

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository: <https://orca.cardiff.ac.uk/id/eprint/161225/>

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

Khan, Sakandar, Younus, Muhammad, Cable, Jo , Hailer, Frank , Idrees, Asif, Imran Rashid, Muhammad and Akbar, Haroon 2023. Epidemiology of Bovine Hydatidosis: Urbanization, dogs, animal care and proximity to slaughterhouses are important risk factors for cattle. *Pakistan Veterinary Journal* 43 (3) , pp. 507-514.
10.29261/pakvetj/2023.055

Publishers page: <https://doi.org/10.29261/pakvetj/2023.055>

Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies. See <http://orca.cf.ac.uk/policies.html> for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.



1 **Research Article**

2

3 **Epidemiology of Bovine Hydatidosis: Urbanization, Dogs, Animal Care and Proximity to**
4 **Slaughterhouses are Important Risk Factors for Cattle**

5 Sakandar Khan¹, Muhammad Younus², Jo Cable¹, Frank Hailer³, Asif Idrees², Muhammad Imran
6 Rashid¹ and Haroon Akbar^{1*}

7 ¹Department of Parasitology, Faculty of Veterinary Science, University of Veterinary and Animal
8 Sciences (UVAS) Lahore, Pakistan

9 ²Department of Pathobiology, College of Veterinary and Animal Sciences, Narowal, Sub-campus,
10 University of Veterinary and Animal Sciences, Pakistan

¹ School of Biosciences, Cardiff University, CF10 3AX, Wales, UK

*Corresponding Author: Email: drharoonakbar@uvas.edu.pk

11 **Statement of novelty:** Only limited data has been available to date on the epidemiology of
12 *Echinococcus granulosus* infecting cattle in the Narowal, Sheikhupura and Sialkot regions of
13 Northern Punjab, Pakistan. We here show that animals (cattle and dogs) kept near slaughterhouses,
14 particularly in urban and semi-urban areas, significantly increased the risk of cystic echinococcosis
15 in cattle.

16

17 **Abstract**

18 Cystic echinococcosis, a neglected tropical disease caused by *Echinococcus granulosus*, is of
19 OneHealth importance. The disease has significant impact on the economy of Pakistan, where
20 livestock is an important pillar of farming. Given the large socio-economic and zoonotic
21 importance of cattle, we explored echinococcosis prevalence in livestock, focussing on three
22 previously littlestudied districts of Punjab (Narowal, Sheikhupura and Sialkot), Pakistan. We
23 screened in total (1168 slaughtered cattle) for presence of hydatid cysts. The collected hydatid cysts
24 were subjected to microscopy, histopathology, and PCR. Overall disease prevalence was 7.7%
25 (n=1168), significantly higher in Narowal (9.6%) than in Sheikhupura (7.6%) and Sialkot (5.7%).
26 The oldest cattle group (>5 years) had significantly higher prevalence (11.8%) than younger
27 animals (6.8% in 3-5-year and 4% in 1-3-year-olds). Females had significantly higher prevalence
28 (9.1%) than male (4.9%) cattle. Significantly more cysts occurred in cattle lungs (71.4%) rather
29 than the liver (28.5%), and the number of fertile cysts was significantly higher in lungs (56.9%)
30 compared to liver (50%). PCR and sequencing of one cyst confirmed the species to be
31 *Echinococcus granulosus*, with phylogenetic analysis clustering our ND1 sequence with the G1-
32 G3 lineages. We found hydatid cysts to cause histopathological changes in the host tissue
33 surrounding the cysts, including atrophy, fibrosis, cell degeneration and leucocytic infiltration.
34 General Linear Models revealed that animals (cattle and dogs) kept near slaughterhouses,
35 particularly in urban and semiurban areas, significantly increased the risk of cystic echinococcosis
36 in cattle. Based on these findings, we recommend a public health campaign to increase awareness
37 of zoonotic infections.

38

39 **Key Words:** *Echinococcus granulosus*, echinococcosis, neglected tropical disease (NTD),
40 zoonoses, cattle disease

41 **Introduction**

42 Livestock are affected by many infectious diseases that adversely reduce their growth and
43 production. Cystic echinococcosis or hydatidosis is one such zoonotic disease, caused by the
44 cestode parasite *Echinococcus granulosus* that infects both animals and humans (Ali, Iqbal, Munir
45 *et al.*, 2015). This is a neglected tropical disease (NTD) of One Health importance. The dog (*Canis*
46 *familiaris*) is the definitive host for *Echinococcus granulosus*, whereas intermediate hosts are
47 herbivores and omnivores including humans. The disease is transmitted to the intermediate host
48 by ingestion of the cestode eggs expelled by the definitive host (Lawson and Gemmell, 1983).
49 Humans act as accidental hosts by acquiring the infection through close contact with dogs, or by
50 ingesting water or food contaminated with parasitic eggs (Nakao, Yanagida, Okamoto *et al.*, 2010).
51 In intermediate hosts, the parasite develops into a fluid filled hydatid cyst, typically in the liver
52 and lungs and rarely in the kidney, brain and bone marrow, resulting in morbidity and mortality
53 (Battelli 2009). In humans, the infection causes bile duct obstruction and pleural fistula disorders
54 (Daali, Fakir, Hssaida *et al.*, 2001).

55 Cystic echinococcosis has a cosmopolitan distribution (Haleem, Niaz, Qureshi *et al.*, 2018),
56 and is highly prevalent in herd keeping areas across the world (Bekele and Butako 2011), ranging
57 from 12% in India (Grakh, Prakash, Mittal *et al.*, 2020), 13.9% in Iran (Vaisi-Raygani,
58 Mohammadi, Jalali *et al.*, 2021) and 22% in Ethiopia (Shumuye, Ohiolei, Gebremedhin *et al.*,
59 2021) to 53.9% in China (Fan, Dong, Ma *et al.*, 2022). Within Pakistan, prevalence in different
60 host species ranges from 2.4 to 65.4% (Tasawar, Naz and Lashari 2014). The parasite is responsible
61 for huge economic losses due to reduced milk and meat production, and condemned meat (Lemma,
62 Abera, Urga *et al.*, 2014). It causes an estimated loss of Rs 26.5 million annually to the livestock
63 sector of Pakistan (Latif *et al.*, 2010), approximately USD 1.65 per organ (Shafiq 2004). X-ray
64 Computed Topography (CT) scans (e.g. Díaz-Menéndez, Pérez-Molina, Norman *et al.*, 2012),
65 ELISA and PCR (Khan *et al.*, 2023) can be exploited for diagnosis of this disease, but to reduce
66 prevalence, further information is needed on the distribution and risk factors associated with *E.*
67 *granulosus*.

68 There is limited data on the epidemiology of cystic echinococcosis in remote areas of
69 Northern Punjab, Pakistan, as well as on the histopathological changes associated with this disease.
70 The present study was thus designed to explore the prevalence, histopathology, and associated risk

71 factors of the causative agent of bovine cystic echinococcosis while sampling slaughtered cattle in
72 three districts of the Punjab Province in Pakistan.

