

ORCA - Online Research @ Cardiff

This is an Open Access document downloaded from ORCA, Cardiff University's institutional repository:https://orca.cardiff.ac.uk/id/eprint/106107/

This is the author's version of a work that was submitted to / accepted for publication.

Citation for final published version:

Vulcano, Rosaria, Pengo, Paolo, Velari, Simone, Wouters, Johan, De Vita, Alessandro, Tecilla, Paolo and Bonifazi, Davide 2017. Toward fractioning of isomers through binding-induced acceleration of azobenzene switching. Journal of the American Chemical Society 139 (50), pp. 18271-18280. 10.1021/jacs.7b09568

Publishers page: http://dx.doi.org/10.1021/jacs.7b09568

Please note:

Changes made as a result of publishing processes such as copy-editing, formatting and page numbers may not be reflected in this version. For the definitive version of this publication, please refer to the published source. You are advised to consult the publisher's version if you wish to cite this paper.

This version is being made available in accordance with publisher policies. See http://orca.cf.ac.uk/policies.html for usage policies. Copyright and moral rights for publications made available in ORCA are retained by the copyright holders.



Toward fractioning of isomers through binding-induced acceleration of azobenzene switching

Rosaria Vulcano,[†] Paolo Pengo,[§] Simone Velari,[#] Johan Wouters,[†] Alessandro De Vita, ^{#,‡} Paolo Tecilla,[§] and Davide Bonifazi^{¥,†,*}

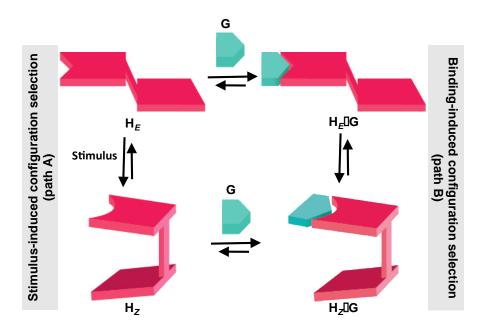
†Department of Chemistry, University of Namur (UNamur), Rue de Bruxelles 61, Namur, 5000 (Belgium); §Department of Chemical and Pharmaceutical Sciences, University of Trieste, Piazzale Europa 1, Trieste, 34127 (Italy); #Department of Engineering and Architecture, University of Trieste, Piazzale Europa 1, Trieste, 34127 (Italy); ‡Department of Physics, King's College London, Strand, London, WC2R 2LS (United Kingdom); *School of Chemistry, Cardiff University, Main Building, Park Place, Cardiff, CF10 3AT (United Kingdom).

KEYWORDS: supramolecular chemistry, azobenzene, molecular switches, host-guest systems, H-bonding interactions, nucleobases, triple H-bonding interactions.

ABSTRACT: The E/Z isomerization process of a uracil-azobenzene derivative in which the nucleobase is conjugated to a phenyldiazene tail is studied in view of its ability to form triply-H-bonded complexes with a suitably complementary 2,6-diacetylamino-4-pyridine ligand. UV-Vis and 1 H-NMR investigations of the photochemical and thermal isomerization kinetics show that the thermal $Z \rightarrow E$ interconversion is four-fold accelerated upon formation of the H-bonded complex. DFT calculations show that the formation of triple H-bonds triggers a significant elongation of the N=N double bond, caused by an increase of its π_g^* antibonding character. This results in a reduction of the N=N torsional barrier and thus in accelerated thermal $Z \rightarrow E$ isomerization. Combined with light controlled $E \rightarrow Z$ isomerization this enables controllable fractional tuning of the two configurational isomers.

