide complexes can elicit T cell clonal expansions *in vivo* (14, 16).

While HLA-DQ8 binds and presents gluten peptides with glutamate at anchor positions P1 and/or P9 (15, 17, 18), HLA-DQ2.5 and HLA-DQ2.2 binds and presents gluten peptides with glutamate residues at anchor positions P4, P6, or P7 (15, 19–23). Three known HLA-DQ2.2–restricted epitopes, namely DQ2.2–glut-L1 (PFSEQEQPV), DQ2.2–glia- α 1 (QGSVQPOQL), and DQ2.2–glia- α 2 (QYSQPEQPI), have sequences similar to HLA-DQ2.5–binding peptides, with the exception that they all carry serine at P3 (14, 24, 25). As seen for HLA-DQ2.5–restricted epitopes, the HLA-DQ2.2–restricted epitopes display a hierarchy with DQ2.2–glut-L1 being the epitope recognized by most T cells (24). In binding experiments with DQ2.2–glut-L1 variant peptides substituted at P3, HLA-DQ2.2 but not HLA-DQ2.5 showed preferential binding of the serine and threonine variants (14). An earlier study had indicated that HLA-DQ2.2, in contrast to HLA-DQ2.5, utilizes a P3 anchor (20), and a P3 anchor selectivity of HLA-DQ2.2 for serine or threonine was demonstrated when comparing the repertoires of endogenous peptide ligands eluted from purified HLA-DQ2.5, HLA-DQ2.2, and HLA-DQ7.5 molecules (25).

The HLA-DQ2 molecules, HLA-DQ2.5 and HLA-DQ2.2, are highly homologous. Previous studies have ascribed the differences in conferred risk associated with CeD as a result of the polymorphism in the DQ α chain between HLA-DQ2.5 and HLA-DQ2.2 (14, 16). In particular, the Tyr22 α to Phe22 α polymorphism is located near the P3 peptide binding pockets of the HLA-DQ molecule. Here, we examined the role of Phe22 α variant in HLA-DQ2.2 that influence peptide binding preferences and how the T cell receptors (TCRs) derived from DQ2.2 CeD patients engage the HLA-DQ2.2:DQ2.2–glut-L1 complex. We provide a molecular basis for these key interactions in HLA-DQ2.2–mediated CeD and demonstrate how this is different from HLA-DQ2.5– and HLA-DQ8–mediated epitope presentation and TCR recognition in CeD.

Results

DQ2.2–glut-L1–Specific TCR Repertoire. We performed high-throughput DNA sequencing of rearranged *TRAV* and *TRBV* genes of the single HLA-DQ2.2:DQ2.2–glut-L1 tetramer binding CD4⁺ T cells isolated from six T cell lines (TCLs) of four CeD patients described in a previous study (14), specifically CD555 (TCL555.A.1.4 and TCL555.A.2.2), CD594 (TCL594.5.2 and TCL594.8.1), CD627 (TCL627.7.3), and CD1005 (TCL1005.1). Following processing of sequencing data, we obtained productive paired TCR β sequences of 549 cells that were categorized into 18 unique clonotypes (Table 1). Cells expressing paired identical nucleotide *TRAV/TRBV* genes are defined as a clonotype. We also obtained five unique clonotypes by sequencing *TRAV/TRBV* gene sequences of the T cell clones generated from the same four patients. Of those five unique clonotypes, cells expressing the same TCR as that of three unique T cell clones were observed as clonally expanded population in the single-cell sequencing data generated by tetramer-based sorting. In total, we generated 20 unique clonotypes of DQ2.2–glut-L1–specific T cells from tetramer-sorted single cells and DQ2.2–glut-L1–specific T cell clones.

We observed that few clonotypes dominate the DQ2.2–glut-L1–specific TCR repertoire in the TCLs derived from HLA-DQ2.2–positive CeD patients. In the majority of the patients, one or two expanded clonotypes dominated the T cell repertoire. However, we cannot exclude that some of this bias is the effect of *in vitro* culture of the TCLs. As the number of unique DQ2.2–glut-L1–specific TCRs is small and the dataset is obtained from TCLs generated by *in vitro* culture, caution should be exercised on generalizing features of the DQ2.2–glut-L1–specific TCR repertoire. In our fairly limited dataset, we observed that while some V genes like *TRAV21*,

TRAV26-1, *TRBV20-1*, and *TRBV7-2/3* genes were expressed in more than one CeD patient, there was no apparent V gene bias, preferential *TRAV:TRBV* pairing or conserved CDR3 features in DQ2.2–glut-L1–specific TCR repertoire. In two patients (CD1005, CD555), we observed T cells expressing TCRs with identical CDR3 α and CDR3 β amino acid sequences. The CDR3 of these public TCRs are encoded by different nucleotides, representing a phenomenon termed convergent recombination (26).

Among these HLA-DQ2.2:DQ2.2–glut-L1–specific T cell clones, four TCRs were expressed and successfully refolded: TCR 555 (*TRAV1-1*01/TRBV6-2*01*), TCR 594 (*TRAV9-2*01/TRBV11-2*01*), TCR 627 (*TRAV3*01/TRBV28*01*), and TCR 1005.2.56 (*TRAV21*02/TRBV7-3*01*) (Table 1 and *SI Appendix, Table S1*). These four HLA-DQ2.2–glut-L1 reactive TCRs do not contain a conserved arginine residue in the CDR loops that underpin the response to gluten antigens in HLA-DQ2.5:DQ2.5–glia- α 2, HLA-DQ8:DQ8–glia- α 1, and HLA-DQ8:DQ8.5–glia- γ 1 reactive TCRs and displayed more diverse *TRAV/TRBV* gene usage (22).

Structural Overview of Two TCR:HLA-DQ2.2:DQ2.2–glut-L1 Complexes.

To establish how TCRs interact with HLA-DQ2.2:DQ2.2–glut-L1, we determined the structure of the TCR 594:HLA-DQ2.2:DQ2.2–glut-L1 and TCR 1005.2.56:HLA-DQ2.2:DQ2.2–glut-L1 ternary complexes at 2.8 Å and 3.0 Å resolution, respectively (Fig. 1 and *SI Appendix, Table S2*). The TCR 594 docked centrally above HLA-DQ2.2:DQ2.2–glut-L1, $\sim 85^\circ$ across the antigen-binding cleft, where the V α and V β chains of the TCR lay above the β -helix and α -helix of HLA-DQ2.2, respectively (Fig. 1A and C). The total buried surface area (BSA) upon ligation of TCR 594 with HLA-DQ2.2:DQ2.2–glut-L1 was $\sim 1,900$ Å². The V α and V β domains of the TCR 594 contributed 45.4% and 54.6%, respectively, to the BSA at the interface (Fig. 1C). Notably 36.5% BSA arose from the CDR3 β loop, which played a dominant role in contacting the epitope and HLA-DQ2.2 (Fig. 1C). There was a relatively even distribution of contacts among the CDR loops toward the HLA-DQ2.2 molecule, with the CDR1 α , CDR2 α , CDR3 α , and CDR2 β loops, contributing 16.4%, 13.6%, 11.2%, and 14.1% BSA to the interface, respectively.

The TCR 594 α -chain interacted with the HLA-DQ2.2 β -chain, spanning from Asp66 β to Arg80 β (Fig. 2A and *SI Appendix, Table S3*). Here, Tyr30 α from the CDR1 α loop made van der Waals (vdw) contacts with Asp76 β and Arg80 β of HLA-DQ2.2, while Thr57 α and Asn58 α from the CDR2 α loop made vdw contacts with Glu69 β , Arg72 β , and Ala73 β (Fig. 2A). The neighboring V α framework residue Lys55 α further stabilizes the CDR2 α –HLA-DQ2.2 interactions by forming a salt bridge with Asp66 β and Glu69 β . Regarding the TCR β -chain, the CDR2 β loop made the majority of the contacts with the HLA-DQ2.2 α -chain (Fig. 2B). Namely, multiple vdw contacts between Gln57 β , Asn58 β , and Gly64 β from the TCR and the HLA-DQ2.2 residues Lys39 α , Gln57 α , and Thr61 α were observed. Moreover, additional vdw contacts were made between the V β framework residue Val66 β and the HLA-DQ2.2 residue Asp55 α (Fig. 2B). Accordingly, the TCR 594 engaged the HLA-DQ2.2 molecule in a standard docking mode, with varied contributions from each of the germ-line–encoded CDR loops.