73 **Materials and Methods**

74 *Ethical Approval and sample size determination*

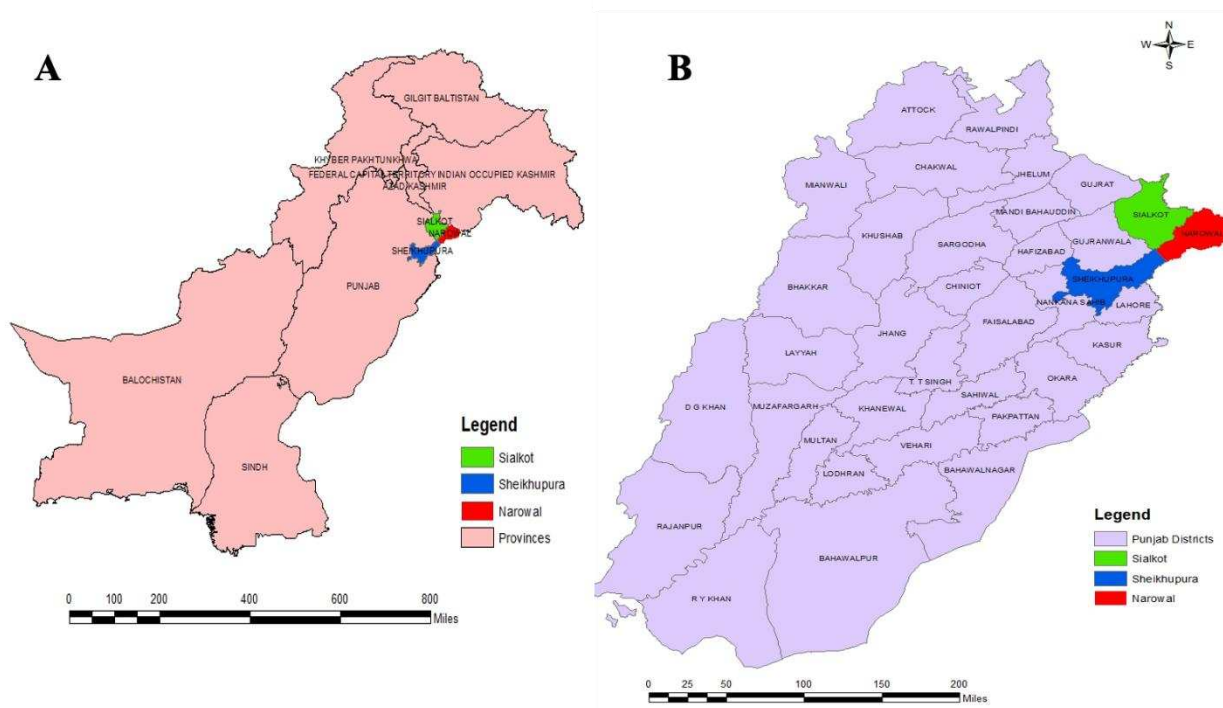
75 All experimental procedures were approved by the Institutional Guidelines of Ethical
76 Review Committee of UVAS, Lahore, vide letter No. 939-1, Dated 05-09-2019.

77 Samples size for determination of *Echinococcus* occurrence was calculated by considering
78 previous prevalence of 45.4% (Shahzad, Abbas, Munir *et al.*, 2014) with 95% confidence interval
79 and 5% absolute precision. The formula used to calculate the sample size (Thrusfield 2018) was:
80 $N = (1.96)^2 P (1-P)/d^2$, where N = required sample size, P is previous prevalence and d is desired
81 absolute precision, such that $N = (3.84) 0.45 (1-0.45) / (0.0025)^2=380$. We collected 400 samples
82 from each district to allow for any samples that might subsequently have to be excluded from data
83 analysis.

84 *Sample collection and questionnaire*

85 From three districts in Northern Punjab, Pakistan (Narowal, Sheikhpura and Sialkot, Fig.
86 1), 1200 cattle (n=400/district) were examined in the main slaughterhouse for each district through
87 random sampling for the presence of hydatid cysts in the liver, lungs, kidneys, and spleen between
88 December 2019 to November 2020. The animals were categorised based on age as <1, 1-3, 3-5
89 and >5 years old. We subsequently excluded animals younger than 1 year from statistical analyses
90 due to the small sample size (31), as cattle of this age are rarely slaughtered. Cysts were collected
91 in sterilized containers with 70% ethanol (for DNA extraction) or 10% neutral buffered formalin
92 (for histology). Samples for histopathology (individual cysts from eight different animals) were
93 processed according to Belina, Demissie, Ashenafi *et al.*, (2015). A questionnaire based on 20
94 simple close-ended questions about owner and animal details was used for risk factor analysis. All
95 1168 farmers who completed the survey (one farmer for each surveyed cattle, and each
96 animal/farmer in this study originated from a different farm) were asked about district of their
97 farm, farm habitat (peri-urban, urban and rural), whether they kept dogs, reason for keeping dogs
98 (guard dog, companion and hunting), if they dewormed their dogs, home slaughtering of animals,
99 disposal of offal (buried, left open or undisclosed), feeding dogs with viscera, disposal of dog
100 faeces, distance from abattoir, deworming of cattle, and animal feeding conditions
101 (confined/mixed/grazing) in a face-to-face discussion at the slaughterhouses. We also recorded the

102 species, age and sex of the animal, and season of the slaughter (spring - March, April, and May;
103 summer - June, July, and August; autumn - September, October, and November; and winter -
104 December, January, and February). Post-slaughter, we recorded the presence of cysts (yes or no),
105 cyst location (liver, lungs or other) and fertility (fertile, sterile, or calcified).



106
107 **Figure 1:** Sampling districts in the current study. (A) Provinces of Pakistan and (B) districts within
108 the province of Punjab. Maps constructed in QGIS software (3.28.1).

109
110 *Molecular parasite identification*

111 Hydatid cysts were characterized according to Haleem *et al.*, (2018). To confirm parasite
112 species, DNA was extracted from one randomly selected cyst sample using a commercially
113 available DNA extraction kit (WizPrep™ gDNA Tissue kit Wizbiosolutions, South Korea,
114 W71060-300). A 226 bp fragment of the mtDNA ND1 gene was targeted by using primers Eg1F81
115 5'-GTT TTT GGC TGC CGC CAGAAC-3', Eg1R83 5'-AAT TAA TGG AAA TAA TAACAA
116 ACT TAA TCA ACA AT-3' (Boufana, Umhang, Qiu *et al.*, 2013). PCR was performed in a T100
117 Thermal Cycler (Bio-Rad, Hercules, CA, USA) as described previously (Mahmood *et al.*, 2022).
118 Briefly, a total reaction volume of 50 µL included 25 µL of 2X AmpMaster™ Taq master mix

119 (GeneAll[®], Exgene[™], catalogue number 541-001), 10 µL Ultrapure[™] DEPC water (Invitrogen,
120 750023), 5 µL each of the forward and reverse primer (50 µM each), and 5 µL DNA extract, with
121 PCR conditions: initial denaturation at 94°C for 3 minutes followed by 28 cycles (denaturation for
122 30 s at 94°C, annealing at 59.8°C for 30 s, extension at 72°C for 1 minute) and final extension step
123 at 72°C for 5 minutes. The PCR product was then run on a 2% agarose gel (1 h at 120 V), stained
124 with SYBR safe DNA (catalogue no. 2291850; Invitrogen, Waltham, MA, USA) and viewed under
125 a transilluminator (Trans Lum SOLO, Biotop China, serial no. 21102053).

