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Investigation of the functional expression of purine and pyrimidine receptors in porcine isolated pancreatic arteries

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Running title: P2 receptors in porcine pancreatic arteries

Receptors for purines and pyrimidines are expressed throughout the cardiovascular system. This study investigated their functional expression in porcine isolated pancreatic arteries. Pancreatic arteries (endothelium intact or denuded) were prepared for isometric tension recording and preconstricted with U46619, a thromboxane A2 mimetic; ADP, UTP and MRS2768, a selective P2Y₂ agonist, were applied cumulatively, while ATP and αβ-meATP response curves were generated from single concentrations per tissue segment. Antagonists/enzyme inhibitors were applied prior to U46619 addition. ATP, αβ-meATP, UTP and MRS2768 induced vasoconstriction, with a potency order of: α,β -meATP > MRS2768 > ATP \geq UTP. Contractions to ATP and α , β -meATP were blocked by NF449, a selective P2X1 receptor antagonist. The contraction induced by ATP, but not UTP, was followed by vasorelaxation. Endothelium removal and DUP 697, a cyclooxygenase-2 inhibitor, had no significant effect on contraction to ATP, but attenuated that to UTP, indicating actions at distinct receptors. MRS2578, a selective P2Y₆ receptor antagonist, had no effect on contractions to UTP. ADP induced endothelium-dependent vasorelaxation which was inhibited by MRS2179, a selective P2Y₁ receptor antagonist, or SCH58261, a selective adenosine A_{2A} receptor antagonist. The contractions to ATP and αβ-meATP were attributed to actions at P2X1 receptors on the vascular smooth muscle, whereas it was shown for the first time that UTP induced an endothelium-dependent vasoconstriction which may involve P2Y2 and/or P2Y₄ receptors. The relaxation induced by ADP is mediated by P2Y₁ and A_{2A} adenosine receptors. Porcine pancreatic arteries appear to lack vasorelaxant P2Y2 and P2Y4 receptors.

Keywords: αβ-meATP, ATP, UTP, ADP, MRS2578, P2Y₁, P2Y₂, P2X1, A_{2A} adenosine receptors, vasoconstriction, relaxation, endothelium.

Abbreviations: αβ-meATP, αβ-methylene-adenosine-5'-triphosphate; ADP, adenosine-5'-diphosphate; ATP, Adenosine-5'-triphosphate; EDCFs, endothelium-derived contractile factors; ENTPDase, ecto-nucleotidase 5'-triphosphate diphosphohydrolase; PPADS, pyridoxal phosphate-6-azo(benzene-2,4-disulfonic acid); UTP, uridine-5'-triphosphate; VSMCs, vascular smooth muscle cells; XAC, xanthine amine congener.

Introduction

The activities of both exocrine and endocrine cells of the pancreas are regulated by autonomic nerves (parasympathetic and sympathetic), as well as by hormones, and autocrine and paracrine mediators. Although the exact mechanisms remain to be established, it is generally agreed that an increase in endocrine cell activity during hormone secretion corresponds with an increase in blood flow, to meet metabolic demand. The role of exogenous purine and pyrimidine nucleotides in controlling the functions of endocrine and exocrine components of the pancreas are well described [1, 2], but little is known about their effects on pancreatic arterial vasocontractility.

There are two main families of P2 purine and pyrimidine receptors, ionotropic P2X and G protein-coupled P2Y receptors. Molecular cloning has identified seven mammalian P2X-receptor subunits: P2X1, P2X2, P2X3, P2X4, P2X5, P2X6 and P2X7 [3], while eight mammalian P2Y receptors have been identified: P2Y1, P2Y2, P2Y4, P2Y6, P2Y11, P2Y12, P2Y13 and P2Y14 [4]. P2X receptors are activated by ATP and its stable, and consequently more potent, analogue αβ-meATP [5, 6]. P2Y receptors can be divided on the basis of their endogenous agonists into adenine nucleotide-preferring (P2Y1, P2Y11, P2Y12 and P2Y13 receptors) and uracil nucleotide or UDP-sugar-preferring (P2Y2, P2Y4, P2Y6 and P2Y14 receptors) [7]. Among the adenine nucleotide group, the human P2Y11 receptor is selectively

activated by ATP and fails to respond to ADP [8], although the dog orthologue responds to both ADP and ATP [9]. P2Y₁, P2Y₁₂, and P2Y₁₃ receptors are activated by ADP, and with lower potency by ATP [10-13]. Among the uracil nucleotide or UDP-sugar receptors, P2Y₂ is equally activated by ATP and UTP, while P2Y₄ receptor is highly selective for UTP over ATP [14]. The P2Y₆ receptor is activated by UDP and UTP, while the P2Y₁₄ receptor is activated by UDP and UDP-sugars [6, 15].

Within the pancreatic vasculature, P2X1, P2X2, P2Y₁ and P2Y₂ receptors were detected by immunohistochemistry [16]. More than two decades ago it was shown that P2X receptors mediate pancreatic vasoconstriction and P2Y receptors mediate vasodilatation in response to ATP [17], and subsequent studies showed an additional involvement of contractile receptors sensitive to UTP (named P2U receptors) [18]. Purine receptor sub-classification has advanced significantly since that time. A re-evaluation of purine receptors in the pancreatic vasculature is clearly warranted. In the current study, we describe the pharmacological characterisation of P2Y₁ and A_{2A} receptor-mediated relaxatory responses, in addition to P2X1, P2Y₂ and P2Y₄ receptor-mediated contractile responses of porcine isolated pancreatic artery preparations. P2Y₂ and/or P2Y₄ receptors appear to be expressed mainly in endothelial cells, while P2X1 and A_{2A} receptors appear to be expressed in smooth muscle cells of the pancreatic arteries. A preliminary account of some of these data has previously been presented to the British Pharmacological Society [19].

