On BH3 Mimetics and Ca²⁺ Signaling

Pawel E. Ferdek 🗅* and Monika A. Jakubowska 🕩

Medical Research Council Group, Cardiff School of Biosciences, Cardiff University, Cardiff, Wales CF10 3AX, United Kingdom

Strategy, Management and Health Policy							
Enabling Technology, Genomics, Proteomics	Preclinical Research	Preclinical Development Toxicology, Formulation Drug Delivery, Pharmacokinetics	Clinical Development Phases I-III Regulatory, Quality, Manufacturing	Postmarketing Phase IV			

ABSTRACT BH3 mimetics are anticancer agents that reproduce the spatial arrangement of the BH3 domain of Bcl-2 family proteins. Just like the BH3-only proteins, these compounds bind to the hydrophobic cleft of the pro-survival Bcl-2 members such as Bcl-2 or Bcl-xL, and disrupt their heterodimerization with pro-apoptotic Bax or Bak, sensitizing cells to chemotherapy. In recent years, it has become clear that Bcl-2 family proteins are engaged in regulation of intracellular Ca²⁺ homeostasis, including Ca²⁺ release from the intracellular stores as well as Ca²⁺ fluxes across the plasma membrane. Given that BH3 mimetics shift the balance between the prosurvival and proapoptotic Bcl-2 members, they might indirectly exert effects on intracellular Ca²⁺ signals. Indeed, it has been reported that some BH3 mimetics release Ca²⁺ from the intracellular stores causing Ca²⁺ overload in the cytosol. Therefore, the effects of any new BH3 mimetics on cellular Ca²⁺ homeostasis should be tested before these compounds progress to clinical trials. Drug Dev Res 78 : 313-318, 2017. © 2017 The Authors Drug Development Research Published by Wiley Periodicals, Inc.

Key words: Bcl-2; BH3 mimetics; calcium; cell signaling; clinical trials; protein-protein interaction

BACKGROUND

The evolutionary conserved Bcl-2 (B-cell lymphoma 2) protein family consists of about 18 members very well known for their role in the process of programmed cell death. Based on their structure and functions these proteins have been categorized into three groups: (1) the prosurvival members, such as Bcl-2 itself, along with Bcl-xL, Bcl-w or Mcl-1; (2) the proapoptotic proteins (Bax, Bak); (3) and a divergent class of the proapoptotic BH3-only proteins, including Bim, Bid, Puma, Noxa, and others. Prosurvival Bel-2 proteins bear four BH (Bcl-2 homology) domains and usually a transmembrane domain at the C-terminus. Bax and Bak have three BH domains (BH1-BH3) but their helix al somewhat resembles the BH4 domain of BclxL [Suzuki et al., 2000]. And the BH3-only proteins have a single BH3 domain [Chipuk and Green, 2008]. The BH3 domain is an amphipathic α -helix, consisting of 9-16 amino acids with conserved residues of leucine

(Leu) and aspartic acid (Asp) [Aouacheria et al., 2015], that is responsible for the interaction with the hydrophobic cleft formed by BH1–BH3 domains of the prosurvival Bcl-2 proteins [Fesik, 2000; Huang & Strasser, 2000].

*Correspondence Pawel E. Ferdek; Email: p.e.ferdek@ gmail.com

The authors contributed equally to this commentary.

Conflict of interest: The authors declare that they have no conflict of interest.

Published online in Wiley Online Library (wileyonlinelibrary. com). DOI: 10.1002/ddr.21405

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

In healthy cells, Bak is already inserted into the outer mitochondrial membrane, whereas Bax is a cytosolic protein, with the capacity for translocation to the mitochondrial and ER (endoplasmic reticulum) membranes upon activation. According to the current dogma, even activated proapoptotic effectors Bax and Bak can be sequestrated and neutralized by the prosurvival Bcl-2 members [Kroemer et al., 2007]. Upon reception of an apoptotic signal, one or more BH3only proteins undergo transcriptional or posttranscriptional activation. Activated BH3-only proteins either antagonize the prosurvival Bcl-2 members ('sensitizers' or 'de-repressors', e.g., Noxa, Puma, Bad) or also directly act on the proapoptotic effectors ('direct activators', e.g., Bid and Bim), resulting in freeing Bax or Bak. The last two undergo conformational changes and/or insertion (Bax) into the outer mitochondrial membrane followed by oligomerization. This leads to MOMP (mitochondrial outer membrane permeabilization), which is the key event in the intrinsic apoptotic pathway [Kroemer et al., 2007; Chipuk & Green, 2008]. As a result, apoptogenic factors, such as cytochrome c, become released from the mitochondria triggering a downstream cascade of events, including caspase activation [Danial & Korsmeyer, 2004].