126

127 *Sequencing and phylogenetics*

128 The PCR product was sequenced by a commercial sequencing facility (1st BASE Pte Ltd.,
129 Singapore) using the forward primer. The resulting chromatogram was trimmed to delete low
130 quality bases, and the resulting 162 bp sequence submitted to GenBank (Accession Number:
131 OM935772). For comparison, we downloaded 13 ND1 sequences of *Echinococcus* from GenBank,
132 covering the principal lineages of *Echinococcus granulosus* and closely related lineages (Bowles,
133 Blair and McManus 1992). The downloaded ND1 sequences were aligned with our new sequence
134 in Genious Prime (version 2022.2.2; www.geneious.com) using the MUSCLE plugin. A maximum
135 likelihood phylogenetic tree was reconstructed using IQ-tree webserver (version 2.2.0)
136 (Trifinopoulos, Nguyen, von Haeseler *et al.*, 2016). The most suitable substitution model for our
137 alignment was determined by the built-in model finder function (yielding the HKY+I model with
138 empirically determined base frequencies, based on the Bayesian Information Criterion)
139 (Kalyaanamoorthy, Minh, Wong *et al.*, 2017). Statistical support for branches was determined from
140 1000 ultrafast bootstrap replicates (Hoang, Chernomor, Von Haeseler *et al.*, 2018).

141

142 *Statistical Analyses*

143 All statistical analyses were performed using RStudio version 4.2.2. To understand the
144 relationship between the chance of cysts being present within cattle and key environmental
145 variables, we developed binomial generalized linear models (GLMs) with a logit link function,
146 with the dependant variable being the presence or absence of cysts in slaughtered animals. We ran
147 three models to analyse the data. In Model 1, the independent variables were district, age of animal
148 and sex. Model 2 included habitat, deworming practice in cattle, farming type, animal feeding,
149 home slaughtering, disposal of offal, seasons, and distance from abattoir as independent variables.

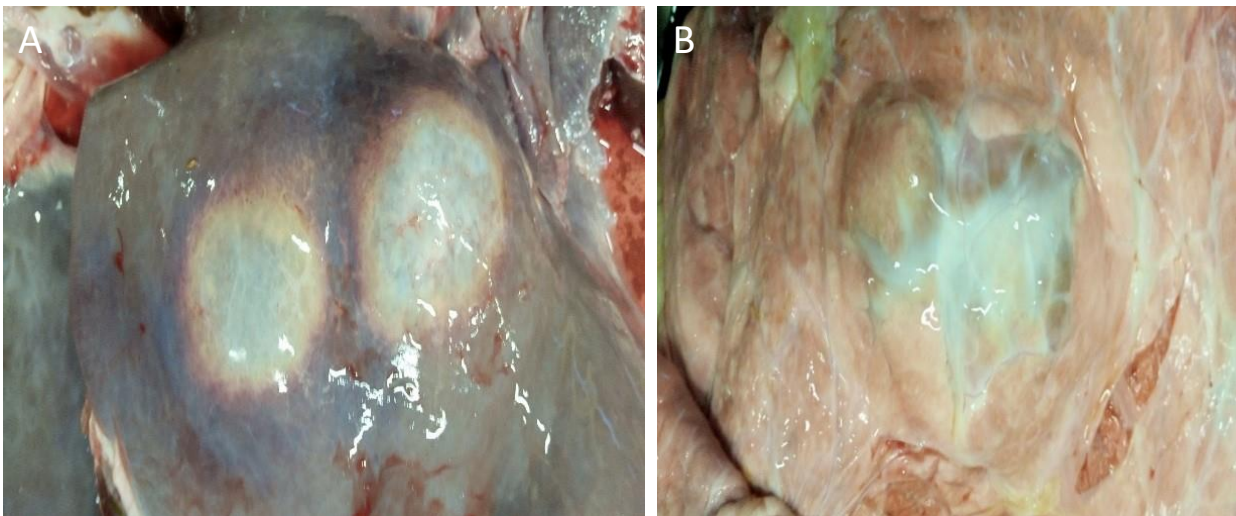
150 Model 3 included keeping of dogs, reasons for keeping dogs, deworming of dogs, feeding dogs
151 with viscera, and disposal of dog faeces. Removal of non-significant terms was performed to
152 ensure model refinement (Thomas *et al.*, 2017). The odds ratios were extracted from GLMs.
153 Finally, Pearson Chi square tests assessed the association between cyst location and fertility. We
154 excluded calcified cysts from the analysis, as only one calcified cyst was recovered.

155

156 **Results**

157 *Epidemiological Study*

158 From a total of 1168 slaughtered cattle from three districts in Pakistan, gross examination
159 revealed 90 (7.7%) were infected with hydatid cysts (Fig. 2A and 2B). Infection rate was
160 significantly higher in the Narowal district (9.6%) followed by Sheikhpura (7.6%) and Sialkot
161 (5.7%) (Table 1). The oldest animals (>5 years) were significantly more prone to infection (11.8%)
162 than those aged 3-5 years (6.8%) and 1-3-years (4%) (Table 1). Males were significantly less
163 infected (4.9%) than females (9.1%) ($p < 0.001$, Table 1). Cysts were more common in lungs
164 (71.4%) compared to liver (28.5%), and cyst fertility was also significantly higher in lungs (56.9%)
165 than liver (50%) (chi-square test, $\chi^2 = 1203.7$, $P < 0.05$). Prevalence was significantly higher in winter
166 (11.2%) compared to autumn (8.0%), spring (6.1%) and summer (5.4%) (Table 1).