Materials and methods

Tissue Preparation

Pancreases from pigs (either sex, age less than 6 mo, wt ~50 kg) were obtained on ice from a local abattoir (G Wood & Sons Ltd, Mansfield). A crude dissection was conducted to isolate

the porcine pancreatic arteries (greater pancreatic artery) which were located in the body of the pancreas. The vessels were dissected out and placed in Krebs'-Henseleit buffer containing 2% (w/v) Ficoll (hydrophilic polysaccharide, type 70) and were refrigerated overnight at 4°C. The next day, a fine dissection was performed on arteries, and the artery segments were cut into rings of about 0.5 cm in length and suspended in Krebs'-Henseleit buffer (gassed, 95% O₂, 5% CO₂).

The endothelium of some arteries was removed by gently rubbing the innermost surface of the artery with forceps on a paper tissue before attaching it to the setup [20]. Successful removal of endothelium was tested using substance P (10 nM). Endothelium-denuded arteries relaxed in response to substance P to less than 10% of the U46619-induced contraction, while in endothelium-intact arteries the relaxation to substance P was $36\% \pm 8$ (n=7, data not shown).

Responses in the porcine isolated pancreatic artery

Arterial rings were mounted onto wires in tissue baths containing warm (37°C), oxygenated Krebs'-Henseleit solution and were connected via isometric force transducers (mechanotransducer MLT 050/D, ADInstruments, Sydney, Australia) to a PC running the computer program, LabChart (ADInstruments, Sydney, Australia). Rings were put under tension (15 g) and allowed to equilibrate for 60 min before assessing viability with two challenges of 75 mM potassium chloride (KCl). The tissues were then allowed to equilibrate for 60 min, after which U46619 (10 - 100 nM), a thromboxane A₂ mimetic, was used to contract the tissues to between 40-80% of the second KCl response. This ensured that if there was a vasodilator component to the response, this could be detected. Once an appropriate level of U46619 response had been achieved, ATP, αβ-meATP, UTP, ADP or MRS2768 were added. Antagonists or enzyme inhibitors were applied 10 min prior to the addition of U46619,

allowing them to be incubated with the tissues for a minimal contact time of 30 min prior to the application of agonists. Some arteries were incubated with 0.1 % (v/v) DMSO (vehicle control).

Reagents and Drugs

Krebs'-Henseleit buffer was composed of the following (mM): NaCl 118, KCl 4.8, CaCl₂.H₂O 1.3, NaHCO₃ 25.0, KH₂PO₄ 1.2, MgSO₄.7H₂O 1.2 and glucose 11.1. Suramin, UTP, ATP, αβmeATP, ADP, U46619, xanthine amine congener (XAC), and SCH58261 (7-(2-phenylethyl)-5-amino-2-(2-furyl)-pyrazolo-[4,3-e]-1,2,4-triazolo[1,5-c]pyrimidine) were purchased from Sigma (Poole, Dorset, UK), while DUP 697 (5-Bromo-2-(4-fluorophenyl)-3-[4-(me-thylsulfonyl)phenyl]-thiophene), pyridoxal phosphate-6-azo(benzene-2,4-disulfonic acid) MRS2578 (N,N''-1,4-butanediyl bis(N'-[3-isothiocynatophenyl)](PPADS), MRS2179 (2'-Deoxy-N⁶-methyladenosine 3',5'-bisphosphate tetrasodium salt), MRS2768 (Uridine-5'-tetraphosphate δ-phenyl ester tetrasodium salt) and substance P were purchased from Tocris Biosciences Ltd. (Bristol, UK). NF449 (4,4',4",4"'-[Carbonylbis(imino-5,1,3-benzenetriyl-bis(carbonylimino))]tetrakis-1,3-benzenedisulfonic acid) was purchased from Calbiochem-Merck4Biosciences. U46619 was dissolved in ethanol at 10 mM stock concentration. PPADS, suramin, αβ-meATP, ATP, ADP, UTP, NF449, MRS2179, MRS2768 and substance P were dissolved in distilled water. DUP 697, XAC, MRS2578 and SCH58261 were dissolved in DMSO at 10 mM stock concentration.

Data Analysis

The contractions to ATP, $\alpha\beta$ -meATP and UTP were measured from the stabilised U46619-induced response and were expressed in g, while the relaxations to ATP and ADP were expressed as a percentage of the U46619-induced contraction. Data were expressed as log concentration-response plots. Values for all figures refer to mean \pm S.E.M with 95% confidence. Results were compared by one-way ANOVA or two-way ANOVA with Bonferroni's post hoc test or unpaired Student's *t*-test (Prism, GraphPad, San Diego, CA, USA). Differences were considered to be significant when the P value was < 0.05. N expresses the number of animals.

Results

Effect of purine and pyrimidine nucleotides on vascular tone in porcine isolated pancreatic arteries

To investigate the effect of purine and pyrimidine nucleotide agonists on porcine pancreatic arteries, $\alpha\beta$ -meATP (10 nM to 100 μ M), ATP (1 μ M to 10 mM), UTP (10 μ M to 1 mM), ADP (1 μ M to 1 mM) and MRS2768 (100 nM to 30 μ M) were applied after preconstriction with U46619. The responses to ATP and $\alpha\beta$ -meATP were found to desensitise rapidly. Therefore, they were applied at single concentrations (one concentration per tissue segment). The responses to UTP, ADP and MRS2768 did not desensitise rapidly, thus cumulative concentration-response curves were generated. ATP, $\alpha\beta$ -meATP, UTP and MRS2768 induced concentration-dependent contraction with a potency order of $\alpha\beta$ -meATP > MRS2768 > ATP \geq UTP (P < 0.001, two-way ANOVA; Fig 1A). The response to ATP was biphasic, since its contraction was followed by a relaxation (Fig 1B) which was equipotent to the concentration-

dependent relaxation produced by ADP (Fig 1A). The efficacies of ATP and $\alpha\beta$ -meATP in inducing contraction were similar, and were greater than that of UTP or MRS2768. The relaxation to ADP and ATP at the highest concentration of the agonists used (1 mM) was similar at 4.5 \pm 0.5g (n=10) and 5.5 \pm 0.2g (n=7) respectively; there was no significant difference between these responses (Fig 1A). UTP, MRS2768 and $\alpha\beta$ -meATP did not elicit vasorelaxation.

