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Single Diastereomers of the Clinical Anti-cancer ProTide Agents NUC-1031 and NUC-3373 Preferentially Target Cancer Stem Cells

In Vitro

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ABSTRACT

A 3'-protected route towards the synthesis of the diastereomers of clinically active ProTides, NUC-1031 and NUC-3373 is described. The *in vitro* cytotoxic activities of the individual diastereomers were found to be similar to their diastereomeric mixtures. In the KG1a cell line NUC-1031 and NUC-3373 have preferential cytotoxic effects on leukaemic stem cells (LSCs). These effects were not diastereomer-specific and were not observed with the parental nucleoside analogues gemcitabine and FUDR, respectively. In addition, NUC-1031 preferentially targeted LSCs in primary AML samples and cancer stem cells in the prostate cancer cell line, LNCaP. Although the mechanism for this remains incompletely resolved, NUC-1031-treated cells

showed increased levels of triphosphate in both LSC and bulk tumour fractions. As ProTides are not dependent on nucleoside transporters, it seems possible that the LSC targeting observed with ProTides may be caused, at least in part, by preferential accumulation of metabolised nucleos(t)ide analogues.

INTRODUCTION

The phosphoramidate (ProTide) approach¹ applied to anti-viral and anti-cancer nucleoside analogues (NAs), and more recently also to non-nucleoside compounds,²⁻⁸ continues to be of significant interest in the drug discovery field. This approach is used to overcome the limitations of NAs that are known to restrict their therapeutic potential. In the oncology arena, the application of ProTide technology to gemcitabine and floxuridine (FUDR) led to the discovery and clinical development of two anti-cancer agents, NUC-1031⁹ and NUC-3373¹⁰ (Figure 1). The first-in-class agent, NUC-1031, was designed to overcome the cancer cell resistance mechanisms that limit the clinical utility of gemcitabine (dFdC). The three key resistance mechanisms involve down-regulation of nucleoside transporter proteins (hENT1); down-regulation of the key initial phosphorylating enzyme (deoxycytidine kinase [dCK]) that is required to activate dFdC to the 5'-monophosphate form dFdCMP; and up-regulation of the catabolising enzyme cytidine deaminase (CDA).¹¹⁻¹³ NUC-1031 has been assessed in four completed clinical studies, including a Phase I study in patients with advanced solid tumors (PRO-001), a Phase Ib study in patients with recurrent ovarian cancer (PRO-002), a Phase Ib study in patients with locally advanced or metastatic biliary tract cancer (BTC) (ABC-08), and a Phase II study in patients with platinum-resistant ovarian cancer (PRO-105). Two clinical studies of NUC-1031 are currently ongoing: a Phase III study in metastatic pancreatic carcinoma (ACELARATE), and a

Phase III study in locally advanced or metastatic BTC (NuTide:121). Results from PRO-001 revealed that NUC-1031 has a favourable pharmacokinetic profile with long plasma half-life (8.3 hours versus 1.5 hours) and can generate intracellular levels of the active anti-cancer metabolite dFdCTP that are 217-times higher than those generated by gemcitabine. Furthermore, NUC-1031 achieved high levels of disease control (78%) in patients who had exhausted all other treatment options, including those with tumours relapsed/refractory to prior chemotherapy with gemcitabine.¹⁴ In June 2019, Acelarin, the NUC-1031_*S_P* isomer enriched API (98% *S_P*-isomer: 2% *R_P*-isomer), received orphan drug designation from the FDA for the treatment of BTC.¹⁵ Currently, the *S_P* isomer of NUC-1031 is being investigated in all NUC-1031 clinical studies.

The second agent, NUC-3373, is a ProTide modification of FUDR, the fluorinated pyrimidine nucleoside derivative of 5-FU. These fluoropyrimidines are established clinical drugs used in the treatment of solid tumours including gastrointestinal, kidney, ovarian and breast cancers.¹⁶ The key resistance mechanisms that limit 5-FU and FUDR involve down-regulation of the activating enzyme, thymidine kinase (TK), required for the first phosphorylation step that transforms FUDR to the active anti-cancer metabolite (FdUMP); up-regulation of thymidylate synthase (TS), which is considered to be the main target for 5-FU and FUDR; up-regulation of the degradative enzyme thymidine phosphorylase (TP); and decreased expression of nucleoside/nucleobase transport proteins.¹⁷ In addition, more than 85% of administered 5-FU is degraded by dihydropyrimidine dehydrogenase (DPD) before it has an opportunity to enter cancer cells and exert any therapeutic effect.¹⁸ *In vitro* data confirmed that NUC-3373 exerts its cytotoxic activity independently of thymidine

kinase in TK-deficient tumour cell lines and it is resistant to degradative action by catabolic enzymes including TP and DPD.¹⁹ In addition, NUC-3373 was found to generate 366-fold higher intracellular levels of the active monophosphate form, FdUMP, in the human colorectal cancer cell line HT29 compared with 5-FU and achieved significantly greater tumour volume reduction than 5-FU in HT29 xenograft studies.²⁰ NUC-3373, as a diastereomeric mixture, is currently being assessed in two ongoing clinical studies. Interim pharmacokinetic data from the first-in-human Phase I study of NUC-3373 in patients with advanced solid tumours (NuTide:301) demonstrated that NUC-3373 has a long plasma half-life (9.7 hours versus 8-14 minutes) and the ability to completely deplete the pool of dTMP in patients PBMCs, compared to 5-FU. A three-part Phase I study of NUC-3373 in combination with standard agents used in colorectal cancer treatment (NuTide:302) is also ongoing and is designed to identify a recommended Phase II dose (RP2D) and schedule for NUC-3373 when combined with other agents.²¹ Recently, a third anti-cancer agent, NUC-7738, the ProTide modification of 3'-deoxyadenosine (3'-dA; also known as cordycepin), entered a Phase I clinical study.²²

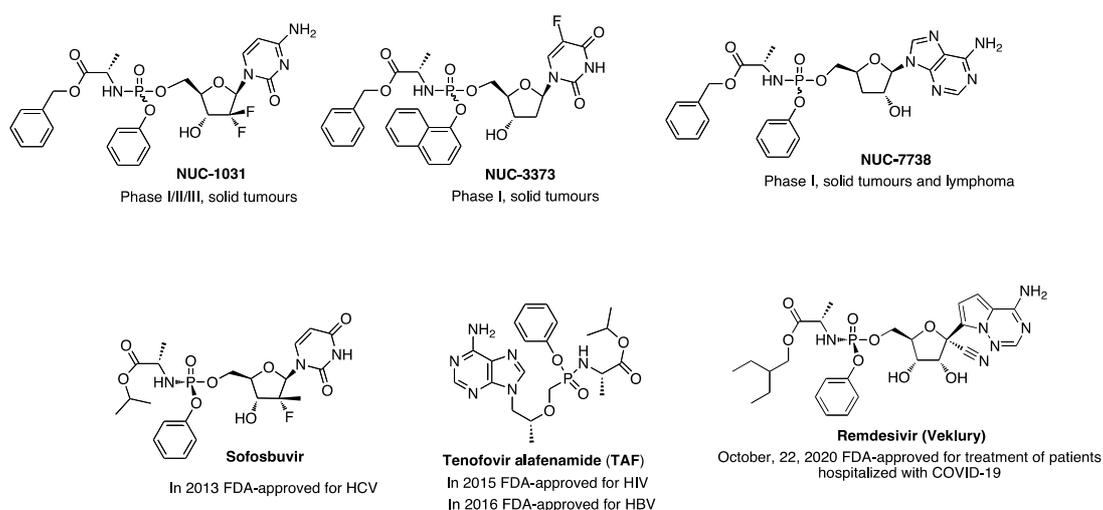


Figure 1. Phosphoramidate transformation of anti-cancer and FDA-approved anti-viral nucleoside monophosph(on)ate analogues in the clinic and ongoing clinical studies.

NUC-1031 and NUC-3373 ProTides were obtained via a coupling reaction between 3'-Boc-protected gemcitabine or FUDR and phosphorylating agent (phosphorochloridate) in the presence of either Grignard reagent (*tert*-butyl magnesium chloride) or *N*-methylimidazole (NMI). In both cases, the formation of a pair of diastereomers at the phosphate centre (1:1 ratio $S_P : R_P$) was achieved and, as such, NUC-1031 and NUC-3373 were further biologically evaluated.^{9,10} Although NMI favours the phosphorylation of the primary hydroxyl group at the nucleoside's 5'-position, the yield of the coupling reaction is usually low. On the contrary, the Grignard reagent is not selective, hence allowing the formation of a 5'-phosphoramidate along with undesired 3'- and 3',5'-phosphoramidates. In order to prevent the formation of these by-products, a selective 3'-protection of the nucleosides is required prior to the coupling reaction. This strategy involves an additional deprotection step at the 3'-positions in the obtained protected-phosphoramidates, as recently reviewed.²³ Because the yield of coupling reaction in the presence of the Grignard reagent is usually moderate to high, we adopted the hydroxyl groups protection as a strategy to prepare NUC-3373 with higher yield. In the case of gemcitabine, 3'-OH protection is more direct and selective due to the presence of fluorine atoms at the 2'-position in the sugar moiety. For FUDR, which lacks any substituents at the 2'-position, an initial protection of both 3'- and 5'-OH and subsequent selective 5'-deprotection was necessary. The 3'-OH protected FUDR was further used for a coupling reaction with a phosphorochloridate in the presence of

the Grignard reagent to improve the overall yield of NUC-3373 synthesis. A variety of hydroxyl protecting groups can be used to prepare 3'-protected ProTides.²³ *Tert*-butyl silyl group (TBDMS) was selected at first and was found to be the most optimal as a clear separation of two diastereomers of 3'-protected NUC-3373 mixture was possible by normal-phase chromatography. Although a regioselective strategy to obtain the desired 5'-ProTides was published,²⁴ during the course of the present study, we decided to proceed with our alternative methodology and to separate a mixture of two diastereomers at the level of 3'-protected ProTides. This alternative approach was of particular interest in light of the notable difference in activity between S_P and R_P isomers discovered for FDA-approved anti-viral ProTide agents sofosbuvir²⁵ (18-fold increase in potency against HCV for the S_P isomer)²⁶ and TAF²⁷ (10-fold increase in potency against HIV for the S_P isomer)²⁸ (Figure 1). In fact, chirality and chiral separation of racemic compounds are considered as an important topic in drug development. Most isomers of chiral drugs, including diastereomers, differ in their pharmacology, pharmacokinetics, metabolism and biological activities.²⁹ With this in mind, the current FDA recommendation is to assess activity of each isomer of racemic therapeutic agents.³⁰ Over the past decade, different diastereoselective methods for the synthesis of *P*-chirogenic phosphoramidates have been developed, including methods using a chiral auxiliary-bearing phosphoramidating reagent,³¹ diastereomerically pure phosphoramidating agent with *p*-nitro-phenyl or 2,3,4,5,6-pentafluorophenyl as leaving aryloxy groups;^{32,33} or method based on the process for the diastereomeric enrichment due to differences in solubility of S_P and R_P diastereomers in alcohols.³⁴

In our laboratory we have focused on the development of diastereoselective synthetic methods *via* copper-catalysed reaction in the presence of different bases.³⁵ Additionally, methods for the separation of 3'-protected ProTides as a diastereomeric mixture, in general being very difficult to isolate as single diastereomers by standard chromatography (reverse-phase chromatography or crystallization), were probed. Herein, we report an alternative synthetic procedure to obtain S_P and R_P isomers of the two clinical anti-cancer agents NUC-3373 and NUC-1031. A selective TBDMS-protection and deprotection manipulation of FUDR and gemcitabine was followed by a coupling reaction to give 3'-protected phosphoramidates which were separated to single diastereomers. Further removal of 3'-TBDMS group in each separated isomer yielded NUC-3373 and NUC-1031 as single isomers. NUC-1031 diastereomers were obtained upon column chromatography in crystalline form and the stereochemistry at the phosphorus centre was assigned via X-ray crystallography. Moreover, for the first time, we disclose *in vitro* biological data for single R_P and S_P isomers of NUC-1031 and NUC-3373 (stereochemistry to be determined), as well as their ability to target cancer stem cells (CSC).

