

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository: <https://orca.cardiff.ac.uk/id/eprint/157914/>

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

Gao, Rongrong, Yan, Mengdi, Chen, Ming, Hayes, Sally , Meek, Keith M. , He, Huanhuan, Chen, Xueyang, Xu, Wenjin, Yan, Shixiang, Huang, Yuyan, Ding, Shengnan, Wang, Qinmei, Li, Junhua and Huang, Jinhai  
2022. The impact of different rose bengal formulations on corneal thickness and the efficacy of rose bengal/green light corneal cross-linking in the rabbit eye. *Journal of Refractive Surgery* 38 (7) , pp. 450-458.  
10.3928/1081597X-20220601-03

Publishers page: <http://dx.doi.org/10.3928/1081597X-20220601-03>

Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies. See <http://orca.cf.ac.uk/policies.html> for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.



1 **The impact of different rose bengal formulations on corneal thickness and the**  
2 **efficacy of rose bengal/green light cross-linking in the rabbit eye**

3 Running head: Rb formulations affect CCT and RGX efficacy

4

5 Rongrong Gao MD PhD,<sup>1#</sup> Mengdi Yan MD,<sup>1#</sup> Ming Chen MD,<sup>1</sup> Sally Hayes PhD,<sup>2</sup>  
6 Keith M. Meek DSc PhD,<sup>2,3</sup> Huanhuan He MD,<sup>1</sup> Xueyang Chen MD,<sup>1</sup> Wenjin Xu MD,<sup>1</sup>  
7 Shixiang Yan MD,<sup>1</sup> Yuyan Huang MD,<sup>1</sup> Shengnan Ding MD,<sup>1</sup> Qinmei Wang MD,<sup>1\*</sup>  
8 Junhua Li MD,<sup>1\*</sup> Jinhai Huang MD PhD<sup>1,3\*</sup>

9

10 <sup>1</sup> School of Ophthalmology and Optometry and Eye Hospital, Wenzhou Medical  
11 University, Wenzhou, Zhejiang, China.

12 <sup>2</sup> Structural Biophysics Research Group, School of Optometry and Vision Sciences,  
13 Cardiff University, Cardiff, UK.

14 <sup>3</sup> Eye Institute and Department of Ophthalmology, Eye & ENT Hospital, Fudan  
15 University; Key Laboratory of Myopia, Chinese Academy of Medical Sciences,  
16 Shanghai, China.

17

18 <sup>#</sup>These authors contributed equally.

19

20 **\*Corresponding authors: e-mail:** Qinmei Wang wqm6@mail.eye.ac.cn; Junhua Li  
21 54064308@qq.com; Jinhai Huang vip999vip@163.com

22 **Mailing address:**

23 Eye Hospital of Wenzhou Medical University, 270 West Xueyuan Road, Wenzhou,  
24 Zhejiang, China, 325027. Tel: 86-577-88068880, Fax: 86-577-88832083.

25 **Grant/financial Support:** This work was supported in part by the Foundation of  
26 Wenzhou City Science & Technology Bureau (Y20210207); the Major Scientific and  
27 Technological Innovation Project of Wenzhou (ZY2021016); Zhejiang Provincial &  
28 Ministry of Health Research Fund for Medical Sciences (WKJ-ZJ-2134); the Natural  
29 Science Foundation of Zhejiang Province (LY21H120003); the Major Fund of  
30 Wenzhou Medical University (YNZD1201903); the Project Launch Fund of Affiliated  
31 Eye Hospital of Wenzhou Medical University (KYQD20190701); and the EYE & ENT  
32 Hospital of Fudan University High-level Talents Program (2021318). The sponsor or  
33 funding organization had no role in the design or conduct of this research.  
34 **Financial Disclosures:** The authors have no proprietary or financial interest in any  
35 materials discussed in this article.

36 **ABSTRACT**

37 **Purpose:** To examine central corneal thickness (CCT) changes during in vivo rose  
38 bengal-green light corneal cross-linking (RGX) and compare the cross-linking efficacy  
39 of different rose bengal (Rb) formulations.

40 **Methods:** After epithelium removal, the right eyes of rabbits were immersed in Rb  
41 solution for 2 or 20 minutes, then the Rb distribution in the corneal stroma was analyzed  
42 by confocal fluorescence detection. During the RGX process, the CCT was measured  
43 at 7 time points. The left eyes served as untreated control group. Corneal enzymatic  
44 resistance and corneal biomechanics were tested to compare the RGX efficacy.

45 **Results:** The Rb infiltration depths were about 100  $\mu\text{m}$  and 200  $\mu\text{m}$  for the 2-minute  
46 and the 20-minute groups, respectively. CCT increased significantly after infiltration,  
47 then decreased significantly in the first 200 seconds of irradiation and decreased slowly  
48 for the next 400 seconds. The CCT of the 20 min groups was significantly higher than  
49 that of the 2 min groups ( $P < 0.0001$ ). All the RGX treatments improved the corneal  
50 enzymatic resistance and corneal biomechanics, with the effects being greater in the 20  
51 min groups. The inclusion of 1.1% hydroxypropyl methylcellulose (HPMC) in the Rb  
52 formulation helped to maintain CCT during irradiation, whilst not affecting either the  
53 infiltration of Rb or the efficacy of RGX.

54 **Conclusions:** Within the range studied, RGX efficacy increase with infiltration time.  
55 The incorporation of a 20-minute infiltration of 0.1% Rb-1.1% HPMC into the RGX

56 procedure may further improve the safety of the treatment and its prospects for clinical

57 use.

