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Exploring the C^N^C theme: synthesis and biological properties of tridentate cyclometalated gold(III) complexes

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Abstract

A family of cyclometalated Au(III) complexes featuring a tridentate C^N^C scaffold has been synthesized and characterized. Microwave assisted synthesis of the ligands has also been exploited and optimized. The biological properties of the thus formed compounds have been studied in cancer cells and demonstrate generally moderate antiproliferative effects. Initial mechanistic insights have also been gained on the gold complex [Au(C^N^C)(GluS)] (**3**), and support the idea that the thioredoxin system may be a target for this family of compounds together with other relevant intracellular thiol-containing molecules.

Keywords
Gold(III) complexes; Cyclometalation; Anticancer agents; Thioredoxin reductase; Thiols.

Introduction

Gold-based organometallic complexes as anti-cancer agents have become increasingly popular in recent years, and several reviews have been published highlighting their structural diversity and biological activity.[1] In fact, organometallic complexes display numerous attractive features. For example, while the organic ligand allows for the introduction of stereospecificity and alteration of the physicochemical properties by choosing different functional groups, the metal–carbon (M–C) bond provides strong *trans* influence and, in the case of π-bonded aromatic arene and cyclopentadienyl ligands, can act both as electron donors and acceptors. Furthermore, variations of the organic moiety are accessible, allowing the "fine-tuning" of the physiochemical properties of the respective metal compound. Finally, by choosing specific targeting moieties, either incorporation into the ligand or tethering to the metal centre can be achieved in a modular approach.

Typical classes of organometallics include metallocenes, metallo-arenes, metallo-carbonyls, metallo-carbenes (e.g. N-heterocyclic carbenes, NHC), alkynyl complexes, etc.[2] featuring both mononuclear and multinuclear scaffolds. All these compound families have been extensively applied in catalysis during the last decades.[3] However, their medicinal use has been considered only much more recently.[1c, 2, 4]

Concerning Au(I) organometallics, in 2008 Berners-Price *et al.* synthesized a series of mononuclear cationic Au(I) biscarbene complexes that show remarkable cytotoxicity *in vitro* and are able to induce mitochondrial damage[5]. Since then, the effects of Au(I) NHC complexes on cell metabolism and their interference with pathways relevant to cancer cell proliferation have been studied in a broad number of cases.[1a, 4b, 6]

In this context, to explore the design of Au(III) compounds for biological applications, cyclometalation is a convenient method to stabilize the otherwise easily reduced Au(III) centre, and numerous cyclometalated scaffolds have been synthesized, including C^N, C^N^C, C^N^N and C^N^S.[1b, 1c, 7] Cyclometalated Au(III) C^N compounds of general formula **[(Au(damp)X_2]** (Fig. 1) with anticancer properties were first investigated by Parish, Buckley *et al.* in 1996,[8]

featuring a 2[dimethylamino)methyl]-phenyl (damp) backbone. The compounds display cytotoxic activity which is comparable to that of cisplatin, against a variety cancer cell lines. This activity is also accompanied by high selectivity and cytotoxicity *in vitro* which translates to moderate activity *in vivo*.

In 1996, the synthesis of an Au(III) 2-benzylpyridine derivative **[Au(py^{b}-H)Cl$_2$]** (py^{b}-H = C^N cyclometalated 2-benzylpyridine) (Fig. 1) was reported by Cinellu *et al.* [9] In 2015 Casini, Cinellu *et al.* synthesized a structural analogue of this complex replacing the chlorido ligand, *trans* to the nitrogen atom, with 1,3,5-triazaphosphaadamantane (PTA).[10] The resulting compound displays good cytotoxic activity against various cancer cell lines such as A2780 (human ovarian adenocarcinoma). Furthermore, dinuclear oxo-bridged Au(III) C^N^N complexes of formula **[(C^N^N)$_2$Au$_2$(μ-O)](PF$_6$)$_2$** (with C^N^N = 6-(1-methylbenzyl)-2,2'-bipyridine or 6-(1,1-dimethylbenzyl)-2,2'-bipyridine) showing moderate cytotoxicity against various cancer cell lines were synthesized.[11] (Fig. 1).

The field of Au(III) C^N^C complexes with anticancer properties has been closely examined by Che and co-workers[1b, 12] For example, dinuclear complexes of the type [Au$_m$(C^N^C)$_m$L]$^{n+}$ (with HC^N^CH=2,6-diphenylpyridine; m =1–3; n=0–3) showed higher cytotoxic activity against various cancer cell lines than their mononuclear counterparts. [12]

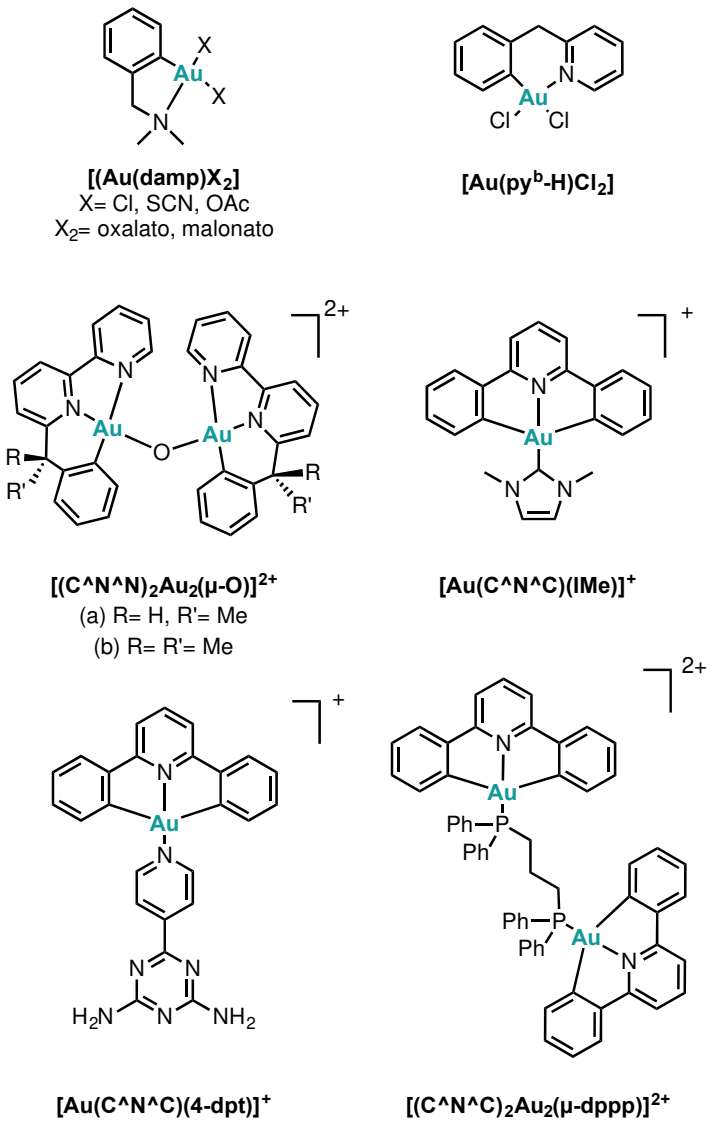

Fig. 1. Representative cyclometalated Au(III) complexes with anticancer properties.