RESULTS AND DISCUSSION

Chemistry

The selective protection and deprotection of hydroxyl groups in nucleoside analogues subjected to ProTide technology and the coupling reaction, represents a solid strategy to improve the overall yield of phosphoramidate synthesis. This alternative approach may be considered as advantageous over the general method in which non-protected nucleoside starting materials are used, as only the desired 5'-phosphorylated ProTides are formed. The alternative procedure for the preparation of clinical anti-cancer

agents NUC-3373 and NUC-1031 from 3'-protected FUDR and gemcitabine, respectively, is depicted in general Scheme 1. In our approach to improve the efficiency of the route to obtain the desired 5'-ProTides, we selected the *tert*-butyldimethyl silyl group reported for its ability to be selectively cleaved from the primary TBDMS hydroxyl groups in the presence of their secondary equivalents.³⁶ Thus, first FUDR (**1**) was treated with TBDMS, imidazole and DMAP at elevated temperature³⁷ to give 5',3'-TBDMS protected FUDR derivative (**3**) in 98% yield. Next, the FUDR intermediate **3** was subjected to different deprotection conditions to selectively remove the 5'-TBDMS moiety. The primary silyloxy groups are known to be cleaved under acidic conditions more efficiently in comparison with their secondary counterparts. Therefore, we investigated a variety of acidic conditions starting with a TFA/H₂O/THF (1:1:4) mixture, reported by Zhu *et al.* as the most suitable for selective 5'-desilylation of multisilylated nucleoside analogous.³⁶ However, when compound **3** was treated under the above-mentioned conditions, 3'-TBDMS-FUDR derivative (**5**), was obtained in low yield (20%). Due to poor selectivity of trifluoroacetic acid toward 5'-desilylation, different acids such as *p*-toluenesulfonic acid (*p*-TsOH) or camphorsulfonic acid (CSA) were further tested as summarised in Table 1. No selectivity was observed when desilylation was performed with 0.6 equivalent of *p*-TsOH in methanol at room temperature. After 1 h of reaction, a mixture of 3'- and 5'-mono silylated products in almost 1:1 ratio were detected along with a significant amount of starting material **3** (Entry 1, Table 1). Decreasing the reaction temperature to -15°C considerably improved the selectivity and the desired product **5** was recovered in 45% yield after 4 h (Entry 2, Table 1). Replacing MeOH with DCM drastically slowed down the reaction and significantly decreased the 5'-desilylation selectivity (data not reported). The yields were estimated by ¹H and

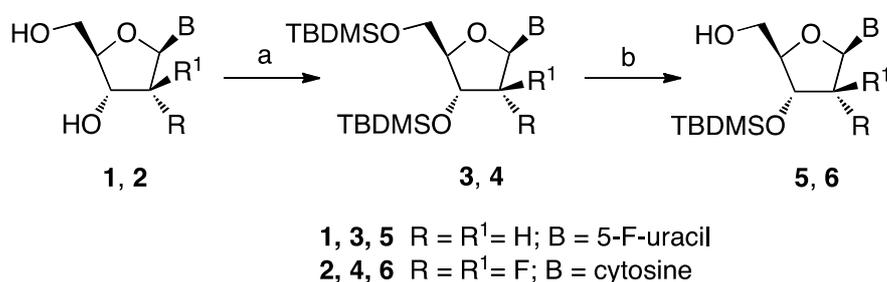
^{19}F NMR of the crude reaction mixture. Given the unsatisfactory result obtained with *p*-TsOH, we turned our attention to a new strategy with 10-camphorsulfonic acid (CSA), which has been reported in a few cases to be able to selectively deprotect primary silyl ether in multi-silylated nucleosides to give free primary alcohol.^{37,38} Thus, when the reaction was carried out in a 1:1 mixture of DCM/MeOH at low temperature (0-5°C), slow deprotection was detected; however, a remarkable selectivity was observed. The desired compound **5** was obtained in 38% crude yield as estimated by ^1H and ^{19}F NMR. By increasing the temperature to 25°C the selectivity toward 5'-desilylation was retained and the reaction was completed within 2.5 h, providing compound **5** with 37% yield (data not shown). A major improvement was obtained when only MeOH was employed as a solvent. In this case the reaction was accomplished much faster and after 1 h the desired compound **5** was formed in 60% crude yield based on ^1H and ^{19}F NMR (Entry 5, Table 1). Performing the reaction under the same conditions but at room temperature increased the rate of the deprotection. However, after 45 minutes, a significant amount of FUDR was formed, suggesting that meticulous monitoring is required (Entry 7, table 1). The best result was obtained when performing the reaction in MeOH with 1 equivalent of CSA at room temperature in 10 minutes. Under these conditions the desired product **5** was recovered after work up in 60% yield (Entry 8, Table 1). Additional attempts to improve the yield of 5'-TBDMS removal by changing solvents were unsuccessful (Entries 9 and 10, Table 1).

Next, the same conditions for protection and selective 5'-deprotection were applied to gemcitabine (**2**) to form the key intermediate **6**. However, instead of 3'-TBDMS-protected gemcitabine **6**, formation of its 5'-TBDMS-protected analogue was

observed along with the starting bis-silylated compound **4**. While an attempt to selectively remove a silyl group from the 5'-position with oxalic acid (up to 3.0 eq.) was found unsuccessful (data not reported), the use of trichloroacetic acid (28 eq.) in a mixture of THF/H₂O (3.5:1) gave the key intermediate **6** with 82% yield (Entry 12, Table 1).

Scheme 1. General synthetic strategy towards 3'-TBDMS-protected nucleosides **5** and

6.



Reagents and conditions: (a) imidazole (5.0 eq.), TBDMSCl (2.2 eq), DMAP (0.3 eq), DMF, 0°C to rt, 1h; b) CSA (1.0 eq.), MeOH, rt, 10 min. or TCA (28.0 eq.), THF, H₂O, rt, 2h.

Table 1. Selective 5'-desilylation of 3',5'-TBDMS-FUDR (**3**) and 3',5'-TBDMS-gemcitabine (**4**): condition optimisation

Entry	Nucleoside Analogue	Acid/eq.	Solvent	Time/Temp	Yield**** (%)
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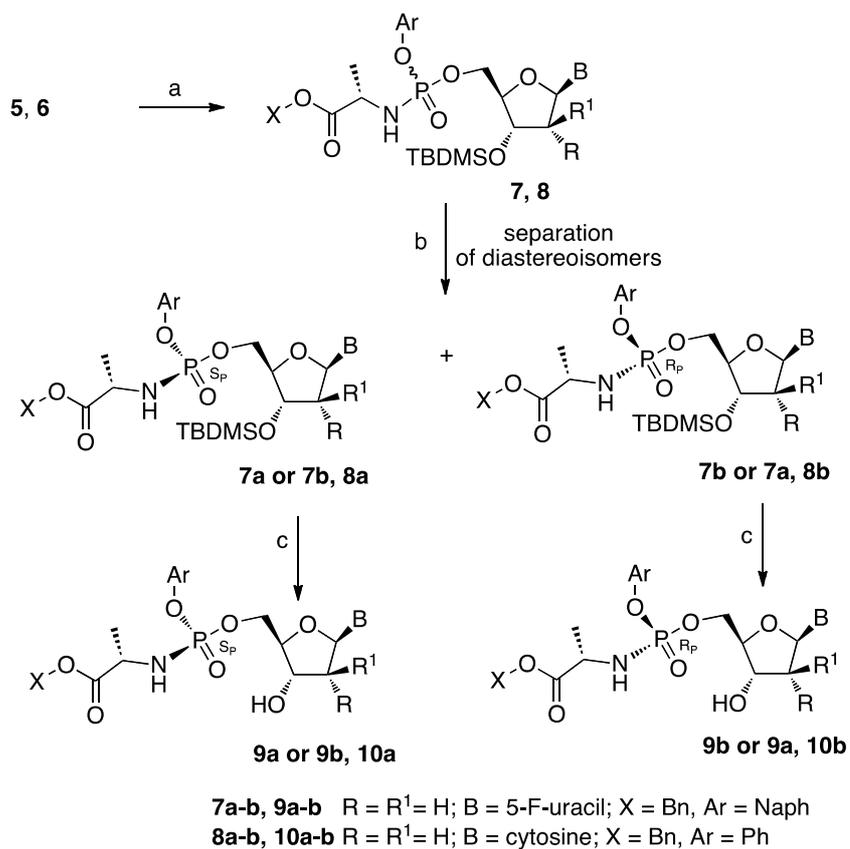
1	FUDR	<i>p</i> -TsOH/0.6	MeOH	1h/rt	-
2	FUDR	<i>p</i> -TsOH/1.0	MeOH	4h/-15°C	45*
3	FUDR	<i>p</i> -TsOH/1.0	DCM	6h/15°C - rt	-
4	FUDR	CSA/0.6	MeOH	30min/0°C	56*
5	FUDR	CSA/0.6	MeOH	1h/0°C	60*
6	FUDR	CSA/0.5	MeOH	25min/rt	50**
7	FUDR	CSA/0.6	MeOH	45min/rt	-
8	FUDR	CSA/1.0	MeOH	10min/rt	60**
9	FUDR	CSA/1.0	AcCN	1h/rt	-
10	FUDR	CSA/1.0	Acetone	10min/rt	-
11	Gemcitabine	CSA/1.0	MeOH	10min/rt	-
12	Gemcitabine	TCA/28.0	THF/H ₂ O	30min/rt	82**

*5'-Desilylated product, calculated yield by ¹H and ¹⁹F NMR, **isolated yield

The key intermediate **5** was further subjected to the standard coupling conditions with L-Ala-OBn-ONaph phosphorochloridate¹⁰ in the presence of the Grignard reagent (Scheme 2). 3'-TBDMS-protected compound **7** obtained as a mixture of diastereomers was purified using normal-phase chromatography to give single isomers: fast eluting (**7_isomer A**, **7a** or **7b**) and slow eluting (**7_isomer B**, **7a** or **7b**). A final deprotection of the 3'-silyl group in isomers **7a** and **7b** was performed with a 4:1:1 mixture of THF/TFA/H₂O at 0°C and yielded single diastereomers of NUC-3373, herein reported as NUC-3373_isoA (**9a** or **9b**) (obtained from **7_isoA**) and NUC-3373_isoB (**9a** or **9b**) (obtained from **7_isoB**) as stereochemistry at the phosphorus atom remained undetermined at the time of preparation of this manuscript. Attempts to crystallise NUC-3373_isoA (**9a** or **9b**) and NUC-3373_isoB (**9a** or **9b**), as well as their 3'-protected precursors (**7_isomer A** and **7_isomer B**)

from various solvents including MeOH, 2-propanol, CHCl₃ and a combination of CHCl₃/Et₂O (1:1), and MeOH/CHCl₃ (3:1), CHCl₃/H₂O (1:1) were unsuccessful. In addition, the vapour diffusion method, using 2-propanol/hexane, MeOH/H₂O and MeOH/petroleum ether as the binary solvent system, did not lead to formation of crystalline form. In parallel, a coupling reaction between L-Ala-OBn-OPh phosphorochloridate⁹ and **6** gave 3'-TBDMS-protected NUC-1031 analogue **8**, further isolated as single isomers **8a** and **8b** via column chromatography. Removal of the 3'-TBDMS-group in separated diastereomers **8a** and **8b** under acidic conditions provided isomers of NUC-1031 **10a** and **10b** (Scheme 2), which after normal-phase chromatography were obtained as crystalline solid. The molecular structure and stereochemistry at the phosphate centre was elucidated by single-crystal X-ray crystallography for the two isomers of NUC-1031 and determined to be *S_P* and *R_P* for diastereomers **10a** and **10b**, respectively.

Scheme 2. General synthetic strategy towards separated diastereomers of **NUC-3373** and **NUC-1031 (9a,b and 10a,b)**.



Reagents and conditions: (a) (ArO)(XOC(O)CHCH₃NH)P(O)Cl (2.0 eq), *tert*-BuMgCl, THF, rt, 16h;
 b) separation of diastereomers; c) THF/TFA/DCM (4:1:1), 0 °C for 2h, then rt for 12h.

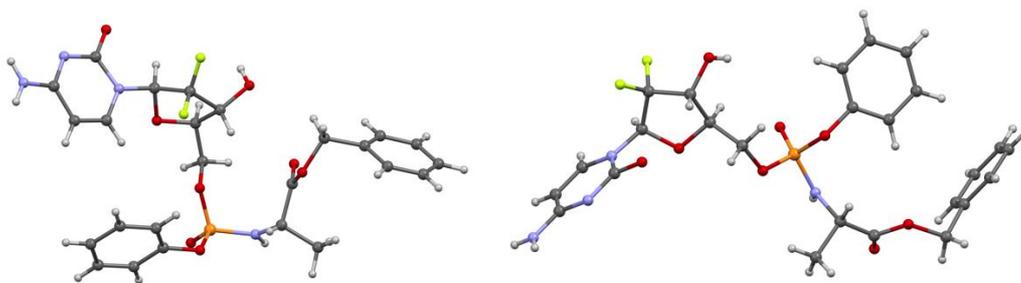


Figure 2: 10a (NUC-1031_{Sp}) and 10b (NUC-1031_{Rp})

In vitro cytotoxic activity

The single isomers of NUC-1031 and NUC-3373 were biologically evaluated for cytotoxic activity in a panel of solid tumour cell lines (Mia-Pa-Ca-2, Bx-PC-3, LNCaP, HT-29, HCT-116, MDA-MB-231 and SW-620), the acute myeloid leukaemia (AML) cell line, KG1a and primary AML blasts. The activity of the ProTides was compared with the corresponding diastereomeric mixtures NUC-3373 and NUC-1031, as well as with the parent nucleosides FUDR (**1**) and gemcitabine (**2**), respectively (Table 2). In the solid tumour cell lines, the LC₅₀ values for both NUC-3373 isomers were comparable to those of NUC-3373 and FUDR and ranged between 0.61–2.37 μM and 0.31–1.46μM in HT-29 and SW-620 for NUC-3373_IsoA (**9a** or **9b**) and NUC-3373_IsoB (**9a** or **9b**), respectively. In comparison to 5-FU, a significant increase in potency was observed and ranged between 13-25-fold and 3-5-fold for NUC-3373_IsoA (**9a** or **9b**) and NUC-3373_IsoB (**9a** or **9b**), respectively in HT-29 and SW-620 cell lines. The cytotoxic activity of the gemcitabine ProTide NUC-1031 and its isomers were similar, except for the NUC-1031_*S*_P isomer (**10a**) in the Mia-Pa-Ca-2 cell line, which was found to be 6-fold more potent than its *R*_P analogue **10b** (0.03μM vs 0.19μM) and 3-fold more potent than its diastereomeric mixture (0.03 μM vs 0.10 μM). In the HT-29 cancer cell line, the sub-micromolar LC₅₀ values for both *S*_P and *R*_P NUC-1031 isomers and NUC-1031 ranged between 0.34-0.74μM, similar to the gemcitabine LC₅₀ value of 0.21μM. NUC-1031 and both isomers were equipotent in the Bx-PC-3 tumour cell line. When considering the KG1a cell line, NUC-1031 was approximately 6-fold more potent than gemcitabine, again, with no significant difference between the diastereomers (Figure 3A-C). We went on to confirm that NUC-1031 was significantly more potent than gemcitabine in primary AML blasts (Figure 3G-H). Similarly, NUC-3373 was approximately 23-fold more potent than FUDR with no significant difference between the diastereomers (Figure 3D-F).