The TCR 1005.2.56 also adopted a canonical docking position at $\sim 60^\circ$ across the antigen-binding cleft (Fig. 1B and D). The total BSA upon ligation of TCR 1005.2.56 with HLA-DQ2.2:DQ2.2–glut-L1 was slightly higher than TCR 594 with $\sim 2,030$ Å², where the contribution of the V α and V β domains of the TCR 1005.2.56 to the BSA at the interface were 43.8% and 56.2%, respectively. The TCR 1005.2.56 CDR loops made relatively even contact across the HLA-DQ2.2:DQ2.2–glut-L1 (Figs. 1D and 3). However, the TCR 1005.2.56 made contact with the DQ2.2–glut-L1 epitope with both germ-line–encoded CDR1 α and nongerm-line–encoded CDR3 β loop (Figs. 1D and

Table 1. DQ2.2-specific TCR sequences

No.	Donor	TRAV	TRAJ	CDR3a	TRBV	TRBD	TRBJ	CDR3b	No. of cells	TCC
1	1005	12-3	34	CAMTLNTDKLIF	4-2	1	1-2	CASSLQGGDYGYTF	1	
2	1005	38-1*01	32*02	CAFMKEDGGATNKLIF	20-1*01	1*02	2-5*01	CSASPNGTGVQAQYF	5	1005.2.54 and 1005.2.60
3	1005	26-1	48	CIVYLSNFGNEKLTF	5-5	2	2-2	CASSLIGGGELFF	1	—
4	1005	29/DV5	42	CAASAHYGGSQGNLIF	12-5	1	1-1	CASGPGTATEAFF	1	—
5	1005	38-2/DV8	58	CAYRSQETSGSRLTF	20-1	1	2-5	CSASWLGFGETQYF	9	—
6	1005	38-2/DV8	58	CAYTPLETSGSRLTF	20-1	1	2-7	CSARAPITGRFYEQYF	4	—
7	1005	21	42	CAVRPGGSQGNLIF	4-2	2	2-5	CASSLTSEETQYF	74	—
		29/DV5	30	CAASADLTNRDDKIIF						
8	1005	38-1	42	CAFMKGSYGGSQGNLIF	7-8	1	2-7	CASSRTANYEQYF	28	—
		40	40	FLLGITSPTYKYIF						
9	1005	26-1	54	CIVRVAEGAQKLVF	20-1		2-5	CSATKETQYF	1	—
10	1005	26-1	54	CIVRVAEGAQKLVF	20-1	1	2-5	CSATKETQYF	3	—
11	1005	21*02	31*01	CAVHTGARLMF	7-3*01	1*01	2-3*01	CASSHGASTDTQYF	NA	1005.2.56, 1005.2.57, 1005.2.58, 1005.2.62, and 1005.2.65
12	555	26-1	54	CIVRVAEGAQKLVF	20-1	—	2-5	CSATKETQYF	3	—
13	555	6	26	CALVRDYGQNFVF	7-2	2	2-7	CASSTSGGGARSYEYF	1	—
14	555	1-1*01	9*01	CAWTGGGGFKTIF	6-2*01	2*01	2-7*01	CASRKPGGDYEYF	80	555A.1.4.38
15	555	21	58	CAVIGTSGSRLTF	7-3	1	2-3	CASSSGAATDTQYF	166	—
16	594	26-1	54	CIVRPAQGAQKLVF	29-1	2	1-2	CSVSSGGEGNGYTF	1	—
17	594	9-2*01	57*01	CASPQGGSEKLVF	11-2*01	1*01	1-1*01	CASSSGWGGTEAFF	62	594.5.2.6 [†]
		27	21	CAGRDDFNKFYF						
18	627	6	39	CALGLNNAAGNMLTF	10-3	1	2-5	CAISPRQAETQYF	1	—
19	627	21	11	CAASGYSTLTF	11-1	1	2-3	CASTHGAGSDTQYF	108	—
20	627	3*01	8*01	CAVSGNTGFQKLVF	28*01	2*02	1-5*01	CASTSRGGNSNQPHF	NA	627.1.3.199
		DV1*01	12*01	CALGATVMDSSYKLIIF						

The table shows the total overview of unique TCRαβ paired sequences generated from high-throughput sequencing of HLA-DQ2.2:glut-L1 tetramer-sorted single T cells and/or by Sanger sequencing of DQ2.2-glut-L1-specific TCCs. Due to uncertainty in the allele assignment in the TCRαβ sequences generated from single-cell TCR sequencing approach, the alleles for the V, D, and J genes are not shown for the TCRαβ sequences observed only in the single-cell TCR sequencing data. Three TCRαβ sequences generated from single-cell TCR sequencing (nos. 2, 14, and 17) were also found to be expressed by the DQ2.2-glut-L1-specific TCCs, which are mentioned in rightmost column. Two of the TCRαβ sequences (nos. 11 and 20) were observed only by Sanger sequencing of DQ2.2-glut-L1-specific TCCs (last column) and not observed in TCR sequencing data of HLA-DQ2.2:glut-L1 tetramer sorted cells.

[†]By performing Sanger sequencing, we observed that TCC594.5.2.6 expresses single TCRα chain (TRAV9-2*01/TRAJ57*01). However, by performing single-cell TCR sequencing, we were able to identify T cells that expressed an additional TCRα chain (TRAV27/TRAJ21).

3D), rather than just the CDR3 residues as seen in the TCR594 ternary complex.

Compared to the TCR 594, the CDR1α loop of the TCR 1005.2.56 shifted closer to the peptide antigen (*SI Appendix, Fig. S24*), where Tyr37α made contacts with Phe58α and Arg77β from HLA-DQ2.2-glut-L1, as well as forming a hydrogen bond with the main chain of the peptide P3-Ser and vdW interactions with the P2-Phe (Fig. 3A and D and *SI Appendix, Tables S4 and S5*). This further allows the CDR3α loop to make vdW contact with the Asp55α and Phe58α from HLA-DQ2.2 (Fig. 3A). The CDR2α loop of the TCR 1005.2.56 made similar contacts with the Glu69β, Arg70β, and Ala73β from HLA-DQ2.2 through vdW interactions (Fig. 3A), comparable to the TCR 594 interactions (Fig. 2A). All three CDRβ loops from the TCR 1005.2.56 also interacted with the HLA-DQ2.2 molecule (Fig. 3B and C). Similar vdW contacts between CDR1β and CDR2β with a stretch of the helix of the HLA-DQ2.2 α-chain were observed (Fig. 3B). The Vβ framework residue Tyr55β, Ala66β, and Asp67β further supported the interactions between CDR1β and CDR2β and the HLA-DQ2.2 by making hydrogen bond with HLA-DQ2.2 residue Gln57α and additional vdW contacts with Thr61α, Ala64α, and His68α (Fig. 3B). The TCR 1005.2.56 CDR3β also made extensive hydrogen bonds with the HLA-DQ2.2 β-chain (Fig. 3C). Overall, all CDR loops from both Vα and Vβ domains of the TCR 1005.2.56 made a fairly even number of contacts

with both the HLA-DQ2.2 as well as the DQ2.2-glut-L1 epitope (Fig. 3E).

DQ2.2-glut-L1 Peptide-Mediated TCR Interactions. The DQ2.2-glut-L1 epitope residues that were exposed for TCR contact were P2-Phe, P5-Gln, and P7-Gln (Fig. 2C and *SI Appendix, Table S5*). The nongerm-line-encoded CDR3 loops of the TCR 594 contacted all three positions, contributing to 22% BSA of the peptide HLA interface (*SI Appendix, Table S5*). Here, the CDR3α loop made limited contact, interacting only with the P2-Phe in the DQ2.2-glut-L1 peptide (Fig. 2C). However, there was an intricate network of polar interactions between the Glu112α residue of CDR3α loop, Arg70β of the HLA-DQ2.2, and the P5-Gln in the DQ2.2-glut-L1 peptide (Fig. 2D). This P5-Gln of the DQ2.2-glut-L1 peptide also formed a hydrogen bond with Gly109β of the CDR3β loop, and all together, appeared as a focal point for the CDR3 α/β-loops of the TCR 594 on to the HLA-DQ2.2:glut-L1 (Fig. 2D).