The highest cytotoxicities were observed for complex **[Au$_2$(C^N^C)$_2$(μ-dppp)](OTf)$_2$** (

Fig. ***1***), relating to the cytotoxicity of the free 1,2-bis(diphenylphosphino)propane (dppp) ligand. By replacing the phosphane moiety with an NHC ligand as in the cationic C^N^C stabilized complexes [Au$_n$(R–C^N^C)$_n$(NHC)]$^{n+}$, a general decrease in cytotoxic effects has been noted,[13] supporting the idea that indeed phosphine ligand-mediated cytotoxicity plays a major role in the overall anticancer properties. Nevertheless, within this series, the mononuclear complex **[Au(C^N^C)(IMe)]CF$_3$SO$_3$** (IMe = 1,3-dimethylimidazol-2-ylidene) (Fig. 1) shows higher cytotoxic activity than its dinuclear analogue, and a high degree of selectivity towards human cancer cells compared to normal lung fibroblasts (CCD-19Lu). Through DNA interaction studies it was demonstrated that the compound induces DNA strand breaks and can cause subsequent cell death through the stabilization of Topoisomerase-linked DNA[14]. Treatment of nude mice bearing PLC tumors (hepatocellular carcinoma) with **[Au(C^N^C)(IMe)]CF$_3$SO$_3$** at 10 mg/kg/week for 28 days significantly suppressed (47%) tumor growth when compared with that of the vehicle control.[13]

Of note from the same group, is the application of supramolecular polymers, self-assembled from cyclometalated Au(III) C^N^C complexes, as anticancer agents. The mononuclear complex **[Au(C^N^C)(4-dpt)]$^+$** (C^N^C =2,6-diphenylpyridine, 4-dpt = 2,4-diamino-6-(4-pyridyl)-1,3,5-triazine) (Fig. 1) was chosen due to the ability of the antiangiogenic 4-dpt ligand to form intramolecular hydrogen bonds and to establish π-π interactions, leading to supramolecular complex formation by self-assembly at ambient temperatures.[15] The supramolecular polymer displays high cytotoxic activity towards B16 cells (murine cancer).

In terms of possible mechanisms of action, cytotoxic Au(III) complexes can interact with protein targets[16], including those constituting the *thioredoxin system*, often overexpressed in tumor cells and involved in maintaining the intracellular redox balance[17]. Among the enzymes included in this system, the seleno-protein thioredoxin reductase (TrxR) contains a cysteine-selenocysteine redox pair at the C-terminal active site, and the solvent-accessible selenolate group, arising from enzymatic reduction, constitutes a likely target for "soft" metal ions such as gold. In addition, Au(III) complexes can interfere with thiols including glutathione, the most abundant intracellular reducing agent.

Within the scope of new medicinally relevant organometallic Au(III) compounds, two new cyclometalated Au(III) complexes of the general formula [Au(III)(C^N^C)X] (with C^N^C = 2,6-diphenylpyridine and X = Cl, thio-*β*-D-glucose-tetraacetate (GluS), 1,3,5-triazaphosphaadamantane (PTA)) were synthesized. The complexes were prepared starting from the literature known chlorido precursor [Au(III)(C^N^C)Cl][18] (Fig. 2, **1**), substituting the chlorido ligand with PTA (**2**) and thio-*β*-D-glucose-tetraacetate (GluSH) (**3**), respectively. The PTA ligand was chosen to increase the water solubility, while the $GluS^-$ ligand was selected as modulator of the hydrophilic/lipophilic character, as well as a possible facilitator of the compound's uptake via interactions with the GLUT1 transporter. A second series of analogous C^N^C complexes - Au(III)[(C^N^R^C)] - was synthesized starting from the corresponding 2,4,6-triarylpyridine ligands, but featuring different substituents in the *para* position of the phenylpyridine (R = OH, F, Br, NO_2) (Fig. 2, **4-7**). Notably, complex **5** has already been described by Che *et al.* in 2013 as a precursor for photoactive functionalized bis-cyclometalated alkynyl gold(III) complexes suitable as phosphorescent organic light-emitting diodes (OLEDs).[19] However, the authors did not evaluate the biological activity of this complex. All the cyclometalated Au(III) complexes reported herein were studied for their antiproliferative effects against different human cancer cell lines, including some that are resistant to cisplatin. Moreover, compounds **1-3** were also tested for their inhibition properties of TrxR on the purified protein and in cell lysates. The effects of the new complexes on the oxidation state of Trx were also investigated by Western blot analysis. Discrimination between the oxidation of the GSH/GSSG system and the Trx system can be very informative in terms of mechanisms of toxicity since different cellular pathways are controlled by GSH and Trx. Thus, estimation of the glutathione content was performed in treated cells.

2. Results and Discussion

2.1 Synthesis and structural characterization

To achieve the synthesis of the new compounds, the transmetalation pathway via the respective Hg(II) precursor was followed as described in Fig. 2. Thus, treatment of 2,6-diphenylpyridine or *para* substituted 2,4,6-triarylpyridines with mercury(II) acetate, followed by salt metathesis with LiCl, affords the organomercury(II) precursors. Subsequent transmetalation with $K[AuCl_4]$ in refluxing acetonitrile, adapting previously reported procedures,[20] yields the respective cyclometalated Au(III) complexes. The synthesis of the cyclometalated precursor [Au(C^N^C)Cl] (C^N^C= 2,6-diphenylpyridine) (**1**) has been reported in 1998 by Che *et al.*[20a], but the biological activity of this complex has not been reported. Afterwards, two novel derivatives, namely the cationic $[Au(C^N^C)(PTA)][PF_6]$ (**2**) and the neutral [Au(C^N^C)(GluS)] (**3**), have been obtained from complex **1** by ligand exchange reactions (Fig. 2) and characterized by means of NMR, ESI-MS and elemental analysis.