58 **Keywords:** corneal cross-linking, rose bengal, 532 nm green light, hydroxypropyl

59 methylcellulose, central corneal thickness

60 **Introduction**

61 Keratoconus is a progressive corneal degenerative disease, characterized by corneal  
62 thinning, irregular astigmatism and secondary visual impairment.<sup>1</sup> Corneal cross-  
63 linking is the main treatment to enhance the biomechanical properties of the cornea and  
64 delay the progress of keratoconus.<sup>2,3</sup>

65 The standard corneal cross-linking protocol (UVX), often referred to as the Dresden  
66 protocol, involves the use of riboflavin and ultraviolet light, and requires a central  
67 corneal thickness (CCT) of no less than 400  $\mu\text{m}$  after de-epithelialization to keep the  
68 irradiation dose within the safe range of the corneal endothelium.<sup>4</sup> However, it is  
69 sometimes difficult to achieve and maintain the required thickness throughout the UVX  
70 process, and the CCT of many patients before the operation is less than 400  $\mu\text{m}$ .<sup>5</sup>  
71 Numerous clinical and laboratory studies have examined the efficacy of different  
72 riboflavin (Rf) formulations on CCT during UVX, and shown that variations in the Rf  
73 carrier solution can lead to considerable variations in the final CCT.<sup>6-9</sup>

74 Rose bengal-green light corneal cross-linking (RGX) is a promising treatment for thin  
75 corneas due to the shallow infiltration of rose bengal (Rb) in the corneal stroma.<sup>10-14</sup>  
76 Since the irradiance and total energy of light is much larger with RGX than UVX (0.25  
77  $\text{W}/\text{cm}^2$  to 0.4  $\text{W}/\text{cm}^2$  in RGX vs. 3  $\text{mW}/\text{cm}^2$  in UVX), we hypothesized that water  
78 evaporation during light exposure might lead to a decrease in the CCT, thus affecting  
79 the safety of endothelial cells. Although RGX performed on laser-made 250  $\mu\text{m}$  thick  
80 rabbit corneas at an illumination intensity of 0.4  $\text{W}/\text{cm}^2$  for 250s (100  $\text{J}/\text{cm}^2$ ) has been  
81 shown to be safe<sup>15</sup>, maintaining a suitable CCT during surgery should further improve  
82 the safety of the technique, making it suitable for more patients. However, unlike UVX  
83 for which a variety of commercial Rf formulations have been developed to enable the  
84 customization of treatments, there is a lack of commercial Rb formulations and studies

85 to date have been limited to the use of a Rb formulation comprising 0.1% Rb in  
86 phosphate buffered saline (PBS). To our knowledge, the efficacy of this Rb formulation  
87 on CCT has not yet been reported.

88 The Rf formulation used in the Dresden UVX protocol comprises 0.1% riboflavin in  
89 20% dextran T500. The dextran increases the solution viscosity and has good film-  
90 forming performance with an average film rupture time of 22 minutes.<sup>16</sup> However, due  
91 its strong hydrophilic hydroxyl groups and hyperosmolarity, its application can cause  
92 corneal dehydration and result in a significant decrease in CCT.<sup>6</sup> The use of  
93 hydroxypropyl methylcellulose (HPMC) as an alternative Rf carrier solution, has some  
94 advantages over dextran in that it offers a longer average film rupture time of 32  
95 minutes and it does not cause significant corneal dehydration or tissue thinning.<sup>6, 17, 18</sup>  
96 However, it is still controversial which is more effective in UVX, the use of Rf solutions  
97 containing HPMC or those containing dextran. Based on a retrospective analysis of 24-  
98 month follow-up data from 33 patients that underwent UVX with either a HPMC Rf  
99 formulation or a dextran Rf formulation, Rapuano et al. concluded that the dextran Rf  
100 formulation may result in significantly better visual acuity compared to the isotonic  
101 HPMC Rf formulation.<sup>19</sup> Contrary to this, Thorsrud et. al's study of 40 patients at 2-  
102 years follow-up showed the opposite, i.e. that UVX with Rf solutions containing HPMC  
103 had a better efficacy on visual outcomes than UVX with Rf solutions containing  
104 dextran.<sup>7</sup> In light of the above, we postulate that HPMC may be appropriate for  
105 maintaining the CCT in the process of RGX, but its efficacy on RGX needs to be  
106 explored.

107 The osmotic pressure of the photosensitizer formulation is another important factor that  
108 affects CCT. In some cases, hypotonic Rf formulations have been used to swell very  
109 thin corneas to ensure that they achieve the minimum thickness required for UVX

110 treatment but this efficacy can be transient and unstable due to the endothelial cell  
111 function and the evaporation of corneal surface water.<sup>17, 18, 20</sup> The Rb formulation used  
112 in previously published RGX studies was 0.1% Rb in PBS and the effect of other Rb  
113 formulations on CCT is as yet unknown.

114 The present study aims to explore the CCT changes in rabbit corneas during in vivo  
115 RGX. We also examine the surgical efficacy of different Rb formulations that vary in  
116 terms of their carrier solution, concentration and infiltration time.

117

## 118 **Materials and Methods**

### 119 **Materials**

120 All chemicals used in the preparation of the different Rb formulations (Table 1), were  
121 purchased from Sigma-Aldrich, including Rb, dextran (»500 kDa) and HPMC. The  
122 concentration of Rb in all of the prepared formulations was 0.1% weight/volume. 0.2%  
123 type II collagenase was also purchased from Sigma-Aldrich, prepared as a 0.2%  
124 weight/volume solution in PBS and kept at -4 °C.

125

### 126 **Experimental Animals**

127 Clean grade male Japanese white rabbits (2.5-3 kg) were supplied by the experimental  
128 animal center of Wenzhou Medical University. No abnormal anterior segment was  
129 observed by slit lamp. The feeding environment was good, the food and water were  
130 supplemented regularly. This experiment was granted by the animal ethics committee  
131 of Wenzhou Medical University (NO. wydW 2021-0056). The welfare and use of the  
132 experimental animals complied with the ARRIVE guidelines and were carried out  
133 following the U.K. Animals (Scientific Procedures) Act, 1986 and associated



134 guidelines, EU Directive 2010/63/EU for animal experiments. After treatment, rabbits  
135 were euthanized by inhaling excessive carbon dioxide.