Given that increased levels of Bcl-2 proteins have been reported in different cancer types correlating with chemotherapy resistance and poor prognosis [Miyashita & Reed, 1993], Bcl-2 proteins have become a viable target for anticancer therapy. Substantial efforts in this field yielded in development of synthetic compounds binding to the hydrophobic cleft of the pro-survival Bel-2 proteins such as Bcl-2 and Bcl-xL, which results in the inhibition of heterodimerization of the prosurvival and proapoptotic Bcl-2 family members. This leads to the release and activation of Bax and Bak, followed by induction of apoptosis (Fig. 1). Those largely terphenylbased compounds have been termed BH3 mimetics as they reproduce the spatial arrangement of key amino acids in the BH3 domain. In contrast to their prototypes, BH3 peptides, BH3 mimetics are characterized by better stability and therefore have a greater therapeutic potential for controlled inhibition of the pro-survival Bcl-2 members [Lessene et al., 2008].

BH3 MIMETICS

The first BH3 mimetic obtained by molecular modeling and computer screening, HA14-1, was able to displace Bax from Bcl-2 and induce apoptosis *in vitro*, characterized by loss of mitochondrial potential and activation of caspases [Wang et al., 2000]. Soon after, two structurally unrelated groups of BH3

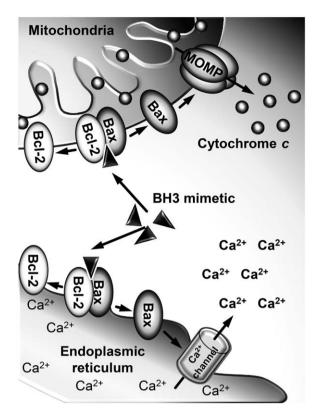


Fig. 1. Schematic illustration of the intracellular effects of BH3 mimetics. BH3 mimetics disrupt the heterodimerization of the prosurvival (e.g. Bcl-2) and pro-apoptotic (e.g. Bax) Bcl-2 members located at different intracellular compartments such as the endoplasmic reticulum or mitochondria. Liberation of the proapoptotic proteins leads to (1) the formation of MOMP (mitochondrial outer membrane permeabilization) followed by the release of cytochrome *c* from mitochondria, (2) as well as the release of Ca²⁺ from the intracellular stores.

inhibitors (BH3Is), derived from BH3I-1 and BH3I-2, were discovered in a fluorescence polarization-based screening [Degterev et al., 2001]. BH3Is were found to displace Bak peptide from Bcl-xL and induce apoptosis characterized by cytochrome *c* release and caspase activation [Degterev et al., 2001]. In the meantime, the anticancer effects of gossypol isolated from the cotton plant (Gossypium) have been attributed to inhibition of Bcl-2, Bcl-xL, and Mcl-1 [Kitada et al., 2003]. In 2005, ABT-737 was developed [Oltersdorf et al., 2005]. This small-molecule inhibitor of Bcl-2, Bcl-xL, and Bcl-w, was two-three orders of magnitude more potent than the previous BH3 mimetics. It did not induce apoptosis on its own, but rather sensitized cells to cell death signals, demonstrating efficacy with chemotherapeutic agents and radiation [Oltersdorf et al., 2005]. The oral bioavailability of this agent was improved even further, resulting in ABT-263 (Navitoclax), a Bad-like BH3 mimetic, capable of triggering Bax translocation, cytochrome c release, and subsequent apoptosis [Tse et al.,

2008]. However, both ABT-737 and ABT-263 were found to induce thrombocytopenia and transient thrombocytopathy that severely hindered their therapeutic use [Schoenwaelder et al., 2011]. Recently, Navitoclax was re-engineered to create a potent, orally bioavailable inhibitor selective for Bcl-2, ABT-199 (Venetoclax) [Souers et al., 2013], which has become the first clinically approved small molecule targeting a protein–protein interaction for treating CLL (chronic lymphocytic leukemia) [Green, 2016]. On-going clinical trials using BH3 mimetics are listed in Table 1.