Characterisation of responses to ATP and $\alpha\beta$ -meATP in U46619-preconstricted porcine isolated pancreatic arteries:

• Effect of suramin, PPADS and αβ-meATP

Responses to ATP and $\alpha\beta$ -meATP were characterised using the non-selective P2 receptor antagonists suramin (100 μ M) and PPADS (10 μ M). Both suramin and PPADS significantly attenuated the contractions-evoked by ATP (1 mM) and $\alpha\beta$ -meATP (1 μ M), (Fig 2A, 2B). These concentrations of ATP and $\alpha\beta$ -meATP were chosen since they produced robust and submaximal responses, and for $\alpha\beta$ -meATP, the concentration was close to the EC₅₀ value (mean EC₅₀ value was 1.6 μ M (95% confidence interval (CI): 1.05 to 2.53 μ M; n=8; Fig 1A). The relaxation to ATP was not affected by suramin or PPADS (Fig 2C). Since $\alpha\beta$ -meATP induces desensitisation of P2X receptors more readily than ATP because it is broken down more slowly than ATP [5], the responses to ATP and $\alpha\beta$ -meATP were studied in the presence of $\alpha\beta$ -meATP, in which $\alpha\beta$ -meATP (1 μ M) was added 10 min prior the addition of U46619. As seen in Fig 2A and 2B, the contractions to ATP and $\alpha\beta$ -meATP were reduced in the presence of the desensitising agent, while the relaxation to ATP was not affected (Fig 2C).

• Effect of NF449, a selective P2X1 receptor antagonist

Contractile responses to $\alpha\beta$ -meATP suggests expression of P2X1 receptors in porcine pancreatic arteries (Fig 2B). In turn, the involvement of P2X1 receptors in contraction to ATP seems likely because contraction was significantly blocked by $\alpha\beta$ -meATP (Fig 2A). The responses to ATP and $\alpha\beta$ -meATP were studied further in the presence of NF449 (10 μ M), a P2X1 receptor selective antagonist. The contractions to ATP and $\alpha\beta$ -meATP were inhibited in the presence of NF449 (Fig 3).

• Effect of endothelium removal

The response to ATP was tested after the endothelium had been removed. The contraction and the relaxation induced by ATP (Fig 4) were statistically not significantly different in the absence or presence of the endothelium. Similarly, removal of the endothelium had no effects on the contractions to KCl, U46619 or $\alpha\beta$ -meATP; for example, the contraction to 75 mM KCl was $9.5 \pm 0.5g$ in endothelium-intact arteries, while it was $9 \pm 0.5g$ in endothelium-denuded arteries (n=7-9). The contraction to 10 - 100 nM U46619 was $5.5 \pm 0.5g$ in endothelium-intact arteries, while it was $5.8 \pm 0.6g$ in endothelium-denuded arteries (n=12-14). The contraction to 1μ M $\alpha\beta$ -meATP was $3.2 \pm 0.6g$ in endothelium-intact arteries, while it was $3 \pm 0.6g$ in endothelium-denuded arteries (n=6); there was no significant difference between these responses.

• Effect of XAC, an adenosine receptor antagonist

The relaxation to ATP was investigated in the presence of a non-selective adenosine receptor antagonist; XAC (10 μ M) had no effect on the contraction-evoked by ATP (Fig 5A), while it reduced significantly the relaxation to ATP (Fig 5B).

Characterisation of response to UTP in U46619-preconstricted porcine isolated pancreatic arteries:

• Effect of suramin, PPADS, $\alpha\beta$ -meATP and MRS2578, a selective P2Y6 receptor antagonist

The contraction to UTP was examined in the presence of suramin (100 μ M), PPADS (10 μ M), $\alpha\beta$ -meATP (1 μ M) and MRS2578 (10 μ M). Suramin and PPADS significantly reduced the contraction to UTP (Fig 6), while the UTP responses were not affected after P2X receptor desensitisation in the presence of $\alpha\beta$ -meATP (1 μ M) or in the presence of a selective P2Y₆ receptor antagonist (MRS2578); for example, the contraction to 1 mM UTP was $1.8 \pm 0.2g$ in the absence of MRS2578 (n=7), while it was $2.1 \pm 0.2g$ in the presence of MRS2578 (n=6); there was no significant difference between these responses.

• Effect of endothelium removal

The effects of UTP were studied after the endothelium had been removed. The contraction induced by UTP was significantly attenuated in the endothelium-denuded arteries (Fig 7).

• Effect of DUP 697, a cyclooxygenase-2 inhibitor

Because the contraction to UTP was largely endothelium-dependent, the contraction was studied in the presence of DUP 697, a cyclooxygenase-2 (COX-2) inhibitor, since COX-2 facilitates the release of agents which are responsible for endothelium-dependent contraction. DUP 697 (3 μ M) diminished the response to UTP (Fig 8) to a similar extent as removal of the endothelium (Fig 7), while DUP 697 did not alter the contraction to U46619 (the preconstriction agent) or the contraction to ATP (data not shown).