Complex **2** contains the water-soluble ligand PTA, which, together with the positive charge of the resulting complex, should improve the overall water solubility while being non-cytotoxic *per se*. As has been reported before, replacement of a *trans* bound chlorido ligand by a tertiary phosphine at the Au(III) center can be achieved by first abstracting the halide to unveil a cationic gold (III) that can then be intercepted by the donor PTA ligand.[10, 18]

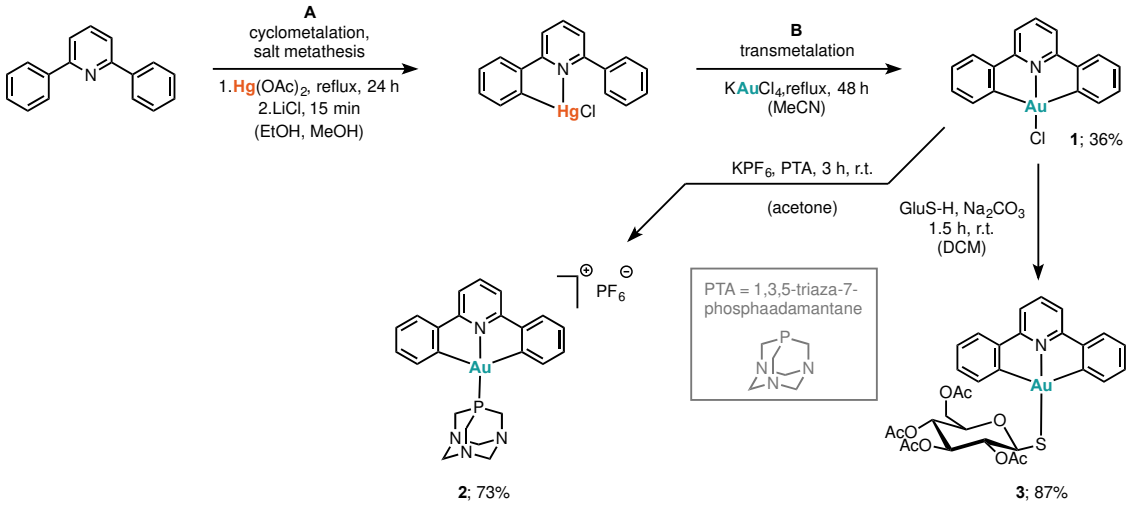

Fig. 2. General synthesis scheme of the obtained cyclometalated Au(III) [C^N^C] complexes **1-3**.

Thus, by reacting **1** with one equivalent of PTA in the presence of KPF_6 in acetone for 3 h at room temperature, complex **2** is obtained in good yields. Single crystals of complex **2** suitable for X-ray diffraction were grown by slow diffusion of *n*-pentane into an acetone solution of the compound. The crystallographic structure of **2** is depicted in Fig. 3. The Au(III) centre is coordinated in a slightly distorted square planar fashion by the C^N^C pincer ligand and PTA with bond lengths of Au1—C1 = 2.106(3) Å, Au1—N1 = 2.025(2) Å, Au1—C17 = 2.091(3) Å and Au1—P1 = 2.2707(7) Å which are comparable to literature-reported values for the Au(III)-C^N^C[19] scaffold and an Au(III) ion binding to PTA.[10, 21] While there is no significant out-of-plane distortion of the square planar arrangement, the geometry of the pincer ligand causes an in-plane distortion illustrated by the C1—Au1—C17 angle of 160.8(1)°. The pincer ligand itself is close to perfect planarity with only the phenyl-side-arm containing C17 slightly being bent out of plane and the C1-C6 phenyl ring being bent even more slightly in the antipodal direction. Fig. S1 in the supplementary material depicts the allocation of the signals for ^{1}H and ^{13}C NMR spectra of the complexes. When comparing the free 2,6-diphenylpyridine ligand to the obtained complexes **1-3**, a significant shift in the ^{1}H NMR spectrum for H_f can be observed (see Fig. S1 in the supplementary material for the allocation of the signals for ^{1}H and ^{13}C NMR spectra of the complexes), resulting from the strong de-shielding caused by the –I effect of the electron-accepting Au-Cl moiety ($\Delta\delta$ =0.48 ppm for complex **1**).

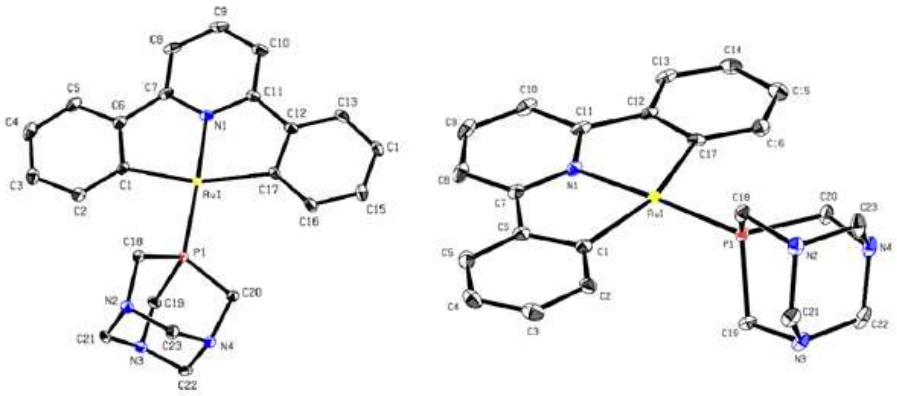

Fig. 3. ORTEP style presentation of the molecular structure of compound **2** in the solid state with ellipsoids given at 50% probability level. Hydrogen atoms, hexafluorophosphate anions and co-crystallized water molecules are omitted for clarity. Selected bond lengths (Å) and angles (°): Au1—C1 2.106(3), Au1—N1 2.025(2), Au1—C17 2.091(3), Au1—P1 2.2707(7), P1—Au1—C1 97.13(7), P1—Au1—C17 102.10(7), N1—Au1—C1 80.32(9), N1—Au1—C17 80.44(9).

Complex **1** was also treated with one equivalent of thio-*β*-D-glucose tetraacetate (GluSH) and sodium carbonate in dichloromethane for 1.5 hours at room temperature to obtain compound **3** in good yields. As this ligand is optically active, the ^{1}H NMR presents less sharp signals than the spectrum for complex **2**. The proton count, however, fits the structure of **3** perfectly and the signal of the thiol proton at 2.20 ppm is not present in the product NMR spectrum, confirming the bond formation between gold and sulfur, while the four acetate groups of the $GluS^-$ ligand appear as sharp singlet signals between 1.96 and 1.77 ppm. Moreover, all protons of the $GluS^-$ are shifted as well, resulting from the strong de-shielding caused by the –I effect of the electron-accepting Au(III) centre. The structure of complex **3** was further assessed by ESI-MS and elemental analysis.

Microwave assisted synthesis can afford several benefits, especially to modular synthesis programmes such as those concerning medicinal chemistry. Such benefits may come in the form of increased product purity and reduced reaction times.[22] The use of robotic handlers and touch-screen interfaces can greatly streamline the preparation of a library of compounds, especially when using an Initiator Robot system. Thus, the para-substituted 2,4,6-triaryl [C^N^R^C] ligands (L**4-7**) for complexes **4-7** (Fig. 4) were obtained by applying microwave assisted ligand synthesis. In detail, microwave irradiated synthesis of the [C^N^R^C] ligands was carried out adapting a previously reported procedure for hydroxylated tri-substituted pyridines.[23] Optimization of the reaction conditions was performed with respect to temperature, power and reaction time of the irradiation. A major benefit of the microwave synthesis is the circumvention of otherwise needed catalyst, such as previously reported $Bi(OTf)_3$,[24] and solvents, thus enabling a metal and solvent free synthesis.