136

#### 137 **Animal grouping**

138 Rabbits were randomly divided into 12 treatment groups, in which the right eye of each  
139 animal was treated with a different combination of the formula of the Rb solution and  
140 infiltration time, and the left eye was de-epithelialized as the untreated control group  
141 (Table 1).

142

#### 143 **Rb infiltration test**

144 Rabbits were anesthetized by intramuscular injection. After topical ocular surficial  
145 anesthesia, the central 8 mm diameter corneal epithelium was removed, and the corneal  
146 surface of each group was completely infiltrated by the corresponding Rb formulation  
147 via a corneal well for either 2 or 20 minutes. After euthanasia, 5 mm diameter central  
148 corneal buttons were trephined and 10  $\mu\text{m}$  frozen sections were cut. Rb fluorescence of  
149 corneal sections were photographed using a Zeiss 710 confocal microscope with an  
150 excitation wavelength of 543 nm and an emission wavelength of 600 nm. ImageJ  
151 v1.51j8 software was used to analyze the Rb fluorescence (n=4).

152

#### 153 **RGX and CCT measurement**

154 After Rb infiltration, the other experimental corneas were irradiated immediately with  
155 0.25 W/cm<sup>2</sup> green light for 600 sec. During this time, a 30 second re-application of the

156 respective Rb formulation was performed at 200 sec and 400 sec of irradiation, and the  
157 cornea was rinsed with PBS at the end of the irradiation procedure. CCTs were  
158 measured with an ultrasound pachymeter (USP; SP-3000, Tomey Corp., Nagoya, Japan)  
159 at the following 7 time points: before de-epithelialization, after de-epithelialization,  
160 after infiltration, after irradiation for 200 sec, 400 sec and 600 sec, and after rinsing.  
161 All CCT measurements were performed 5 times by one well experienced operator and  
162 the average value recorded. After RGX, the rabbits were euthanized and used for further  
163 experiments as follows.

164

#### 165 **Corneal enzymatic resistance test**

166 After euthanasia, an 8 mm diameter central corneal button was trephined from each eye  
167 and digested in 0.2% type II collagenase at a constant temperature of 37 °C. The  
168 undigested corneal buttons were photographed every 2 hours until complete digestion.  
169 The sample areas were calculated using ImageJ software, and area versus time curves  
170 were drawn (n=4).

171

#### 172 **Corneal biomechanics test**

173 After animal euthanasia, the central vertical 3mm width corneal strips with 3mm sclera  
174 were cut with a double-edged knife and placed in a universal testing machine (Model  
175 3343, Instron Corp., Canton, Mass., USA). The strips' initial lengths were set as 10 mm  
176 and the extension rate was set as 2 mm/min. The strips were stretched to a displacement

177 of 1 mm, then returned to displacement of 0, and this was cycled three times with a  
178 recovery of 30 sec between cycles. Finally, the strips were stretched to 20%  
179 deformation. The stress-strain curves were drawn, and the slopes of the curves (i.e. the  
180 Young's modulus) at different strains were calculated by the instrument's software  
181 (n=4).

182

### 183 **Statistical analysis**

184 The data and statistical charts were processed by GraphPad Prism v8.2.1 software (San  
185 Diego, USA). Single factor analysis of variance and multi factor analysis of variance  
186 were used.  $P < 0.05$  indicated statistical significance.

187

## 188 **Results**

### 189 **Rb infiltration test**

190 The presence of HPMC did not affect the infiltration of Rb. The infiltration depths were  
191 about 120  $\mu\text{m}$  in the 2 min groups and 200  $\mu\text{m}$  in the 20 min groups. Both the infiltration  
192 depth and the areas under the fluorescence versus depth curves (AUCs) increased  
193 significantly with the extension of infiltration time (Figure 1 and Table 2). The groups  
194 containing dextran demonstrated the minimal AUCs, with the values being about 10%  
195 that of the other treatment groups with the same infiltration time, and thus were not  
196 included in the follow-up experiments. There was no significant difference of the AUCs  
197 among the other groups with the same infiltration time (Figure 1).

198

199 **CCT changes during RGX**

200 The CCTs (recorded at specific time points during each treatment) minus the CCTs  
201 after de-epithelialization were recorded as  $\Delta$ CCTs. Table 3 and Table 4 show the  
202 CCTs and the  $\Delta$ CCTs of each group at different time points. The average initial CCT  
203 (before de-epithelialization) ranged from 364  $\mu$ m to 372  $\mu$ m (Table 3), and the average  
204 corneal epithelial thickness ranged from 45  $\mu$ m to 55  $\mu$ m among groups (Table 4). There  
205 was no significant difference among the groups ( $P>0.05$ ).

206 As shown by Figure 2, the overall trend in the CCT variation during the RGX process  
207 was that the CCTs increased significantly in all groups after infiltration (about 70  $\mu$ m  
208 in the 2 min groups and 170  $\mu$ m in the 20 min groups) with the exception of the  
209 hypotonic 0.1% Rb groups which showed only a slight increase in CCT (about 30  $\mu$ m)  
210 (Figure 2 A1, A2, B1). The CCT of all the groups decreased significantly during the  
211 first 200 sec of irradiation (Figure 2 A1, A2, B2), and then decreased slowly during the  
212 last 400 sec of irradiation (Figure 2 A1, A2, B3, B4). The groups with HPMC  
213 concentration of 1.1% and 1.7% maintained larger CCTs during irradiation than groups  
214 with other Rb formulations ( $P < 0.05$ ) (Figure 2 A1, A2, B2-4). In all treatment groups,  
215 the CCTs increased after rinsing (Figure 2 A1, A2).