The TCR 594 interacted with P7-Gln mostly via its CDR3β loop (Fig. 2C and *SI Appendix, Table S5*). The small glycine residues in the CDR3β loop, namely Gly109β, Gly110β, and Gly112β which flanked the Trp111β, permitted close contact with the peptide and formed hydrogen bond with the side chain of P5-Gln and P7-Gln (Fig. 2C and D). The CDR3β loop of the TCR 594 also displayed extensive interactions with the surrounding residues, where Trp111β formed a hydrogen bond with the side

chain of Asn62 on the HLA-DQ2.2 α -chain, while Glu116 β formed a hydrogen bond with Asp66 on the HLA-DQ2.2 β -chain (Fig. 2D). Trp111 β of the CDR3 β loop made additional vdw

contact with HLA-DQ2.2 α -chain at residues Phe58 α , Thr61 α , and Val65 α . Further vdw interactions were made between the Gly112 β , Gly113 β , and Thr115 β of the CDR3 β loop and the HLA-DQ2.2

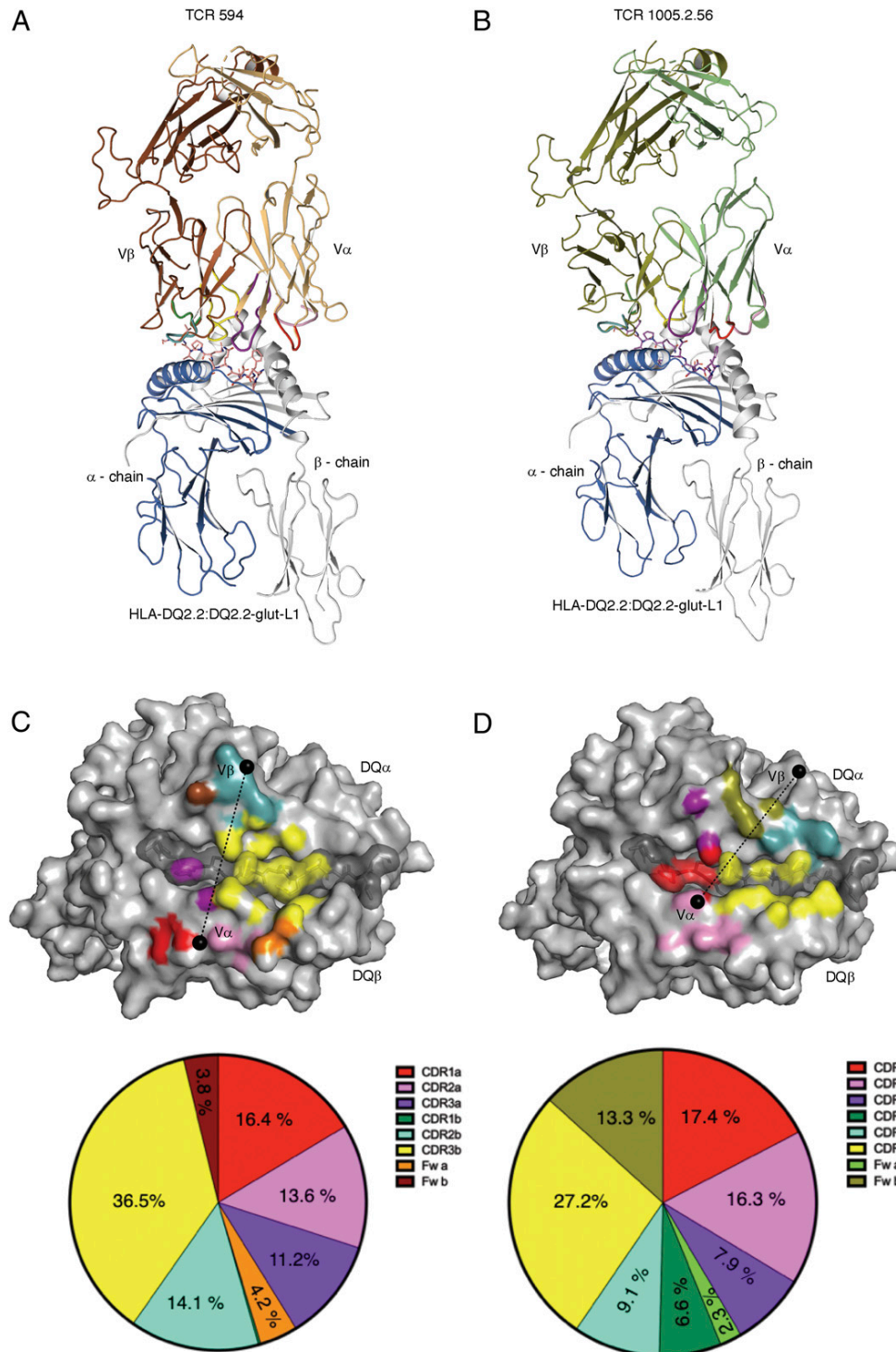


Fig. 1. Structures of TCR-HLA-DQ2.2:glut-L1 ternary complexes. TCR 594 (A) and TCR 1005.2.56 (B). Footprint of the CDR loops of the TCR 594 (C) and TCR 1005.2.56 (D) on the HLA-DQ2.2:glut-L1. CDR1 α , CDR2 α , and CDR3 α are colored in red, pink, and purple, respectively, while the CDR1 β , CDR2 β , and CDR3 β are colored in green, light teal, and yellow, respectively. The TCR framework for each V α - and V β -chain is colored orange and brown, respectively, for TCR 594, and colored lime and deep-olive, respectively, for TCR 1005.2.56. The pie chart below show percentage of the CDR loops contributing to the buried surface area.

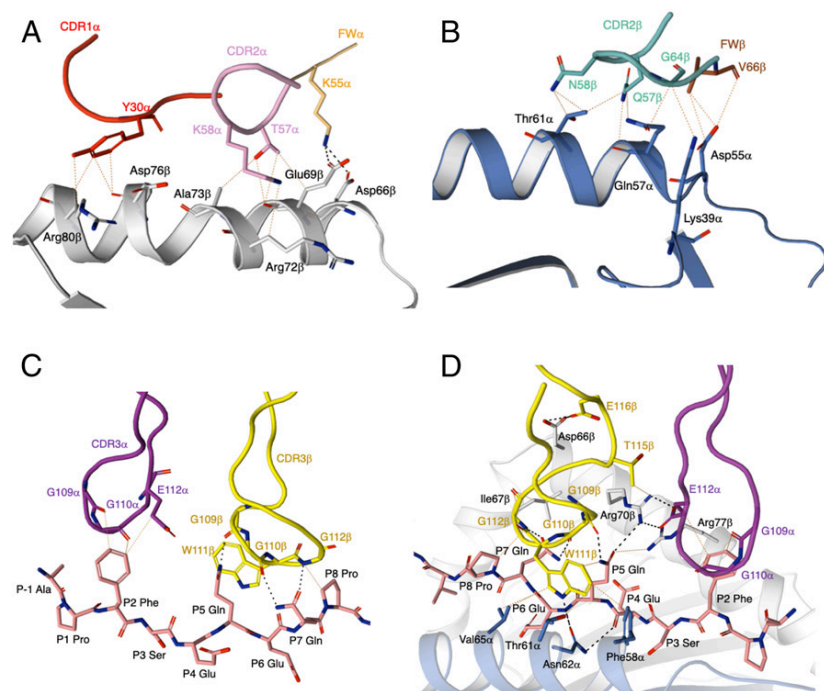


Fig. 2. TCR 594 CDR loops in contact with the HLA-DQ2.2:glut-L1. (A) The CDR1 α and CDR2 α loop contacting HLA-DQ2.2 β -chain. (B) The CDR2 β contacts with HLA-DQ2.2 α -chain. (C) CDR3 α and CDR3 β interactions with the DQ2.2-glut-L1 peptide. (D) Key interaction around the P5-Gln residue of the DQ2.2-glut-L1 epitope.