CALCIUM SIGNALING

Ca²⁺ signaling is one of the most important types of intracellular communication implicated in a wide variety of biological processes, including cell proliferation [Borowiec et al., 2014], migration [Wei et al., 2012], adhesion [Sheng et al., 2013], fertilization [Armant, 2015], muscle contraction [Bers, 2002], neuronal physiology and signal transmission [Brini et al., 2014], exocytosis [Petersen, 1992] and cell death [Criddle et al., 2007]. Therefore, it is not at all surprising that in the past two decades substantial evidence has accumulated for the role of Bcl-2 proteins in the regulation of multiple aspects of the intracellular Ca^{2+} homeostasis [Vervliet et al., 2016]. These proteins have been found not only at the mitochondrial membranes, but are also present in the cytosol, at the nuclear envelope as well as at the ER, the main intracellular Ca^{2+} store [Akao et al., 1994]. They directly interact with Ca^{2+} channels and pumps affecting Ca^{2+} release and the steady state ER Ca^{2+} levels. For example, depending on the site of interaction, Bcl-2 can act either as a direct inhibitor or sensitizer of endoplasmic IP₃Rs (inositol triphosphate receptors) [Rong et al., 2009; Monaco et al., 2012]. The sensitizing effect is also shared by Bcl-xL and Mcl-1 [White et al., 2005; Eckenrode et al., 2010]. Further, Bcl-2 and Bcl-xL can directly bind to RyRs (ryanodine receptors) and inhibit RyR-mediated Ca^{2+} release from the ER [Vervliet et al., 2014; Vervliet et al., 2015]. Bcl-2 may either protect the function of SERCA (sarco/endoplasmic reticulum Ca²⁺-ATPase) [He et al., 1997], or destabilize it [Dremina et al., 2006]. At the mitochondrial membranes, Bcl-2 and Bcl-xL have been demonstrated to directly inhibit mitochondrial Ca²⁺ uptake via VDAC1 (voltage-dependent anion channel 1), a large conductance channel permeable to ions and metabolites [Arbel and Shoshan-Barmatz, 2010; Arbel et al., 2012]; whereas Mcl-1 was shown to have the opposite effect [H. Huang et al., 2014]. Bcl-2 may also inhibit (Na^{+}/Ca^{2+}) NCX mitochondrial exchanger),

increasing Ca^{2+} retention in this organelle [Zhu et al., 2001]. Finally, Bcl-2 can suppress PMCA (plasma membrane Ca^{2+} -ATPase)-mediated Ca^{2+} extrusion with important implications for cell fate [Ferdek et al., 2012].

BH3 MIMETICS AND CALCIUM

Given the above, it might be expected that pharmacological inhibition of the pro-survival Bcl-2 proteins by BH3 mimetics could, in principle, affect the intracellular Ca²⁺ homeostasis. Indeed, the research has demonstrated that the early mimetics, HA14-1 and BH3I-2', caused a slow and complete release of Ca²⁺ from the ER, followed by a sustained elevation of cytosolic Ca2+ concentration in pancreatic acinar cells [Gerasimenko et al., 2010]. Although this effect might be beneficial in cancer, in healthy cells Ca²⁺ overload is undesirable as it promotes cell death, particularly necrosis [Criddle et al., 2007]. This Ca^{2+} release was shown to be attenuated, but not completely blocked, by inhibition of IP₃Rs and RyRs as well as substantially reduced by strong intracellular Ca2+ buffering. Importantly, inhibition of IP₃Rs and RvRs dramatically reduced BH3I-2'elicited apoptosis, indicating that Ca²⁺ release from the ER contributed to cell death induction by this BH3 mimetic [Gerasimenko et al., 2010]. Similar effects of Ca²⁺ deregulation by HA14-1 were also demonstrated in platelets, HeLa and HEK-293T cells [Akl et al., 2013]. A recent study has shed new light on this phenomenon by showing that Ca^{2+} responses induced in pancreatic acinar cells by HA14-1, BH3I-2' and gossypol were largely diminished in the absence of Bax, but not Bak or Bcl-2 [Ferdek et al., 2017], suggesting a regulatory role for Bax in Ca^{2+} release from the intracellular stores (Fig. 1). Of note is that BH3 mimetics in this study caused not only apoptosis, but also substantial levels of necrosis in pancreatic acinar cells, both of which were inhibited by strong Ca²⁺ buffering, again pointing towards a Ca²⁺-dependent component in the mechanism of BH3 mimetic-induced killing. Since global and sustained Ca2+ signals are associated with induction of necrosis, fine tuning of these signals could be useful in shifting unfavorable necrosis towards more physiological apoptosis and thus limiting the side effects of a BH3 mimetic therapy. This has been achieved by CALPs (Ca²⁺-like peptides), which, by binding to the EF-hand motifs, mimic the effects of Ca^{2+} , preactivating various Ca²⁺-sensitive intracellular targets such as calmodulin and Ca2+ channels and pumps [Villain et al., 2000]. CALPs partially reduced Ca^2 responses induced by BH3 mimetics resulting in