Characterisation of response to ADP in U46619-preconstricted porcine isolated pancreatic arteries:

• Effect of MRS2179, a P2Y₁ receptor selective antagonist, and of endothelium removal

The relaxation to ADP in pancreatic arteries was studied in the presence of MRS2179 (10 μ M), and after the endothelium had been removed. The relaxation to ADP was reduced slightly but significantly in the presence of MRS2179 (Fig 9A) and in the endothelium-denuded arteries (Fig 9B), which indicates the involvement of P2Y₁ receptors and the endothelium in ADP-mediated relaxation of porcine pancreatic arteries.

• Effect of XAC, an adenosine receptor antagonist, and SCH58261, a selective adenosine A_{2A} receptor antagonist

The relaxation to ADP was investigated in the presence of XAC ($10 \mu M$). The relaxation to ADP was largely reduced in the presence of this inhibitor which indicates the involvement of adenosine receptors (Fig 10). To find out about the adenosine subtype involved in the relaxation to ADP, the response to ADP was investigated in the presence of SCH58261, a selective

adenosine A_{2A} receptor antagonist. This antagonist significantly inhibited the relaxation to ADP, to a similar extent as seen with XAC (Fig 10). This showed that the relaxation to ADP involved A_{2A} adenosine receptors.

Discussion

The current report has provided evidence for the functional expression of contractile P2X1, $P2Y_2$ and $P2Y_4$ receptors, and vasorelaxant $P2Y_1$ and A_{2A} adenosine receptors in porcine pancreatic arteries. These receptors are sensitive to the extracellular nucleotides ATP (P2X1), UTP (P2Y₂ and P2Y₄) and ADP (P2Y₁ and A_{2A}). The contraction to ATP was endothelium-independent, while UTP induced an endothelium-dependent contraction which may involve $P2Y_2$ and/or $P2Y_4$ receptors. The relaxation to ADP involved the endothelium and $P2Y_1$ receptors and A_{2A} adenosine receptors.

A vasoconstrictor response elicited by ATP has been reported in a number of different arteries [21-23]. ATP may also induce vasorelaxation depending on the experimental conditions (level of pre-tone) and relative expression of relevant vasocontractile and vasorelaxant receptors [24, 25]. In porcine pancreatic arteries, ATP induced a biphasic response consisting of a contraction followed by a relaxation (Fig 1B). Since the contraction to ATP was rapidly desensitising, non-cumulative concentration response curves were investigated. The contractions to ATP and $\alpha\beta$ -meATP were reduced in the presence of suramin, PPADS, $\alpha\beta$ -meATP (a desensitiser of P2X1 receptors) and NF449 (a P2X1 selective antagonist) (Fig 2A, 2B, 3A, 3B), which indicates that a large part of the contraction to ATP could be attributed to the activation of P2X1 receptors. Moreover, the contractile effect of $\alpha\beta$ -meATP is consistent with expression of P2X1 receptors in porcine pancreatic arteries. $\alpha\beta$ -meATP was more potent than ATP in eliciting vasoconstriction most likely due to its greater stability [5]. Since the contraction to ATP was

not changed after the endothelium had been removed (Fig 4A), the expression of P2X1 receptors was shown to be on the vascular smooth muscle cells (VSMCs). This is consistent with the abundant expression of P2X1 receptors on VSMCs of most tissues [7].

ATP-induced vasorelaxation was not affected after the endothelium had been removed, or in the presence of suramin or PPADS, which suggests that the relaxation to ATP was not due to its action at P2Y receptors. However, the relaxation to ATP was significantly inhibited in the presence of XAC, which suggested an involvement of adenosine receptors expressed on VSMCs of the pancreatic arteries; it is likely that this is due to the activity of adenosine derived from ATP metabolism by ecto-nucleotidase 5′-triphosphate diphosphohydrolase (ENTPDases) enzymes followed by the activity of CD37 and ecto-5′nucleotidase enzymes [26]. Similarly, in rat coronary arteries, the relaxation to ATP involved P1 receptors, although there was an additional involvement of P2Y receptors [24]. In the current study, further investigation of the adenosine receptor subtypes involved in the relaxation to ATP is required. We and others have shown previously a slow relaxation in response to $\alpha\beta$ -meATP in rat mesenteric arteries, subsequent to contraction [27-29], but we did not observe this in the present study in the porcine pancreatic arteries.

The vasoconstriction to UTP did not desensitise quickly, therefore, cumulative concentration response curves were used to study the effect of UTP on pancreatic arteries. This contraction was significantly inhibited by suramin and PPADS (Fig 6), and there was a reduction of the response after the removal of the endothelium (Fig 7). That would indicate for the first time an endothelium-dependent vasoconstriction-evoked by UTP. UTP is known to be active at P2Y₂, P2Y₄ and P2Y₆ receptors [30]. The expression of these receptors in the endothelium and the smooth muscle of vessels has been reported [31]. Since MRS2578 was not able to alter the

contraction to UTP, this indicates that UTP had no action at P2Y₆ receptors. There are currently no commercially available selective antagonists for either P2Y₂ or P2Y₄ receptors. However, we believe that UTP acted at P2Y₄ receptors since the contraction to UTP was significantly inhibited by both endothelium removal and in the presence of DUP 697, but responses to ATP were unaffected. UTP induced-contraction may also be mediated by P2Y₂ receptors, since MRS2768 which is a selective agonist at P2Y₂ receptors and displays no affinity for P2Y₄ or P2Y₆ receptors was able to evoke a contraction in pancreatic arteries [32] (Fig 1A).

UTP-induced vasoconstriction has been documented in a number of arteries including rat pulmonary arteries in which the contraction was attributed to P2Y₂ receptors, and in rabbit basilar arteries in which the contraction to UTP was due to action of P2Y₄ receptors [33, 34]. UTP produced an endothelium-dependent relaxation in rabbit pulmonary arteries and in rat mesenteric arterial bed, but the receptor subtypes were undefined [22, 35]. In bovine middle cerebral arterial strips, UTP had a dual response, it induced a contraction in endothelium-denuded arteries but a relaxation in endothelium-intact arteries [36]. The absence of endothelium-dependent or -independent relaxation to UTP and some other nucleotides in rat renal arteries was reported [37], which is consistent with the current study since there was no evidence of a UTP-mediated relaxation in porcine pancreatic arteries. Hence, porcine pancreatic arteries appear not to express relaxant P2Y₂ and/ or P2Y₄ receptors.