β -chain including residues Asp66 β , Thr61 β , and Ile67 β . Interestingly, the positioning of the Trp111 β in the CDR3 β of the TCR 594 is analogous to that of the conserved Arg109 found in the CDR3 β loop of the HLA-DQ2.5:glut-L1 reactive TCRs (*SI Appendix, Fig. S1*), making similar contact to HLA-DQ2.2 residues Phe58 α , Thr61 α , and Asn62 α , as well as gluten epitope residues at position P5 and P7 (Fig. 2 C and D).

Gly109 α and Thr112 β from the TCR 1005.2.56 also interacted with gluten epitope residues at position P5 (Fig. 3 D and E). However, the overall TCR:peptide interaction was further extended by contacts between the germ-line-encoded CDR loops and the DQ2.2-glut-L1 peptide residues P2-Phe, P3-Ser, P6-Glu, and P8-Pro, including Tyr37 α from the CDR1 α loop, Gln57 α from the CDR2 α loop, and Thr37 α from the CDR1 α loop (Fig. 3D). Notably, the P7-Gln residue in the TCR1005.2.56:HLA-DQ2.2:glut-L1 ternary complex was orientated toward the antigen-binding cleft and formed a hydrogen bond with Arg70 β from HLA-DQ2.2 (Fig. 3E) and was not involved in any direct TCR contact. In TCR 594:HLA-DQ2.2:glut-L1, Arg70 β forms a salt bridge with the Glu112 α from the CDR3 α loop and made polar contacts with the P5-Gln of the DQ2.2-glut-L1 (Fig. 2D). In the TCR1005.2.56:HLA-DQ2.2:glut-L1 ternary complex, however, the HLA-DQ2.2 residue Arg70 β had rotated toward the P7 peptide binding pocket, forming hydrogen bond with P7-Gln, pulling it inwards, while making hydrogen bond with Thr112 β and Asp113 β from the CDR3 β loop of TCR 1005.2.56 (Fig. 3 D and E). Overall, the TCR 1005.2.56 contacts with the HLA-DQ2.2:glut-L1 are more well distributed along the peptide compared to TCR 594 whose interactions focused on the central region of the peptide.

Comparison of HLA-DQ2.2:glut-L1 and HLA-DQ2.5:glut-L1. The DQ2.2-glut-L1 peptide (PFSESEQPV) used in this study was derived from low molecular weight glutenin protein subsequent to TG2 mediated deamidation of the P4 and P6 glutamine residues in

the native peptide sequence (PFSQQQPV) to glutamate (14, 24). Crystal structures of HLA-DQ2.5:glut-L1 and HLA-DQ2.2:glut-L1 reveal that lysine at position 71 of the HLA-DQ2.5 β -chain is involved in binding of the P4-Glu and P6-Glu (22, 23, 27). The electron density maps of the DQ2.2-glut-L1 peptide recognized by TCR 594 (Fig. 4A) and TCR 1005.2.56 (Fig. 4B) were unambiguous. A more comprehensive hydrogen bonding network is shown from the view of a TCR for these two ternary complexes in Fig. 4 C and D, corresponding to P1 to P9 residues of the DQ2.2-glut-L1 peptide shown in Fig. 4 A and B. Similar HLA-peptide contacts were observed in HLA-DQ2.2:glut-L1, where the P4-Glu and P6-Glu are buried deep into the binding pockets, acting as anchor residues, interacting with Tyr9 β , Ser30 β , and Lys71 β of the HLA-DQ2.2 (Fig. 4 C and D).

Upon superposition, HLA-DQ2.2:glut-L1 and HLA-DQ2.5:glut-L1 showed no major structural differences, with an rmsd of 0.50 Å. In their membrane distal domains the HLA-DQ2.2 and HLA-DQ2.5 have identical DQ β chains and differ by 10 residues in their DQ α -chains (Fig. 4 E–G). None of these polymorphic residues of HLA-DQ2.2 made direct contact with the DQ2.2-glut-L1 epitope, or with the TCR in both of the TCR 594:HLA-DQ2.2:glut-L1 and TCR 1005.2.56:HLA-DQ2.2:glut-L1 complexes solved in this study (Fig. 4 E and F). The only key polymorphic residue that is in contact with the DQ2.2-glut-L1 peptide is the Phe/Tyr substitution in position 22 of the DQ α -chain. Here, we clearly show that the presence of P3-Ser in HLA-DQ2.2:glut-L1 forms a hydrogen bond to side chain of His24 α and main chain of Tyr9 α of the HLA-DQ2.2 molecule (Fig. 4 E and F). By contrast, in HLA-DQ2.5, Tyr22 α hydrogen bonds with the His24 α (Fig. 4G) (22, 23). Moreover, these two HLA-DQ2.2:glut-L1 complexes displayed relatively conserved electrostatic-potential surfaces for TCR interaction (Fig. 4 H and I). Hence, such selective P3-Ser peptide binding preferences is attributed to a hydrogen-bonding network whose functionality is differentially influenced by tyrosine and

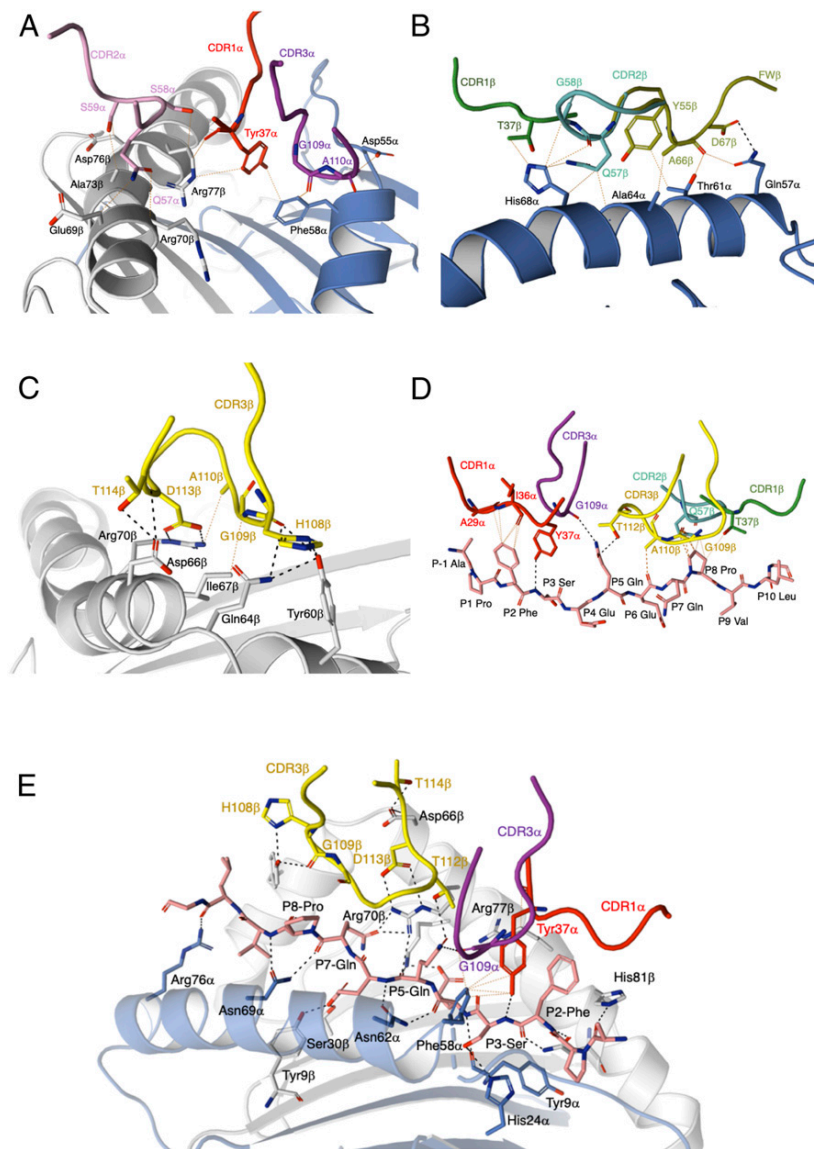


Fig. 3. TCR 1005.2.56 CDR loops in contact with the HLA-DQ2.2:glut-L1. (A) The CDR1 α , CDR2 α , and CDR3 α loops contact the HLA-DQ2.2 α -chain and β -chain. The CDR1 β , CDR2 β (B), and CDR3 β (C) interactions with the HLA-DQ2.2 α -chain and β -chain, respectively. (D) The CDR1 α and CDR3 β loops of the TCR 1005.2.56 that interacted the DQ2.2-glut-L1 peptide. The CDR3 α , CDR1 β , and CDR2 β loop contacts with the peptide. (E) Key interaction around the P5-Gln and P7-Gln residue of the DQ2.2-glut-L1 epitope.

phenylalanine. While the polymorphic Phe22 α is located at the base of the HLA peptide binding cleft and has no direct contact with the TCR, it provides a basis for understanding why P3-Ser binds more favorably to HLA-DQ2.2 compared to HLA-DQ2.5. These structural data are in keeping with a model previously presented (14).