Introduction

Isomerization is the process by which one molecule is transformed into another molecule that has exactly the same atoms, but with a different arrangement.^{1,2} In artificial systems, it is usually an external stimulus (*e.g.*, light, electrochemical or thermal inputs, cf. path A, Scheme 1) that triggers such configuration interconversion.^{3–6} A typical example is the *E/Z* photoisomerization of the azobenzene scaffold, for which the single isomer can be selectively produced by the relevant light excitation^{3,7–9} or electrochemical¹⁰ input. This is certainly one of the most studied functional groups allowing spatiotemporal control of the conformational and overall structural properties of molecular systems.^{11–13} Notable applications include the fabrication of photoresponsive architectures in materials science^{3,14–30} and biology.^{31–35} Isomerization also plays a fundamental role in a number of naturally occurring biochemical processes.^{36–38} For example, the *cis-trans* isomerization of amide bonds involving proline is an important step in the folding of proteins. Yet, being relatively slow, it is often rate-limiting.^{39,40} To hasten it, Nature provides specific enzymes referred as "peptidyl-prolyl isomerases" that through binding of the relevant amino acids trigger the switching at the *N*-terminal amide bond of proline (path B, Scheme 1).³⁹

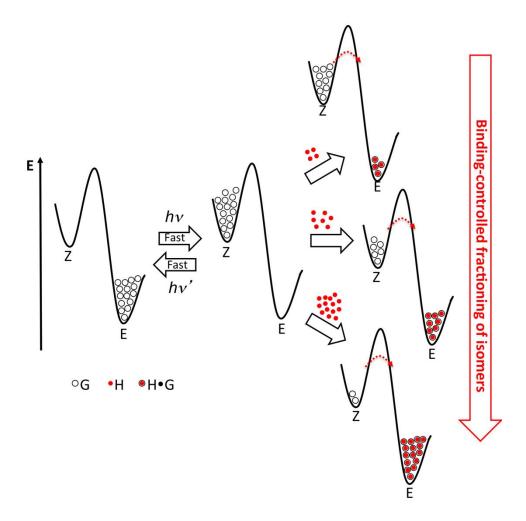


Scheme 1. General scheme for a dual stimuli- and binding-triggered isomerization system. H and G indicate the host and guest molecules, respectively, with the host adopting two different configurations (H_Z and H_E).

Systems that respond to both stimuli- and binding-induced mechanisms of isomerization or switching are, however, more difficult to find (Scheme 1). Hecht and co-workers⁴¹ developed a triply H-bonding motif whose association strength could be modulated by the ring-closed switching state of a photochromic bis(thiazol-4-yl)maleimides. Switchable H-bonded host-guest complexes containing an azobenzene were developed by the groups of Goswami,⁴² Rotello,⁴³ Gadhiri,⁴⁴ Hecht,⁴⁵ and Ballester⁴⁶. The possibility to further modulate the isomerization process through non-covalent bonding was not explored in these previous works and only examples describing the use of gold nanoparticles^{47–49} have been described. This would be appealing, as it could give access to responsive architectures that in the presence of a binder might

enable a controlled structural selection protocol capable of achieving a well-defined target complex structure in a desired concentration. This possibility is explored here, where our specific goal is to develop a complete dual stimuli- and binding-triggered isomerization cycle.

The concept: toward fractioning of isomers. Namely, we wish to investigate the possibility of controlling the relative molar fraction χ and $(1-\chi)$ of the two configuration isomers of a molecule, with the goal of being able to tune these concentrations to arbitrary complementary values in the o-1 range. We note that a range of χ values consistent with thermodynamic equilibrium could be in principle obtained by simply varying the temperature of the system and allowing the system equilibrate, provided that the two isomers have different free energies. This however could be disruptive for the system if the temperatures required for the desired concentration were too high, while this approach would in all cases never achieve a higher concentration for the isomer of higher energy, which we wish to be able to do here. A second possibility would be to use complexation of the isomers to tune the energy levels of the two complexes, so to alter their equilibrium concentration without temperature adjustment. However, full tunability of χ in the o-1 range would be lost in this case, unless an "almost continuum" range of complexes could be devised for this scope.



Scheme 2. Schematic Gibbs free energy landscape illustrating the general principle of the E/Z isomer fractioning through non-covalent complexation.