A previously described procedure reports on the use of a conventional microwave with unfocused irradiation at 400 Watts.[23] We found this high power in our focussed reactor to cause pressure build up within in the system, leading to the termination of the microwave through triggering of the inbuilt safety features. A basis set (100 W, 120 °C, 30 minutes) was chosen, from which on the single parameters were optimized. The irradiation power has been altered as shown in Table S1 in the supplementary material, with the other parameters being fixed at basic

conditions. The maximum product yield was found for a power of 175 W. The yield performance drops significantly after the power exceeds 200 W, which is believed to be contributed to beginning decomposition of the formed triarylpyridines.

The next parameter that could be altered is the temperature as depicted in Table S2 in the supplementary material. The maximum product yield was found for a temperature of 180 °C, while yield performance dropped significantly after the temperature exceeds 200 °C. The reaction time was the last parameter of the basis set to be altered as can be seen in Table S3. As expected, the yield reaches a plateau after a defined period of time (80 min and upwards). Overall, the combined optimized parameters for the microwave irradiated synthesis of the 2,4,6-triarylpyridine ligands were found to be at 180 °C, 175 W and 80 minutes.

Following the successful development of a microwave protocol for the preparation of the ligands, the cyclometalated Au(III) complexes were obtained via transmetalation of the respective [Hg(C^N^R^C)Cl] precursors with K[$AuCl_4$]. This was achieved by heating the reaction mixture to reflux in acetonitrile for 24 hours (Fig. 4). Likewise, to the 2,6-diphenylpyridine systems, a significant shift in the ^{1}H NMR spectrum for H_1 can be observed when comparing the free 2,4,6-triaryl ligands to their respective complexes **4-7**, resulting from the strong de-shielding caused by the –I effect of the electron-accepting Au-Cl moiety ($\Delta\delta$ =0.39 ppm for complex **4**).

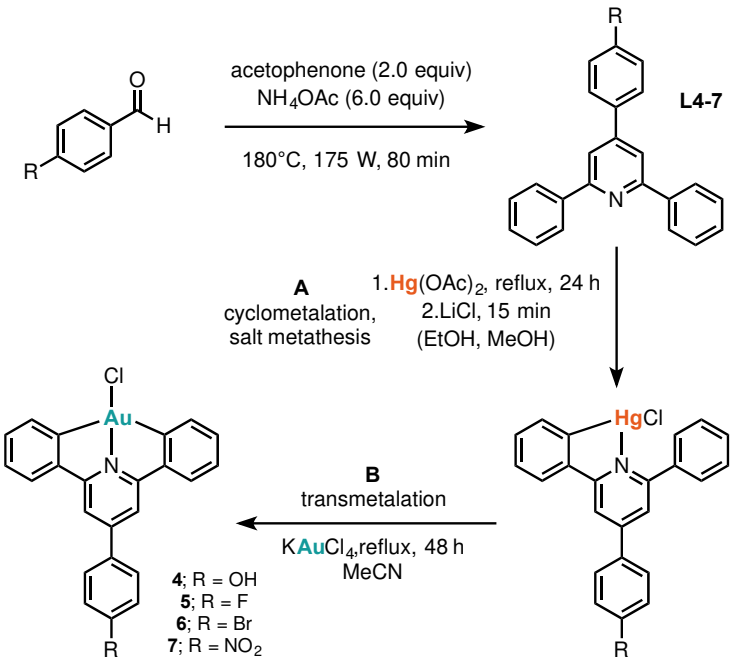

Fig. 4. Synthesis of [Au(III)(C^N^R^C)] (C^N^R^C = para-substituted 2,4,6-triarylpyridine) complexes.

The UV-visible absorption of complex **1** and various derivatives has been previously reported and shows a characteristic absorption with bands between 300−400 nm.[12] The characteristic absorption was tentatively assigned to a metal-perturbed intraligand (IL) transition. We have obtained similar results in the case of complex **3**. Furthermore, treatment of the gold(III) compound with excess glutathione (GSH, 1 mM) in PBS solution caused no significant UV-visible spectral changes over a 24 h period at room temperature (Figure S25).

2.2 Antiproliferative activity

The gold complexes were tested for their cytotoxic activity against human cancer cell lines A549 (lung adenocarcinoma) SKOV-3 (ovarian adenocarcinoma) and 2008 (human ovarian cancer), in comparison to their ligands, using a classical MTT assay as reported in the Experimental section. It is worth mentioning that A549 and SKOV-3 cells were chosen since they manifest resistance to cisplatin treatment. Overall, compounds **4-7** show modest activity in both A549 and SKOV-3 cell lines, with IC_{50} values > 50 μM (Table 1). Similarly, the ligands **L4-L7** show very

moderate effects. The highest antiproliferative effects are found for complexes **1** and **3** in 2008 cells, with IC_{50} values of 29 μM ± 3 and 7.0 μM ± 1.2, respectively.

Table 1. IC_{50} values (μM) of the Au(III) compounds **1-7**, ligands (**L4-L7**) and cisplatin against A549 and SKOV-3 (72 h incubation) and 2008 (48 h incubation).

compound	A549	SKOV-3	2008
1	35 ± 5	39 ± 7	29 ± 3
2	> 50	48.1 ± 2.3	> 50
3	30.4 ± 1.3	13.0 ± 0.9	7.0 ± 1.2
4	> 50	> 50	-
5	> 50	> 50	-
6	> 50	> 50	-
7	> 50	> 50	-
L4	30.0 ± 4.7	28.8 ± 5.5	-
L5	42.3 ± 5.3	44.5 ± 7.9	-
L6	30.8 ± 4.5	59.5 ± 9.7	-
L7	>100	>100	-
cisplatin	12 ± 0.5	16.3 ± 1.7	-

2.3 Thioredoxin reductase inhibition

Since TrxR is also a potential target for gold complexes, *in vitro* inhibition of purified rat liver cytosolic and mitochondrial TrxR by compounds **1-3** was studied and compared with the activity of auranofin, a well-known TrxR inhibitor, using established protocols as described in the Experimental section. The results are summarized in Table 2. All compounds are good inhibitors of cytosolic thioredoxin reductases (TrxR1), showing IC_{50} values in the nanomolar range of concentrations, with complex **3** being the most potent. Conversely, the mitochondrial isoform of the enzyme is slightly inhibited by the three complexes after incubation in the same experimental conditions, revealing a different inhibitory capacity on the two isoforms. Further studies

demonstrated that **3** is also able to inhibit the TrxR closely related, but selenium-free, enzyme glutathione reductase (GR) although 60-fold less efficiently than in the case of TrxR (Table 2).

Table 2. IC_{50} values of the inhibition of TrxR1, TrxR2 and GR on the isolated enzymes.