Role of P3-Ser as an Anchor in Peptide Binding to HLA-DQ2.2. To further investigate the role of P3-ser as an anchor residue for HLA-DQ2.2, we performed a competitive peptide binding assay and compared the binding of DQ2.2-glut-L1 and 10 different P3-Ser substituted peptide variants (*SI Appendix, Table S6*) to HLA-DQ2.2 (Fig. 5). We contrasted the results with testing the same peptides for binding to HLA-DQ2.5 (Fig. 5).

Among all of the 11 peptides with different amino acids at P3 of the DQ2.2-glut-L1 epitope, the peptide containing P3-Ser

displayed the best binding to HLA-DQ2.2. Between the 10 P3-Ser-substituted peptides, threonine substitution had the least impact. The variant with cysteine also was a reasonable binder. A possible explanation why threonine and cysteine could serve as P3 anchors for HLA-DQ2.2 could be that their side chain can establish the above-mentioned hydrogen bond to the side chain of His24 α and the main chain of Tyr9 α . Further to this notion, α -aminobutyric acid, which is structurally similar to serine except for lacking the hydroxyl group, bound less well than the serine variant. Notwithstanding, α -aminobutyric acid was better than many of the other amino acids tested, indicating additional importance of size and hydrophobicity for optimal anchoring into the P3 pocket of HLA-DQ2.2. The patterns of binding of the 11 DQ2.2-glut-L1 variants to HLA-DQ2.5 was very different for that of HLA-DQ2.2 with P3-Ser not being preferred. Taken together, the binding experiments substantiate the notion of P3-Ser

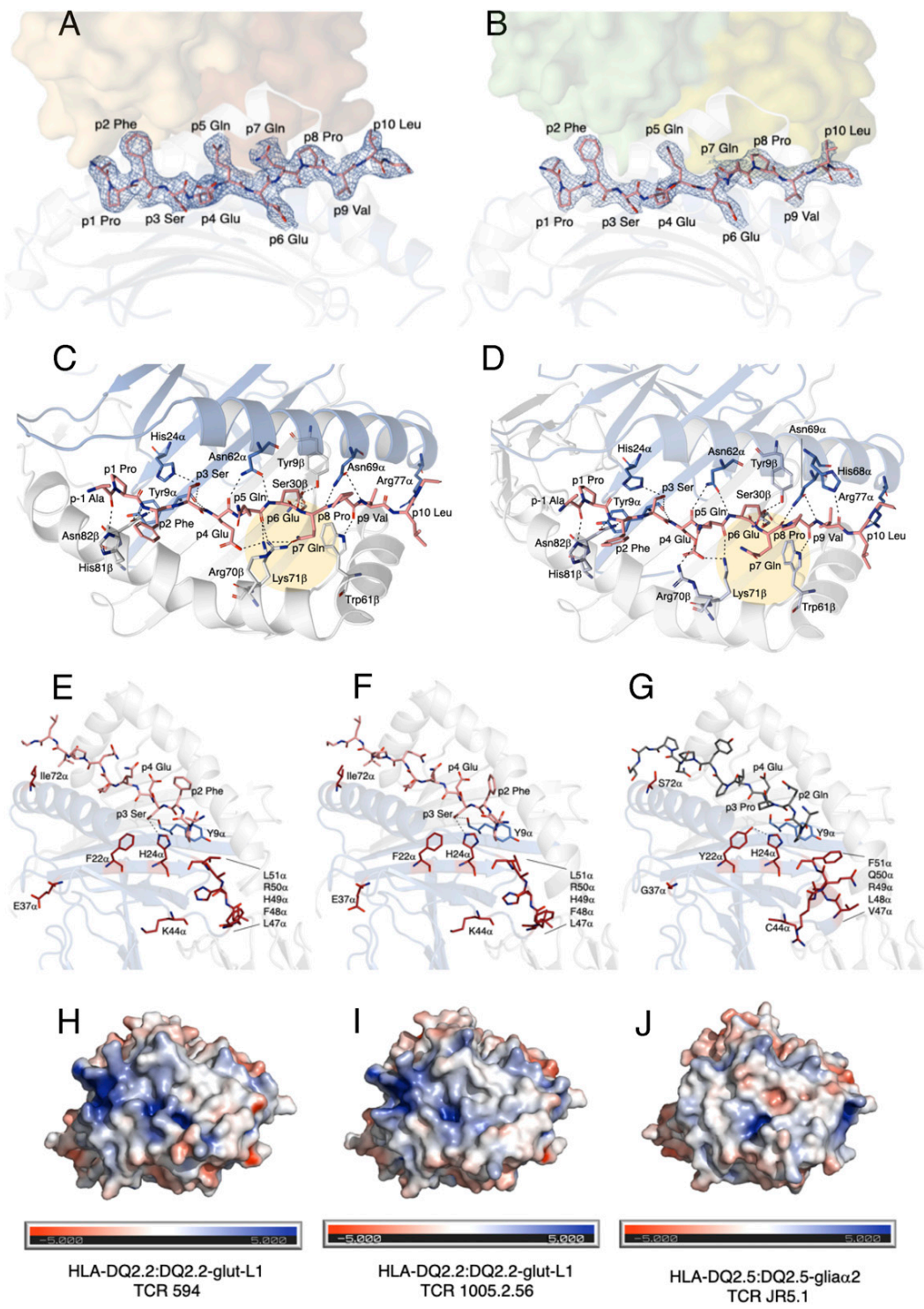


Fig. 4. Comparison of HLA-DQ2.2:glut-L1 and HLA-DQ2.5 in complexed with gluten epitopes. The DQ2.2:glut-L1 peptide is bound in the peptide-binding groove, with carbon colored in salmon, nitrogen colored in blue, and oxygen colored in red. The TCR 594 (A) and TCR 1005.2.56 (B) docked on top of the HLA-DQ2.2:glut-L1 are shown as solid surface, and each of the TCR $\alpha\beta$ pair were colored in brown/orange and green/lime, respectively. The peptide's 2Fo-Fc electron density map is shown in blue and contoured to 1 σ . Hydrogen bond interactions between HLA-DQ2.2 and the DQ2.2:glut-L1 epitope in the TCR 594:HLA-DQ2.2:glut-L1 ternary complex (C) and in the TCR 1005.2.56:HLA-DQ2.2:glut-L1 ternary complex (D) are shown. Comparison of the DQ2.2:glut-L1 peptide bound to HLA-DQ2.2 in the two ternary complexes solved showed minor movement in P7-Gln residues, highlighted in yellow circle. Polymorphic residues on the ectodomain of HLA-DQ differing between HLA-DQ2.2 (E and F) and HLA-DQ2.5 (G) are represented in stick and colored in firebrick. The His24 α interacts with P3-Ser in DQ2.2:glut-L1 (E and F) but with Tyr22 α in HLA-DQ2.5 (G). The solvent-accessible electrostatic potential was calculated for HLA-DQ2.2:glut-L1 in complexed with TCR 594 (H) or TCR 1005.2.56 (I) and HLA-DQ2.5:glut-L1 in complexed with TCR JR5.1 (J). Electrostatic calculations were performed using APBS (± 5 kT/e).

as a key anchor for peptide binding to HLA-DQ2.2 with the hydroxyl group of the serine side being functionally important.