A different, simpler, and more interesting possibility is to achieve concentrations at "metastable" values outside thermodynamic equilibrium.⁵ A necessary condition for this route to be allowed is that the two isomer states must correspond to local energy minima separated by energy barriers high enough to make any equilibrating transition negligibly slow. In this respect, the E/Z isomerization of azobenzene is certainly one of the most accessible reactions³. The (stable) E and the (metastable) E isomers have separated energies, so that at equilibrium for all temperatures of interest the E isomer is the only present species. Furthermore, if some E concentration is externally induced (E0, by light), the thermal process bringing the system back to equilibrium occurs on a slow (hour) time scale, E1, meets a "slow kinetics" requirement. Under these circumstances, a shift of concentration out of the equilibrium value can be achieved by suitably photoactivating the $E\to E$ 1 (after which, $E\to E$ 1 is also viable), using an appropriate light frequency. Any E2 concentration could in principle be achieved by this way by careful dosing the photon flux and the irradiation time.

We can however conjecture that an alternative route to control the isomeric composition could be achieved by photo-inducing a complete $E \rightarrow Z$ transition, and accelerating the $Z \rightarrow E$ back-transition process (Scheme 2, red arrow) only for a controllable fraction of the Z isomers. If the fast $Z \rightarrow E$ decaying isomers could be those that form a non-covalent complex with appropriately designed guest molecules, their fraction would be easily controlled by tuning the guest concentration. Once the fast, fractional Z to E back-conversion has occurred, all that is needed to "pin" the system indefinitely at the remaining (target) Z isomer fraction, is to correct for the slow drift towards thermodynamic equilibrium. This can be easily achieved by further, timesparse "refreshing" photoactivation steps.

Building on these hypotheses, here we report the first study of an azobenzene-like system whose isomerization properties are responsive to both a luminous stimulus and a non-covalent H-bonded recognition event. The idea is that a perturbation caused by the formation of a multiple H-bonded interaction should trigger a change of the photoisomerisation behavior of the phenyldiazene moiety by modifying the energy barriers associated with isomers interconversion.

Results and Discussion.

Molecular design and solid-state recognition investigations. To engineer such a system, one needs to consider a structure in which the isomerizing unit is in direct conjugation with the recognition site. In this way, any changes occurring at one of the two moieties, is then reflected to the other. Capitalizing on our earlier achievements on the H-bonded uracil•2,6-diamidopyridine complex,^{50–56} we therefore designed and synthesize a photoswitchable uracil-azobenzene derivative presenting direct anchoring of the phenyldiazenyl group to the nucleobase either at the 5- or 6-position (5AUP and 6AUP, respectively Fig. 1).

Figure 1. Molecular structures of the uracil-azobenzene derivatives.

This bridging was selected to favor the establishment of an effective electronic communication between the two moieties. As far as the synthesis is concerned, molecules **5AUP** and **6AUP** were prepared in a two-steps pathway (see SI). To improve the solubility of the molecular scaffold in organic solvents, we equipped the aryl and uracil rings with *tert*-butyl and N1-propyl substituents, respectively. *N*-dialkylated analogue **5AUPP** (Fig. 1) was also prepared as a reference molecule. To probe the H-bonding recognition properties of **5AUP** and **6AUP** we have used **2**,6-diacetylamino-4-[(trimethylsilyl)ethynyl]pyridine (**DAP**) as complementary recognition unit (Fig. 1).50-52,56

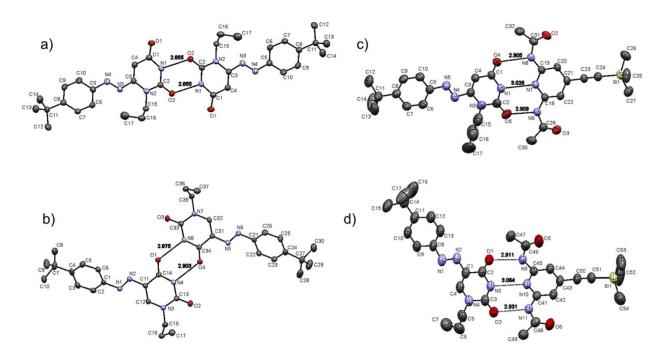


Figure 2. ORTEP representation of the X-ray structures of (a) *E*-6AUP, (b) *E*-5AUP, (c) *E*-6AUP·DAP and (d) *E*-5AUP·DAP. The space groups are P -1, P $_{2_1}$ /n, P -1, and P $_{2_1}$ /c for a), b), c) and d), respectively.