Table 2. Cytotoxicity of NUC-3373 and NUC-1031 as a diastereomeric mixture, and single isomers *R_P* and *S_P* reported as LC₅₀ (μM) values.

Compound	LC ₅₀ (μM) ±SEM in different cell lines							
	Mia-Pa-Ca-2*	Bx-PC-3*	LNCaP	HT-29	HCT-116	SW-620*	MDA-MB-231	KG1a
5-FU	-	-		5.5 ±0.8	3.7 ±0.7	7.98	12.6 ±1.1	1.4±0.5
FUDR	-	-		0.8±0.3	1.6±0.4	1.55	2.4±0.7	1.18±0.4
NUC-3373	-	-		1.7 ±0.6	3.1±0.8	1.84	2.8±0.9	0.053±0.02
NUC-3373_IsoA (9a or 9b)	-	-		2.1±0.6	4.3±1.0	2.37	3.6±0.6	0.052±0.01
NUC-3373_IsoB (9a or 9b)	-	-		1.5 ±0.4	2.6±0.9	1.46	2.5±0.6	0.055±0.02
Gemcitabine	0.01	0.74	2.8±0.5	0.7±0.2		-		1.8±0.3
NUC-1031	0.1	2.29	3.1±0.6	1.3±0.5		-		0.28±0.07
NUC-1031_ <i>S_P</i> (10a)	0.03	2.44	2.6 ±0.7	1.5±0.3		-		0.31±0.1
NUC-1031_ <i>R_P</i> (10b)	0.19	1.29	3.4±0.5	1.0±0.4		-		0.25±0.1

LC₅₀ Values (the concentration of drug required to induce 50% cell death) were calculated using *an MTS assay performed in duplicate and tested in 9 serial concentrations from 198 μM to 0.0199 μM (data expressed as mean LC₅₀ values) or an Annexin V /7-AAD assay performed in triplicate (data expressed as mean LC₅₀ values±SEM).

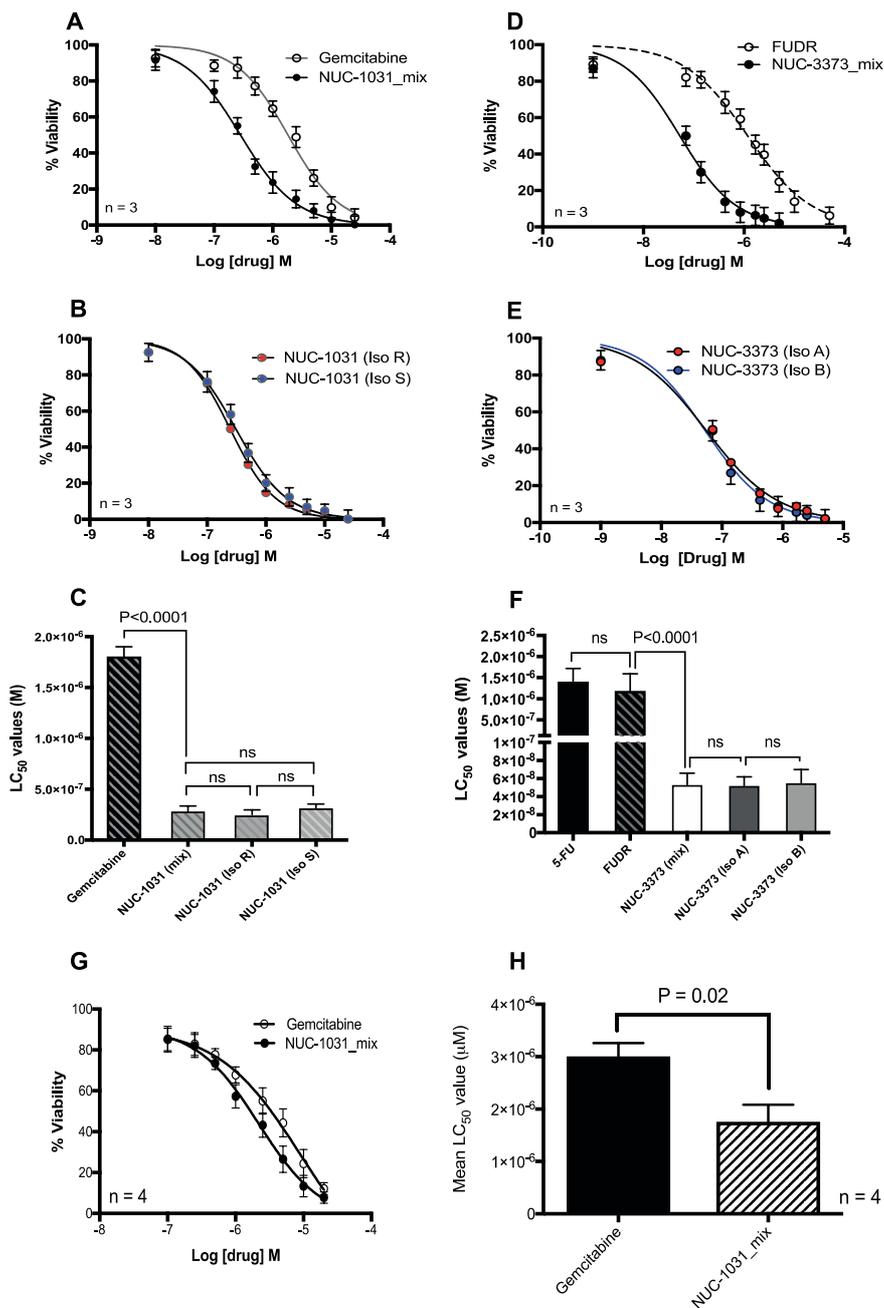


Figure 3. Overlaid sigmoidal dose-response curves. (A) Comparison of gemcitabine and NUC-1031_mix (diastereomeric mixture) and (B) the separated diastereomers, NUC-1031_{SP} (10a) NUC-1031_{RP} (10b). (C) Shows the comparison of the LD₅₀ values for gemcitabine, NUC-1031_mix, NUC-1031_{SP} (10a) NUC-1031_{RP} (10b). (D) Comparison of FUDR and NUC-3373_mix (diastereomeric mixture) and (E) the separated diastereomers NUC-3373_{isoA} (9a or 9b) and NUC-3373_{isoB} (9a or 9b). (F) Shows the comparison of the LD₅₀ values for 5-FU, FUDR, NUC-3373_mix, NUC-3373_{isoA} (9a or 9b) and NUC-3373_{isoB} (9a or 9b). All assays carried out using KG1a cells are presented as mean (±SEM) of five independent experiments. (G-H) Relative sensitivity of primary AML blasts to gemcitabine and NUC-1031_mix (n=4).

The metabolic stability of NUC-3373, NUC-1031 and the separated diastereomers (**9a**, **9b** and **10a,10b**) was investigated by incubation with human hepatocytes. The half-lives and percentage of compounds remaining after 1h were measured and are outlined in Table 3. In human hepatocytes, NUC-3373_IsoA (**9a** or **9b**) was found to be slightly more stable than NUC-3373_IsoB (**9a** or **9b**) and its diastereomeric mixture NUC-3373, with a half-life of 53 minutes. In the case of the gemcitabine family, NUC-1031_*S_P* (**10a**) was found to have a shorter half-life of 37 minutes in human hepatocytes compared with NUC-1031_*R_P* (**10b**), which had a half-life >120 min. This intriguing discrepancy in half-life of both NUC-1031 isomers in human hepatocytes could potentially be explained by different carboxylesterases (CESs) substrate specificity also towards isomers. Mammalian carboxylesterases (CES1 and CES2), the crucial enzymes involved in the metabolism of endogenous esters and ester or amide-containing xenobiotics, may exhibit different hydrolysis rate between the R and S-isomers. Thus, their reactivity might be affected by the conformational orientation into the active pocket of CESs.³⁹

Table 3. Metabolic stability of ProTides incubated with human hepatocytes.

Intrinsic clearance (cryopreserved human hepatocytes)		
Compound	Half-Life (min)	Compound remain % after 1h
NUC-3373	34	30
NUC-3373_IsoA (9a or 9b)	53	40
NUC-3373_IsoB (9a or 9b)	47	40
NUC-1031	76	58
NUC-1031_<i>S_P</i> (10a)	37	33
NUC-1031_<i>R_P</i> (10b)	>120	97

Biological activation: Carboxypeptidase Y Assay

The mechanism of activation of NUC-1031 and NUC-3373 in their diastereomeric forms was previously reported^{9,10} and was confirmed to follow the commonly accepted putative route (Scheme 3). The activation pathway involves (i) carboxypeptidase-mediated hydrolysis of the ProTide ester moiety to form the intermediate **A**, (ii) displacement of the aryl moiety and spontaneous cyclization of **A** to form metabolite **B**, and (iii) spontaneous hydrolysis of cyclic anhydride **B** to the intermediate **C**. The latter metabolite would then be converted to the monophosphate **D** via P-N bond cleavage catalysed by a phosphoramidase-type enzyme. In this work, we carried out an enzymatic assay for the two separated isomers NUC-3373_isoA (**9a** or **9b**) and NUC-3373_isoB (**9a** or **9b**) and for the gemcitabine ProTide NUC-1031_S_P (**10a**) and NUC-1031_R_P (**10b**) in the presence of carboxypeptidase Y according to the standard procedure.⁴⁰ Within 10 minutes of incubation (Figure 4a), NUC-3373_isoA (**9a** or **9b**) was fully processed to metabolite **A**, as indicated by lack of the single peak at δ 4.04 ppm ascribed to the NUC-3373_isoA (**9a** or **9b**), and appearance of a single peak at δ 5.16 ppm (metabolite **A**). During this time, a single peak at δ 6.80 ppm corresponding to achiral metabolite **C** was also detected. Slightly slower processing to metabolites **A** and **C** was observed for the isomer NUC-3373_isoB (**9a** or **9b**) within the first 10 minutes of incubation, during which 28% of NUC-3383_isoB remained unprocessed by the enzyme (Supporting Information, Figure 4b).

In the case of the gemcitabine ProTide NUC-1031, only S_P isomer (**10a**) was processed to its metabolites **A** (δ 4.51 ppm, 68%) and **C** (δ 6.80 ppm, 18%) within the first 10 minutes of incubation with carboxypeptidase Y. NUC-1031_R_P (**10b**) was

converted to metabolite **C** (without metabolite **A** detection) within approximately 120 minutes (Supportive Information, Figures 4c,d).

Scheme 3. The general metabolic activation pathway for NUC-3373 and NUC-1031 ProTides.