BH3-mimetic (Alternative name)	Protein target	Disease target (Additional agent)	Active clinical trial stage			
			I	П		Estimated completion
ABT-199	Bcl-2	AML (Cytarabine)	+			2019
(Venetoclax*,**)		AML (Cobimetinib or Idasanutlin)	+	+		2019
		AML (Azacitidine or Decitabine)	+			2020
		AML (Azacitidine)			+	2022
		Amyloid light chain amyloidosis (Dexamethasone)	+			2021
		B-cell lymphoma (Ibrutinib and Rituximab)	+			2020
		B-cell lymphoma (Obinutuzumab)		+		2020
		B-cell N-HL (Lenalidomide and Obinutuzumab)	+			2021
		CLL (Bendamustine and Obinutuzumab or Bendamustine and Rituximab)	+			2020
		CLL or SLL (Ibrutinib)	+	+		2021
		CLL (Ibrutinib and Obinutuzumab)	+	+		N/A
		CLL (Allopurinol and Ibrutinib)		+		2022
		CLL (–)			+	2022
		CLL (multiple)		+		2023
		CLL (-)		+		2024
		CLL or SLL (Ibrutinib)		+		2024
		Expanded access program for AML, CLL, MM, N-HL (–)				N/A
		FL (Obinutuzumab)	+			2020
		FL (Ibrutinib)	+	+		2021
		FL (Obinutuzumab and Polatuzumab Vedotin)	+	+		2021
		MDS (Azacitidine)		+		2019
		MDS (Azacitidine)	+			2020
		MM (Bortezomib and Dexamethasone)			+	2020
		MM (multiple)	+			2021
		MM (Carfilzomib and Dexamethasone)		+		2021
		N-HL (Ibrutinib)		+		2018
		N-HL (multiple)	+	+		2019
		N-HL (-)	+	+		2019
		Waldenstrom macroglobulinemia (–)		+		2023
ABT-263	Bcl-2	Advanced or metastatic solid tumors (Trametinib)	+	+		N/A
(Navitoclax)	Bcl-xL	CLL or N-HL (Rituximab)	+			2018
	Bcl-w	CLL (-)		+		2018
		Melanoma or solid tumors (Dabrafenib or Trametinib)	+	+		N/A
		Non-small cell lung carcinoma (Osimertinib)	+			N/A
		Ovarian cancer (–)		+		2018
AT-101	Bcl-2	CLL (Lenalidomide)	+	+		2018
(R-(-)-Gossypol acetic acid)	Bcl-xL	Laryngeal cancer (multiple)		+		2018
	McI-1	MM (Dexamethasone and Lenalidomide)	+	+		2021
PNT2258	Bcl-2	B-cell lymphoma (–)		+		2018
S 055746	Bcl-2	AML or MDS (-)	+			2018

TABLE 1. Clinical Trials on BH3 Mimetics (https://clinicaltrials.gov)

AML acute myeloid leukemia; CLL chronic lymphocytic leukemia; FL follicular lymphoma; MDS myelodysplastic syndromes; MM multiple myeloma; N/A not available on May 29, 2017; N-HL Non-Hodgking lymphoma; SLL small lymphocytic lymphoma.

*New drug Venxlexta for CLL in patients with a specific chromosomal abnormality **, approved by The US Food and Drug Administration on April 11, 2016; **an orphan drug designation.

necrosis inhibition or a significant shift in cell death towards apoptosis [Ferdek et al., 2017]. This demonstrates that even a nonspecific inhibition of intracellular Ca^{2+} fluxes can attenuate pathophysiological Ca^{2+} responses and influence the cell death mode and thus may improve the outcome of anticancer therapies.