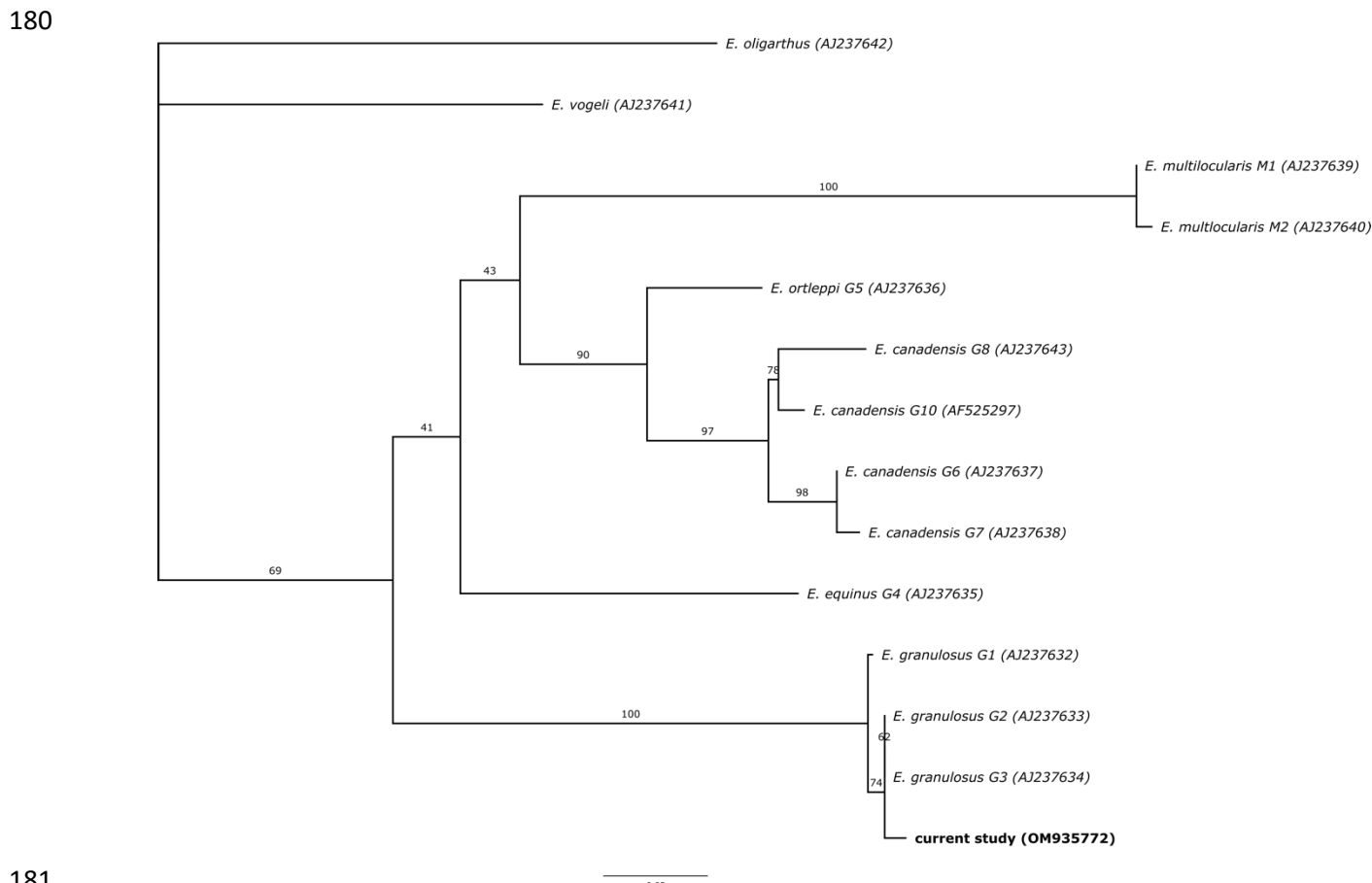
167

168 **Figure 2:** Photographs showing multiple hydatid cysts present in the (A) liver and (B) lungs of
169 cattle, encountered during gross examination at slaughterhouses.

170

171 The species of *Echinococcus* was confirmed through PCR and Sanger sequencing of one
172 sample. Phylogenetic analysis (Fig. 3) revealed that the obtained ND1 sequence clustered with

173 high statistical support (ultrafast bootstrap support: UF=100) within the wider G1/G3 lineage,
 174 identifying it as *E. granulosus sensu stricto* (Vuitton *et al.*, 2020). Our short alignment provided
 175 only limited resolution about clustering within this G1/G3 lineage, suggesting clustering of our
 176 sequence with moderate support (UF=74) with the G2/G3 types. The alignment however contained
 177 one diagnostic site that allowed us to distinguish between G1 and G3, and our sequence grouped
 178 with G3 (at site 82 of our submitted GenBank sequence, G1 had a C, and all other sequences
 179 included in the alignment, including G3 and our sequence, had a T).



181
 182 **Figure 3:** Maximum likelihood phylogeny of ND1 sequences of main lineages of *Echinococcus*
 183 *granulosus (s.l.)* plus outgroups with GenBank accession numbers in brackets, along with the
 184 sequence obtained in the current study. Numbers on branches denote ultrafast bootstrap support
 185 values for the inferred groupings.

186
 187 *Sociodemographic survey regarding disease risk factors*

188 Infection was significantly higher in urban (29.4%), and peri-urban (25.4%) areas
 189 compared to rural locations (5.4%; Table 1). Infection in cattle was significantly higher where

190 farmers kept dogs at home or with other animals; ($p < 0.001$) and higher when these were hunting
191 (30.7%; $p < 0.27$) or companion (69.3%; $p < 0.001$) dogs compared to guard dogs (7.9%), and if the
192 owners dewormed their dogs this decreased prevalence ($p < 0.001$). The practice of home
193 slaughtering did not increase the risk of disease ($p < 0.18$), whereas farmers who improperly
194 disposed of offal ($p < 0.001$) and/or fed dogs with viscera increased the risk of disease ($p < 0.001$).
195 The improper disposal of dog faeces did not increase the infection ($p < 0.98$). Proximity of the home
196 or dairy farm to an abattoir enhanced the risk of disease ($p < 0.001$). There was a higher infection
197 among confined cattle (23/192; 11.9%) compared to those which were kept in mixed conditions
198 (65/917; 7.0%; $p < 0.001$) or grazing (2/59; 3.3%; $p < 0.79$). Surprisingly, deworming of cattle did
199 not significantly ($p < 0.12$) impact prevalence of cystic echinococcosis.

200

200 **Table 1: GLM analyses of prevalence of hydatidosis of slaughtered cattle and risk factors**
 201 **based on data from sociodemographic survey from farmers completed in parallel to sample**
 202 **collection.**