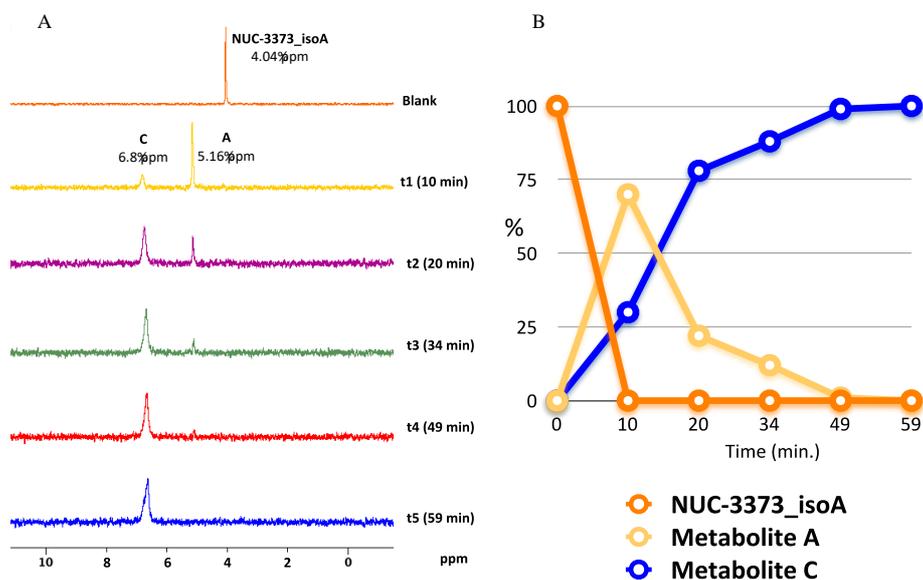
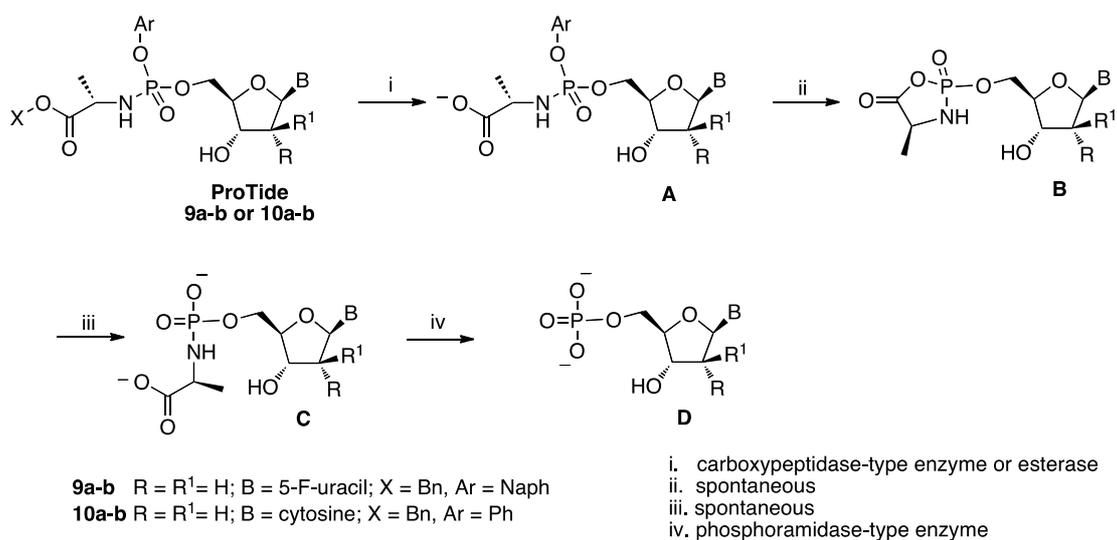


Figure 4a. (A) ³¹P NMR spectra of NUC-3373_isoA (**9a** or **9b**) over time after enzymatic incubation with carboxypeptidase Y. (B) Chart representation of percentage of NUC-3373_isoA (**9a** or **9b**), metabolites **A** and **C** over time after enzymatic incubation with carboxypeptidase Y. Conditions 202 MHz, 25°C, *d*₆-acetone, 0.05 M Trizma buffer, pH 7.6

Cancer stem cell targeting

The putative existence of cancer stem cells (CSCs) has been suggested in many human cancers, including leukaemias and solid tumours. The cancer stem cell hypothesis postulates that only a small sub-population of tumour cells are responsible for tumour formation, maintenance of the bulk of the tumour, as well as its invasion and metastasis. In 1994, Lapidot and co-workers demonstrated that only a small percentage of acute myeloid leukaemia cells had the capability to initiate leukaemia in mice.⁴¹ These leukaemic stem cells (LSCs) were shown to express similar cell surface markers (CD34⁺/CD38⁻) to normal haematopoietic stem cells implying that a similar hierarchical organisation may occur in tumours. Subsequently, CSCs have been found in a wide range of solid tumours including breast, lung, colon, prostate, ovarian, skin, and pancreas.⁴² CSCs are also linked with the development of resistance to therapy and recurrence of cancer, as reported for human colon cancer cell resistance to 5-FU *in vitro*⁴³ and for human pancreatic adenocarcinoma resistance to gemcitabine.⁴⁴

Preferential targeting of cancer stem cells by ProTides

Given the growing interest in the treatment of cancer by targeting tumour stem cells, we assessed the relative abilities of NUC-1031_mix and NUC-3373_mix, their single diastereomers and the parental nucleoside analogues to preferentially kill LSCs (as defined by the phenotype CD34⁺CD38⁻) in the acute myeloid leukaemia cell line, KG1a, and in primary AML blasts.⁴⁵ Figures 5A and 5B show that gemcitabine did not significantly alter the percentage of LSCs in KG1a cell cultures at concentrations $\leq 2.5 \times 10^{-6}$ M. In contrast, the NUC-1031 diastereomers significantly reduced the LSC fraction at concentrations $\geq 2.5 \times 10^{-7}$ M; both diastereomers had a similar effect (Figure 5B). Importantly, we were also able to demonstrate that NUC-1031_mix

preferentially depleted LSCs in primary AML blasts (Figure 5C). Furthermore, consistent with the results generated with gemcitabine, neither 5-FU nor FUDR showed any preferential toxic effect on LSCs when compared to the bulk tumour at concentrations $\leq 5 \times 10^{-6} \text{M}$ (Figure 5D). In contrast, both NUC-3373 diastereomers preferentially depleted the LSC compartment at concentrations $\geq 7 \times 10^{-8} \text{M}$ (Figure 5D). In order to establish whether this phenomenon was specific for KG1a cells and AML blasts, the prostate cancer cell line, LNCaP was employed as an alternative cancer stem cell model. In this case, the cancer stem cells were identified by a $\text{CD44}^+\text{CD133}^+$ phenotype, this population of cells accounted for just 1.4% of the cell line. Although the LC_{50} values for gemcitabine and NUC-1031 were not significantly different ($P = 0.24$), NUC-1031 showed a preferential effect on cancer stem cell population when compared to gemcitabine at concentrations $\geq 2.5 \times 10^{-6} \text{M}$ (Figure 5E).

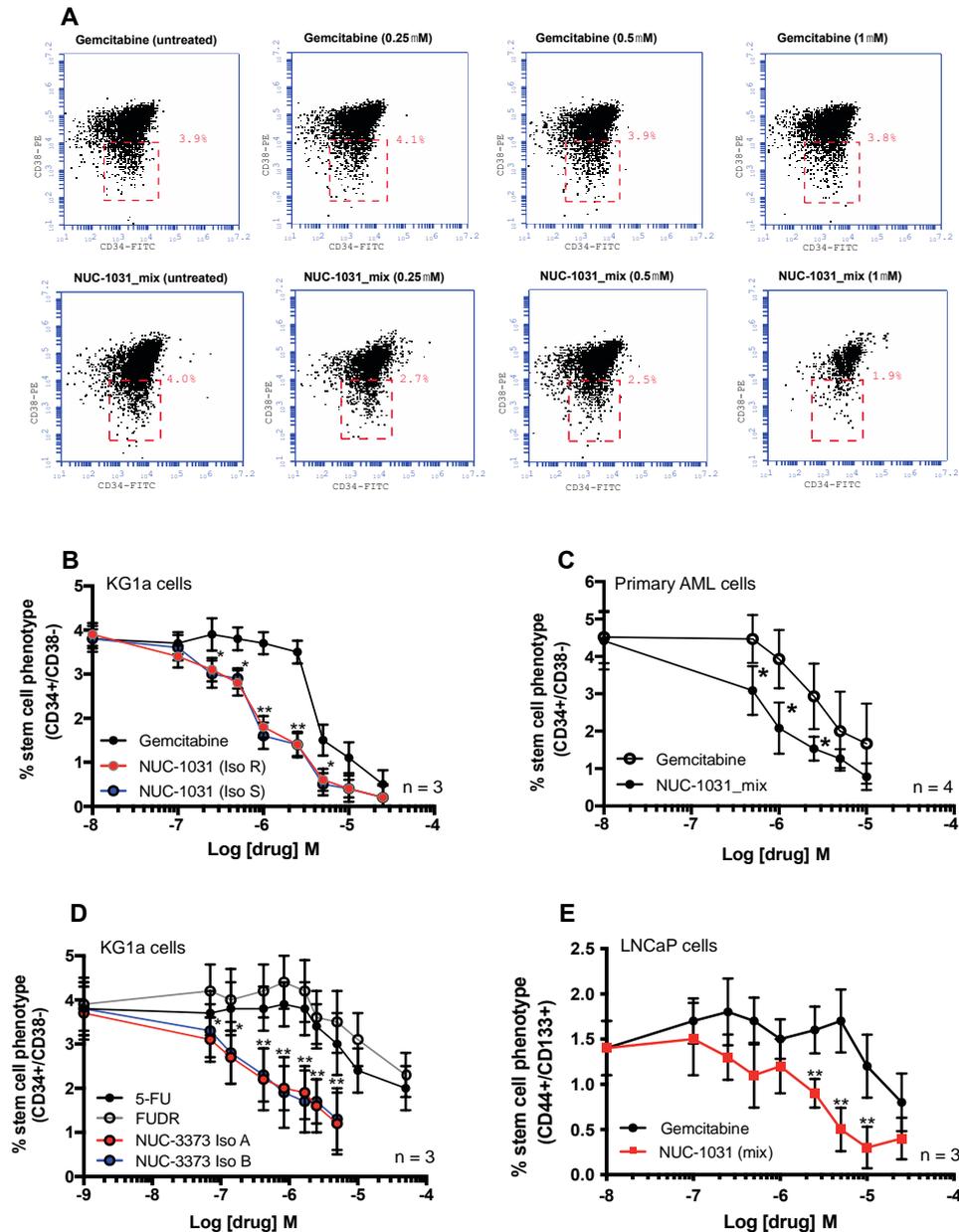


Figure 5. Differential cytotoxic effects of nucleosides and ProTides on cancer stem cells. (A) Comparison of the viable fraction of KG1a cells exposed to gemcitabine and NUC-1031_mix for 48h. (B) Both diastereomers of NUC-1031 caused a dose-dependent, statistically significant, reduction in the LSC (CD34⁺/CD38⁻) fraction of KG1a cells. In contrast, gemcitabine had no significant effect on the LSC compartment at concentrations $\leq 2.5 \times 10^{-6}$ M. (C) The diastereomeric mixture of NUC-1031 induced a dose-dependent decrease in the LSC fraction of primary AML blasts. (D) Both diastereomers of NUC-3373 caused a dose-dependent, statistically significant, reduction in the LSC (CD34⁺/CD38⁻) fraction of KG1a cells. In contrast, 5-FU and FUDR had not significant effect on the LSC compartment at concentrations $\leq 5 \times 10^{-6}$ M. * $P < 0.05$, ** $P < 0.01$. (E) A similar effect was observed in the prostate cancer cell line, LNCaP. NUC-1031 preferentially depleted the cancer stem cells (CD44⁺/CD133⁺) at concentrations $\geq 2.5 \times 10^{-6}$ M when compared with gemcitabine.

Intracellular triphosphate accumulation and ABC transporter expression

LSC and bulk tumour fractions (5×10^6) were isolated by high-speed cell sorting and were subsequently cultured for 1h with either $1 \mu\text{M}$ gemcitabine or $1 \mu\text{M}$ NUC-1031_mix. The level of intracellular triphosphate was assessed using mass spectrometry and quantify by running defined concentrations of nucleoside through the mass spectrometer in order to construct a standard curve (Figure 6A). In both fractions, NUC-1031 resulted in higher intracellular accumulation of the triphosphate when compared to the parental gemcitabine nucleoside (Figure 6B). Furthermore, the LSC fraction retained high levels of the triphosphate following treatment with NUC-1031_mix, which may contribute to the preferential depletion of these cells when exposed to ProTide. In contrast, LSCs treated with gemcitabine showed approximately a 30% reduction in intracellular triphosphate when compared with bulk tumour cells. Based on the intracellular triphosphate data, we next evaluated the expression of ABC transporters in LSC and bulk tumour fractions both at the level of protein and mRNA transcript. Figure 6C shows that LSCs show a trend towards higher expression of ABCB1 and significantly higher expression of ABCG2 in LSCs versus bulk tumour cells. Furthermore, at the level of transcript ABCB1, ABCG2, ABCC4 and ABCC5 all showed significantly increased expression in LSCs versus bulk tumour cells (Figure 6D). A number of these transporters have been previously implicated in gemcitabine resistance.^{46,47}