216 There was no significant CCT difference between the 2 min and 20 min infiltration  
217 protocol of 0.1% Rb-water ( $P > 0.05$ ), and the CCTs of the two 0.1% Rb-water groups  
218 were both lower than that of other groups during RGX. Except for these two groups,

219 the CCTs of other formulations in the 20 min groups were significantly greater than 2  
220 min groups during RGX ( $P < 0.05$ ) (Figure 2).

221

### 222 **Corneal enzymatic resistance test**

223 Figure 3 shows groups of photos taken every two hours. The untreated corneas were  
224 digested most rapidly, being completely digested within 6 to 8 hours. The digestion  
225 times in all experimental groups were longer than the untreated control group ( $P < 0.05$ ).  
226 The average digestion time varied from 11.5 to 14 hours in the 2 min groups, and 17 to  
227 19.5 hours in the 20 min groups. Overall, the digestion time of the 20 min groups were  
228 about 5 to 6 hours longer than the 2 min groups with the same formulation ( $P < 0.05$ ,  
229 Table 5). There was no significant difference in the digestion time among experimental  
230 groups with the same infiltration time. Separation of the anterior and posterior stroma  
231 during the enzyme digestion was observed between 6 to 8 hours in some RGX-treated  
232 corneas (Figure 3 A2). Their anterior stroma was able to be maintained in collagenase  
233 solution for a long time, while the posterior stroma was completely digested at a rapid  
234 rate once separated.

235

### 236 **Corneal biomechanics test**

237 According to the above results, cross-linking in the 0.1% Rb-1.1% HPMC 20 min group  
238 showed a good combination of a thick CCT and excellent enzyme resistance, so it was  
239 chosen for the corneal biomechanics test. The 0.1% Rb-PBS group and the untreated

240 group were also included as a routine control and a negative control respectively.  
241 Although hypotonic groups resulted in significant improvements in the resistance of  
242 the cornea to enzyme digestion, they were abandoned because of the steep decline of  
243 CCT during the irradiation procedure. Table 6 and Figure 4 show the Young's modulus  
244 of corneal strips at different strains. At 10% strain, the untreated group had the smallest  
245 average Young's modulus with a value of  $18.95 \pm 2.12$  MPa. The 0.1% Rb-PBS 2 min  
246 group, 0.1% Rb-PBS 20 min group and 0.1% Rb-1.1% HPMC 20 min groups' Young's  
247 moduli were  $32.55 \pm 2.31$  MPa,  $39.80 \pm 1.53$  MPa and  $38.72 \pm 4.50$  MPa, respectively,  
248 i.e. 1.72, 2.10 and 2.04 fold the value of the untreated group, respectively ( $P < 0.05$ ).  
249 0.1% Rb-PBS 2 min was significantly lower than that of 0.1% Rb-PBS 20 min and 0.1%  
250 Rb-1.1% HPMC 20 min ( $P < 0.05$ ). There was no significant difference in Young's  
251 modulus between the last two groups ( $P > 0.05$ ).

252

### 253 **Discussion**

254 UVX cross-links the anterior 250 to 300  $\mu\text{m}$  of the corneal stroma, and increases corneal  
255 stiffness by about 3-fold.<sup>21</sup> However, many keratoconus patients with thin corneas do  
256 not meet the traditional UVX requirement that the de-epithelialized CCT should be  
257 greater than 400  $\mu\text{m}$  to ensure that the UVA irradiance of endothelial cells remains  
258 lower than the toxicity threshold of  $0.35 \text{ mW/cm}^2$ .<sup>22</sup>

259 In a small pilot study, Mark et al.<sup>9</sup> compared UVX with different formulations of Rf  
260 which varied in their type and concentration of carrier solution. They found that the  
261 mean post-treatment CCTs were 1.72, 1.83 and 1.70 folds of the preoperative values in  
262 Rf formulations which contained 0.5%, 1.0% and 1.7% HPMC respectively, while CCT

263 reduced to 0.80 of its initial value when a Rf-10% dextran formulation was used.  
264 Thorsrud et al.<sup>7</sup> found that although the maximum corneal curvature ( $K_{max}$ ) and best-  
265 corrected visual acuity (BCVA) of patients treated with Rf-dextran remained stable at  
266 2-years follow-up, those treated with Rf-HPMC showed significant improvements in  
267 both parameters, suggesting that UVX using Rf-HPMC can produce a deeper stromal  
268 effect. Hammer et al.<sup>23</sup> found in rabbits that the corneal Rf concentration of the Rf-  
269 HPMC groups was 4 to 18 times higher than that of Rf-dextran groups. Similar results  
270 were obtained by Ehmke et al.<sup>24</sup> in porcine corneas.

271 Rb is a halogenated xanthene dye that is often used as a diagnostic agent for corneal  
272 surface damage and is approved by FDA.<sup>25</sup> Both Rb and Rf have been used as oxidative  
273 photosensitizers for photosensitized protein cross-linking. Although their  
274 photophysical properties are similar, Rb associates tightly with collagen whereas Rf  
275 diffuses freely,<sup>25,26</sup> and the effect of formulation components on the permeation of Rb  
276 may be different from that of Rf. We observed that the Rb infiltration depths were about  
277 120  $\mu\text{m}$  after a 2 min infiltration and 200  $\mu\text{m}$  after a 20 min infiltration (Table 2). The  
278 depth of the 2 min group was 20 $\mu\text{m}$  deeper than a previous report,<sup>10</sup> and the depth of  
279 the 20 min group also differed from Wang et. al who found that most Rb was localized  
280 within the superficial 120  $\mu\text{m}$  of the rabbit corneal stroma.<sup>15</sup> This discrepancy is likely  
281 due to differences in the application method, as Wang et. al applied 0.1% Rb at 5 min  
282 intervals over a period of 20-minutes, and then allowed the tissue to absorb it for a  
283 further 10 minutes in the dark, while in this study Rb was applied via a corneal well to  
284 ensure continuous soaking of the corneal surface for 20 minutes without further  
285 absorption. Since the Rb formulation can easily flow away, the more continuous contact  
286 is conducive to its penetration into the cornea. We confirmed that the presence of  
287 HPMC did not affect the infiltration of Rb, while the groups that contained dextran

288 demonstrated the shallowest penetration depth (Figure 1). We speculate that Rb may  
289 bind to dextran physically or chemically, thus hindering its penetration into the cornea.  
290 The specific mechanism needs to be verified by more studies in the future.