	IC_{50} (nM)		
Complex	**TrxR1**	**TrxR2**	**GR**
auranofin	0.9 ± 0.3	3 ± 1	>10 000
1	10 ± 1	59 ± 5	219 ± 18
2	10 ± 1	213± 20	439 ± 7
3	3± 1	60 ± 4	180 ± 22

The effect of compounds **1-3** on TrxR and GR activities was also evaluated in cell lysates. For this purpose, 2008 cells, where the three compounds showed markedly different cytotoxic effects, were pre-treated for 48 h with **1-3** at different concentrations (Fig. 5A). Complex **3** was the most effective in inhibiting TrxR, followed by compound **2**. Complex **1** was scarcely active. In addition, GR activity was also determined and, as shown in Fig. 5B, the enzyme was not inhibited by the compounds even at 50 µM concentration.

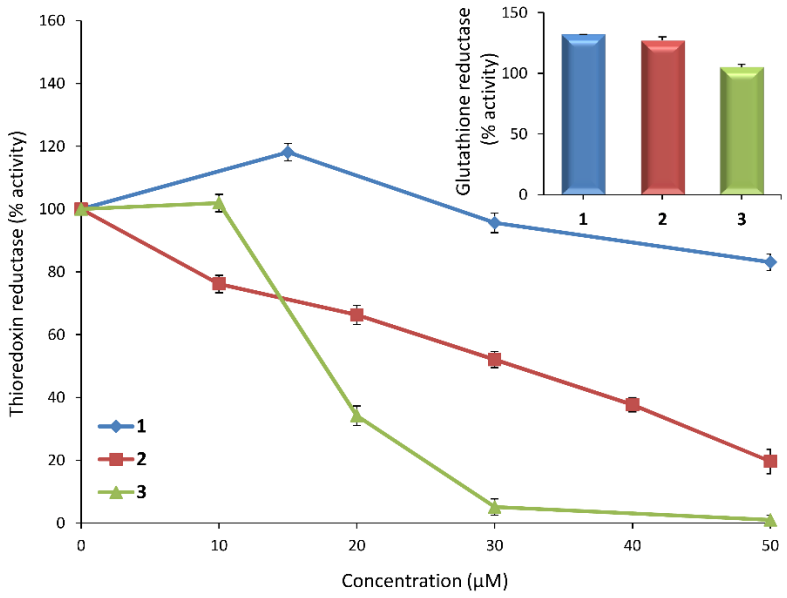

Fig. 5. Thioredoxin reductase and glutathione reductase (insert) activities in 2008 ovarian cancer cells after 48 h treatment with **1**-**3**. For TrxR activity determinations, cells (1 x 10^6) were treated with the compounds at the indicated concentration between 0-40 µM. GR activity was evaluated after cells treatment with 50 µM of each of the three compounds, as reported in the Experimental section.

2.4 Effects on cellular redox state

Furthermore, the cellular redox state was evaluated after the incubation of 2008 cells with **1**-**3**. For the determination of the total thiols in cell lysates, an assay was conducted according to the protocol described in the Experimental section. The results presented in Fig. 6 show that the total amount of thiols is unaltered upon treatment with **1** and **2**, indicating that these complexes have no effect on the redox state of the intracellular sulfhydryl groups, while at 50 μM, **3** shows a decrease of about 40% of the total thiols. This suggests that **3** determines a redox imbalance interacting with thiols inside the cells.

Since the glutathione redox pair (GSH/GSSG) is another fundamental component of the cell redox regulation, an analysis of the total glutathione content and of the GSH/GSSG ratio was performed in 2008 cells, after treatment with **1-3** for 48 hours. The obtained results are elucidated in Fig. 6B. After the treatment of 5 x 10^5 ovarian cancer cells, an important increase of oxidized glutathione, suggesting thiols oxidation, can be observed only in the presence of **3** (50 μM), while **1** and **2** do not affect the glutathione system since total and oxidized levels were comparable to the control cells (Fig. 6B).

Therefore, the redox state of Trx1 and Trx2 in 2008 cells treated for 48 h with **1** and **3** was studied (Fig. S26, supplementary material) to confirm that the inhibition of TrxR determines an increase of oxidation of the principal substrates. Interestingly, complex **3** induces a marked oxidation of Trx1 and Trx2, evidenced by the shift of the bands relative to the two proteins towards their oxidized form by western-blot analysis, providing indirect evidence that TrxR is a pivotal target of this compound in cells.

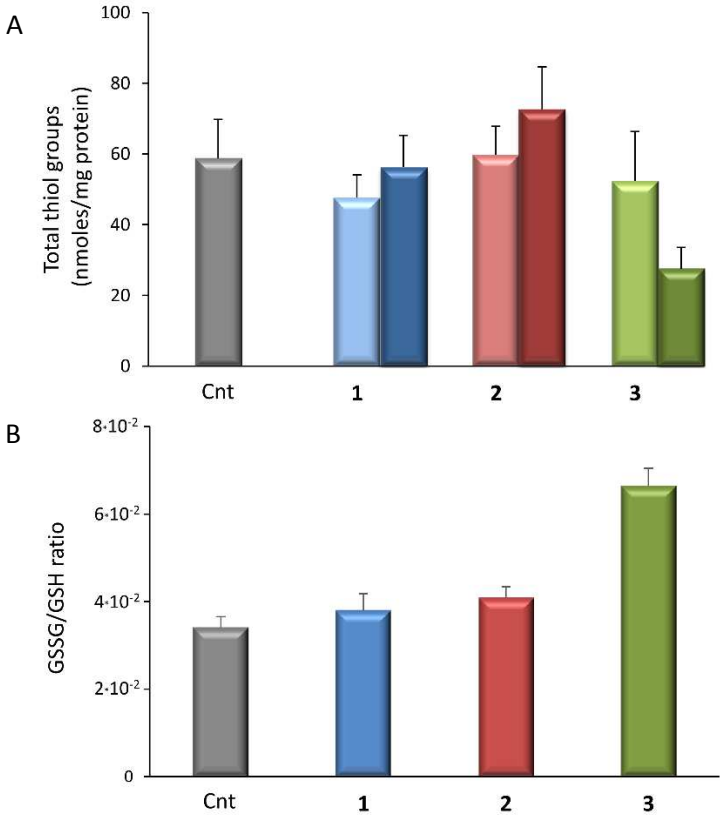

Fig. 6. Total thiols: (A) in 2008 ovarian cancer cells after the treatment with **1-3**. A: cells (5 x 10^5) were treated with the compounds at the indicated concentrations and then subjected to total thiol determination (light bars: 25 µM; dark bars: 50 µM). (B) Glutathione redox state ratio (GSSG/GSH) after cell treatment with 50 µM of the three compounds.

3. Conclusions

Two new Au(III)[C^N^C] complexes, [Au(C^N^C)(PTA)][PF_6] (**2**) and [Au(C^N^C)(GluS)] (**3**), and three new Au(III)[C^N(R)^C] complexes with R= Ph-*p*-OH (**4**), Ph-*p*-Br (**6**), Ph-*p*-NO_2 (**7**) have been synthesized. Microwave assisted ligand synthesis for the *para* substituted 2,4,6-triaryl ligands for complex **4-7** was optimized concerning applied temperature, power and reaction time. The compounds have been examined for their antiproliferative effects in a small panel of human cancer cells. In general, the compounds are poorly toxic, with the exception of compounds **1** and **3**, which were active in the 2008 ovarian cancer cells. Initial mechanistic studies have shown that **1-3** are nanomolar inhibitors of isolated cytosolic and mitochondrial TrxRs, while they scarcely affect the activity of GR, supporting the idea that they could bind preferentially TrxRs. These results are confirmed also by the increase of the oxidized form of Trxs, upon metallodrugs' treatment. Furthermore, the observed effects on the overall redox state

of the cells of complex **3** suggest that this derivative is particularly effective in disrupting the thiol redox homeostasis of the cell.