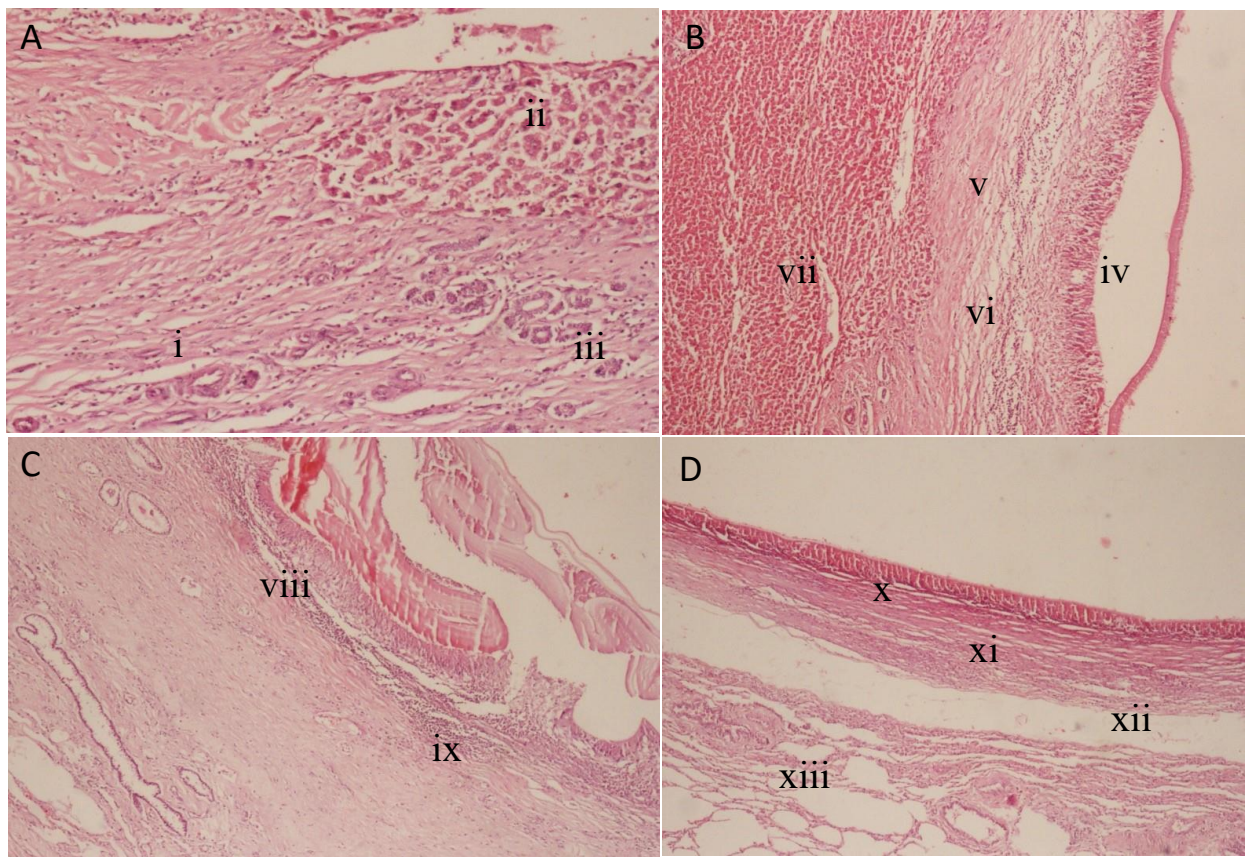
Risk Factors	Response	No. of Positive/ Total (%)	SE	Z. value	Odds ratios	P Value
Models 1 and 2: Risk factors related to location and cattle. Reference categories Narowal and >5 years old, periurban, confined, 10-20 km, buried and autumn.						
Districts	Narowal	38/395 (9.6)				
	Sheikhupura	30/393 (7.6)	0.34	1.01	1.41	0.31
	Sialkot	22/380 (5.7)	0.40	0.55	1.24	0.58
Age (years)	1-3	11/272 (4.04)	0.43	-3.70	0.20	0.001
	3-5	38/541 (6.7)	0.35	-2.38	0.43	0.01
	>5	42/356 (11.79)				
Sex	Male	20/401 (4.97)	0.26	-3.23	0.42	0.001
	Female	70/767 (9.12)				
Habitat	Peri-urban	29/114 (25.43)				
	Rural	56/1037 (5.40)	0.40	-6.79	0.06	0.001
	Urban	5/17 (29.41)	0.67	-1.10	0.47	0.27
Feeding	Confined	23/192 (11.97)				
	Mixed	65/917 (7.08)	0.39	-3.15	0.28	0.001
	Grazing	2/59 (3.38)	0.98	-0.26	0.77	0.79
Deworming cattle	Yes	61/779 (7.83)	0.31	-1.55	0.61	0.12
	No	29/389 (7.45)				
Home slaughtering	Yes	49/409(11.98)	0.31	1.32	1.51	0.18
	No	41/759 (5.40)				
Distance from abattoir	3-5 km	25/183 (13.66)	0.31	3.55	3.01	0.001
	5-10km	12/177 (6.77)	0.50	-2.43	0.29	0.01
	10-20km	53/808 (6.55)				
Disposal of offal	Buried	15/280 (5.35)				
	Left open	34/129 (26.35)	0.33	5.66	6.50	0.001
	Unknown	41/759 (5.40)	0.31	0.02	1.0	0.97
Season of slaughter	Autumn	23/286 (8.04)				
	Spring	18/295 (6.10)	0.36	0.38	1.15	0.70
	Summer	16/293 (5.46)	0.38	-0.14	0.94	0.88
	Winter	33/294 (11.22)	0.33	2.07	2.01	0.03
Model 3: Risk factors related to dogs. Reference categories for reason of keeping dog was guard dog, no dog for deworming and feeding viscera to dogs and unknown for faeces disposal.						
Keeping of dogs	Yes	67/427(15.69)	0.24	7.10	5.89	0.001
	No	23/741 (3.10)				
Reasons for keeping dogs	Guard dog	29/365 (7.94)				
	Companion	34/49 (69.38)	1.29	4.99	6.34	0.001
	Hunting	4/13 (30.76)	2.36	1.09	1.88	0.27
	No dog	23/741 (3.10)	900.88	0.02	2.88	0.97
Deworming of dogs	Yes	41/363 (11.29)	1.33	3.85	5.79	0.001
	No	26/64 (40.62)				
	No dog	23/741 (3.10)				

Feeding dogs with viscera	Yes	52/80 (65)	1.15	4.10	1.12	0.001
	No	15/347 (4.32)				
	No dog	23/741 (3.10)				
Disposal of dog faeces	Yes	16/348 (4.59)	900.8	0.01	4.19	0.98
	No	51/79 (64.55)	900.8	0.02	1.84	0.98
	Unknown	23/741 (3.10)				

203

203 *Histopathology of liver and lungs*

204 In the liver, fibrosis, atrophy of hepatocytes and bile duct hyperplasia were common, and at
205 the cyst lining, infiltration of mononuclear inflammatory cells, proliferation of fibrous connective
206 tissue and atrophy were seen (Fig. 4A and B). In the lungs, cellular degeneration, leucocytic
207 infiltration, proliferation of fibrous connective tissue, infiltration of mononuclear inflammatory
208 cells, cystic wall and atelectasis were observed (Fig. 4C and D).