Suitable single crystals for X-ray diffraction analysis were grown by slow evaporation of a toluene solution containing a 1:1 mixture of the relevant uracil-azobenzene derivative and **DAP**. As expected, the X-ray structure (Fig. 2) reveals the presence of the H-bonded complex, in which the *E* isomer of **5AUP** (*E*-**5AUP**)

and **6AUP** (*E*-**6AUP**) engages triple H-bonding interactions with **DAP** (relevant distances for *E*-**5AUP·DAP**: 2.911; 3.064 and 2.931 Å for O1····N9, N3····N10 and O2····N11, respectively). These values are comparable to those previously reported for similar H-bonded complexes.^{57,58} Any attempts to obtained the X-ray structure for the *Z*-**5AUP·DAP** and *Z*-**6AUP·DAP** complexes failed.

E/Z isomerization studies in solution for the uracil derivatives. The electronic absorption spectrum of 5AUP in toluene is depicted in Figure 3a. The molecule exhibits two UV absorption bands at 360 nm and 442 nm, which are attributed to π - π * (of the E isomer) and n- π * (of the Z isomer) electronic transitions, respectively. ⁵⁹⁻⁶¹ When irradiated at 360 nm, remarkable changes are observed in the absorption spectra of 5AUP (Fig. 3b), with an intense and fast increase of a new band centered at around 300 nm and a substantial increase of the intensity of the n- π * band at 442 nm with a concomitant disappearance of the π - π * absorption. Neat isosbestic points are maintained throughout the photoirradiation experiments, showing the clean occurrence of the nearly quantitative $E \rightarrow Z$ isomerization. Thermal $Z \rightarrow E$ interconversion producing the most thermodynamically-stable E isomer occurs very slowly as shown in Fig. 3c (see also below). On the other hand, irradiation at 442 nm causes a fast $Z \rightarrow E$ isomerization, resulting in decreasing of the n- π * absorption and a full restoring of the π - π * band (Fig. 4a). Repetitive photoinduced switching cycles evidenced an excellent fatigue resistance and remarkable photostability of the system (Fig. 4b). Taken all together, these data suggest that the substitution of the phenyl ring with the nucleobase has little influence on the photochemical properties, with 5AUP showing the typical E/Z switching behavior of the archetypal azobenzene. ^{61,62}

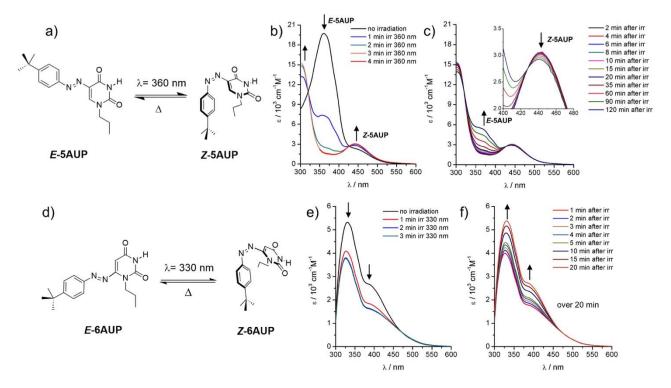


Figure 3. E/Z Isomerization equilibria for **5AUP** (a) and **6AUP** (d) in toluene. Photo $E \rightarrow Z$ (b and e) and thermal $Z \rightarrow E$ (c and f) isomerization of **5AUP** (2.5 ·10⁻⁵ M) and **6AUP** (5.4·10⁻⁵ M) in toluene at 25 °C. Inset: zoom in the n- \mathbb{Z} absorption region. The thermal $Z \rightarrow E$ isomerization was followed over 120 and 20 min period for 5UAP and 6AUP, respectively.

When looking at the photoisomerization of **6AUP**, the characteristic azobenzene bands are no longer distinguishable in the absorption spectrum of this regioisomer (Fig. 3e). Likely in **6AUP**, the azobenzene unit is involved in a 1,4-type conjugation with the carbonyl group of the uracil moiety. When irradiated at 360 nm, the absorption band in the 300-400 nm spectral region decreases, reaching saturation after 3 min of irradiation. The weak features of the *Z* isomer seem to be substantially masked within the strong absorption envelope of the conjugated molecule. When left into the dark, full recovery of the initial absorption spectra is obtained after 20 min. Together with the possibility of reaching nearly quantitate two-way isomerization, these data suggest that the slower reaction dynamics of **5AUP** make this regioisomer the best candidate to intercept both configurational isomers and study their binding properties with **DAP** in solution with classical spectroscopic techniques like ¹H-NMR. For this reason, only regioisomer **5AUP** has been used to perform the binding studies.