291 The green light irradiation energy used in RGX ( $150 \text{ J/cm}^2$  in the current study) is much  
292 higher than the energy of the ultraviolet rays used in UVX. It was reported that the  
293 corneal surface temperature increased by less than  $8 \text{ }^\circ\text{C}$  during the irradiation period<sup>27</sup>.

294 Water evaporation may lead to a significant reduction of CCT during the process of  
295 irradiation, especially in the first 200 seconds. The CCTs of the 0.1% Rb-PBS 2 min  
296 group and the 20 min group were respectively  $(40 \pm 19) \mu\text{m}$  and  $(74 \pm 13) \mu\text{m}$  thinner  
297 after irradiation for 600 sec than after de-epithelialization (Figure 2). The significant

298 reduction may lead to potential safety hazards associated with RGX. HPMC is a non-  
299 ionic cellulose polymer often used as a lubricant in ophthalmology. Wollensak et al.<sup>16</sup>  
300 measured the thickness of the Rf film formed by different Rf formulations on the  
301 corneal surface, and found that the thicknesses were  $300 \mu\text{m}$ ,  $70 \mu\text{m}$  and  $40 \mu\text{m}$  for  
302 Rf-HPMC, Rf-dextran and Rf-saline (Medio-Cross hypotonic solution)solutions,

303 respectively. The good film-forming property of HPMC can prevent water evaporation  
304 from the corneal tissue and the consequent reduction of CCT during irradiation. The  
305 results of our study revealed that Rb-HPMC produced the same RGX efficacy as 0.1%  
306 Rb-PBS formulation whilst also maintaining the thickness of the cornea during  
307 irradiation. These findings indicate that the use of Rb-HPMC may be seen as a  
308 promising modification to the RGX treatment to improve patient safety. Another  
309 important finding of this study was that the groups treated with a hypotonic Rb  
310 formulation had CCTs significantly lower than all other groups during the whole  
311 infiltration and irradiation process; the difference was as high as  $110 \mu\text{m}$  after  
312 irradiation for 600 sec, thus it is not recommended for RGX.

**Commented [SH1]:** Suggest changing the terminology slightly so that it more closely matches what has been used in the introduction.



313 Cherfan et al.<sup>10</sup> showed that an RGX treatment (0.1% Rb-PBS application for 2 min,  
314 150 J/cm<sup>2</sup>) increased the corneal Young's modulus 4.4 fold compared with the  
315 untreated group (16.3±4.08 MPa vs. 3.72 6 ± 1.69 MPa, *P* < 0.05) in fresh young rabbit  
316 eyes. Due to factors such as corneal edema in vitro, the stiffness of their in vitro  
317 untreated group was found to be lower than that of the in vivo untreated corneas. Zhu  
318 et al.<sup>27</sup> found RGX in vivo using the same protocol increased the Young's modulus of  
319 rabbit corneas by a factor of 1.72 on day 1 compared with control untreated corneas  
320 (10.9 ± 3.37 N/mm<sup>2</sup> vs. 6.33 ± 1.38 N/mm<sup>2</sup>, *P* < 0.05). We carried out the biomechanical  
321 testing immediately after RGX, and the increase in Young's modulus was also 1.72-  
322 fold in the 0.1% Rb-PBS 2 min group, consistent with the Zhu et al. study. Besides, the  
323 current study showed that the 0.1% Rb-PBS 20 min group and 0.1% Rb-1.1% HPMC  
324 20 min groups improved the corneal stiffness to 2.10 and 2.04 folds of the untreated  
325 group respectively at 10% strain (*P* < 0.05), and their slight difference was not  
326 statistically significant (Table 6, Figure 4). Our findings suggest that the RGX efficacy  
327 of Rb soaking for 20 minutes was better than for 2 minutes, and the addition of HPMC  
328 did not affect the outcome of surgery.

329 Appropriate intraoperative corneal thickness needs to consider the balance between  
330 safety and efficacy of photosensitized protein cross-linking. Some studies on UVX  
331 suggested that an increase in corneal thickness may deteriorate the cross-linking  
332 efficacy since the percentage of the cross-linked cornea was decreased.<sup>28, 29</sup> However,  
333 the HPMC maintained the CCT (even thicker than before cross-linking) without  
334 blocking Rb penetration or weakening the efficacy of RGX. We speculate that the  
335 reasons may be the high penetration of green light and/or the collagen binding  
336 properties of Rb. The cross-linking was located in the anterior part of the cornea,  
337 confirmed by the fact that the un-cross-linked posterior stroma was easily digested,

338 while the corneal thickening may mainly occur in the middle and posterior part of the  
339 cornea.

340 Unexpectedly, the efficacy of HPMC in maintaining CCT did not increase with the  
341 increase of its concentration, the maximum efficacy was observed at 1.1%  
342 concentration. Furthermore, increasing the HPMC concentration to 1.7% decreased the  
343 Rb infiltration depth and the resulted in a lower CCT during irradiation than that  
344 achieved with the 1.1% concentration. Similar results were found in UVX by Mark et.  
345 al.<sup>9</sup> who increased the HPMC concentration in the Rf drops from 0.5% to 1.0% and  
346 1.7%, with final CCTs of 172%, 183% and 170% in the patient cornea. What is more,  
347 an exorbitant increase in HPMC concentration raises the viscosity of the formulation,  
348 thus reducing its practicality.