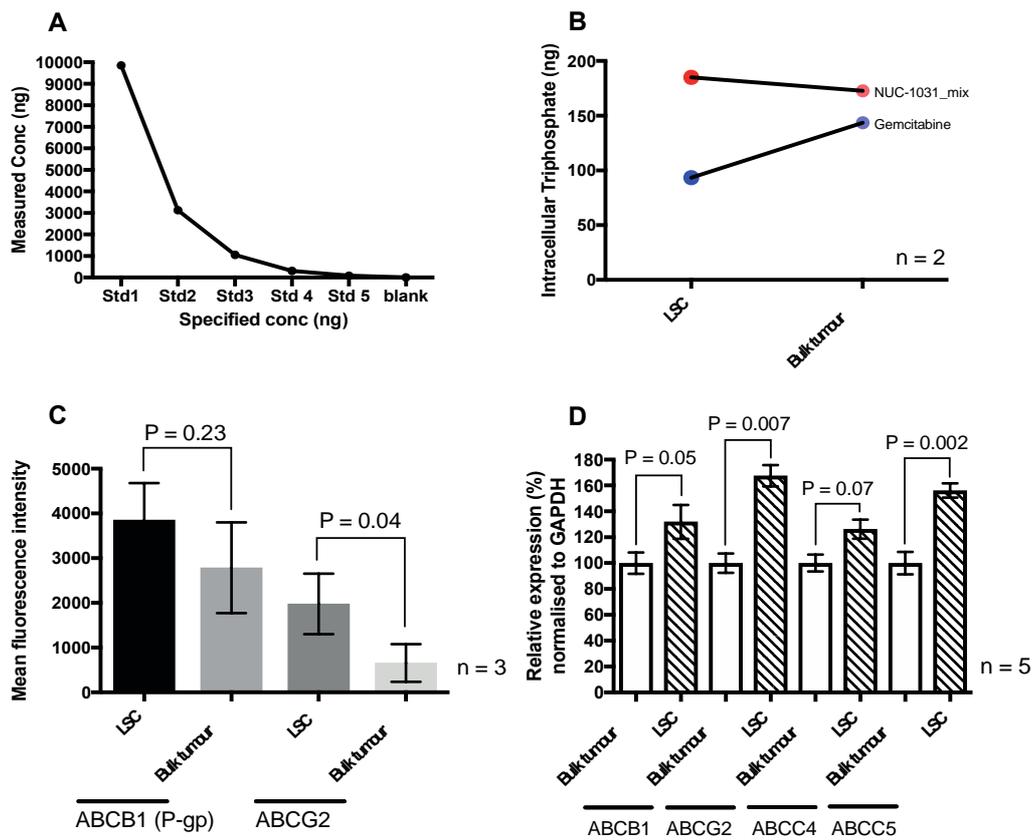


Figure 6. Measurement of intracellular triphosphate ABC transporters in LSC and bulk tumour cell fractions from KG1a cells. (A) Defined concentrations of nucleoside were run through the mass spectrometer in order to construct a standard curve (B) Purified LSCs and bulk tumour cells were treated with 1 μ M of gemcitabine or NUC-1031_mix for 1h prior to cell lysis and mass spectrometry. Both LSC and bulk tumour fractions showed increased intracellular triphosphate following treatment with NUC-1031_mix. (C) Protein expression of ABCB1 and ABCG2 on purified bulk tumour (CD34⁺/CD38⁺) and LSCs (CD34⁺/CD38⁻). Cell sorted LSCs showed significantly higher levels of ABCG2 when compared with bulk tumour cells. (D) Relative gene transcription of ABC transporter genes in purified bulk tumour cells (solid bars) and LSCs (hatched bars). LSCs showed significantly increased relative transcription of ABCB1, ABCG2 and ABCC5 when compared with bulk tumour cells. All data are the mean (\pm SEM) of five independent experiments, performed in triplicate.

CONCLUSION

The biological potencies and activities of single enantiomers or diastereomers of chiral compounds such as phosphoramidates (ProTides), with their stereocentre at the phosphorus atom, may differ markedly. In light of the FDA recommendation to assess activity of each isomer of racemic or diastereomeric mixtures, it became even more important to develop either stereoselective synthetic strategies or separation methods to obtain the desired isomers. Herein, we described an alternative synthesis of the two clinical anti-cancer agents NUC-1031 and NUC-3373 via a 3',5'-hydroxyl group protection and 5'-deprotection strategy, which allowed separation of the single S_P and R_P diastereomers. Two diastereomers of NUC-1031 were obtained in crystalline form and their molecular structure and stereochemistry at the phosphate centre was assigned via X-ray crystallography. The cytotoxic activities of R_P and S_P isomers of NUC-1031 were similar and there was no significant difference in the *in vitro* potency when compared to NUC-1031 as a diastereomeric mixture. Similarly, the *in vitro* potency of NUC-3373 and its single isomers NUC-3373_isoA and NUC-3373_isoB was comparable in the cancer cell lines tested. Attempts to crystallise two diastereoisomers of NUC-3373 were unsuccessful at the time of preparation of this manuscript. Both NUC-1031_mix and NUC-3373_mix and their separated diastereoisomers were shown to preferentially target leukaemic cancer stem cells, whereas the parent nucleosides gemcitabine and FUDR did not. Mechanistically, increased intracellular accumulation of triphosphate may contribute to the preferential killing of CSCs by the ProTide in contrast to the parental analogue but other factors may also be involved. Given that one of the gemcitabine resistance mechanisms has been associated with ABC transporter expression,^{46,47} we investigated their expression in LSC and bulk tumour cells. LSCs showed significantly increased expression of

ABCB1, ABCG2, ABCC4 and ABCC5 at the level of transcript and increased expression of ABCG2 at the level of protein. We postulate that the increased intracellular triphosphate in ProTide-treated cells is caused, at least in part, by the differential bioavailability of the ProTide versus the parental nucleoside analogue. As LSCs preferentially express ABC transporters, the increased intracellular accumulation of triphosphate in these cells, following treatment with ProTide, results in increased cell death of these LSCs. Importantly, the same effect was observed in cancer stem cells from a prostate cancer cell line (LNCaP). As such, ProTides may prove to be an important addition to the therapeutic repertoire, particularly in the context of eradication of cancer stem cells.

EXPERIMENTAL SECTION.

Materials and methods

Cell lines. KG1a, HT-29, HCT-116, LNCaP and MDA-MB-231 cell lines were all purchased from DSMZ (Germany). All lines were initially expanded and then frozen down in aliquots (10^7 cells) at low passage number. Cell lines were never expanded beyond 8-10 passages and were routinely confirmed as mycoplasma-free to ensure the integrity of each cell line.

MTS Cell Viability Assay. The assay was contracted and carried-out by WuXi AppTec (Shanghai) Co., Ltd. The tumour cell lines Mia-PaCa-2, Bc-PC3, HT-29 and SW620 were seeded at cell densities of 0.5 to 100×10^3 cells/well in a 96-well plate the day before drug incubation. Then the plates were incubated for 72 hours with the different concentrations of compound to be tested. After the incubation period, 50 μ L of MTS was added and the tumour cells were incubated for 4 h at 37°C. The data were read and collected by a Spectra Max 340 Absorbance Microplate Reader. The

compounds were tested in duplicate with 9 serial concentrations (3.16-fold titrations with 198 μM as the highest concentration), and the data were analysed by XL-fit software.

Annexin V/7-AAD cell viability assay. Cell lines were grown in T175 flasks until confluent. Cells were then aliquoted (10^5 cells/ $100\mu\text{L}$) into 96-well plates and were incubated at 37°C in a humidified 5% CO_2 atmosphere for 72 h in the presence of tested compounds at the concentrations that were experimentally determined for each compound. In addition, control cultures were set up to which no drug was added. Cells were subsequently harvested by centrifugation and were analysed by flow cytometry using the Annexin V/7-AAD assay. All experiments were performed in triplicate. LC_{50} values (the concentration of compound required to kill 50% of the cells in culture) were calculated by non-linear regression modelling using Graphpad Prism software and are shown as mean \pm SEM for each replicate dataset.

Metabolic Stability (cryopreserved hepatocytes, human) Assay. The assay was contracted and performed by Cerep (Seattle, WA, USA Laboratories, 15318 N.E 95th Street Redmond, WA, 98052, USA) according to the published procedure (Cerep Ref. 1432). Pooled cryopreserved hepatocytes were thawed, washed and re-suspended in Krebs-Heinslet buffer (pH 7.3). The reaction was initiated by adding the test compound (1 μM final concentration) into cell suspension and incubated in a final volume of 100 μL on a flat-bottom 96-well plate for 0 minute and 60 minutes, respectively, at $37^\circ\text{C}/5\%\text{CO}_2$. The reaction was stopped by adding 100 μL of acetonitrile into the incubation mixture. Samples were then mixed gently and briefly on a plate shaker, transferred completely to a 0.8 mL V-bottom 96-well plate, and centrifuged at 2550 $\times g$ for 15 minutes at room temperature. Each supernatant (150

μL) was transferred to a clean cluster tube, followed by HPLC-MS/MS analysis on a Thermo Electron triplequadrupole system.

Carboxypeptidase Y (EC 3.4.16.1) Assay. The experiment was carried out by dissolving ProTides (3.5 mg) in acetone- d_6 (0.15 mL) followed by addition of 0.30 mL of Trizma buffer (pH 7.6). After recording the control ^{31}P NMR at 25°C, a previously defrosted carboxypeptidase Y (0.1 mg dissolved in 0.15 mL of Trizma) was added to the sample, which was then immediately submitted to the ^{31}P NMR experiments (at 25°C). The spectra were recorded with 64 scans every 7 minutes unless otherwise stated. ^{31}P NMR recorded data were processed and analysed with the Bruker Topspin 2.1 program.

Cell culture conditions. The acute myeloid leukaemia (AML) KG1a cell line and the prostate cancer cell line, LNCaP, were maintained in RPMI medium (Invitrogen, Paisley, UK) supplemented with 100 units/mL penicillin, 100 $\mu\text{g}/\text{mL}$ streptomycin and 20% foetal calf serum. The colorectal cancer cell lines, HT-29 and HCT-116 and the breast cancer cell line, MDA-MB-231 were maintained in Dulbecco's modified essential medium with high glucose supplemented with 100 units/mL penicillin, 100 $\mu\text{g}/\text{mL}$ streptomycin and 10% foetal calf serum.

Measurement of in vitro apoptosis. Cultured cells were harvested by centrifugation and resuspended in 195 μL of calcium-rich buffer. Subsequently, 5 μL of Annexin V was added to the cell suspension and cells were incubated in the dark for 10 min prior to washing. Cells were finally resuspended in 190 μL of calcium-rich buffer together with 10 μL of propidium iodide. Apoptosis was assessed by dual-colour immunofluorescent flow cytometry⁴⁸ as described previously. Subsequently LC_{50} values (the doses required to kill 50% of the cells in culture) were calculated for each nucleoside analogue and ProTide.

Immunophenotypic identification of the cancer stem cells. KG1a cells or LNCaP cells were cultured for 72 h in the presence of a wide range of concentrations of each nucleoside analogue and their respective ProTides. Cells were then harvested and KG1a cells were labelled with anti-CD34 (FITC) and anti-CD38 (PE) and LNCaP cells were labelled with anti-CD44 (APC) and anti-CD133 (Alexa488). The sub-populations expressing a cancer stem cell (CSC) phenotype were subsequently identified and were expressed as a percentage of all viable cells left in the culture. The percentages of stem cells remaining were then plotted on a dose-response graph and the proportion of cancer stem cells was compared across the various concentrations of ProTides and their respective parental nucleoside in order to determine if they preferentially target cancer stem cells.

Cell sorting. KG1a cells were cultured until confluent in order to generate 10^8 cells. Subsequently, 5×10^6 of CD34⁺/CD38⁻ LSCs and CD34⁺/CD38⁺ bulk tumour cells were purified by high-speed cell sorting using a FACS Melody cell sorter (Becton Dickenson) and were placed back into culture prior to the addition of 1 μ M gemcitabine or the ProTide NUC-1031_mix.

Mass spectrometry assessment of intracellular nucleoside/ProTide. Defined concentrations of nucleoside were run through the mass spectrometer in order to construct a standard curve (Figure 6A). This was used to quantify the levels of nucleoside/ProTide triphosphate in each of the samples provided. KG1a cells (5×10^6), cultured in DMEM + 10% FCS, were exposed to 1 μ M of gemcitabine or NUC-1031_mix for 1h prior to harvesting by centrifugation. Cells were then FACS sorted into LSC (CD34⁺/CD38⁻) and bulk tumour cell fractions (CD34⁺/CD38⁺). Purified cells were then pelleted prior to lysis and mass spectrometry analysis for intracellular triphosphate content of gemcitabine or NUC-1031.

Immunopheotypic expression of ABCB1 and ABCG2. KG1a cells were harvested from culture and labelled with anti-CD34 (FITC) and anti-CD38 (PE) to identify the LSC and bulk tumour fractions. In addition, they were labelled with anti-ABCB1 (Alexa 647) and anti-ABCG2 (APC). Subsequently, the LSC and bulk tumour fractions were gated and the expression levels of the two ABC transporters were quantified using mean fluorescence intensity values.

Transcription of ABC transporter genes. 1mL TRIzol was added to each pellet of 2×10^5 harvested bulk tumour or LSC cells. 200 μ L of chloroform was added and centrifuged at 8,000xg for 15 minutes at 4°C. The aqueous layer was gently removed and added to an equal volume of 70% ethanol. 700 μ L of the sample was then added to an RNeasy spin column and the RNeasy Mini Kit protocol was followed. The elution step used 50 μ L RNase-free water and RNA quantification was assessed using a Nanodrop:1000 Spectrophotometer. Duplicate 1 μ L samples were evaluated and the RNA concentration, A260/280 ratio and A260/A230 ratio obtained. Aliquots of 0.5 μ g of RNA were then added to 0.5 mL microfuge tubes containing 10 μ L master mix and made up to 20 μ L with sterile RNase-free water. Samples were added to thermal cycler and run as per the kit's protocol. Once the run was complete, 20 μ L of newly converted cDNA was diluted 5:1 with sterile DNA and RNase-free water to give a working concentration of 2.5 ng/ μ L. Samples were stored at -20°C. The TaqMan Fast Advanced Master Mix kit protocol was followed. Samples were run in triplicate, including a minus RT sample and a minus cDNA sample. Samples were run on a 96-well plate with a running volume 10 μ L per well. This was composed of 2.5 μ L of thawed cDNA, 0.5 μ L of forward and reverse primers (ABC transporter gene or GAPDH as a housekeeping control; see Table 4, Supportive Information), 5 μ L TaqMan master mix and 2 μ L of RNase-free water. Samples were run on a ViiA7

Real-Time PCR system using the TaqMan FAST programme. Results were analysed using the Thermo Fisher Cloud software using the Relative Quantification qPCR application.