It is worth noting that not all BH3 mimetics can affect Ca^{2+} homeostasis. A few studies were unable to

demonstrate any significant Ca^{2+} release induced by ABT-737 in platelets and cell lines [Schoenwaelder & Jackson, 2012; Akl et al., 2013] or by ABT-199 in various *in vitro* models [Vervloessem et al., 2017]. It remains unclear why some BH3 mimetics trigger Ca^{2+} release from the intracellular stores, whereas others do not share this effect. Given the strong dependence of Ca^{2+} responses on the presence of Bax, it is rather

unlikely that off-target effects of early BH3 mimetics are entirely responsible for this phenomenon.

CONCLUSION

In conclusion, extensive research on inhibitors of the prosurvival Bcl-2 members yielded a new class of anticancer agents, showing promise particularly against leukemia and lymphoma. Initial excitement, however, slightly faded when the early compounds showed marked side effects. Some of these effects have been attributed to deregulated intracellular Ca²⁺ homeostasis. Despite that, the efforts continued to tailor the specificity of BH3 mimetics in order to preserve the anticancer activity and reduce the undesirable effects. This resulted in ABT-199, the first clinically approved drug targeting a protein-protein interaction [Green, 2016]. Current clinical trials attempt to combine BH3 mimetics with existing chemotherapeutic agents (Table 1). Nevertheless, it might become essential to establish whether any new BH3 mimetic deregulates intracellular Ca²⁺ release in healthy cells. What is more, in order to increase the safety and efficacy of BH3 mimetic drugs, simultaneous application of agents that regulate intracellu- Ca^{2} lar homeostasis might be taken into consideration.

ACKNOWLEDGMENTS

The authors were supported by a Medical Research Council Program Grant MR/J002771/1.

REFERENCES

- Akao Y, Otsuki Y, Kataoka S, Ito Y, Tsujimoto Y. 1994. Multiple subcellular localization of bcl-2: detection in nuclear outer membrane, endoplasmic reticulum membrane, and mitochondrial membranes. Cancer Res 54:2468–2471.
- Akl H, Vandecaetsbeek I, Monaco G, Kauskot A, Luyten T, Welkenhuyzen K, Hoylaerts M, De Smedt H, Parys JB, Bultynck G. 2013. HA14-1, but not the BH3 mimetic ABT-737, causes Ca²⁺ dysregulation in platelets and human cell lines. Haematologica 98:e49–e51.
- Aouacheria A, Combet C, Tompa P, Hardwick JM. 2015. Redefining the BH3 death domain as a 'short linear motif'. Trends Biochem Sci 40:736–748.
- Arbel N, Ben-Hail D, Shoshan-Barmatz V. 2012. Mediation of the antiapoptotic activity of Bcl-xL protein upon interaction with VDAC1 protein. J Biol Chem 287:23152–23161.
- Arbel N, Shoshan-Barmatz V. 2010. Voltage-dependent anion channel 1-based peptides interact with Bcl-2 to prevent antiapoptotic activity. J Biol Chem 285:6053–6062.
- Armant DR. 2015. Intracellular Ca²⁺ signaling and preimplantation development. Adv Exp Med Biol 843:151–171.
- Bers DM. 2002. Cardiac excitation–contraction coupling. Nature 415:198–205.