349 There were some limitations in the present study. First of all, previous studies of rabbit  
350 corneas at 1 and 28 days after RGX have shown that the corneal stiffness continues to  
351 increase after treatment,<sup>27</sup> but here we only evaluated the immediate efficacy after RGX  
352 without follow-up. Secondly, the Rb infiltration times examined were limited to just 2  
353 and 20-minutes. Although a 20-minute infiltration time of Rb-HPMC resulted in the  
354 greatest RGX efficacy, further studies are warranted to determine the optimal  
355 infiltration time in terms of maximizing the RGX efficacy and minimizing the patient  
356 treatment time.

357

### 358 **Conclusion**

359 In RGX, the CCT increased after infiltration but decreased significantly during  
360 irradiation, especially over the first 200 sec. The addition of HPMC in the Rb  
361 formulation slowed down the reduction of CCT during RGX without affecting either  
362 the infiltration of Rb into the cornea or the cross-linking efficacy. 0.1% Rb-1.1%

363 HPMC infiltration for 20 minutes RGX is likely to have considerable potential for  
364 future clinical applications.

365 **References**

- 366 1. Sharif R, Bak-Nielsen S, Hjortdal J, Karamichos D. Pathogenesis of Keratoconus:  
367 The intriguing therapeutic potential of Prolactin-inducible protein. *Prog Retin Eye Res.*  
368 2018;67:150-67.
- 369 2. Beloshevski B, Shashar S, Mimouni M, Novack V, Malyugin BE, Boiko M,  
370 Knyazer B. Comparison between three protocols of corneal collagen crosslinking in  
371 adults with progressive keratoconus: Standard versus accelerated CXL for keratoconus.  
372 *Eur J Ophthalmol.* 2021;31(5): 2200-05.
- 373 3. Niyazmand H, McKelvie J, Li Y, McLintock C. Comparison of Visual and  
374 Tomographic Outcomes of Epithelium-On and Epithelium-Off Accelerated Corneal  
375 Crosslinking: A Longitudinal Study. *Cornea.* 2021;40(5):643-7.
- 376 4. Spoerl E, Hoyer A, Pillunat LE, Raiskup F. Corneal cross-linking and safety issues.  
377 *Open Ophthalmol J.* 2011;5:14-6.
- 378 5. Vinciguerra R, Pagano L, Borgia A, Montericcio A, Legrottaglie EF, Piscopo R,  
379 Rosetta P, Vinciguerra P. Corneal Cross-linking for Progressive Keratoconus: Up to 13  
380 Years of Follow-up. *J Refract Surg.* 2020;36(12):838-43.
- 381 6. Oltulu R, Satirtav G, Donbaloglu M, Kerimoglu H, Ozkagnici A, Karaibrahimoglu  
382 A. Intraoperative corneal thickness monitoring during corneal collagen cross-linking  
383 with isotonic riboflavin solution with and without dextran. *Cornea.* 2014;33(11):1164-  
384 7.
- 385 7. Thorsrud A, Hagem AM, Sandvik GF, Drolsum L. Superior outcome of corneal

386 collagen cross-linking using riboflavin with methylcellulose than riboflavin with  
387 dextran as the main supplement. *Acta Ophthalmol.* 2019;97(4):415-21.

388 8. Vetter JM, Brueckner S, Tubic-Grozdanis M, Vossmerbaumer U, Pfeiffer N, Kurz  
389 S. Modulation of central corneal thickness by various riboflavin eyedrop compositions  
390 in porcine corneas. *J Cataract Refract Surg.* 2012;38(3):525-32.

391 9. Mark T, Ngounou F, Tamon J, Marx-Gross S, Preussner PR. Modulatory effect of  
392 different riboflavin compositions on the central corneal thickness of African  
393 keratoconus corneas during collagen crosslinking. *Middle East Afr J Ophthalmol.*  
394 2014;21(1):66-71.

395 10. Cherfan D, Verter EE, Melki S, Gisel TE, Doyle FJ, Jr., Scarcelli G, Yun SH,  
396 Redmond RW, Kochevar IE. Collagen cross-linking using rose bengal and green light  
397 to increase corneal stiffness. *Invest Ophthalmol Vis Sci.* 2013;54(5):3426-33.

398 11. Germann JA, Martinez-Enriquez E, Martinez-Garcia MC, Kochevar IE, Marcos S.  
399 Corneal Collagen Ordering After In Vivo Rose Bengal and Riboflavin Cross-Linking.  
400 *Invest Ophthalmol Vis Sci.* 2020;61(3):28.

401 12. Zarei-Ghanavati M. Rose Bengal-Green Light for Collagen Cross-linking. *J*  
402 *Ophthalmic Vis Res.* 2017;12(2):241-2.

403 13. Bekesi N, Kochevar IE, Marcos S. Corneal Biomechanical Response Following  
404 Collagen Cross-Linking With Rose Bengal-Green Light and Riboflavin-UVA. *Invest*  
405 *Ophthalmol Vis Sci.* 2016;57(3):992-1001.

406 14. Gumus K. A New Alternative to Riboflavin/Ultraviolet-A: Collagen Cross-Linking

407 With Rose Bengal/Green Light. *Invest Ophthalmol Vis Sci.* 2016;57(3):1002.

408 15. Wang T, Zhu L, Zhu J, Peng Y, Shen N, Yu Y, Yao M. Subacute effects of rose  
409 Bengal/Green light cross linking on rabbit thin corneal stability and safety. *Lasers Surg*  
410 *Med.* 2018;50(4):324-32.