Statistical analysis. The data obtained in these experiments were evaluated using one-way ANOVA. All data was confirmed as Gaussian or a Gaussian approximation using the omnibus K2 test. LC₅₀ values were calculated from the non-linear regression and line of best-fit analysis of the sigmoidal dose-response curves. Values are shown as mean \pm SEM from each replicate dataset. All statistical analyses were performed using Graphpad Prism 6.0 software (Graphpad Software Inc., San Diego, CA).

Crystal structure determination. Single-crystal XRD data were collected on an Agilent SupaNova Dual Atlas diffractometer with a mirror monochromator using Cu ($\lambda = 1.5418$ Å) XRD data were collected on an Agilent cooling apparatus. Crystal structures were solved and refined using SHELX (see Table 5, Supportive Information).^{49,50} Geometric restraints were applied during refinement of orientationally disordered components. Non-hydrogen atoms were refined with anisotropic displacement parameters. Hydrogen atoms were inserted in idealized positions, and a riding model was used. CCDC 2049153 and 2049154 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/structures.

General methods. All solvents and reagents were used as obtained from commercial sources unless otherwise indicated. All reactions were performed under a nitrogen atmosphere. The ¹H and ¹³C NMR spectra were recorded on a Varian spectrometer operating at 500 MHz for ¹H and 125 MHz for ¹³C. Deuterated chloroform was used as the solvent for NMR experiments, unless otherwise stated. ¹H chemical shifts

values (δ) are referenced to the residual non-deuterated components of the NMR solvents ($\delta = 7.26$ ppm for CHCl_3 , etc.). The ^{13}C chemical shifts (δ) are referenced to CDCl_3 (central peak, $\delta = 77.0$ ppm). Fluorine chemical shifts are referenced to CFCl_3 . Mass spectra were measured in positive mode electrospray ionization (ESI). TLC was performed on silica gel 60 F254 plastic sheets. Column chromatography was performed using silica gel (35–75 mesh). Purity of prepared compounds was determined by HPLC-UV analysis (Thermo HPLC connected with UV detector). The purity of all final compounds was determined to be >95% by RP-HPLC using the eluents water (eluent A), acetonitrile (eluent B), and the following conditions: Varian Pursuit XS, 4.6 mm \times 150 mm, 5.0 μm , 1.0 mL/min, gradient 30 min 10% \rightarrow 100% eluent B in eluent A (method 1). Purity of intermediates was >90%, unless otherwise stated.

General procedure 1 for the preparation of 3',5'-TBDMS protected FUDR (3) and of 3',5'-TBDMS protected gemcitabine (4). TBDMSCl (2.2 eq.) was added to a stirred solution of nucleoside (1.0 eq), imidazole (5.0 eq.) and DMAP (0.3 eq.) in DMF (10 mL) at 0 $^\circ\text{C}$ and the mixture was allowed to warm to 25 $^\circ\text{C}$ and stirred for 1h. The reaction was quenched with saturated aqueous solution of ammonium chloride, followed by extraction with EtOAc (3 x 5 mL). The combined organic layers were dried over Na_2SO_4 and the solvent was removed under vacuum to give a crude product as an oil used for the next step without further purification.

2'-Deoxy-5-fluoro-3',5'-bis(*O*-tert-butyldimethylsilyl-uridine (3) obtained from FUDR (2.0 g, 8.10 mmol), imidazole (40.62 mmol, 2.76 g), DMAP (2.43 mmol, 0.297 g) and TBDMSCl (17.87 mmol, 2.70 g) in DMF (15 mL) as an oil. Yield, 83% (3.20 g). ^1H NMR (500 MHz, CD_3OD): δ_{H} 8.02 (d, $J = 6.5$ Hz, 1H, $H-6$), 6.30 (t, $J = 7.50$, 1H, $H-1'$), 4.40 (q, $J = 3.5$ Hz, 1H, $H-3'$), 3.97 – 3.86 (m, 2H, $H-4'$, $H-5'$), 3.74

(dd, 1H, $J = 12.0, 3.5$ Hz, $H-5'$), 2.40 – 2.31 (m, 1H, $H-2'$), 2.10 – 2.00 (m, 1H, $H-2'$), 0.93, 0.92 (2 x s, 18H, $C(CH_3)_3$), 0.12, 0.11 (2 x s, 12H, $Si(CH_3)_2$); ^{13}C NMR (125 MHz, $CDCl_3$): δ_C 157.23 (d, $^2J_{C-F} = 26.0$ Hz, $C-4$), 149.17 ($C-2$), 141.54 (d, $^1J_{C-F} = 235.0$ Hz, $C-5$), 124.12 (d, $^2J_{C-F} = 33.7$ Hz, $C-6$), 87.98 ($C-4'$), 85.46 ($C-1'$), 71.38 ($C-3'$), 62.58 ($C-5'$), 41.73 ($C-2'$), 25.65 ($C(CH_3)_3$), 18.37, 18.06 ($C(CH_3)_3$), -4.67, -4.93 ($Si(CH_3)_2$).

2'-Deoxy-2',2'-difluoro-3',5'-bis(*O*-tert-butylidimethylsilyl-D-cytidine (4) obtained from gemcitabine (2.0 g, 7.59 mmol), imidazole (2.58 g, 37.95 mmol), DMAP (0.278 g, 2.27 mmol), and TBDMSCl (2.52 g, 16.69 mmol) in DMF (15 mL) as an oil. Yield 82% (3.05 g). 1H NMR (500 MHz, CD_3OD): δ_H 7.74 (d, $J = 8.0$ Hz, 1H, $H-6$), 6.27 (dd, $J_{H-F} = 9.50, 5.50$ Hz, 1H, $H-1'$), 5.77 (d, $J = 7.5$ Hz, 1H, $H-5$), 4.38 – 4.28 (m, 1H, $H-3'$), 4.02 (dd, 1H, $J = 11.5, 3.5$ Hz, $H-5'a$), 3.92 – 3.88 (m, 1H, $H-4'$), 3.80 (dd, 1H, $J = 2.0, 11.5$ Hz, $H-5'b$), 0.93, 0.92 (2 x s, 18H, $C(CH_3)_3$), 0.12, 0.11 (2 x s, 12H, $Si(CH_3)_2$).

2'-Deoxy-5-fluoro-3'-(*O*-tert-butylidimethylsilyl-uridine (5). CSA (6.74 mmol, 1.56 g) was added to 2'-deoxy-5-fluoro-3',5'-bis(*O*-tert-butylidimethylsilyl-uridine (3) (6.74 mmol, 3.20 g) dissolved in MeOH (15 mL), at 0 °C and the reaction mixture was stirred under an argon atmosphere for 10 min. After the reaction was completed, the solvent was removed under reduced pressure to yield the crude residue that was purified by silica gel column chromatography with a gradient of MeOH (1% to 10%) in DCM as an eluent to afford **5** as a white solid. Yield, 60% (1.45 g). 1H NMR (500 MHz, $DMSO-d_6$): δ_H 8.22 (d, $J_{H-F} = 8.0$ Hz, 1H, $H-6$), 6.25 (td, $J = 7.5, 2.0$ Hz 1H, $H-1'$), 5.20 (t, $J = 3.0$ Hz, 1H, $5'-OH$), 4.53 (q, $J = 3.5$ Hz, 1H, $H-3'$), 3.92 (qr, $J = 3.5$ Hz, 1H, $H-4'$), 3.80 (dd, $J = 12.0, 3.5$ Hz, 1H, $H-5'$), 3.73 (dd, $J = 12.0, 3.5$ Hz, 1H, $H-5'$), 2.26 – 2.21 (m, 2H, $H-2'$), 0.94 (s, 9H, $C(CH_3)_3$), 0.15 (s, 6H, $Si(CH_3)_2$); ^{13}C

NMR (125 MHz, CD₃OD): δ_C 159.46 (d, $^2J_{C-F}$ = 26.2 Hz, C-4), 150.83 (C-2), 141.87 (d, $^1J_{C-F}$ = 231.0 Hz, C-5), 126.24 (d, $^2J_{C-F}$ = 35.0 Hz, C-6), 89.58 (C-4'), 86.83 (C-1'), 73.46 (C-3'), 62.38 (C-5'), 41.99 (C-2'), 26.41 (C(CH₃)₃), 18.84 (C(CH₃)₃), -4.65 (d, $^1J_{C-Si}$ = 18.7 Hz, (Si(CH₃)₂)).

MS (MSI+): m/z calcd for C₁₅H₂₅FN₂O₅Si: 360.45 [M]⁺; found 383.149 [M + Na]⁺.

2'-Deoxy-2',2'-difluoro-3'-(*O*-*tert*-butyldimethylsilyl)-D-cytidine (6). A solution of trichloroacetic acid (113.88 mmol, 11.42 mmol) in 8.7 mL of water was added to a solution of compound **4** (2.0 g, 4.06 mmol) in THF (38 mL) at 0 °C. The reaction mixture was stirred for 2h at rt, and then the reaction solution was neutralised with solid NaHCO₃ until evolution of gas was finished. Next, 100 mL of water was added to the reaction mixture, followed by extraction with EtOAc (3 x 50 mL). The volatiles were concentrated under reduced pressure to afford the title compound **6** as a white solid. Yield, 82% (1.25 g). ¹H NMR (500 MHz, DMSO-*d*₆): δ_H 7.54 (d, J = 7.5 Hz, 1H, *H*-6), 7.55, 7.53 (2 x bs, 2H, NH₂), 6.03 (t, J_{H-F} = 8.0 Hz, 1H, *H*-1'), 5.68 (d, J = 7.5 Hz, 1H, *H*-5), 5.15 (bs, 1H, *OH*-5'), 4.25 – 4.18 (m, 1H, *H*-3'), 3.71 – 3.64 (m, 2H, *H*-4', *H*-5'a), 3.46 (dd, J = 12.5, 3.5 Hz, 1H, *H*-5'b), 0.77 (s, 9H, C(CH₃)₃), 0.00, -0.02 (2 x s, 6H, Si(CH₃)₂). ¹³C NMR (125 MHz, CD₃OD): δ_C 165.99 (C-NH₂), 156.16 (C=O base), 140.43 (CH-base), 122.61 (t, $^1J_{C-F}$ = 256 Hz, CF₂), 95.16 (CH-base), 84.35 (t, $^2J_{C-F}$ = 31.7 Hz, C-1'), 80.67 (t, $^3J_{C-F}$ = 4.0 Hz, C-4'), 68.39 (t, $^2J_{C-F}$ = 22.8 Hz, C-3'), 60.35 (C-5'), 25.09, 24.92 (C(CH₃)₃), 17.92 (C(CH₃)₃), -6.65 (d, $^1J_{C-Si}$ = 12.9 Hz, Si(CH₃)₂).

General procedure 2 for a preparation of 3'-TBDMS-protected ProTides. *Tert*-BuMgCl (1.0 M solution in THF, 1.2 eq.) was added to a solution of 3'-*O*-(*tert*-butyldimethylsilyl)-protected nucleoside (1.0 eq.) in dry THF (10 mL), in one portion followed by addition of an appropriate phosphorochloridate (2.0 eq.) dissolved in

anhydrous THF (3 mL). The reaction mixture was stirred for 16 h and then evaporated under reduced pressure to give a crude residue that was purified either by silica gel column chromatography, eluting with a gradient of MeOH (0 to 2%) in DCM or with a gradient of MeOH (3 to 6%) in a mixture of EtOAc (15%) and DCM as an eluent, followed by TLC preparative purification unless stated otherwise to afford 3'-TBDMS-protected ProTides as single diastereoisomers.

2'-Deoxy-5-fluoro-3'-O-tert-butyl-dimethylsilyl-D-uridine-5'-O-[naphthyl-(benzyloxy-L-alaninyl)] phosphate (7) prepared according to general procedure 2 from 2'-deoxy-5-fluoro-3'-(*O*-tert-butyl-dimethylsilyl-uridine (**5**) (2.77 mmol, 1.0 g) and L-Ala-OBn-ONaph phosphorochloridate¹⁰ (5.55 mmol, 2.24 g), (3.33 mmol, 3.33 mL) in THF (15 mL). The crude mixture was purified by silica column chromatography with a slow gradient of MeOH (0% to 2%) in DCM as an eluent to give the fast eluting isomer A (**7a**, *S_P*) or (**7b**, *R_P*) and the slow eluting isomer B (**7a**, *S_P*) or (**7b**, *R_P*).