To investigate the mechanism underlying the contraction mediated by UTP in pancreatic arteries, the response to UTP was examined in the presence of DUP 697. As seen in Fig 8, the endothelium-dependent contraction was attenuated in the presence of the selective COX-2 inhibitor. Endothelial cells can release endothelium-derived contractile factors (EDCFs), which may include thromboxane A_2 , prostaglandin $F_{2\alpha}$, leukotrienes and endothelin-1. Thromboxane A_2 and prostaglandin $F_{2\alpha}$ are released from the endothelium due to the activity of COX-2 [38,

39]. The reduction of the contraction to UTP in the presence of DUP 697 indicated the involvement of thromboxane A₂ and prostaglandins in the contraction to UTP. These agents, after being released from the endothelium, may act on their receptors on VSMCs to cause contraction [39]. The different effect of DUP 697 on responses to UTP and ATP further suggests that they are acting on different receptors.

The relaxation to ADP did not desensitise rapidly, therefore, cumulative concentration response curves were used to study the effect of ADP on pancreatic arteries. The relaxation was significantly attenuated by MRS2179, a selective P2Y₁ receptor antagonist (Fig 9A). In addition, the relaxation to ADP was reduced after the endothelium had been removed, by a similar extent as observed in presence of the MRS2179 (Fig 9B). This may suggest that P2Y₁ receptors are expressed on the endothelium. Indeed, a number of reports show that P2Y₁ receptors are expressed on the endothelium and are responsible for the relaxation of arteries, including rat thoracic aortic and porcine mesenteric arteries [40, 41]. The relaxation to ADP in our study was largely reduced in the presence of XAC and SCH58261 (adenosine receptor antagonists). Adenosine receptors may be expressed on the endothelium or the vascular smooth muscle [42]. Since XAC and SCH58261 produced a greater reduction in the relaxation to ADP than the inhibition induced by removal of the endothelium (Fig 10), this suggests that relaxation to ADP involves A2A adenosine receptors expressed, at least in part, on VSMCs. The mechanism by which ADP would produce adenosine to act at the adenosine receptors is still to be elucidated. The simplest explanation is that it is broken down by ENTPDases and by CD37enzymes to adenosine [26]. Alternatively, as suggested in porcine coronary arteries, ADP mediates a relaxation via a mechanism that involves ADP-evoked adenosine release and the subsequent activation of A_{2A} receptors [20]. In contrast to the porcine pancreatic vessels, ADP in rat pancreatic arteries induced a contraction at a high concentration (1 mM); this contraction was similar to that produced by ATP and was much lower than the contraction induced by $\alpha\beta$ -meATP [43]. Further investigation is required to determine the involvement of endothelium-derived relaxing factors or endothelium-derived hyperpolarising factors released from the endothelium in the ADP-induced relaxation.

Reduction in pancreatic blood flow has been observed in acute and chronic pancreatitis and some other pancreatic diseases [44, 45], implicating pancreatic tissue perfusion as an important factor in pathogenesis of pancreatic diseases and symptoms. There is increasing evidence for the role of purinergic signalling in the pathophysiology of the pancreas [2]. Hence, drugs designed to target specific components of purinergic system may be of relevance to the management of pancreatitis, cystic fibrosis, pancreatic cancer and diabetes.

In summary, the functional expression of P2X1 and A_{2A} adenosine receptors on VSMCs, and P2Y₂ and/or P2Y₄ receptors on the endothelium of porcine pancreatic arteries was indicated in the current study. Activation of P2X1 receptors by ATP or $\alpha\beta$ -meATP induced a vasoconstriction, and UTP acts at P2Y₂ and/or P2Y₄ receptors to induce a contraction. ADP and ATP activate A_{2A} adenosine receptors to induce relaxation, together with an action of ADP on P2Y₁ receptors. Pancreatic arteries appear to lack vasorelaxant P2Y₂ and/or P2Y₄ receptors.