209
210 **Figure 4: Light microscope images of cattle tissue infected with *Echinococcus granulosus***
211 **stained with haematoxylin and eosin (40X magnification). (A) Liver showing (i) fibrosis, (ii)**
212 **atrophy of hepatocytes and (iii) bile duct hyperplasia. (B) Liver with (iv) cyst lining, (v) infiltration**
213 **of mononuclear inflammatory cells, (vi) proliferation of fibrous connective tissue and (vii) atrophy.**
214 **(C) Lung showing (viii) degeneration of cells and (ix) leucocytic infiltration. (D) Lung with (x)**
215 **proliferation of fibrous connective tissue, (xi) infiltration of mononuclear inflammatory cells, (xii)**
216 **cystic wall and (xiii) atelectasis.**

217

218 Discussion

219 The livestock industry is threatened by many infectious diseases, including cystic
220 echinococcosis that causes significant animal losses (Khan *et al.*, 2023). In Pakistan, favourable
221 socio-economic conditions for hydatidosis and high-level of infection in cattle mean that this is one
222 of the most important diseases for cattle in the area (Fikire *et al.*, 2012). We confirmed with NDI
223 sequencing that *Echinococcus granulosus* is present in cattle in Northern Punjab, Pakistan.
224 Phylogenetic analysis revealed clustering within the G1/G3 lineage (which includes the
225 microvariant G2; Vuitton *et al.*, 2020), with one substitution favouring a clustering with G3 rather
226 than G1. Our findings indicate that the utilized primer pair, which was designed to be specific to
227 the G1 lineage (Boufana *et al.*, 2013), may also amplify the G2/G3 lineages, consistent with
228 lineages G1, G2 and G3 being considered a single clade by Latif *et al.*, (2010). We note, however,
229 the limited phylogenetic resolution provided by our short fragment, so this finding should be
230 reassessed with longer sequences. A recent phylogenetic study showed that although G1 and G3
231 are closely related, the two lineages are clearly diagnoseable with the resolution provided by whole
232 mitogenome DNA sequences (Zhao, Gesang, Wan *et al.*, 2022). The G3 lineage has previously been
233 reported in buffalo from India and China (Bowles *et al.* 1992; Guo *et al.* 2023), and the G1/G3
234 lineages in cattle from Pakistan (Mehmood *et al.*, 2020), camels in Nigeria (Samari *et al.*, 2022),
235 and dogs, sheep, and humans in Uzbekistan (Kim *et al.*, 2020). This highlights that the G1 and G3
236 lineages (at least those identified so far) are present in a variety of intermediate hosts and across a
237 wide geographical distribution.

238 Within Pakistan, 0.71 million cattle suffer from echinococcosis in three North-East districts
239 of the Punjab Province (Narowal, Sheikhupura and Sialkot), but at a lower prevalence (8%) than in
240 North-West (Khyber Pakhtunkhwa; KPK) areas (Haleem *et al.*, 2018; Khan *et al.*, 2021; see Table
241 2). The Punjab is warmer than the KPK, and echinococcosis infection is negatively correlated with
242 temperature; hence there is a lower risk of disease in warmer areas (Piarroux *et al.* 2015). The
243 Punjab is also situated at a low altitude, and altitude is positively correlated with disease occurrence
244 (Giraudoux *et al.*, 2013). We speculate that the increased grassland area in the Punjab with lower
245 cattle densities may result in the lower infection levels compared with the KPK. Other climatic
246 variables and variable landscape features might also contribute to the difference in infection levels,

247 but at a local level contact between animals and animal products is the most important risk factor
248 (Hegglin and Deplazes, 2013).

250 **Table 2: Prevalence of *Echinococcus granulosus* in cattle from two provinces (Punjab and 251**
KPK=Khyber Pakhtunkhwa) and different districts of Punjab, Pakistan.

	Positive/total animals	Prevalence (%)	Reference	252 253
Province				
Punjab	90/1168	7.7	Current study	254
KPK	85/538	15.8	Haleem <i>et al.</i> 2018	255
KPK	41/189	21.7	Khan <i>et al.</i> 2021	
District				256
Narowal	38/395	9.6	Current study	257
Sheikhupura	30/393	7.6	Current study	
Sialkot	22/380	5.7	Current study	258
Multan	105/1179	8.9	Mehmood <i>et al.</i> 2020	259
Sargodha	48/857	5.6	Mehmood <i>et al.</i> 2020	
Islamabad & Rawalpindi	132/3845	3.4	Khan <i>et al.</i> 2020	260 261

262
 263 The risk factors identified in this study indicate that keeping cattle close to a slaughterhouse
 264 enhances the risk of *E. granulosus* infection. Waste in the form of infected offal from the 265
 slaughterhouses can contaminate the surrounding environment for both final and intermediate 266 hosts
 (Otero-Abad and Torgerson, 2013). Cattle from urban and peri-urban areas are also more 267 likely to be
 infected with *E. granulosus* than those from rural habitats (Acosta-Jamett *et al.*, 2010). 268 This may be
 a result of farmers in urban and peri-urban areas living in close contact with canines, 269 the definite host
 for this parasite. Dogs in urban/peri-urban habitats have greater opportunity to 270 ingest infected organs.
 Many butchers discard infected tissues (liver and lungs) inappropriately, 271 increasing the risk for canids
 consuming this meat (Buishi *et al.*, 2006). The rate of infection is 272 higher in dogs whose owners feed
 them with viscera (Otero-Abad and Torgerson, 2013). Also, just 273 keeping dogs with livestock increases
 the chance of echinococcosis (current study; Khan *et al.* 274 2020). Ingestion of eggs from contaminated
 soil is the primary route of echinococcosis infection 275 for intermediate hosts (Shaikenov *et al.*, 2003).
 In the current study, cattle maintained under mixed 276 feeding or confined feeding conditions had a

higher prevalence of disease, probably due to the 277 higher risk of environmental contamination by dogs, compared to grazing cattle.

278 The liver and especially lungs were the most infected organs of cattle (current study; Khan 279 *et al.*, 2023). These highly vascularised organs are ideal for parasite growth, and the rich spongy 280 nature of the lungs is probably more permissive for establishment and maintaining fertility of the 281 oncosphere (Abunna *et al.*, 2012). Not surprisingly, hydatid cysts cause histopathological changes

281 to surrounding host tissues inducing atrophy, degeneration, and inflammatory cell infiltrations
282 (current study; Beigh *et al.*, 2017). Older animals (>5 years) had more cysts and higher prevalence
283 (Khan *et al.* 2023), likely reflecting increased exposure to the parasite over time. In agreement to
284 (Mousa *et al.*, 2015), the seasonal prevalence was higher in winter. The higher prevalence in female
285 compared to male animals might be linked to pregnancy, parturition, and lactation, sometimes
286 leading to malnutrition and transient immunosuppression thus enhancing their susceptibility to
287 infection (Haleem *et al.*, 2018).

288 Deworming of cattle did not appear to reduce the infection rate in the current study. This
289 may be due to use of inappropriate drugs, incorrect administration, drug resistance (Gemmell,
290 Roberts, Beard *et al.*, 2001), or cost. To reduce levels of echinococcosis, it is important to educate
291 farmers about the timing and dosage of the correct cattle dewormers, and to ensure that they have
292 access to effective, affordable dewormers. In agreement with (Mahmood *et al.*, 2022), performing
293 slaughtering at home did not increase the risk of infection in dogs, and reduced offal feeding to
294 dogs reduced infection in cattle (current study; Wilson *et al.* 2019). Deworming of dogs can help
295 control the disease in dogs (current study), which reduces environmental spread of *Echinococcus*
296 eggs (Hegglin and Deplazes, 2013). Seminars/workshops should be arranged for dog owners and
297 other members of the public to increase awareness of echinococcosis and other zoonotic infections.