TCR Affinity Measurements for the HLA-DQ2.2:DQ2.2-glut-L1 Complex.

As determined by surface plasmon resonance (SPR), the steady-state affinity (K_D) of the TCR 555, TCR 594, and TCR 1005.2.56 for the HLA-DQ2.2:DQ2.2-glut-L1 were 26.14 ± 2.64 , 20.95 ± 4.06 , and 22.17 ± 3.68 μM , respectively (Fig. 6). These values corroborate previous T cell proliferation studies (14). None of these TCRs tested showed any binding to HLA-DQ2.5:CLIP1 or HLA-DQ2.2:DQ2.2-glia- α_2 , thereby highlighting the lack of cross-reactivity of these TCRs.

To determine the extent of peptide sensitivity of these TCRs, we generated three peptide mutants corresponding to the solvent-exposed residues at position P2, P5, and P7 by replacing them with an alanine. The impact of these substitutions was measured by SPR. The PheP2Ala mutation reduced the affinity by at least 10-fold ($K_D > 200$ μM), whereas the GlnP5Ala mutation abolished TCR binding to HLA-DQ2.2-glut-L1. This is consistent with the crystal structure where P5-Gln played a central role in mediating TCR 594 and TCR 1005.2.56 interaction in HLA-DQ2.2:DQ2.2-glut-L1 (Figs. 2 C and D and 3 D and E). The GlnP7Ala mutation abolished the binding of TCR 555 and TCR 594, whereas the TCR 1005.2.56 showed weak binding ($K_D > 200$ μM) to this mutant, suggesting a different mode of interaction with this TCR. Indeed, as shown in the crystal structure, the P7-Gln in the TCR1005.2.56:HLA-DQ2.2:DQ2.2-glut-L1 ternary complex did not make any direct contact with the TCR, contrary to the TCR594:HLA-DQ2.2:DQ2.2-glut-L1 ternary complex (Fig. 3 D and E).

Discussion

HLA-DQ2.2 differs from HLA-DQ2.5 and HLA-DQ8 in gluten peptide-binding preferences and CeD risk association. Immuno-dominant gluten epitopes that binds stably to HLA-DQ2.5 were found to bind unstably to HLA-DQ2.2 and, furthermore, did not raise T cell responses in vivo in CeD patients carrying HLA-DQ2.2 (14, 16). Rather T cell clones from CeD patients expressing HLA-DQ2.2 recognized a unique set of gluten epitopes that commonly had a serine residue at position P3 (14, 16). Here, we provide a molecular basis for the T cell-mediated response to an HLA-DQ2.2-restricted immunodominant gluten epitope. We show that, in contrast to HLA-DQ2.5- and HLA-DQ8-mediated CeD, a diverse T cell repertoire underpins the response to HLA-DQ2.2 presenting the glut-L1 epitope. Moreover, we provide a structural basis underpinning this diverse TCR usage, by determining the crystal structures of two TCR:HLA-DQ2.2:DQ2.2-glut-L1 complexes.

Several studies have provided insight into TCR recognition of gluten epitopes presented by HLA-DQ2.5, HLA-DQ8, and HLA-DQ8.5. For instance, the HLA-DQ8:DQ8-glia- α_1 reactive TCRs exhibit biased *TRAV26-2/TRBV9-1* gene usage (18), and a nongerm-line-encoded arginine residue in either the CDR3 α or CDR3 β loop (23, 28). Moreover, *TRAV20+/TRBV9+* TCRs reactive with HLA-DQ8:DQ8-glia- α_1 and HLA-DQ8:DQ8.5-glia- γ_1 also possessed a key arginine residue in their CDR1 α loops (28, 29). All *TRAV26-1/TRBV7-2+* TCRs specific for DQ2.5-glia- α_2 had a nongerm-line-encoded arginine at position 109 of the CDR3 β loop, which forms key interactions with the P5-Leu and P7-Tyr residues found within the gliadin epitopes (22). The HLA-DQ2.5:DQ2.5-glia- α_1 reactive TCRs also shows biased TCR gene usage, *TRAV4* and *TRBV20-1* or *TRBV29-1* (22, 30),

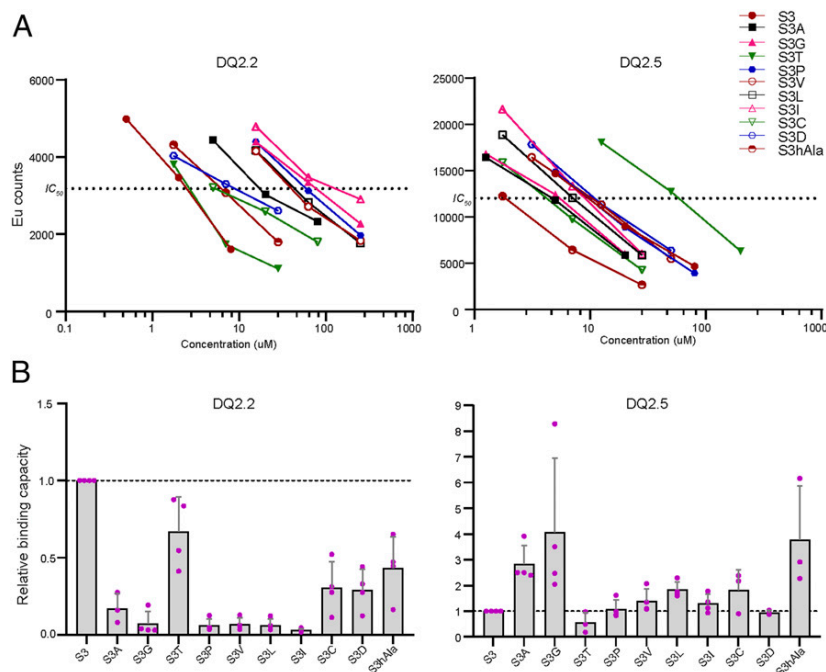


Fig. 5. Binding of unsubstituted and P3-substituted variants of the DQ2.2-glut-L1 epitope to HLA-DQ2.2 and HLA-DQ2.5. Synthetic peptides containing the DQ2.2-glut-L1 epitope (Ac-QQPFSE $\underline{\text{EQPVL}}\text{PQ}$, nine amino acid core sequence underlined), and variants substituted at P3 serine were tested as competitor peptides. The inhibitory effect of the competitor peptides is shown as IC_{50} values. One 10-fold titration experiment and three 4-fold titration experiments were performed. (A) Representative data showing results of one of the three fourfold titration experiments. (B) Results from all four independent experiments depicted as relative binding capacities (i.e., compared to the unsubstituted DQ2.2-glut-L1 epitope). The relative binding values from each experiment are shown as dots with bars representing mean values. A missing dot for some peptides is due to the titration curve not reaching its relevant IC_{50} value. Error bars, SD.