Overall, the obtained results demonstrate that the reported Au(III) complexes need to be further optimized for biological applications, especially in terms of solubility and potency on cancer cells. Moreover, a central role of the ancillary ligands in the observed anticancer effects has been highlighted, in line with previous studies by Che *et al*. Nevertheless, these initial results provide some indications on the possible cellular targets for this family of compounds.

In terms of possible derivatization of the organometallic scaffold, Au(C^N^C) systems replacing the central pyridine ring with a pyrazine have been reported recently.[25] As they display intense thermally activated delayed fluorescence (TADF), these Au(C^N^C) complexes could lead to a convenient way to monitor the uptake of anticancer compounds via fluorescence microscopy.

4. Experimental Section

4.1 General remarks

Reactions were carried out under purified argon using standard Schlenk techniques. Solvents were dried, degassed and stored over molecular sieve and under argon before use. Microwave reactions were carried out in a CEM Focused Microwave™ Synthesis System or a Biotage® Initiator Robot Eight and Sixty. All physico-chemical analytics were performed at the Technische Universität München and the University of Cardiff. UV−vis spectra were recorded on a Perkin-Elmer Lambda 650 UV/vis spectrophotometer. Electrospray ionization mass spectra (ESI-MS) were obtained on a High-Resolution Waters LCT TOF Bruker HR-QTOF maXisPlus operating in positive ionization mode. The samples were dissolved in 1 ml of the appropriate solvent and 10 µl were injected directly in to the mass spectrometer. The mass range was scanned between 100 and 2000 and Leucine Enkephalin was used as an online calibrant. NMR spectra were recorded on a Bruker Avance DPX-300, 400 or 500 at 298 K. All deuterated solvents were purchased from Deutero GmbH and Sigma Aldrich. Chemical shifts are given in δ (ppm) and refer to the residual ^{1}H and $^{13}C(^{1}H)$ signals of the respective solvent. In allocating the signals the following abbreviations have been used: s- singlet, d- doublet, t -triplet, q-quartet, m- multiplet. Coupling constants J are given in Hz. X-ray intensity data were measured on a Bruker D8 Venture Duo IMS system equipped with a Helios optic monochromator and a Mo IMS microsource (λ=0.71073 Å). All mercury precursors were prepared according to Constable *et al.*[20b]. The commercially available ligand 2,6-diphenylpyridine (CAS 3558-69-8) and Au(III) precursor $K[AuCl_4]$ (98%, CAS 13682-61-6) were purchased from Sigma-Aldrich and utilized without further purification. Crystallographic Details are reported in the Supporting Information.

4.2 Synthesis of [Au(III)(C^N^C)]$^{n+}$ (C^N^C= 2,6-diphenylpyridine) complexes

[Au(C^N^C)Cl] (1). Preparation was carried out with slight changes to the synthesis by Che *et al.*[20a]

Hg(C^N^CH)Cl (1eq, 0.51 g, 1.09 mmol) and $K[AuCl_4]$ (1eq, 0.41 g, 1.09 mmol) are refluxed in acetonitrile (20 mL) under argon for 24 h at 80°C forming a greenish precipitate. The crude solid is filtered, washed with diethyl ether and *n*-pentane and dried under reduced pressure to afford an electrostatic light-yellow, solid (0.18 g, 0.39 mmol, 36% yield). ^{13}C NMR spectra have not been reported in the literature so far.

^{1}H NMR (400 MHz, DMSO-d6) δ 8.19 (t, *J* = 8.0 Hz, 1H, H_a), 7.98 (d, *J* = 8.1 Hz, 2H,H_f), 7.88(d, *J* = 7.7 Hz, 2H, H_c), 7.69 (d, *J* = 7.2 Hz, 2H,H_b), 7.43 (t, *J* = 7.3 Hz, 2H, H_d), 7.32 (t, *J* = 7.6 Hz, 2H, H_e). $^{13}C(^{1}H)$ NMR (101 MHz, DMSO) δ 169.5 (C-N), 164.1 (C-C), 148.4 (C-H_a), 142.3 (C-Au), 133.0 (C-H_c), 131.7 (C.H_e), 127.6 (C-H_d), 126.1 (C-H_f), 118.6 (C-H_b). Positive ESI-MS (acetone): m/z = 462.21 $[M+H]^+$ (calcd for $C_{17}H_{12}AuClN$: 462.03). Anal. Calc. for $C_{17}H_{11}AuClN$: C, 44.22; H, 2.40; N, 3.03 %. Found: C, 44.16; H, 2.18 ; N, 3.23 %

[Au(C^N^C)(PTA)][PF_6] (2). Complex **1** (1 eq., 100 mg, 0.22 mmol) is added to KPF_6 (5 eq., 202.47 mg, 1.10 mmol) in acetone (25 mL). PTA (1 eq., 34.57 mg, 0.22 mmol) is added to the suspension at room temperature under vigorous stirring. The reaction is stirred at room temperature for 3 hours. Acetone is subsequently partly removed, dichloromethane was added and the solution was filtered through Celite®. The volatiles are removed under reduced pressure and the resulting white solid is washed with ice-cold acetone, *n*-pentane and diethylether (114.7 mg, 0.16 mmol, 73% yield).

^{1}H NMR (400 MHz, Acetone-d6) δ 8.28 (t, *J* = 8.0 Hz, 1H, H_a), 8.01 (m, *J* = 8.5 Hz, 6H, $H_{b,c,f}$), 7.56 (t, *J* = 7.5 Hz, 2H, H_d), 7.46 (t, *J* = 7.5 Hz, 2H, H_e), 5.21 (d, *J* = 2.1 Hz, 6H, N-CH_2), 4.99 (d, *J* = 13.1 Hz, 3H, P-CH_aH_b), 4.80 (d, *J* = 13.3 Hz, 3H, P-CH_aH_b). ^{13}C NMR (101 MHz, Acetone-d6) δ 169.5 (C-N), 164.7 (C-C), 151.4 (C-H_a), 146.4 (C-Au), 137.6 (C-H_c), 133.5 (C-H_e), 129.1 (C-H_d), 128.0 (C-H_f), 119.6 (C-H_b), 73.1 (N-CH_2), 51.3 (P-CH_2). Positive ESI-MS (acetone): m/z = 583.110 $[M]^+$ (calcd for $C_{23}H_{23}AuN_4P$: 583.126). Anal. Calc. for $C_{23}H_{23}AuF_6N_4P_2$: C, 37.93; H, 3.18; N, 7.69 %. Found: C, 37.79; H, 3.24; N, 7.38 %.