411 16. Wollensak G, Aurich H, Wirbelauer C, Sel S. Significance of the riboflavin film in  
412 corneal collagen crosslinking. *J Cataract Refract Surg.* 2010;36(1):114-20.

413 17. Raiskup F, Spoerl E. Corneal cross-linking with hypo-osmolar riboflavin solution  
414 in thin keratoconic corneas. *Am J Ophthalmol.* 2011;152(1):28-32 e1.

415 18. Zaheer N, Khan WA, Khan S, Khan MAM. Comparison of Changes in Central  
416 Corneal Thickness During Corneal Collagen Cross-Linking, Using Isotonic Riboflavin  
417 Solutions With and Without Dextran, in the Treatment of Progressive Keratoconus.  
418 *Cornea.* 2018;37(3):340-6.

419 19. Rapuano PB, Mathews PM, Florakis GJ, Trokel SL, Suh LH. Corneal collagen  
420 crosslinking in patients treated with dextran versus isotonic hydroxypropyl  
421 methylcellulose (HPMC) riboflavin solution: a retrospective analysis. *Eye Vis (Lond).*  
422 2018;5:23.

423 20. Kaya V, Utine CA, Yilmaz OF. Intraoperative corneal thickness measurements  
424 during corneal collagen cross-linking with hypoosmolar riboflavin solution in thin  
425 corneas. *Cornea.* 2012;31(5):486-90.

426 21. Chang SH, Mohammadvali A, Chen KJ, Ji YR, Young TH, Wang TJ, Willoughby  
427 CE, Hamill KJ, Elsheikh A. The Relationship Between Mechanical Properties,

428 Ultrastructural Changes, and Intrafibrillar Bond Formation in Corneal UVA/Riboflavin  
429 Cross-linking Treatment for Keratoconus. *J Refract Surg.* 2018;34(4):264-72.

430 22. Jacob S, Kumar DA, Agarwal A, Basu S, Sinha P, Agarwal A. Contact lens-assisted  
431 collagen cross-linking (CACXL): A new technique for cross-linking thin corneas. *J*  
432 *Refract Surg.* 2014;30(6):366-72.

433 23. Hammer A, Rudaz S, Guinchard S, Kling S, Richoz O, Hafezi F. Analysis of  
434 Riboflavin Compounds in the Rabbit Cornea In Vivo. *Curr Eye Res.* 2016;41(9):1166-  
435 72.

436 24. Ehmke T, Seiler TG, Fischinger I, Ripken T, Heisterkamp A, Frueh BE.  
437 Comparison of Corneal Riboflavin Gradients Using Dextran and HPMC Solutions. *J*  
438 *Refract Surg.* 2016;32(12):798-802.

439 25. Redmond RW, Kochevar IE. Medical Applications of Rose Bengal- and  
440 Riboflavin-Photosensitized Protein Crosslinking. *Photochem Photobiol.*  
441 2019;95(5):1097-115.

442 26. Alarcon EI, Poblete H, Roh H, Couture JF, Comer J, Kochevar IE. Rose Bengal  
443 Binding to Collagen and Tissue Photobonding. *ACS Omega.* 2017;2(10):6646-57.

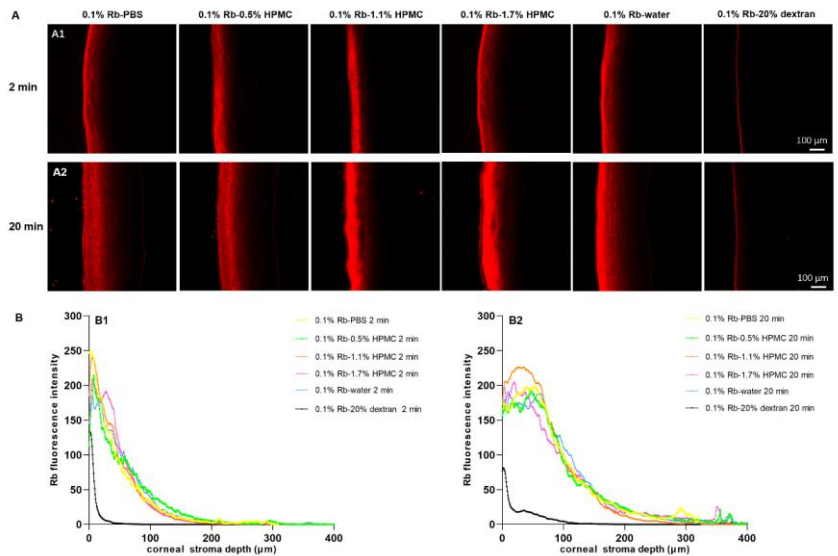
444 27. Zhu H, Alt C, Webb RH, Melki S, Kochevar IE. Corneal Crosslinking With Rose  
445 Bengal and Green Light: Efficacy and Safety Evaluation. *Cornea.* 2016;35(9):1234-41.

446 28. Ozek D, Kemer OE, Ozer PA. Corneal stromal depth of the demarcation line in  
447 'accelerated corneal cross-linking' with different concentrations of riboflavin solutions.  
448 *Int Ophthalmol.* 2019;39(6):1329-35.