2'-Deoxy-5-fluoro-3'-O-(tert-butyl-dimethylsilyl)-D-uridine-5'-O-[naphthyl-(benzyloxy-L-alaninyl)] phosphate (7_Isomer A) the fast eluting fraction obtained as a white solid. Yield, 9% (0.19 g) ³¹P NMR (202 MHz, CD₃OD): δ_P 4.64. ¹H NMR (500 MHz, CD₃OD): δ_H 8.15 (d, *J* = 8.0 Hz, 1H, Ar-*H*), 7.89 (d, *J* = 8.0 Hz, 1H, *H*-Ar), 7.73 (d, *J*_{H-F} = 8.0 Hz, 1H, *H*-6), 7.65 (d, *J* = 6.5 Hz, 1H, Ar-*H*), 7.54 – 7.50 (m, 3H, Ar-*H*), 7.43 (t, *J* = 8.0 Hz, 1H, Ar-*H*), 7.34 – 7.28 (m, 5H, Ar-*H*), 6.08 (t, *J* = 6.5 Hz, 1H, *H*-1'), 5.06 (ABq, *J*_{A-B} = 12.0 Hz, 2H, OCH₂Ph), 4.30 – 4.28 (m, 3H, 2 x *H*-5', *H*-3'), 4.15 – 4.11 (m, 1H, CHCH₃), 3.97 – 3.95 (m, 1H, *H*-4'), 1.96 (ddd, *J* = 13.5, 5.5, 2.5 Hz, 1H, *H*-2'), 1.55 – 1.50 (m, 1H, *H*-2'), 1.38 (d, *J* = 7.0 Hz, 3H, CHCH₃), 0.89 (s, 9H, C(CH₃)₃), 0.01 (d, ²*J*_{Si-H} = 5.5 Hz, 6H, Si(CH₃)₂); MS (MSI⁺): *m/z* calcd for C₃₅H₄₃FN₃O₉PSi: 727.78 [M]⁺; found 750.25 [M+Na]⁺.

2'-Deoxy-5-fluoro-3'-O-(*tert*-butyl-dimethylsilyl)-D-uridine-5'-O-[naphthyl-(benzyloxy-L-alaninyl)] phosphate (7_Isomer B) the slow eluting fraction obtained as a white solid. Yield, 12% (0.25 g). ^{31}P NMR (202 MHz, CD_3OD): δ_{P} 4.28. ^1H NMR (500 MHz, CD_3OD): 8.17 – 8.14 (m, 1H, *H*-Ar), 7.88 – 7.86 (m, 1H, *H*-Ar), 7.70 (d, $J_{\text{H-F}} = 6.5$ Hz, 1H, *H*-6), 7.60 (d, $J = 8.5$ Hz, 1H, *H*-Ar), 7.54 – 7.50 (m, 3H, *H*-Ar), 7.37 (t, $J = 8.0$ Hz, 1H, *H*-Ar), 7.33 – 7.27 (m, 5H, *H*-Ar), 6.06 (t, $J = 7.5$ Hz, 1H, *H*-1'), 5.06 (apparent s, 2H, OCH_2Ph), 4.32 – 4.27 (m, 2H, *H*-3', *H*-5'), 4.23 – 4.19 (m, 1H, *H*-5'), 4.15 – 4.11 (m, 1H, CHCH_3), 3.99 – 3.97 (m, 1H, *H*-4'), 1.98 (ddd, $J = 13.5, 5.5, 2.5$ Hz, 1H, *H*-2'), 1.59 – 1.54 (m, 1H, *H*-2'), 1.38 (d, $J = 7.0$ Hz, 3H, CHCH_3), 0.87 (s, 9H, $\text{C}(\text{CH}_3)_3$), 0.04 (d, $^2J_{\text{Si-H}} = 5.5$ Hz, 6H, $\text{Si}(\text{CH}_3)_2$); ^{13}C NMR (125 MHz, CD_3OD): δ_{C} 173.08 (d, $^3J_{\text{C-P}} = 4.8$ Hz, $\text{C}=\text{O}$, ester), 157.95 (d, $^2J_{\text{C-F}} = 26.2$ Hz, *C*-4), 149.12 (*C*-2), 146.41 (d, $^2J_{\text{C-P}} = 7.8$ Hz, OC-Naph), 140.32 (d, $^1J_{\text{C-F}} = 232.0$ Hz, *C*-5), 135.74, 134.90 (*C*-Ar), 128.19, 127.96, 127.89, 127.60, 126.51 (CH-Ar), 126.47, 126.42 (*C*-Ar), 126.20, 125.17, 124.84 (CH-Ar), 124.28 (d, $^2J_{\text{C-F}} = 34.1$ Hz, *C*-6), 121.20 (CH-Ar), 115.17 (d, $^3J_{\text{C-P}} = 3.2$ Hz, CH-Ar), 85.77 (d, $^3J_{\text{C-P}} = 7.7$ Hz, *C*-4'), 85.55 (*C*-1'), 72.10 (*C*-3'), 66.63 (OCH_2Ph), 66.11 (d, $^2J_{\text{C-P}} = 5.5$ Hz, *C*-5'), 50.46 (CHCH_3), 39.80 (*C*-2'), 24.81 ($\text{C}(\text{CH}_3)_3$), 18.99 (d, $^3J_{\text{C-P}} = 6.9$ Hz, CHCH_3), 17.34 ($\text{C}(\text{CH}_3)_3$), -6.06 (d, $^1J_{\text{C-Si}} = 14.2$ Hz, $\text{Si}(\text{CH}_3)_2$). MS (MSI+): m/z calcd for $\text{C}_{35}\text{H}_{43}\text{FN}_3\text{O}_9\text{PSi}$: 727.78 $[\text{M}]^+$; found 750.25 $[\text{M}+\text{Na}]^+$.

2'-Deoxy-2',2'-difluoro-3'-O-(*tert*-butyl-dimethylsilyl)-D-cytidine-5'-O-[phenyl(benzyloxy-L-alaninyl)] phosphate (8) obtained from 3'-O-(*tert*-butyldimethylsilyl)-gemcitabine **6** (2.66 mmol, 1.0 g) in dry THF (14 mL), *tert*-BuMgCl (2.90 mmol, 2.9 mL) and L-Ala-OBn-OPh phosphorochloridate⁹ (3.97 mmol, 1.40 g) dissolved in anhydrous THF (5 mL). The reaction mixture was stirred

for 16 h and then evaporated in vacuum to give a crude residue that was purified by column chromatography on silica gel, eluting with a gradient of DCM/EtOAc/MeOH (82%/15%/3%) to (79%/15%/6%) to afford **8b** as the fast eluting fraction and **8a** as the slow eluting fraction.

2'-Deoxy-2',2'-difluoro-3'-tert-butylsilyl-D-cytidine-5'-O-[phenyl(benzyloxy-L-alaninyl)] phosphate (8a) the slow eluting fraction obtained as a white solid. Yield, 30% (0.55 g). ^{31}P NMR (202 MHz, CD_3OD): δ_{P} 3.56. ^1H NMR (500 MHz, CD_3OD): δ_{H} 7.40 (d, $J = 7.5$ Hz, 1H, $H-6$), 7.22 – 7.16 (m, 7H, Ar- H), 7.09 – 7.04 (m, 3H, Ar- H), 6.07 (t, $J_{\text{H-F}} = 8.5$ Hz 1H, $H-1'$), 5.71 (d, $J = 7.5$ Hz, 1H, $H-5$), 5.00, 4.96 (AB q, $J_{\text{AB}} = 12.0, 9.0$ Hz, 2H, OCH_2Ph), 4.31 – 4.27 (m, 1H, $H-3'$), 4.22 – 4.16 (m, 1H, $H-4'$), 4.13 – 4.08 (m, 1H, $H-5'$), 3.89 – 3.83 (m, 2H, $H-5'$, CHCH_3), 1.23 (dd, $J = 7.0, 1.5$ Hz, 3H, CHCH_3), 0.78 (s, 9H, $\text{C}(\text{CH}_3)_3$), 0.0 (s, 6H, $\text{Si}(\text{CH}_3)_2$). ^{13}C NMR (125 MHz, CD_3OD): δ_{C} 173.07 (d, $^3J_{\text{C-P}} = 5.3$ Hz, $\text{C}=\text{O}$, ester), 166.26 (C-NH_2), 156.28 ($\text{C}=\text{O}$ base), 150.68 (d, $^2J_{\text{C-P}} = 3.3$ Hz, C-Ar), 141.11 (CH-base), 135.79 (C-Ar), 129.50, 128.22, 127.98, 127.88, 124.94 (CH-Ar), 122.83 (d, $^1J_{\text{C-F}} = 258$ Hz, CF_2), 120.02, 119.98 (CH-Ar), 95.28 (CH-base), 79.39 (broad signal, $\text{C-1}'$), 79.35 ($\text{C-4}'$), 71.05 (apparent t, $^2J_{\text{C-F}} = 26.1$ Hz, $\text{C-3}'$), 66.60 (OCH_2Ph), 64.02 (d, $^2J_{\text{C-P}} = 4.9$ Hz, $\text{C-5}'$), 50.29 (CHCH_3), 24.63 ($\text{C}(\text{CH}_3)_3$), 19.02 (d, $^3J_{\text{C-P}} = 6.8$ Hz, CHCH_3), 17.47 ($\text{C}(\text{CH}_3)_3$), -6.20 (d, $^1J_{\text{C-Si}} = 29.1$ Hz, $\text{Si}(\text{CH}_3)_2$). MS (ES+) m/z calcd for $\text{C}_{31}\text{H}_{41}\text{F}_2\text{N}_4\text{O}_8\text{PSi}$: 694.73 [M^+]; found 717.45 [$\text{M} + \text{Na}$] $^+$.

2'-Deoxy-2',2'-difluoro-3'-tert-butylsilyl-D-cytidine-5'-O-[phenyl(benzyloxy-L-alaninyl)] phosphate (8b), the fast eluting fraction obtained as a white solid. Yield, 25% (0.45 g). ^{31}P NMR (202 MHz, CD_3OD): δ_{P} 3.71. ^1H NMR (500 MHz, CD_3OD): δ_{H} 7.45 (d, $J = 7.5$ Hz, 1H, $H-6$), 7.24 – 7.18 (m, 7H, Ar- H), 7.09 – 7.07 (m, 3H, Ar- H), 6.12 (t, $J_{\text{H-F}} = 8.5$ Hz 1H, $H-1'$), 5.78 (d, $J = 7.5$ Hz, 1H, $H-5$), 5.04, 5.01 (AB q,

$J_{AB} = 12.5, 3.0$ Hz, 2H, OCH_2Ph), 4.39 – 4.36 (m, 1H, $H-3'$), 4.23 – 4.13 (m, 2H, $H-5'$, $H-4'$), 3.93 – 3.87 (m, 2H, $H-5'$, $CHCH_3$), 1.24 (dd, $J = 7.0, 1.5$ Hz, 3H, $CHCH_3$), 0.80 (s, 9H, $C(CH_3)_3$), 0.0 (s, 6H, $Si(CH_3)_2$). ^{13}C NMR (125 MHz, CD_3OD): δ_C 173.35 (d, $^3J_{C-P} = 4.2$ Hz, $C=O$, ester), 166.26 ($C-NH_2$), 156.26 ($C=O$ base), 150.63 (d, $^2J_{C-P} = 2.4$ Hz, $C-Ar$), 140.89 (CH -base), 135.82 ($C-Ar$), 129.47, 128.21, 127.98, 127.92, 125.57 ($CH-Ar$), 123.90 (d, $^1J_{C-F} = 266$ Hz, CF_2), 120.11, 120.07 ($CH-Ar$), 95.38 (CH -base), 79.24 (broad signal, $C-1'$), 75.23 (broad signal, $C-4'$), 70.94 (broad signal, $C-3'$), 66.59 (OCH_2Ph), 63.59 (d, $^2J_{C-P} = 5.0$ Hz, $C-5'$), 50.47 ($CHCH_3$), 24.62 ($C(CH_3)_3$), 18.82 (d, $^3J_{C-P} = 7.7$ Hz, $CHCH_3$), 17.47 ($C(CH_3)_3$), -6.21 (d, $^1J_{C-Si} = 28.1$ Hz, $Si(CH_3)_2$). MS (ES+) m/z calcd for $C_{31}H_{41}F_2N_4O_8PSi$: 694.73 [M^+]; found 717.45 [$M + Na$] $^+$.

General procedure 3 for the 3'-desilylation of 3'-TBDMS-protected ProTides 7a,b and 8a,b. The separated diastereoisomer of 3'-TBDMS-protected ProTide (**7a**), (**7b**), (**8a**) and (**8b**) was treated with a mixture of THF/TFA/DCM (4:1:1) at 0°C. The resulting reaction mixture was stirred at 0 °C for 2 h and then at room temperature for 12 h. After the reaction was completed, the solvents were evaporated, and the residue was purified on silica gel with gradient of MeOH (2% to 8%) in DCM as an eluent.