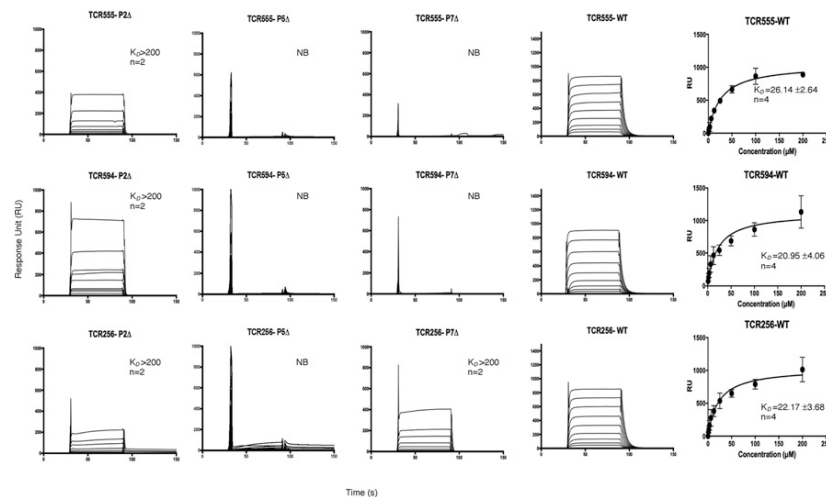


Fig. 6. Affinity measurement of TCR 555, 594, and 1005.2.56. Affinity for HLA-DQ2.2:glut-L1 WT and the P2-Ala, P5-Ala, and P7-Ala epitope mutants were measured via SPR. Two to four independent experiments (with the number of replicates shown as n numbers) were carried out for each of the TCR against each of HLA-DQ2.2:glut-L1 WT and epitope mutants. Binding curves shown represent an independent experiment of TCR 555, TCR 594, TCR 1005.2.56 (labeled as TCR256) binding to HLA-DQ2.2:glut-L1. All data were combined for each TCR and a one-site specific-binding model was used for curve fitting. HLA-DQ2.5-CLIP was used as negative control and acted as baseline reference value. Error bars, SD. NB, no binding.

but they do not carry a conserved arginine that is essential for recognition (31, 32). Ensuing structural studies have identified specific structural features that provided a basis for the selection of biased TCR usage and the preferential usage of the Arg residue (22, 23).

The TCR594:HLA-DQ2.2:glut-L1 interactions resembled some features of both TCR S2 and TCR S16 that recognized HLA-DQ2.5:glut-L1 and HLA-DQ2.5:glut-L1, respectively. Like the S2 TCR, the germ-line-encoded CDRs on the TCR 594 V α possess a tyrosine that make similar interactions with the HLA-DQ2 β -chain helix. Contrary to TCR S2, the TCR 594 interaction with the P5-Gln and P7-Gln was reminiscent of TCR S16 and JR5.1 recognition of the HLA-DQ2.5:glut-L1 epitope. Here, although the TCR594 did not possess a conserved arginine residue within the CDR3 β loop, Trp111 β in the CDR3 β loop of the TCR 594 played an analogous role, forming extensive interactions with the HLA-DQ2.2:glut-L1 complex (*SI Appendix, Fig. S1*). The nature of the contacts between the TCR 594 and the HLA-DQ2.2:glut-L1 also provided a basis for understanding the lack of cross-reactivity toward HLA-DQ2.5:glut-L1. Indeed, the HLA-DQ2.2:glut-L1 reactive TCRs studied here appeared to be epitope-specific and epitope-driven.

However, the TCR1005.2.56:HLA-DQ2.2:glut-L1 interactions resembled TCR Bel502 and Bel602 that recognized HLA-DQ8-glia- α 1 and HLA-DQ8-glia- γ 1, respectively (28, 29) (*SI Appendix, Fig. S2*). Here, TCR Bel502 and Bel602 possess a conserved Arg37 in the CDR1 α loop that interacts with the backbone of the gliadin peptide near the N terminus. The TCR 1005.2.56, like TCR 594, did not bear an arginine in the CDR3 β loop nor in the CDR1 α loop. However, the Tyr37 α in the CDR1 α loop of the TCR1005.2.56 sat atop of the N terminus of the DQ2.2:glut-L1 epitope in the same manner as the conserved Arg37 α in the CDR1 α loop of TCR Bel502 and Bel602 (*SI Appendix, Fig. S2B*).

Despite HLA-DQ2.2 compensating for the substitution of Tyr22 α by selectively binding to gluten epitopes with P3-Ser, it bears a lower risk of CD in comparison to HLA-DQ2.5. A higher chance of proteolytic degradation and stricter binding requirements are likely what limits the repertoire of potential HLA-DQ2.2 epitopes. In addition, the DQ2.2:glut-L1 has a chymotrypsin cleavage site between its P2 and P3 residues, making it

more susceptible to proteolytic degradation. The requirement of the deamidation of glutamine residues at P4 and/or P6 for optimal binding to HLA-DQ2.2 further also limits the number of candidate antigenic epitopes in gluten for individuals with this allotype. These restrictions likely limit the bioavailability of epitopes that could bind to HLA-DQ2.2.

In summary, we have investigated the structural basis of how HLA-DQ2.2-restricted and CeD-derived TCRs bind to HLA-DQ2.2 complexed with the immunodominant DQ2.2:glut-L1 epitope. The single polymorphic residue in HLA-DQ2.2 (Phe22 α) influences the hydrogen bond network within the peptide-binding groove of HLA-DQ2.2 and selectively bind to gluten epitopes that carry a P3-serine as anchor residue. What is remarkable is that while the TCR-accessible features of the antigen-binding clefts of HLA-DQ2.5 and HLA-DQ2.2 are very similar, the nature of the bound epitopes generate markedly different responding TCR repertoires. In HLA-DQ2.5-mediated CeD, the response is characterized by biased TCR usage, while in HLA-DQ2.2-mediated CeD, a diverse TCR response is generated. Thus, relatively subtle differences in the peptide-centric atomic landscape can have profound effects on the ensuing T cell response, which is correlated with disease pathogenesis.

Methods

Patient Material. We characterized T cells isolated from gut biopsy-derived T cell lines of four DQ2.2⁺ subjects that have been described previously (14). According to the Oslo definitions of CeD and related terms (33), three of these subjects (CD555, CD594, and CD627) fulfilled the criteria of CeD while one subject (CD1005) fulfilled the criteria of potential CeD having normal gut histology but highly positive anti-TG2 serology. The subjects gave written informed consents for participation in the previous study, and updated written informed consent forms were obtained for this study. The responsible clinicians had access to sensitive patient data, whereas the research was done on coded material. The study protocol was approved by the Regional Ethical Committee (South East) for Medical and Health Research Ethics (present project ID: 6544). From the T cell lines, we sorted single T cells with HLA-DQ tetramers for TCR sequencing and TCRs of already-established T cell clones were sequenced (TCC627.1.3.199, TCC594.5.2.6, and TCC555A.1.4.38 from ref. 14 and TCC1005.2.54, TCC1005.2.56, TCC1005.2.57, TCC1005.2.58, TCC1005.2.60, TCC1005.2.62, and TCC1005.2.65 from ref. 34).

Tetramer Staining, Surface Antibody Staining, and Fluorescence-Activated Cell Sorting. Tetramers of recombinant HLA-DQ2.2 covalently linked with DQ2.2-glut-L1 peptide (QPPFSEQEQPVL, underlined 9-mer core amino acid sequence) and CLIP1 (PVSKMRMATPLLMQA, negative control for nonspecific binding) were produced as per Dorum et al. (24) and conjugated with phycoerythrin (PE)-labeled streptavidin (Invitrogen) and allophycocyanin (APC)-labeled streptavidin (ProZyme), respectively. Cryopreserved T cell lines were thawed and stained with PE-conjugated HLA-DQ2.2:DQ2.2-glut-L1 (10 µg/mL) and APC-conjugated HLA-DQ2.2: CLIP1 (10 µg/mL) for 2 h at 37 °C. Following tetramer staining, the cells were stained with the mix of surface antibodies for fluorescence-activated cell sorting (FACS). We sorted live singlets that were CD3⁺, CD8⁻, CD4⁺, HLA-DQ2.2:DQ2.2-glut-L1 tetramer⁺, and HLA-DQ2.2:CLIP1 tetramer⁻ (SI Appendix, Fig. S3). The cells were sorted in 96-well plate containing lysis buffer for single-cell TCR sequencing. The sorting was performed with a FACS Aria II instrument (BD Biosciences) at the Flow Cytometry Core Facility (Oslo University Hospital), and the flow cytometry data were analyzed with FlowJo software (FlowJo LLC). The following antibodies were used in the study: CD3-Superbright 600 (eBioscience), CD4-APC-H7 (BD Biosciences), and CD8-PerCP (eBioscience). LIVE/DEAD marker fixable violet stain (Thermo Fischer Invitrogen) was used to exclude dead cells.

TCR Sequencing of T Cell Clones. Sequences of rearranged TCR- α and TCR- β genes of T cell clones were obtained by Sanger sequencing using a protocol as described (35).