449 29. Hafezi F. Limitation of collagen cross-linking with hypoosmolar riboflavin  
450 solution: failure in an extremely thin cornea. *Cornea*. 2011;30(8):917-9.  
451



452 **Figures**



453

454 **Figure 1.** Rb fluorescence distribution in the corneal stroma (n = 4). A) Fluorescence  
455 photos of corneal sections. Rb fluorescence was red, magnification: 10X, scale: 100  
456 μm. B) Rb fluorescence distribution curves of different Rb formulations with the same  
457 infiltration time of 2min (B1) and 20min (B2)

458

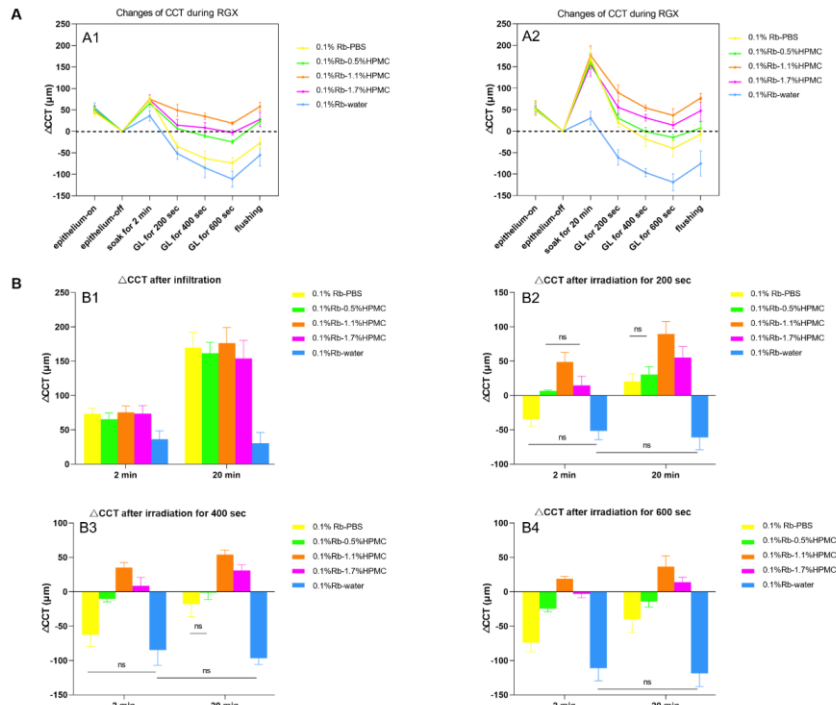
459

460

461

462

463



464

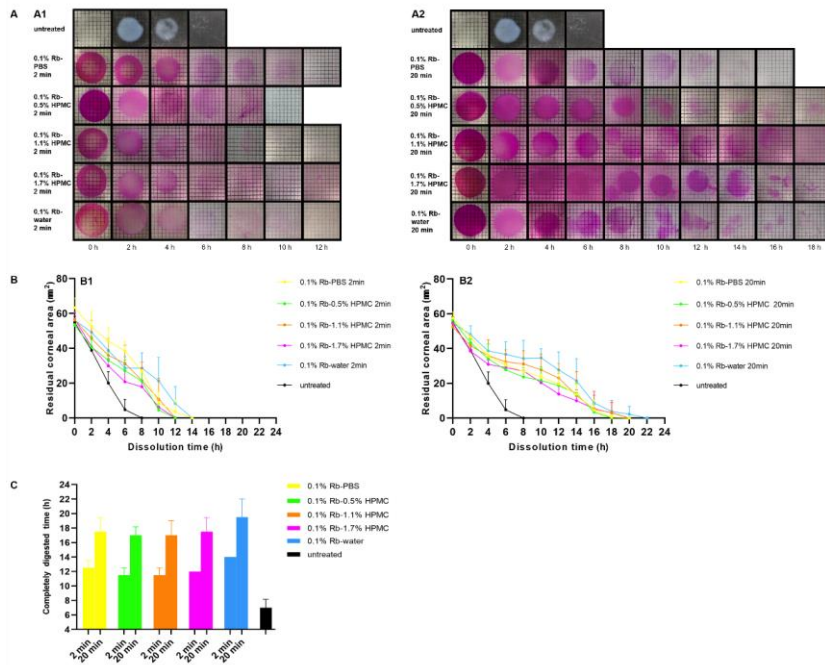
465 **Figure 2.** Changes of CCT during RGX (n = 5). A)  $\Delta$ CCT at different time points  
 466 during RGX in groups with infiltration time of 2 minutes (A1) and 20 minutes (A2). B)  
 467 The average  $\Delta$ CCT of each group after infiltration (B1), after irradiation for 200sec  
 468 (B2), 400sec (B3) and 600sec (B4).

469

470

471

472



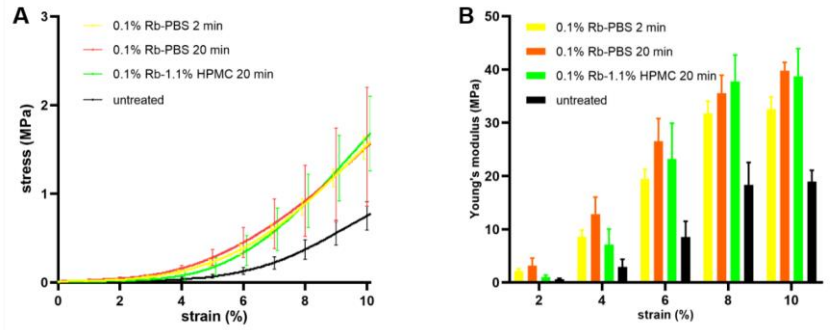
473

474 **Figure 3.** Corneal enzymatic resistance (n = 4). A) Photos of corneal buttons at 2-hour  
 475 intervals. (A1) Photos of 2 min groups and the untreated control group; (A2) Photos of  
 476 20 min groups and the untreated control group. B) Average residual corneal button area  
 477 for each treatment group decreased with time. (B1) Corneal digestion curves of 2 min  
 478 groups and the untreated group; (B2) Corneal digestion curves of 20 min groups and  
 479 the untreated group. C) Comparison of times required for complete digestion (mean ±  
 480 SD).

481

482

483



484

485 **Figure 4.** Corneal biomechanics (n = 4). A) Stress-strain curves of corneas treated with

486 RGX. B) The Young's modulus of corneal strips at different strains.

487

488