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References

- 1. Novak I (2008). Purinergic receptors in the endocrine and exocrine pancreas. Purinergic Signal 4(3): 237-253
- 2. Burnstock G, Novak I (2012). Purinergic signalling in the pancreas in health and disease. Journal of Endocrinology 213(2): 123-141
- 3. Khakh BS, Burnstock G, Kennedy C, King BF, North RA, Séguéla P, et al. (2001). International Union of Pharmacology. XXIV. Current Status of the Nomenclature and Properties of P2X Receptors and Their Subunits. Pharmacological Reviews 53(1): 107-118
- 4. Abbracchio MP, Burnstock G, Boeynaems J-M, Barnard EA, Boyer JL, Kennedy C, et al. (2006). International Union of Pharmacology LVIII: Update on the P2Y G Protein-Coupled Nucleotide Receptors: From Molecular Mechanisms and Pathophysiology to Therapy. Pharmacol Rev 58(3): 281-341
- 5. Kasakov L, Burnstock G (1982). The use of the slowly degradable analog, α,β-methylene ATP, to produce desensitisation of the P2-purinoceptor: Effect on non-adrenergic, non-cholinergic responses of the guinea-pig urinary bladder. European Journal of Pharmacology 86(2): 291-294
- 6. Burnstock G (2006). Pathophysiology and Therapeutic Potential of Purinergic Signaling. Pharmacological Reviews 58(1): 58-86
- 7. Kügelgen I (2008). Pharmacology of mammalian P2X and P2Y receptors. Biotrend Reviews, No. 3.
- 8. Communi D, Govaerts C, Parmentier M, Boeynaems J-M (1997). Cloning of a Human Purinergic P2Y Receptor Coupled to Phospholipase C and Adenylyl Cyclase. Journal of Biological Chemistry 272(51): 31969-31973
- 9. Qi A-D, Zambon AC, Insel PA, Nicholas RA (2001). An Arginine/Glutamine Difference at the Juxtaposition of Transmembrane Domain 6 and the Third Extracellular Loop Contributes to the Markedly Different Nucleotide Selectivities of Human and Canine P2Y₁₁ Receptors. Molecular pharmacology 60(6): 1375-1382
- 10. Léon C, Hechler B, Vial C, Leray C, Cazenave J-P, Gachet C (1997). The P2Y₁ receptor is an ADP receptor antagonized by ATP and expressed in platelets and megakaryoblastic cells. FEBS letters 403(1): 26-30
- 11. Bodor ET, Waldo GL, Hooks SB, Corbitt J, Boyer JL, Harden TK (2003). Purification and Functional Reconstitution of the Human P2Y₁₂ Receptor. Molecular pharmacology 64(5): 1210-1216
- 12. Marteau F, Le Poul E, Communi D, Communi D, Labouret C, Savi P, et al. (2003). Pharmacological Characterization of the Human P2Y₁₃ Receptor. Molecular pharmacology 64(1): 104-112
- 13. Waldo GL, Harden TK (2004). Agonist Binding and Gq-Stimulating Activities of the Purified Human P2Y₁ Receptor. Molecular pharmacology 65(2): 426-436
- 14. Nicholas RA, Lazarowski ER, Watt WC, Li Q, Boyer J, Harden TK (1996). Pharmacological and second messenger signalling selectivities of cloned P2Y receptors. Journal of autonomic pharmacology 16(6): 319-323
- 15. Carter RL, Fricks IP, Barrett MO, Burianek LE, Zhou Y, Ko H, et al. (2009). Quantification of G_i-mediated inhibition of adenylyl cyclase activity reveals that UDP is a potent agonist of the human P2Y₁₄ receptor. Molecular pharmacology 76(6): 1341-1348

- 16. Coutinho-Silva R, Parsons M, Robson T, Lincoln J, Burnstock G (2003). P2X and P2Y purinoceptor expression in pancreas from streptozotocin-diabetic rats. Mol Cell Endocrinol 204(1-2): 141-154
- 17. Hillaire-Buys D, Chapal J, Petit P, Loubatières-Mariani M-M (1991). Dual regulation of pancreatic vascular tone by P2X and P2Y purinoceptor subtypes. Eur J Pharmacol 199(3): 309-314
- 18. Hillaire-Buys D, Dietz S, Chapal J, Petit P, Loubatieres-Mariani MM (1999). [Involvement of P2X and P2U receptors in the constrictor effect of ATP on the pancreatic vascular bed]. Journal de la Societe de biologie 193(1): 57-61
- 19. Alsaqati M, Chan SLF, Ralevic R, (2011). Characterisation of the response to ADP in porcine isolated pancreatic arteries, http://www.pa2online.org/abstracts/vol8issue1abst028p.pdf
- 20. Rayment SJ, Ralevic V, Barrett DA, Cordell R, Alexander SPH (2007). A novel mechanism of vasoregulation: ADP-induced relaxation of the porcine isolated coronary artery is mediated via adenosine release. The FASEB Journal 21(2): 577-585
- 21. Kügelgen I, Häussinger D, Starker K (1987). Evidence for a vasoconstriction-mediating receptor for UTP, distinct from the P2 purinoceptor, in rabbit ear artery. Naunyn-Schmiedeberg's Archives of Pharmacology 336(5): 556-560
- 22. Ralevic V, Burnstock G (1991). Effects of purines and pyrimidines on the rat mesenteric arterial bed. Circulation Research 69(6): 1583-1590
- 23. Rubino A, Burnstock G (1996) Evidence for a P2-purinoceptor mediating vasoconstriction by UTP, ATP and related nucleotides in the isolated pulmonary vascular bed of the rat. British Journal of Pharmacology 118(6):1415-20
- 24. Korchazhkina O, Wright G, Exley C (1999). Intravascular ATP and coronary vasodilation in the isolated working rat heart. British Journal of Pharmacology 127(3): 701-708
- 25. Ralevic V, Burnstock G (1996). Relative contribution of P2U- and P2Y-purinoceptors to endothelium-dependent vasodilatation in the golden hamster isolated mesenteric arterial bed. Br J Pharmacol 117(8): 1797-1802
- 26. Zimmermann H (2000). Extracellular metabolism of ATP and other nucleotides. Naunyn Schmiedebergs Arch Pharmacol 362(4-5): 299-309
- 27. Stanford, S.J. & Mitchell, J.A. (1998). ATP-induced vasodilatation in the rat isolated mesenteric bed exhibits two apparent phases. Br. J. Pharmacol., 125, 94P
- 28. Ralevic V (2001). Mechanism of prolonged vasorelaxation to ATP in the rat isolated mesenteric arterial bed. Br J Pharmacol 132(3): 685-692
- 29. Ralevic V (2002). The involvement of smooth muscle P2X receptors in the prolonged vasorelaxation response to purine nucleotides in the rat mesenteric arterial bed. Br J Pharmacol 135(8): 1988-1994
- 30. Burnstock G, Williams M (2000). P2 purinergic receptors: modulation of cell function and therapeutic potential. The Journal of pharmacology and experimental therapeutics 295(3): 862-869
- 31. Burnstock G (2007). Purine and pyrimidine receptors. Cellular and molecular life sciences: CMLS 64(12): 1471-1483
- 32. Ko H, Carter RL, Cosyn L, Petrelli R, de Castro S, Besada P, et al. (2008). Synthesis and potency of novel uracil nucleotides and derivatives as P2Y₂ and P2Y₆ receptor agonists. Bioorganic & medicinal chemistry 16(12): 6319-6332
- 33. Hartley SA, Kato K, Salter KJ, Kozlowski RZ (1998). Functional evidence for a novel suramin-insensitive pyrimidine receptor in rat small pulmonary arteries. Circ Res 83(9): 940-946