- Borowiec AS, Bidaux G, Pigat N, Goffin V, Bernichtein S, Capiod T. 2014. Calcium channels, external calcium concentration and cell proliferation. Eur J Pharmacol 739:19–25.
- Brini M, Cali T, Ottolini D, Carafoli E. 2014. Neuronal calcium signaling: function and dysfunction. Cell Mol Life Sci 71:2787–2814.
- Chipuk JE, Green DR. 2008. How do BCL-2 proteins induce mitochondrial outer membrane permeabilization? Trends Cell Biol 18:157–164.
- Criddle DN, Gerasimenko JV, Baumgartner HK, Jaffar M, Voronina S, Sutton R, Petersen OH, Gerasimenko OV. 2007. Calcium signalling and pancreatic cell death: apoptosis or necrosis? Cell Death Differ 14:1285–1294.
- Danial NN, Korsmeyer SJ. 2004. Cell death: critical control points. Cell 116:205–219.
- Degterev A, Lugovskoy A, Cardone M, Mulley B, Wagner G, Mitchison T, Yuan J. 2001. Identification of small-molecule inhibitors of interaction between the BH3 domain and Bcl-xL. Nat Cell Biol 3:173–182.
- Dremina ES, Sharov VS, Schoneich C. 2006. Displacement of SERCA from SR lipid caveolae-related domains by Bcl-2: a possible mechanism for SERCA inactivation. Biochemistry 45: 175–184.
- Eckenrode EF, Yang J, Velmurugan GV, Foskett JK, White C. 2010. Apoptosis protection by Mcl-1 and Bcl-2 modulation of inositol 1,4,5-trisphosphate receptor-dependent Ca²⁺ signaling. J Biol Chem 285:13678–13684.
- Ferdek PE, Gerasimenko JV, Peng S, Tepikin AV, Petersen OH, Gerasimenko OV. 2012. A novel role for Bcl-2 in regulation of cellular calcium extrusion. Curr Biol 22:1241–1246.
- Ferdek PE, Jakubowska MA, Nicolaou P, Gerasimenko JV, Gerasimenko OV, Petersen OH. 2017. BH3 mimetic-elicited Ca^{2+} signals in pancreatic acinar cells are dependent on Bax and can be reduced by Ca^{2+} -like peptides. Cell Death Dis 8:e2640.
- Fesik SW. 2000. Insights into programmed cell death through structural biology. Cell 103:273–282.
- Gerasimenko J, Ferdek P, Fischer L, Gukovskaya AS, Pandol SJ. 2010. Inhibitors of Bcl-2 protein family deplete ER Ca²⁺ stores in pancreatic acinar cells. Pflugers Arch 460:891–900.
- Green DR. 2016. A BH3 mimetic for killing cancer cells. Cell 165:1560.
- He H, Lam M, McCormick TS, Distelhorst CW. 1997. Maintenance of calcium homeostasis in the endoplasmic reticulum by Bcl-2. J Cell Biol 138:1219–1228.
- Huang DC, Strasser A. 2000. BH3-only proteins-essential initiators of apoptotic cell death. Cell 103:839–842.
- Huang H, Shah K, Bradbury NA, Li C, White C. 2014. Mcl-1 promotes lung cancer cell migration by directly interacting with VDAC to increase mitochondrial Ca²⁺ uptake and reactive oxygen species generation. Cell Death Dis 5:e1482.
- Kitada S, Leone M, Sareth S, Zhai D, Reed JC, Pellecchia M. 2003. Discovery, characterization, and structure–activity relationships studies of proapoptotic polyphenols targeting Bcell lymphocyte/leukemia-2 proteins. J Med Chem 46:4259– 4264.
- Kroemer G, Galluzzi L, Brenner C. 2007. Mitochondrial membrane permeabilization in cell death. Physiol Rev 87:99– 163.
- Lessene G, Czabotar PE, Colman PM. 2008. BCL-2 family antagonists for cancer therapy. Nat Rev Drug Discov 7:989–1000.