[Au(C^N^C)(GluS)] (3). Complex **1** (1 eq., 50 mg, 0.115 mmol), is added to thio-*β*-D-glucose tetraacetate (1 eq., 41.9 mg, 0.115 mmol) and Na_2CO_3 (5 eq., 61 mg, 0.575 mmol) in dichloromethane (20 mL). The suspension is stirred for 1.5 hours at room temperature. The solution is filtered through Celite® and concentrated under reduced pressure. Upon cooling and subsequent addition of pentane a light yellow precipitate is formed which is filtered, washed with diethylether and *n*-pentane and dried under reduced pressure (78.96 mg, 0.10 mmol, 87% yield).

^{1}H NMR (500 MHz, Acetone-d6) δ 8.34 –7.69 (m, 5H, $H_{a,b,c}$), 7.61 – 7.14 (m, 6H, $H_{d,e,f}$), 5.20 – 5.10 (m, 2H, 2 CH), 4.98 (m, 1H, CH), 4.89 (t, *J* = 9.4 Hz, 1H, CH), 4.79 (t, *J* = 9.7 Hz, 1H, CH), 4.15 (dd, *J* = 12.3 Hz, 2.1 Hz, 1H, CH_2-sugar), 4.02 (dd, *J* = 12.3 Hz, 5.6 Hz, 1H, CH_2-sugar), 1.96 (s, 3H, OAc), 1.94 (s, 3H, OAc), 1.86 (s, 3H, OAc), 1.77 (s, 3H, OAc). Positive ESI-MS (acetone): m/z = 790.1442 $[M+H]^+$ (calcd for $C_{31}H_{31}AuNO_9S$: 790.1385). Anal. Calc. for $C_{31}H_{30}AuNO_9S$: C, 47.15; H 3.83; N, 1.77; S, 4.06 ;%. Found: C, 47.01; H, 3.68; N, 1.85; S, 3.75 %.

4.3 Synthesis of [Au(III)(C^N^R^C)] (C^N^R^C=substituted 2,4,6-triarylpyridine) complexes

4.3.1 General procedure

C^N^R^C ligand synthesis **(L1-L7)**: A mixture of the respective *para*-substituted benzaldehyde (1eq), acetophenone (2.0 eq) and ammonium acetate (6.0 eq) is subjected to microwave irradiation of 180 W at 175 °C for 80 minutes. The resulting biphasic reaction mixture is then transferred to a separating funnel, treated with equal volumes of ice-water and diethyl ether and shaken well. The aqueous phase containing excess ammonium acetate is extracted three times with diethyl ether. The organic phases are combined and concentrated under reduced pressure to afford the crude product. Recrystallization from ethanol and subsequent drying under reduced pressure delivers the respective C^N(R)^C ligand. Column chromatography over silica gel (hexane/ethyl acetate = 1.5/1) can be performed if required.

[Au(III)(C^N^R^C)] complex synthesis: The respective Hg(C^N^R^CH)Cl precursor (1eq, 100 µmol) and K[$AuCl_4$] (1eq, 100 µmol) are added to acetonitrile (20 mL) using standard Schlenk techniques. Under vigorous stirring the suspension is kept at 100°C for 48 h under reflux upon which solubilisation of the educts occurs. The solution is then allowed to cool to room temperature leading to the precipitation of a solid. The crude product is filtered and then washed with ice-cold diethyl ether and *n*-pentane. Any remaining solvent is removed under reduced pressure to afford the complexes as solids.

[Au(III)(C^N^{OH}^C)Cl] (4) The compound is obtained as a pale yellow solid (38,77 mg, 70 µmol, 70% yield). ^{1}H NMR (400 MHz, DMSO-d6) δ 9.76 (s, 1H, OH), 8.27 (d, *J* = 6.8 Hz, 4H, $H_{1,4}$), 8.06 (d, *J* = 13.7 Hz, 4H,$H_{6,7}$), 7.60 – 7.53 (m, 4H, $H_{2,3}$), 7.51 – 7.46 (m, 2H,H_5). ^{13}C NMR (101 MHz, $CDCl_3$) δ 157.5 (2 C-N), 151.2 (C-OH), 150.0 (C_{para}-N), 140.1 (C-Au), 129.0, 128.9, 128.8, 128.0, 127.3, 127.3, 126.2, 116.8, 116.2, 112.6. Positive ESI-MS (acetone/water): m/z = 571.0382 $[M+H_2O]^+$ (calcd for $C_{23}H_{17}AuClNO_2$: 571.0613). Anal. Calc. for $C_{23}H_{15}AuClNO$: C, 49.88; H, 2.73; N, 2.53;%. Found: C, 49.63; H, 2.55; N, 2.57;%.

[Au(III)(C^N^F^C)Cl] (5) The compound is obtained as a pale yellow solid (35.01 mg, 63 µmol, 63% yield)**.** ^{1}H NMR (300 MHz, DMSO-d6) δ 8.32 (s, 2H, H_5), 8.25 (m, 2H, H_7), 8.12 (d, *J* = 7.7 Hz, 2H, H_4), 7.70 (d, *J* = 7.2 Hz, 2H, H_1), 7.54 – 7.41 (m, 4H, $H_{2,3}$), 7.34 (m, 2H, H_6). ^{13}C NMR (101 MHz, DMSO) δ 189.1 (C-F), 142.7 (C-Au), 137.4 (2 C-N), 134.0 (C_{para}-N), 133.3, 131.9, 131.3, 130.9, 128.9, 128.6, 128.1, 124.1, 122.8**.** Positive ESI-MS (acetone): m/z = 555.0463 $[M]^+$ (calcd for $C_{23}H_{17}AuClNO_2$: 555.0464)**.** Anal. Calc. for $C_{23}H_{14}AuClFN$: C, 47.15; H 3.83; N, 1.77 %. Found: C, 47.04; H 3.68; N, 1.79 %.

[Au(III)(C^N^{Br}^C)Cl] (6) The compound is obtained as a pale yellow solid (40.08 mg, 65 µmol, 65% yield)**.** ^{1}H NMR (300 MHz, DMSO-d6) δ 8.32 (s, 2H, H_5), 8.12 (m, 4H, $H_{4,1}$), 7.85 (d, *J* = 6.0 Hz, 2H, H_7), 7.71 (d, *J* = 8.0 Hz, 2H, H_6), 7.50 – 7.41 (m, 2H, H_2), 7.40 – 7.29 (m, 2H, H_3). ^{13}C NMR (75 MHz, DMSO) δ 157.1 (2 C-N), 148.8 (C_{para}-N), 139.1 (C-Au), 137.3, 132.4, 130.0, 129.8, 129.2, 127.4 (C-Br), 124.4, 117.0. Positive ESI-MS (acetone): m/z = 616.9659 $[M]^+$ (calcd for $C_{23}H_{17}AuClNO_2$: 614.9664)**.** Anal. Calc. for $C_{23}H_{14}AuBrClN$: C, 44.80; H, 2.29; N, 2.27 %. Found: C, 44.61; H, 2.24; N, 2.36 %.