- 34. Miyagi Y, Zhang JH (2004). α,β-Methylene ATP enhances P2Y₄ contraction of rabbit basilar artery. American Journal of Physiology Heart and Circulatory Physiology 286(4): H1546-H1551
- 35. Qasabian RA, Schyvens C, Owe-Young R, Killen JP, Macdonald PS, Conigrave AD, et al. (1997). Characterization of the P2 receptors in rabbit pulmonary artery. Br J Pharmacol 120(4): 553-558
- 36. Miyagi Y, Kobayashi S, Nishimura J, Fukui M, Kanaide H (1996). Dual regulation of cerebrovascular tone by UTP: P2U receptor-mediated contraction and endothelium-dependent relaxation. British Journal of Pharmacology 118(4): 847-856
- 37. Knight GE, Oliver-Redgate R, Burnstock G (2003). Unusual absence of endothelium-dependent or -independent vasodilatation to purines or pyrimidines in the rat renal artery. Kidney international 64(4): 1389-1397
- 38. Mombouli JV, Vanhoutte PM (1993). Purinergic endothelium-dependent and independent contractions in rat aorta. Hypertension 22(4): 577-583
- 39. Wong SL, Leung FP, Lau CW, Au CL, Yung LM, Yao X, et al. (2009). Cyclooxygenase-2-derived prostaglandin F2alpha mediates endothelium-dependent contractions in the aortae of hamsters with increased impact during aging. Circ Res 104(2): 228-235
- 40. Dol-Gleizes F, Mares AM, Savi P, Herbert JM (1999). Relaxant effect of 2-methyl-thio-adenosine diphosphate on rat thoracic aorta: effect of clopidogrel. Eur J Pharmacol 367(2-3): 247-253
- 41. Alefishat E, Alexander S, Ralevic V, (2010). Effect of palmitoyl CoA on ADP-evoked vasorelaxations in porcine isolated coronary and mesenteric arteries http://www.fasebj.org/cgi/content/meeting_abstract/24/1_MeetingAbstracts/lb426
- 42. Schulte G, Fredholm BB (2003). Signalling from adenosine receptors to mitogenactivated protein kinases. Cellular signalling 15(9): 813-827
- 43. Chapal J, Loubatieres-Mariani MM (1983). Evidence for purinergic receptors on vascular smooth muscle in rat pancreas. Eur J Pharmacol 87(4): 423-430
- 44. Satoh A, Shimosegawa T, Satoh K, Ito H, Kohno Y, Masamune A, et al. (2000). Activation of adenosine A₁-receptor pathway induces edema formation in the pancreas of rats. Gastroenterology 119(3): 829-836
- 45. Nguyen NC, Taalab K, Osman MM (2010). Decreased blood flow with increased metabolic activity: a novel sign of pancreatic tumor aggressiveness. Clinical cancer research: an official journal of the American Association for Cancer Research 16(1): 367; author reply 567

Figure Legends

Fig 1. (A) Concentration-dependent contraction of ATP, $\alpha\beta$ -meATP, UTP and MRS2768, a selective P2Y₂ agonist, and relaxation of ADP and ATP in U46619-preconstricted porcine pancreatic arteries (n=7-12). (B) Typical trace showing the biphasic response to ATP (contraction followed by relaxation). Data are presented as mean \pm SEM.

Fig 2. Effect of suramin (100 μM), PPADS (10 μM) and desensitisation by $\alpha\beta$ -meATP (1 μM) on contractions to (A) ATP (1 mM), (B) $\alpha\beta$ -meATP (1 μM), and (C) on the relaxation to ATP in U46619-preconstricted porcine pancreatic arteries. PPADS, suramin and $\alpha\beta$ -meATP reduced the contractions of (A) ATP and (B) $\alpha\beta$ -meATP (**P < 0.01, ***P < 0.001, one-way ANOVA with Bonferroni's post hoc test, responses of ATP or $\alpha\beta$ -meATP vs their responses in the presence of PPADS, suramin or $\alpha\beta$ -meATP, n=6-9). (C) The relaxation to ATP was not significantly different in the absence or in the presence of PPADS, suramin or $\alpha\beta$ -meATP (n=7). Data are presented as mean \pm SEM.

Fig 3. Effect of NF449 (10 μM), a selective P2X1 receptor antagonist, on contractions to (A) ATP (1 mM), (B) $\alpha\beta$ -meATP (1 μM), in U46619-preconstricted porcine pancreatic arteries. NF449 reduced the effects of (A) ATP and (B) $\alpha\beta$ -meATP (***P < 0.001, unpaired Student's *t*-test, n=10-13). Data are presented as mean ± SEM.

Fig 4. Effect of removal of the endothelium on (A) contraction, (B) relaxation to ATP (1 mM) in U46619-preconstricted porcine pancreatic arteries. The effect of the removal of endothelium

on the contraction or relaxation of ATP was not significantly different (n=9-11). Data are presented as mean \pm SEM.

Fig 5. Effect of XAC (10 μ M) on (A) contraction, (B) relaxation to ATP (1 mM) in U46619-preconstricted porcine pancreatic arteries. (A) XAC had no effect on the contraction to ATP (n=8-10), (B) XAC reduced the relaxation to ATP (***P < 0.001, unpaired Student's *t*-test, n=8-10). Data are presented as mean \pm SEM.