298 *Conclusion*

299 Cystic echinococcosis, caused by *E. granulosus*, including lineage G1/G3, is prevalent in
300 Northern Punjab, Pakistan (5.7-9.6%) and causes histopathological changes in vital organs. The
301 disease shows significant association with host age and sex, district, and homes and dairy farms
302 close to slaughterhouses. This study also highlights the disease risk of *E. granulosus* transmission
303 between animals and humans if dogs are kept at home or with other animals. The zoonotic impact
304 of echinococcosis needs urgent attention by governments and stakeholders to reduce livestock loss
305 and safeguard public health. A policy on dog keeping and handling, including registration advice
306 on treatment, is needed, as well as control of stray dogs. Slaughterhouses with appropriate disposal
307 pits and obligatory meat inspections are also highly recommended to reduce the prevalence of this
308 common, but preventable, zoonotic disease.

309

310 **Authors' Contribution**

311 HA, MIR and MY designed the experiment. SK performed the field and laboratory experiments,
312 analysed the data, and drafted the manuscript, with input from HA and JC. AI helped with sample
313 collection. FH conducted phylogenetic analyses. HA, JC, and FH reviewed the manuscript.

314 **Acknowledgement**

315 We thank Dr. Ghulam Mustafa for histopathology assistance and Dr. Numair Masud, School of
316 Biosciences, Cardiff University, Wales, UK, for statistical advice and help.

317 **Conflict of interest**

318 The authors declared no conflict of interest.

319 **Funding**

320 S.K. received funding from Higher Education Commission (HEC) under the International
321 Research Support Initiative Program (1-8/HEC/HRD/2022/12612/IRSIP 51 Agri 20); M.Y.
322 received funding from HEC-National Research Program for Universities (NRPU-7018); H.A. and
323 M.I.R. have funding from HEC-Grand Challenge Fund (GCF-273), Pakistan Agriculture Research
324 Board (PARB-18-476) and the Punjab Higher Education Commission (PHEC/ARA/PIRCA/2020-
325 6/8).

326

327 **References**

- 328 Ali I, Iqbal A, Munir I, et al., 2015. Molecular characterization of echinococcus species in Khyber
329 pakhtunkhwa, pakistan. *Acta Sci Vet* 43: 1-7.
- 330 Abunna F, Fentaye S, Megersa B, *et al.*, 2012. Prevalence of bovine hydatidosis in Kombolcha
331 ELFORA abattoir, North Eastern Ethiopia. *Sci Res* 2: 1-6.
- 332 Acosta-Jamett G, Cleaveland S, Barend M, *et al.*, 2010. *Echinococcus granulosus* infection in
333 domestic dogs in urban and rural areas of the Coquimbo region, north-central Chile. *Vet*
334 *Parasitol* 169: 117-122.
- 335 Basinger SC, Khan A, Ahmed H, et al., 2021. Estimation of the monetary burden of treated human
336 cystic echinococcosis in Pakistan. *Acta Trop* 222: 106026.
- 337 Battelli G 2009. Echinococcosis: costs, losses and social consequences of a neglected zoonosis.
338 *Vet Res Commun* 33: 47-52.
- 339 Beigh AB, Darzi MM, Bashir S, *et al.*, 2017. Gross and histopathological alterations associated
340 with cystic echinococcosis in small ruminants. *J Parasit Dis* 41: 1028-1033.

- 341 Bekele J, Butako B 2011. Occurrence and financial loss assessment of cystic echinococcosis
342 (hydatidosis) in cattle slaughtered at Wolayita Sodo municipal abattoir, Southern Ethiopia.
343 Trop Anim Health Prod 43: 221-228.
- 344 Belina D, Demissie T, Ashenafi H, et al., 2015. Comparative pathological study of liver fluke
345 infection in ruminants. Indian J Vet Pathol 39: 113-120.
- 346 Boufana B, Umhang G, Qiu J, et al., 2013. Development of three PCR assays for the differentiation
347 between *Echinococcus shiquicus*, *E. granulosus* (G1 genotype), and *E. multilocularis* DNA
348 in the co-endemic region of Qinghai-Tibet plateau, China Am J Trop Med 88: 795-802.
- 349 Bowles J, Blair D, McManus DP 1992. Genetic variants within the genus *Echinococcus* identified
350 by mitochondrial DNA sequencing. Mol Biochem Parasitol 54: 165-173.
- 351 Buishi I, Njoroge E, Zeyhle E, et al., 2006. Canine echinococcosis in Turkana (north-western
352 Kenya): a coproantigen survey in the previous hydatid-control area and an analysis of risk
353 factors. Ann Trop Med Parasitol 100: 601-610.
- 354 Daali M, Fakir Y, Hssaida R, et al., editors. Annales de chirurgie 2001. DOI: 10.1007/s00268-
355 0047516-z
- 356 Díaz-Menéndez M, Pérez-Molina JA, Norman FF, et al., 2012. Management and outcome of
357 cardiac and endovascular cystic echinococcosis. PLOS Negl Trop Dis 6: 1-8.
- 358 Fan S, Dong H, Ma H, et al., 2022. Meta-analysis on the prevalence of bovine hydatid disease in
359 China from 2000 to 2021. Microb Pathog 168: 1-11.
- 360 Fikire Z, Tolosa T, Nigussie Z, et al., 2012. Prevalence and characterization of hydatidosis in
361 animals slaughtered at Addis Ababa abattoir, Ethiopia. J Parasitol Vector Biol 4: 1-6.
- 362 Gemmell M, Roberts M, Beard T, et al., 2001. Control of echinococcosis. WHO/OIE manual on
363 echinococcosis in humans and animals: a public health problem of global concern. Manual
364 : 286
- 365 Giraudoux P, Raoul F, Pleydell D, et al., 2013. Drivers of *Echinococcus multilocularis*
366 transmission in China: small mammal diversity, landscape or climate? PLOS Negl Trop
367 Dis 7: 1-12
- 368 Grakh K, Prakash A, Mittal D, et al., 2020. Epidemiology, Risk Factors and Economics of
369 Echinococcosis in India: A Review. Int J Livest Res 10: 1-10.

370 Guo B, Zhao L, Zhao L, *et al.*, 2023. Survey and Molecular Characterization of *Echinococcus*
371 *granulosus sensu stricto* from Livestock and Humans in the Altai Region of Xinjiang,
372 China. *Pathogens* 12: 1-134.

373 Haleem S, Niaz S, Qureshi NA, *et al.*, 2018. Incidence, risk factors, and epidemiology of cystic
374 echinococcosis: a complex socioecological emerging infectious disease in Khyber
375 Pakhtunkhwa, Province of Pakistan. *Biomed Res Int* 1-15

376 Hegglin D, Deplazes P 2013. Control of *Echinococcus multilocularis*: Strategies, feasibility and
377 cost–benefit analyses. *Int J Parasitol* 43: 327-337.