- Miyashita T, Reed JC. 1993. Bcl-2 oncoprotein blocks chemotherapy-induced apoptosis in a human leukemia cell line. Blood 81:151–157.
- Monaco G, Decrock E, Akl H, Ponsaerts R, Vervliet T, Luyten T, De Maeyer M, Missiaen L, Distelhorst CW, De Smedt H, et al. 2012. Selective regulation of IP3-receptor-mediated Ca²⁺ signaling and apoptosis by the BH4 domain of Bcl-2 versus Bcl-Xl. Cell Death Differ 19:295–309.
- Oltersdorf T, Elmore SW, Shoemaker AR, Armstrong RC, Augeri DJ, Belli BA, Bruncko M, Deckwerth TL, Dinges J, Hajduk PJ, et al. 2005. An inhibitor of Bcl-2 family proteins induces regression of solid tumours. Nature 435:677–681.
- Petersen OH. 1992. Stimulus-secretion coupling: cytoplasmic calcium signals and the control of ion channels in exocrine acinar cells. J Physiol 448:1–51.
- Rong YP, Bultynck G, Aromolaran AS, Zhong F, Parys JB, De Smedt H, Mignery GA, Roderick HL, Bootman MD, Distelhorst CW. 2009. The BH4 domain of Bcl-2 inhibits ER calcium release and apoptosis by binding the regulatory and coupling domain of the IP3 receptor. Proc Natl Acad Sci USA 106:14397–14402.
- Schoenwaelder SM, Jackson SP. 2012. Bcl-xL-inhibitory BH3 mimetics (ABT-737 or ABT-263) and the modulation of cytosolic calcium flux and platelet function. Blood 119:1320–1321 (author reply 1321–1322).
- Schoenwaelder SM, Jarman KE, Gardiner EE, Hua M, Qiao J, White MJ, Josefsson EC, Alwis I, Ono A, Willcox A, et al. 2011. Bcl-xL-inhibitory BH3 mimetics can induce a transient thrombocytopathy that undermines the hemostatic function of platelets. Blood 118:1663–1674.
- Sheng L, Leshchyns'ka I, Sytnyk V. 2013. Cell adhesion and intracellular calcium signaling in neurons. Cell Commun Signal 11:94.
- Souers AJ, Leverson JD, Boghaert ER, Ackler SL, Catron ND, Chen J, Dayton BD, Ding H, Enschede SH, Fairbrother WJ, et al. 2013. ABT-199, a potent and selective BCL-2 inhibitor, achieves antitumor activity while sparing platelets. Nat Med 19:202–208.
- Suzuki M, Youle RJ, Tjandra N. 2000. Structure of Bax: coregulation of dimer formation and intracellular localization. Cell 103: 645–654.

- Tse C, Shoemaker AR, Adickes J, Anderson MG, Chen J, Jin S, Johnson EF, Marsh KC, Mitten MJ, Nimmer P, et al. 2008. ABT-263: a potent and orally bioavailable Bcl-2 family inhibitor. Cancer Res 68:3421–3428.
- Vervliet T, Decrock E, Molgo J, Sorrentino V, Missiaen L, Leybaert L, De Smedt H, Kasri NN, Parys JB, Bultynck G. 2014. Bcl-2 binds to and inhibits ryanodine receptors. J Cell Sci 127:2782–2792.
- Vervliet T, Lemmens I, Vandermarliere E, Decrock E, Ivanova H, Monaco G, Sorrentino V, Nadif Kasri N, Missiaen L, Martens L, et al. 2015. Ryanodine receptors are targeted by anti-apoptotic Bcl-XL involving its BH4 domain and Lys87 from its BH3 domain. Sci Rep 5:9641.
- Vervliet T, Parys JB, Bultynck G. 2016. Bcl-2 proteins and calcium signaling: complexity beneath the surface. Oncogene 35: 5079–5092.
- Vervloessem T, Ivanova H, Luyten T, Parys JB, Bultynck G. 2017. The selective Bcl-2 inhibitor venetoclax, a BH3 mimetic, does not dysregulate intracellular Ca²⁺ signaling. Biochim Biophys Acta 1864:968–976.
- Villain M, Jackson PL, Manion MK, Dong WJ, Su Z, Fassina G, Johnson TM, Sakai TT, Krishna NR, Blalock JE. 2000. *De novo* design of peptides targeted to the EF hands of calmodulin. J Biol Chem 275:2676–2685.
- Wang JL, Liu D, Zhang ZJ, Shan S, Han X, Srinivasula SM, Croce CM, Alnemri ES, Huang Z. 2000. Structure-based discovery of an organic compound that binds Bcl-2 protein and induces apoptosis of tumor cells. Proc Natl Acad Sci USA 97: 7124–7129.
- Wei C, Wang X, Zheng M, Cheng H. 2012. Calcium gradients underlying cell migration. Curr Opin Cell Biol 24:254–261.
- White C, Li C, Yang J, Petrenko NB, Madesh M, Thompson CB, Foskett JK. 2005. The endoplasmic reticulum gateway to apoptosis by Bcl-X_L modulation of the $InsP_3R$. Nat Cell Biol 7: 1021–1028.
- Zhu L, Yu Y, Chua BH, Ho YS, Kuo TH. 2001. Regulation of sodium–calcium exchange and mitochondrial energetics by Bcl-2 in the heart of transgenic mice. J Mol Cell Cardiol 33:2135– 2144.