2'-Deoxy-5-fluoro-D-uridine-5'-O-[naphthyl-(benzyloxy-L-alaninyl)] phosphate (9a or 9b, NUC-3373_IsoA): obtained from the fast eluting **7_isomer A** (**7a** or **7b**) (0.19 g, 0.26 mmol) as a white solid. Yield, 75% (0.12 g). ^{31}P NMR (202 MHz, CD_3OD): δ_P 4.60. ^{19}F NMR (470 MHz, CD_3OD): δ_F -167.05. 1H NMR (500 MHz, CD_3OD): δ_H 8.14 (dd, $J = 8.5, 2.0$ Hz, 1H, $Ar-H$), 7.87 (d, $J = 7.0$ Hz, 1H, $H-Ar$), 7.72 (apparent s, 1H, $Ar-H$), 7.69 (d, $J_{H-F} = 6.5$ Hz, 1H, $H-6$), 7.54 – 7.49 (m, 3H, $Ar-H$), 7.42 (t, $J = 8.0$ Hz, 1H, $Ar-H$), 7.34 – 7.28 (m, 5H, $Ar-H$), 6.14 (t, $J = 6.0$ Hz 1H, $H-1'$), 5.10 (ABq, $J_{A-B} = 12.5$ Hz, 2H, OCH_2Ph), 4.35 – 4.33 (m, 2H, $H-3'$, $H-5'$), 4.30 –

4.28 (m, 1H, *H*-5'), 4.14 – 4.08 (m, 1H, *CHCH*₃), 4.07 – 4.05 (m, 1H, *H*-4'), 2.11 (ddd, *J* = 14.0, 6.0, 3.0 Hz, 1H, *H*-2'), 1.73 – 1.68 (m, 1H, *H*-2'), 1.33 (dd, *J* = 7.5, 1.5 Hz, 3H, *CHCH*₃). ¹³C NMR (125 MHz, CD₃OD): δ_C 174.92 (d, ³*J*_{C-P} = 3.9 Hz, C=O, ester), 159.27 (d, ²*J*_{C-F} = 25.9 Hz, C-4), 150.52 (C-2), 147.90 (d, ²*J*_{C-P} = 7.3 Hz, OC-Naph), 141.73 (d, ¹*J*_{C-F} = 232.3 Hz, C-5), 137.19, 136.29 (C-Ar), 129.59, 129.36, 128.91, 127.90 (CH-Ar), 127.85, 127.80 (C-Ar), 127.60 (CH-Ar), 126.50 (d, ³*J*_{C-P} = 1.4 Hz, CH-Naph), 126.18 (CH-Ar), 125.53 (d, ²*J*_{C-F} = 34.0 Hz, C-6), 122.64 (CH-Ar), 116.30 (d, ³*J*_{C-P} = 3.12 Hz, CH-Ar), 86.93 (C-1'), 86.88 (d, ³*J*_{C-P} = 8.0 Hz, C-4'), 72.18 (C-3'), 68.10 (OCH₂Ph), 67.86 (d, ²*J*_{C-P} = 5.2 Hz, C-5'), 51.96 (*CHCH*₃), 40.84 (C-2'), 20.24 (d, ³*J*_{C-P} = 7.5 Hz, *CHCH*₃). MS (MSI+): *m/z* calcd for C₂₉H₂₉FN₃O₉P: 613.53 [M]⁺; found 636.15 [M + Na]⁺. Reverse HPLC eluting with (H₂O/AcCN from 100/0 to 0/100) in 30 min., *t*_R 16.61 min.

2'-Deoxy-5-fluoro-D-uridine-5'-O-[naphthyl-(benzyloxy-L-alaninyl)] phosphate (9a or 9b, NUC-3373_IsoB) obtained from the slow eluting **7_isomer B (7a or 7b)** (0.24 g, 0.36 mmol) (0.19 g, 0.26 mmol) as a white solid. Yield, 75% (0.15 g). ³¹P NMR (202 MHz, CD₃OD): δ_P 4.26. ¹⁹F NMR (470 MHz, CD₃OD): δ_F -167.31. ¹H NMR (500 MHz, CD₃OD): δ_H 8.17 – 8.14 (m, 1H, Ar-*H*), 7.90 – 7.87 (m, 1H, Ar-*H*), 7.72 (apparent s, 1H, Ar-*H*), 7.69 (d, *J*_{H-F} = 6.0 Hz, 1H, *H*-6), 7.54 – 7.48 (m, 3H, Ar-*H*), 7.38 (t, *J* = 8.0 Hz, 1H, Ar-*H*), 7.34 – 7.28 (m, 5H, Ar-*H*), 6.10 (t, *J* = 6.0 Hz 1H, *H*-1'), 5.12 (s, 2H, OCH₂Ph), 4.36 – 4.25 (m, 3H, *H*-3', 2 x *H*-5'), 4.14 – 4.08 (m, 1H, *CHCH*₃), 4.05 (apparent q, *J* = 2.5 Hz, 1H, *H*-4'), 2.14 (ddd, *J* = 14.0, 6.0, 3.0 Hz, 1H, *H*-2'), 1.75 – 1.69 (m, 1H, *H*-2'), 1.36 (dd, *J* = 7.0, 0.5 Hz, 3H, *CHCH*₃). ¹³C NMR (125 MHz, CD₃OD): δ_C 174.57 (d, ³*J*_{C-P} = 4.5 Hz, C=O, ester), 159.38 (d, ²*J*_{C-F} = 26.0 Hz, C-4), 150.48 (C-2), 147.80 (d, ²*J*_{C-P} = 7.3 Hz, OC-Naph), 141.67 (d, ¹*J*_{C-F} = 232.0 Hz, C-5), 137.15, 136.27 (C-Ar), 129.63, 129.40, 129.36, 128.97, 127.89 (CH-Ar),

127.86, 127.81 (C-Ar), 127.59 (CH-Ar), 126.56 (d, $^3J_{C-P} = 1.4$ Hz, CH-Naph), 126.22 (CH-Ar), 125.62 (d, $^2J_{C-F} = 34.1$ Hz, C-6), 122.63 (CH-Ar), 116.56 (d, $^3J_{C-P} = 3.0$ Hz, CH-Ar), 86.98 (C-1'), 86.68 (d, $^3J_{C-P} = 7.6$ Hz, C-4'), 72.02 (C-3'), 68.08 (OCH₂Ph), 67.85 (d, $^2J_{C-P} = 5.5$ Hz, C-5'), 51.84 (CHCH₃), 40.90 (C-2'), 20.44 (d, $^3J_{C-P} = 6.7$ Hz, CHCH₃). MS (MSI+): m/z calcd for C₂₉H₂₉FN₃O₉P: 613.53 [M]⁺; found 636.15 [M + Na]⁺. Reverse HPLC eluting with (H₂O/AcCN from 100/0 to 0/100) in 30 min., t_R 16.03 min.

2'-Deoxy-2',2'-difluoro-D-cytidine-5'-O-[phenyl(benzyloxy-L-alaninyl)]

phosphate (10a, NUC-1031_Sp) obtained from **8a** (0.55 g, 0.79 mmol) as a white, crystalline solid. Yield, 76% (0.35 g). ³¹P NMR (202 MHz, CD₃OD): δ_P 3.66. ¹⁹F NMR (470 MHz, CD₃OD): δ_F - 118.0 (d, $J = 241$ Hz, *F*), - 120.24 (broad d, $J = 241$ Hz, *F*). ¹H NMR (500 MHz, CD₃OD): δ_H 7.58 (d, $J = 7.5$ Hz, 1H, *H*-6), 7.38 – 7.32 (m, 7H, Ar-*H*), 7.26 – 7.20 (m, 3H, Ar-*H*), 6.24 (t, $J = 7.5$ Hz, 1H, *H*-1'), 5.84 (d, $J = 7.5$ Hz, 1H, *H*-5), 5.20 (AB q, $J_{AB} = 12.0, 7.2$ Hz, 2H, OCH₂Ph), 4.46 – 4.43 (m, 1H, *H*-5'), 4.36 – 4.31 (m, 1H, *H*-5'), 4.25 – 4.19 (m, 1H, *H*-3'), 4.07 – 4.00 (m, 2H, *H*-4', CHCH₃), 1.38 (d, $J = 7.2$ Hz, 3H, CHCH₃). ¹³C NMR (125 MHz, CD₃OD): δ_C 174.61 (d, $^3J_{C-P} = 5.0$ Hz, C=O, ester), 167.64 (C-NH₂), 157.74 (C=O base), 152.10 (d, $^2J_{C-P} = 7.0$ Hz, C-Ar), 142.41 (CH-base), 137.22 (C-Ar), 130.91, 129.64, 129.40, 129.33, 126.33 (CH-Ar), 124.51 (d, $^1J_{C-F} = 257$ Hz, CF₂), 121.47, 121.44 (CH-Ar), 96.67 (CH-base), 86.08 (broad signal, C-1'), 80.32 (C-4'), 71.27 (apparent t, $^2J_{C-F} = 23.7$ Hz, C-3'), 68.04 (OCH₂Ph), 65.74 (d, $^2J_{C-P} = 5.30$ Hz, C-5'), 51.67 (CHCH₃), 20.44 (d, $^3J_{C-P} = 6.25$ Hz, CHCH₃). (ES+) m/z , found: (M + Na⁺) 603.14. C₂₅H₂₇F₂N₄O₈NaP required: (M⁺) 580.47. Reverse HPLC, eluting with H₂O/MeOH from 100/0 to 0/100 in 35 min, $t_R = 18.32$ min.

2'-Deoxy-2',2'-difluoro-D-cytidine-5'-O-[phenyl(benzyloxy-L-alaninyl)]

phosphate (10b, NUC-1031_ RP) was obtained from **8b** (0.45 g, 0.65 mmol) as a white, crystalline solid. Yield, 78% (0.29 g). ^{31}P NMR (202 MHz, CD_3OD): δ_{P} 3.83. ^{19}F NMR (470 MHz, CD_3OD): δ_{F} - 118.3 (d, $J = 241$ Hz, F), - 120.38 (broad d, $J = 241$ Hz, F). ^1H NMR (500 MHz, CD_3OD): δ_{H} 7.56 (d, $J = 7.5$ Hz, 1H, $H-6$), 7.38 – 7.31 (m, 7H, Ar- H), 7.23 – 7.19 (m, 3H, Ar- H), 6.26 (t, $J = 7.5$ Hz, 1H, $H-1'$), 5.88 (d, $J = 7.5$ Hz, 1H, $H-5$), 5.20 (s, 2H, OCH_2Ph), 4.49 – 4.46 (m, 1H, $H-5'$), 4.38 – 4.34 (m, 1H, $H-5'$), 4.23 – 4.17 (m, 1H, $H-3'$), 4.07 – 4.01 (m, 2H, $H-4'$, CHCH_3), 1.38 (d, $J = 7.2$ Hz, 3H, CHCH_3). ^{13}C NMR (125 MHz, CD_3OD): δ_{C} 174.65 (d, $^3J_{\text{C-P}} = 5.0$ Hz, $\text{C}=\text{O}$, ester), 167.65 (C-NH_2), 157.75 ($\text{C}=\text{O}$ base), 152.10 (d, $^2J_{\text{C-P}} = 7.0$ Hz, C-Ar), 142.28 (CH-base), 137.50 (C-Ar), 130.86, 129.63, 129.40, 129.32, 126.31 (CH-Ar), 123.73 (d, $^1J_{\text{C-F}} = 252$ Hz, CF_2), 121.44, 121.40 (CH-Ar), 96.67 (CH-base), 85.90 (broad signal, $\text{C-1}'$), 80.27 ($\text{C-4}'$), 71.02 (apparent t, $^2J_{\text{C-F}} = 23.7$ Hz, $\text{C-3}'$), 68.04 (OCH_2Ph), 65.52 (d, $^2J_{\text{C-P}} = 5.30$ Hz, $\text{C-5}'$), 51.85 (CHCH_3), 20.23 (d, $^3J_{\text{C-P}} = 7.5$ Hz, CHCH_3). (ES+) m/z , found: ($\text{M} + \text{Na}^+$) 603.14. $\text{C}_{25}\text{H}_{27}\text{F}_2\text{N}_4\text{O}_8\text{NaP}$ required: (M^+) 580.47. Reverse HPLC, eluting with $\text{H}_2\text{O}/\text{MeOH}$ from 100/0 to 0/100 in 35 min, $t_{\text{R}} = 19.15$ min.

General method for crystallization of 7_isoB from the column purified amorphous 7_isoB using the modified procedure.²⁶ The chromatographed fraction containing the compound **7_isoB** (50 mg, 97%) was dissolved in 5 mL of 2-propanol and diluted with hexane until cloudy. The solution was seeded and stirred at room temperature for 5 h. The resulting solid was filtered, washed with hexane (2 x 2 mL), and dried under the high vacuum. The obtained solid was not suitable for single crystal X-ray analysis. Multiple attempts to crystallize **7_isoA**, **7_isoB**, **9_isoA** and **9_isoB** by slow evaporation of MeOH, 2-propanol, CHCl_3 and a combination of

CHCl₃/Et₂O (1/1, v/v), and MeOH/CHCl₃ (3/1, v/v), CHCl₃/H₂O (1/1, v/v) also failed to give crystals of sufficient quality for single crystal X-ray analysis

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at

Carboxypeptidase Y assay for NUC-3373_isoB, 10a and 10b (PDF)

List of primers used for RT-PCR (PDF)

Compound purity analyses (PDF)

Table 4. List of Primers used for RT-PCR (PDF)

Table 5. Crystal data and structure refinement for the compounds 10a and 10b (PDF)

Compound molecular formula strings (CSV)

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ABBREVIATIONS

BTC, biliary tract cancer; CDA, cytidine deaminase; CSA, camphorsulfonic acid; dCK, deoxycytidine kinase; DPD, dihydropyrimidine dehydrogenase; hENT1, human equilibrative nucleoside transporter 1; TP, thymidine phosphorylase; TS, thymidylate synthase;

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CONFLICT OF INTEREST

The authors all confirm that they have no relevant conflicts of interest.

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