[Au(III)(C^N^{NO_2}^C)Cl] (7) The compound is obtained as a pale yellow solid (35.55 mg, 61 µmol, 61% yield)**.** ^{1}H NMR (400 MHz, DMSO-d6) δ 8.44 (s, 2H, H_5), 8.32 (d, J = 8.6 Hz, 2H, H_7), 8.23 (d, J = 8.7 Hz, 2H, H_6), 7.87 – 7.74 (m, 4H, $H_{1,4}$), 7.58 – 7.51 (m, 2H, H_3), 7.50 – 7.40 (m, 2H, H_2). ^{13}C NMR (75 MHz, DMSO) δ 157.2 (2 C-N), 148.3 (C-NO_2), 147.8 (C_{para}-N), 144.6, 144.1, 138.9 (C-Au), 130.0, 129.9, 129.3, 129.2, 127.5, 124.5, 117.4. Positive ESI-MS (acetone): m/z = 582.0400 $[M]^+$ (calcd for $C_{23}H_{17}AuClNO_2$: 582.0409). Anal. Calc. for $C_{23}H_{14}AuClN_2O_2$: C, 47.40; H, 2.42; N, 4.81 %. Found: C, 47.34; H, 2.37; N, 4.83 %.

4.4 Antiproliferative assays

The human cancer cell lines A549 (lung adenocarcinoma) and SKOV-3 (ovarian adenocarcinoma) were cultured in DMEM containing GlutaMax-I supplemented with 10% FBS and 1% penicillin/streptomycin, at 37 °C in a humidified atmosphere of 95% of air and 5% CO_2 while 2008 ovarian cancer cells were grown in RPMI with 10% FBS and 1% penicillin/streptomycin in the same conditions. Cells in an exponential growth rate were seeded in 96-well plates) at a concentration of 8 x 10^3 cells/well, and grown for 24 h in complete medium. Solutions of the compounds were prepared by diluting a stock solution in DMSO (10^{-2} M) of the corresponding compound in culture media (the percentage of DMSO in the culture medium never exceeded 0.5%). Cisplatin was dissolved in aqueous solution (10^{-3} M) to avoid ligand exchange reactions with DMSO. Subsequently, intermediate dilutions of the compounds were added to the wells to obtain a final concentration ranging from 0.5 to 100 µM. Following 72 h (A549, SKOV-3) or 48 h (2008) drug exposure, medium was removed and 3 (4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) in PBS/10mM glucose was added to the cells at a final concentration of 0.50 mg/mL and incubated for 2.5 h. Afterwards, the medium was removed and the violet formazan crystals dissolved in DMSO. The optical density of each treatment was quantified in quadruplicate at 540 nm, using a multi-well plate reader (ThermoMax microplate reader, Molecular devices, US), and the percentage of surviving cells was calculated from the ratio of absorbance between treated and untreated cells. The IC_{50}

value was calculated as the concentration reducing the proliferation of the cells by 50% and is presented as a mean (± SD) of at least three independent experiments.

4.5 Enzyme inhibition assays on isolated enzymes

Highly purified aliquots of cytosolic and mitochondrial rat liver thioredoxin reductases[26 27] were utilized. Thioredoxin reductases activity was determined in 0.2M NaKPi buffer, 5 mM EDTA, pH 7.4, by estimating the DTNB reduction in the presence of 0.25 mM NADPH after 5 min incubation with increasing concentration of **1**-**3**. The reaction was started with 1 mM DTNB and followed spectrophotometrically at 412 nm for about 10 min at 25 °C. Yeast glutathione reductase activity was measured in 0.2 M Tris–HCl buffer (pH 8.1), 1 mM EDTA, and 0.25 mM NADPH, with the various compounds. The assay was initiated by addition of 1 mM GSSG and followed spectrophotometrically at 340 nm.

4.6 Analysis of thioredoxin reductase and glutathione reductase activities in cell lysates

2008 ovarian cancer cells, grown in completed RPMI medium, were treated in the presence of **1**-**3** in various conditions. Briefly, 2008 cells (1×10^6) were incubated with increasing concentrations of the three compounds for 48 h. Then, cells were harvested, washed with PBS and lysed with a modified RIPA buffer: 150 mM NaCl, 50 mM Tris-HCl, 1 mM EDTA, 0.1% SDS, 0.5% DOC, 1 mM NaF, supplemented with an antiprotease cocktail ("Complete"Roche, Mannheim, Germany) and 0.1 mM PMSF. After 40 min of incubation at 4 °C, lysates were centrifuged at 14000g for 5 min and aliquots (50µg) of the supernatants were utilized for enzyme activities as described above.

4.7 Total thiols estimation in 2008 cell lysates

After the treatment with 25 or 50 µM of the gold compounds for 24 h, cells were washed in PBS and dissolved with 7.2M guanidine in a buffer containing 0.2M Tris-HCl, 5 mM EDTA, pH 8.1.

3 mM DTNB was then added to titrate the free thiol groups and the absorbance increase was monitored at 412 nm for 5 min.

4.8 Total and oxidized glutathione estimation in cell lysates

First, 5×10^5 2008 cells, were incubated for 48 h with 50 μM of **1** - **3**. Then cells, washed twice with cold PBS, were rapidly lysed and deproteinized with 6% meta-phosphoric acid. After 10 min at 4 °C, samples were centrifuged and supernatants were neutralized with 15% Na_3PO_4 and assayed for total glutathione, essentially as described in[28]

To determine the amount of GSSG, aliquots (0.2 mL) of the obtained samples were derivatized with 2-vinylpyridine in order to block reduced glutathione, and oxidized glutathione was then estimated.[29]

4.9 Redox Western blot analysis of Trx1 and Trx2

The redox state of Trx1 and 2 was detected using a modified Western blot analysis[30] utilizing the protocol describe in ref.[31] Briefly, 2008 ovarian cancer cells incubated with the compounds for 48h, were lysed with 150 μL of urea lysis buffer (100 mM Tris-HCl, pH 8.3) containing 1 mM EDTA, 8 M urea supplemented with 10 mM iodoacetamide (IAM) to derivatize free thiols. The incubation was carried out for 20 min at 37 °C. Subsequently, after the treatment with 3.5 mM DTT for 30 min at 37 °C to reduce the oxidized thiols, cell lysates were derivatized with iodoacetic acid (IAA) (30 mM final concentration) for 30 min at 37 °C in order to give a negative charge. Proteins (0.01 mg) were separated by urea-PAGE gel (7% acrylamide/bis(acrylamide) in 7 M urea) in non-reducing conditions and blotted using TurboSystem (Bio-Rad Laboratories, Hercules, CA, USA). Membranes were probed with the primary antibodies respectively for Trx1 (FL 105) and for Trx2 (H75) (Santa Cruz Biotechnology, Santa Cruz, CA, USA).

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