Single-Cell TCR Gene Sequencing and Processing of TCR Gene Sequences. We used a previously published protocol, based on nested PCR amplification using multiplex TRAV and TRBV primers, for single-cell TCR $\alpha\beta$ sequencing (3, 36). The processing of the raw sequences generated from Illumina NGS was carried out as described (3). Sequencing was performed using Illumina MiSeq (250 bp PE) platform at the Norwegian Sequencing Centre (Oslo University Hospital).

Competitive Peptide Binding Assays. The competitive peptide binding assay was performed as described previously (14, 37). In brief, the Epstein-Barr virus (EBV)-transformed B-cell lines VAVY (#9023) and MOU (#9050) were used as the source of HLA-DQ2.5 and HLA-DQ2.2 molecules. Microtiter plates (96 well, Nunc) were coated with an anti-DQ mAb (mAb and SPV-L3; 2 µg per well) as capture antibody for overnight at 4 °C. The wells were then blocked with phosphate buffered saline/gelatin (0.2%) at 37 °C for 5 h. The plates were washed, and cell lysate of EBV cells (equivalent of 4 million cells) was added to wells and incubated at 4 °C overnight. The plates were washed and biotinylated indicator peptide P418 (EPRAPWIEQEGPEYVW-biotin, 0.5 µM) together with unlabeled competitor peptides (GL Biochem) were added in binding buffer with protease inhibitors (Complete ethylenediaminetetraacetic acid [EDTA]-free tablets, Roche Molecular Biochemicals) and 1 mM dithiothreitol followed by 48-h incubation at 37 °C. The plates were washed and incubated with streptavidin-europium diluted (1000x dilution) in DELFIA assay buffer (Wallac) for 45 min at room temperature. The plates were washed and incubated with DELFIA enhancement solution (Wallac) for 15 min at room temperature. Binding of the indicator peptide was then measured as Eu counts in a time-resolved fluorometer (Wallac 1420 multilabel counter). IC₅₀ values (the concentration of the competitor peptide required for half-maximal inhibition of the binding of the indicator peptide) were established for each peptide. One 10-fold titration experiment and three 4-fold titration experiments were performed. The relative binding capacity was calculated as the ratio of the IC₅₀ value of the DQ2.2-glut-L1 to the IC₅₀ value of the P3-substituted peptide variants.

Protein Expression and Purification. cDNAs (Genscript) encoding the individual extracellular domains of TCR α -chain and β -chain, with an engineered interchain disulfide linkage in the constant domains, were cloned into the pET-30 vector and expressed in *Escherichia coli* BL21DE3. The TCR proteins were purified from inclusion bodies, as described previously (38, 39). The TCR $\alpha\beta$ heterodimers were refolded in the presence of 5 M Urea, followed by dialysis and further purification through anion exchange chromatography, size-exclusion chromatography, and hydrophobic-interaction chromatography as described previously (18).

For SPR and structural experiments, the extracellular domains of the HLA-DQ2.2 α -chain and HLA-DQ2.2-glut-L1 β -chain (with glut-L1 epitope covalently linked to the HLA-DQ2.2 β -chain) were coexpressed as soluble protein in High Five insect cells (*Trichoplusia ni* BTI-TN-5B1-4 cells; Invitrogen), via a baculovirus expression system as described previously (17). Briefly, baculovirus HLA-DQ2.2 expression constructs had enterokinase-

cleavable Fos and Jun zippers at the C-terminal ends of the α -chain and β -chain, respectively, to promote dimerization. The C terminus of the β -chain also included a BirA recognition sequence for biotinylation as described previously (18), as well as a histidine tag for purification using immobilized metal ion affinity chromatography. Before crystallization, the Fos and Jun zippers were removed by enterokinase (Genscript) digestion, followed by separation using anion exchange chromatography (HiTrap Q HP, GE Healthcare). Purified HLA-DQ2.2:DQ2.2-glut-L1 was mixed in a 1:1 ratio with purified TCR 594 or TCR 1005.2.56, and the protein complex was then purified by size exclusion chromatography (Superdex 200; GE Healthcare). Three HLA-DQ2.2-glut-L1 epitope mutants were generated by replacing the glut-L1 WT core sequence (PFSEQEQPV) with glut-L1-P2 Δ (PASEQEQPV), glut-L1-P5 Δ (PFSEAEQPV), and glut-L1-P7 Δ (PFSEQEAPV) separately in the HLA-DQ2.2 β -chain and coexpressed with HLA-DQ2.2 α -chain via a baculovirus expression system as described above.

Crystallization, Data Collection, and Processing. The TCR 594/TCR 1005.2.56 and the HLA-DQ2.2:DQ2.2-glut-L1 complex, in 10 mM Tris (pH 8), 150 mM NaCl, were concentrated to 6 mg/mL. Crystallization was carried out using the hanging drop vapor diffusion method at 17 °C. For TCR594:HLADQ2.2:DQ2.2-glut-L1, diffraction quality crystal was obtained in conditions 0.1 M 2-(N-morpholino)ethanesulfonic acid pH 6.5, 5% vol/vol MPD, and 15% wt/vol PEG6K, and crystallization droplets were set up by adding 1 µL of the mother liquor to 1 µL of protein solution. Crystals typically appeared within 2–7 d. For TCR1005.2.56:HLA-DQ2.2:DQ2.2-glut-L1, diffraction quality crystal was obtained in an optimized Morpheus screen conditions 0.1 M Mops/Hepes pH 7.5, 12.5% vol/vol PEG1K, 12.5% vol/vol MPD, 12.5% wt/vol PEG 3350, and 0.04 M nitrate phosphate sulfate mixture (40), and crystallization droplets were set up by adding 1 µL of the mother liquor to 1 µL of protein solution and with the addition of 0.2 µL of seed stock. Crystals typically appeared within 2–3 wk. Protein crystals were exposed to cryoprotectant, 20% glycerol for 30 s before being flash-cooled in liquid nitrogen. Datasets were collected on the MX2 beamline of the Australian Synchrotron. Data were integrated with XDS and scaled and merged in Aimless. Phases were obtained using molecular replacement in PhaserMR, CCP4suite (41). Protein Data Bank (PDB) ID code 4OZH (chain A and B, without glycans or peptides for HLA; chain G and H for TCR) was used as search model for HLA-DQ2.2:DQ2.2-glut-L1 and TCR 594 for molecular replacement. The data collected for the TCR594:HLADQ2.2:DQ2.2-glut-L1 was highly anisotropic, thus ellipsoidal truncations and anisotropic scaling were carried out at the Diffraction Anisotropic Server (42). The structure was built and refined in Coot and PHENIX (43, 44). Final model was validated in Molprobitry (44).

Surface Plasmon Resonance Measurement and Analysis. Surface plasmon resonance experiments were conducted on a BiAcCore T200 system (GE Healthcare Life Sciences) instrument using HBS buffer (10 mM Hepes-HCl, pH 7.4, 150 mM NaCl, and 0.005% surfactant P20). The biotinylated HLA-DQ2.5:CLIP2, HLA-DQ2.2:DQ2.2-glut-L1, and respective peptide mutant DQ2.2-glut-L1-P2 Δ , DQ2.2-glut-L1-P5 Δ , DQ2.2-glut-L1-P7 Δ were immobilized on a streptavidin-coated Biacore Series S Sensor Chip SA (SA Chip; GE Healthcare Life Sciences), creating a surface density of ~800–1,000 response units (RU). The TCR 555, TCR 594, and TCR 1005.2.56 (one in two serial dilutions from the maximum concentration of 200 µM) were injected over the chip at 10 µL/min for 2 min. The TCR response to HLA-DQ2.5:CLIP2 alone was subtracted from the response to HLA-DQ2.2:DQ2.2-glut-L1 and respective epitope mutants. Data in the main text are representative of two or four independent experiments. The equilibrium dissociation constant, K_D , was generated with nonlinear regression analysis of the combined data using Prism (GraphPad Software).

Data Availability. The protein structural data that support the findings of this study have been deposited in the RCSB PDB with the ID codes 6PX6 and 6PY2. All of the single-cell TCR sequencing raw data generated in this study are available in the European Genome-phenome Archive (EGAS00001003673).

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