Fig 6. Effect of suramin (100 μ M) and PPADS (10 μ M) on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. With suramin and PPADS, effect of UTP concentration (F=16.77, 12.38 respectively, ***P < 0.001); suramin and PPADS reduced the contraction-evoked by UTP (F=14.47, 12.48 respectively, ***P < 0.001, two-way ANOVA; n=9-12). Data are presented as mean \pm SEM.

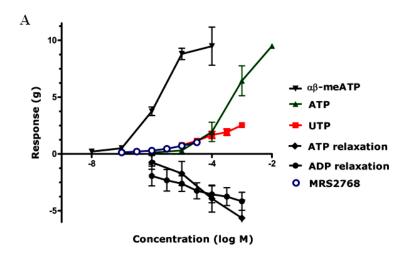
Fig 7. Effect of removal of the endothelium on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. Effect of UTP concentration (F=11.91, ***P<0.001); removal of endothelium reduced the contraction-evoked by UTP (F=43, ***P<0.001, two-way ANOVA; n=10-12). Data are presented as mean \pm SEM.

Fig 8. Effect of DUP 679 (3 μ M), a cyclooxygenase-2 inhibitor, on contraction to UTP in U46619-preconstricted porcine pancreatic arteries. Effect of UTP concentration (F= 8.48, ***P < 0.001); DUP 679 reduced the contraction-evoked by UTP (F=50.8, ***P < 0.001, two-way ANOVA; n=8-12). Data are presented as mean \pm SEM.

Fig 9. Effect of (A) MRS2179 (10 μ M), (B) the removal of the endothelium on relaxation to ADP in U46619-preconstricted porcine pancreatic arteries. With MRS2179 and in endothelium-denuded arteries, effect of ADP concentration (F=21.42, 16.77 respectively, ***P < 0.001); MRS2179 and removal of endothelium reduced the contraction-evoked by ADP (F=21.42, F=32.04 respectively, ***P < 0.001, two-way ANOVA; n=10-12). Data are presented as mean \pm SEM.

Fig 10. Effect of XAC (10 μ M), a non-selective adenosine receptor antagonist, and SCH58261 (1 μ M), a selective adenosine A_{2A} receptor antagonist, on relaxation to ADP in U46619-preconstricted porcine pancreatic arteries. With XAC and SCH58261, effect of ADP concentrations (F=7.14, 6.08 respectively, ***P < 0.001); XAC and SCH58261 reduced the relaxation-evoked by ADP (F=71.19, 58.16 respectively, ***P < 0.001, two-way ANOVA; n=9-14). Data are presented as mean \pm SEM.

Fig 1



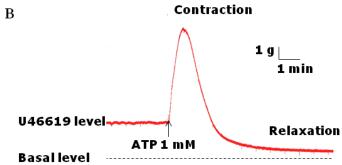
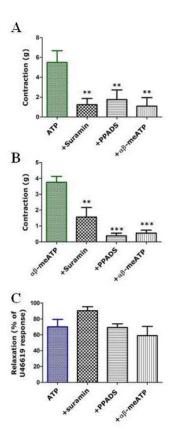


Fig 2



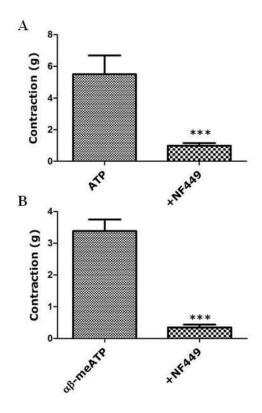
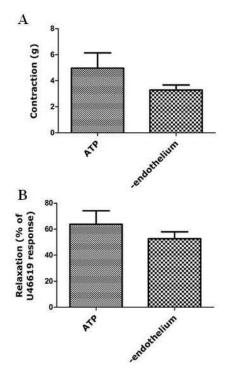
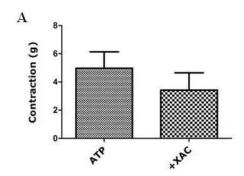


Fig 4





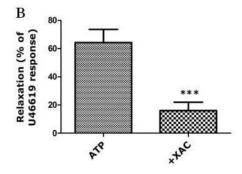


Fig 6

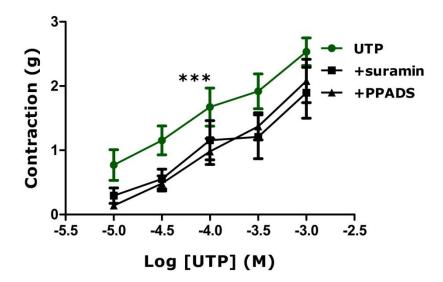


Fig 7

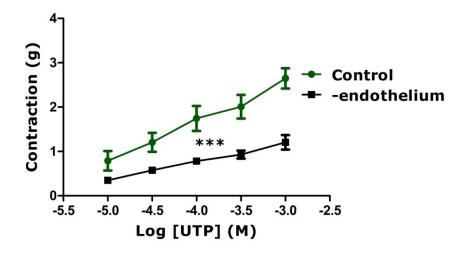


Fig 8

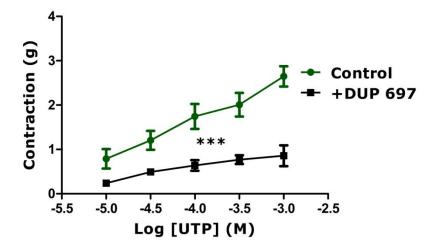


Fig 9

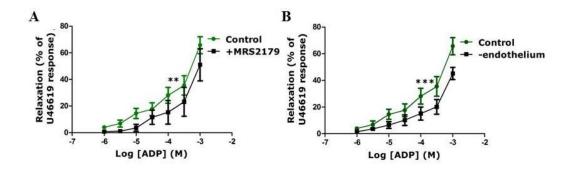


Fig 10