378 Hoang DT, Chernomor O, Von Haeseler A, *et al.*, 2018. UFBoot2: improving the ultrafast bootstrap
379 approximation. *Mol Biol Evol* 35: 518-522.

380 Kalyaanamoorthy S, Minh BQ, Wong TK, *et al.*, 2017. ModelFinder: fast model selection for
381 accurate phylogenetic estimates. *Nat Methods* 14: 587-589.

382 Khan A, Ahmed H, Simsek S, *et al.*, 2020. Spread of cystic echinococcosis in Pakistan due to stray
383 dogs and livestock slaughtering habits: research priorities and public health importance.
384 *Public Health Front* 7: 407-412.

385 Khan J, Basharat N, Khan S, *et al.*, 2021. Prevalence and molecular characterization of cystic
386 echinococcosis in livestock population of the Malakand division, Khyber Pakhtunkhwa,
387 Pakistan. *Front Vet Sci* 8: 1-10.

388 Khan S, Cable J, Younus M, *et al.*, 2023. IEg67kDa Bovine Hydatid Cyst Antigen: a candidate for
389 developing sero-diagnostic assays for cystic echinococcosis, a disease of One Health
390 importance. *Animals* 13: 866.

391 Kim H-J, Yong T-S, Shin MH, *et al.*, 2020. Phylogenetic characteristics of *Echinococcus*
392 *granulosus sensu lato* in Uzbekistan. *Korean J Parasitol* 58: 199-205.

393 Latif AA, Tanveer A, Maqbool A, *et al.*, 2010. Morphological and molecular characterisation of
394 *Echinococcus granulosus* in livestock and humans in Punjab, Pakistan. *Vet Parasitol* 170:
395 44-49.

396 Lemma B, Abera T, Urga B, *et al.*, 2014. Prevalence of bovine hydatidosis and its economic
397 significance in Harar municipality abattoir, eastern Ethiopia. *AEJSR* 9: 143-149.

398 Lawson JR, Gemmell M 1983. Hydatidosis and cysticercosis: the dynamics of transmission. *Adv*
399 *in Parasitol* 22: 261-308.

400 Mahmood Q, Younus M, Sadiq S, *et al.*, 2022. Prevalence and Associated Risk Factors of Cystic

401 Echinococcosis in Food Animals-A Neglected and Prevailing Zoonosis. Pak Vet J 42. 5964
402 Mehmood N, Arshad M, Ahmed H, *et al.*, 2020. Comprehensive account on prevalence and
403 characteristics of hydatid cysts in livestock from Pakistan. Korean J Parasitol 58: 121-127.
404 Mousa W, Mahdy O, Abdel-Wahab A, *et al.*, 2015. Epidemiological and serological
405 studies on cystic echinococcosis among camels in Egypt. J. Parasitol. Photon.
406 105: 212-218.

407 Nakao M, Yanagida T, Okamoto M, *et al.*, 2010. State-of-the-art Echinococcus and Taenia:
408 phylogenetic taxonomy of human-pathogenic tapeworms and its application to molecular
409 diagnosis. Infect Genet Evol 10: 444-452.

410 Piarroux M, Gaudart J, Bresson-Hadni S, *et al.*, 2015. Landscape and climatic characteristics
411 associated with human alveolar echinococcosis in France, 1982 to 2007. Euro Surveill 20:
412 21118.

413 Samari H, Laurimäe T, Reghaissia N, *et al.*, 2022. Molecular characterization of *Echinococcus*
414 *granulosus sensu lato* genotypes in dromedary camels from extreme Sahara of Algeria
415 based on analysis of nad2 and nad5 genetic markers. Act Trop 234: 106616.

416 Shafiq T, A., Athar, M 2004. Epidemiology and economical aspects of hydatidosis in different
417 animals, man and its control in sheep with indigenous plants; Ph.D. Thesis. University of
418 the Punjab. 1-653.

419 Shahzad W, Abbas A, Munir R, *et al.*, 2014. A PCR analysis of prevalence of *Echinococcus*
420 *granulosus* genotype G1 in small and large ruminants in three districts of Punjab, Pakistan.
421 Pak J Zoo 46: 1541-1544.

422 Shaikenov B, Torgerson P, Usenbayev A, *et al.*, 2003. The changing epidemiology of
423 echinococcosis in Kazakhstan due to transformation of farming practices. Act Trop 85:
424 287-293.

425 Shumuye NA, Ohiolei JA, Gebremedhin MB, *et al.*, 2021. A systematic review and meta-analysis
426 on prevalence and distribution of *Taenia* and *Echinococcus* infections in Ethiopia. Parasit
427 Vectors 14: 1-22.

428 Tasawar Z, Naz F, Lashari M 2014. The prevalence of hydatidosis in sheep and buffaloes at Multan,
429 Punjab, Pakistan. Glob Vet 12: 332-335.

430 Thrusfield, M 2018. *Veterinary Epidemiology*, 4th edition; John Wiley & Sons: Edinburgh, UK.
431 pp.887

432 Thomas, R., Vaughan, I., Lello, J., 2017. Data analysis with R statistical software: a guidebook for
433 scientists. Newport: Eco explore.

434 Torgerson PR, Macpherson CN 2011. The socioeconomic burden of parasitic zoonoses: global
435 trends. *Vet Parasitol* 182: 79-95.

436 Trifinopoulos J, Nguyen L-T, von Haeseler A, *et al.*, 2016. W-IQ-TREE: a fast online phylogenetic
437 tool for maximum likelihood analysis. *Nucleic Acids Res* 44: 232-235.

438 Vaisi-Raygani A, Mohammadi M, Jalali R, *et al.*, 2021. Prevalence of cystic echinococcosis in
439 slaughtered livestock in Iran: a systematic review and meta-analysis. *BMC Infec Dis* 21:
440 1-10.

441 Vuitton DA, McManus DP, Rogan MT, *et al.*, 2020. International consensus on terminology to be
442 used in the field of echinococcoses. *Parasite* 27: 41.

443 Wilson CS, Jenkins DJ, Brookes VJ, *et al.*, 2019. An eight-year retrospective study of hydatid
444 disease (*Echinococcus granulosus sensu stricto*) in beef cattle slaughtered at an Australian
445 abattoir. *Prev Vet Med* 173: 104806.

446 Yang X-B, Meng X-Z, Zhao Y, *et al.*, 2022. Meta-analysis of the prevalence of bovine cystic
447 echinococcosis in China during decade. *Res Vet Sci* 152: 465-475.

448 Zhao Y, Gesang D, Wan L, *et al.*, 2022. *Echinococcus* spp. and genotypes infecting humans in
449 Tibet Autonomous Region of China: a molecular investigation with nearcomplete/complete
450 mitochondrial sequences. *Parasites & Vectors* 15: 75.