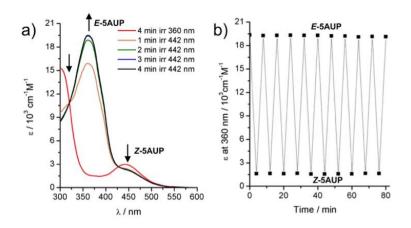


Figure 4. a) $Z \rightarrow E$ photoisomerization of **5AUP** (2.5 ·10⁻⁵ M); b) repetitive photoinduced switching cycles of **5AUP** obtained by alternating irradiation at 360 and 442 nm.

E/Z Isomerization and binding studies for the supramolecular complexes in solution. To shed further light on the recognition properties of 5AUP in solution, ¹H-NMR titration and Job's plot analysis were performed for both Z and E isomers to estimate the association constants (K_i) and the binding stoichiometry in solution. In both cases, increasing concentration of **DAP** caused a progressive downfield shift of the uracil NH proton under a regime of fast equilibrium exchange (Fig. 5). As evidenced at the solid state, the Job's plots analysis confirmed the 1:1 binding stoichiometry in solution for the 5AUP-DAP complex (see SI). The association constant (K_E , calculated through the data fitting with Dynafit software ⁶³) for the E-5AUP-DAP complex was found to be 6710 \pm 800 M⁻¹, which is half of that measured for the reference Upr-DAP complex under the same experimental conditions (see SI). This is in line with the literature findings obtained for other uracil-derived molecular scaffolds in toluene. ⁵⁰

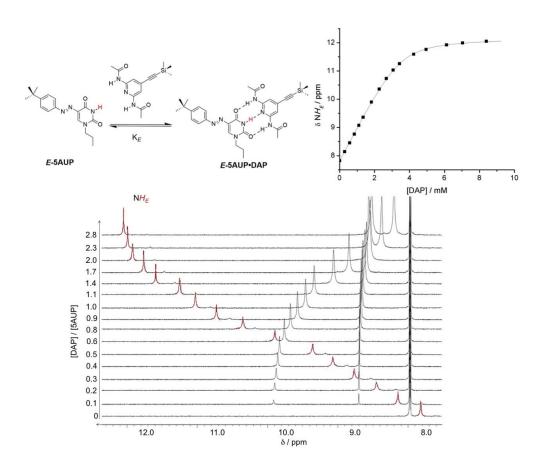


Figure 5. Dark ¹H-NMR titrations of **E-5AUP** (3.0 mM in toluene- d_8) upon addition of increasing amounts of **DAP** at 25 °C. The red signal corresponds to the imide proton of the uracil unit in free **E-5AUP** (δ_{NH} = 7.8 ppm) and in **E-5AUP·DAP** complex (δ_{NH} = 12.1 ppm after addition of 2.8 equiv of **DAP**), respectively.

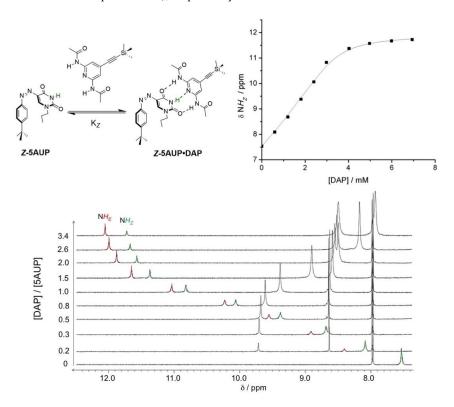


Figure 6. Dark ¹H-NMR titrations of **Z-5AUP** obtained after irradiation at 360 nm of the **5AUP** sample (3.8 mM in toluene- d_8) upon addition of increasing amounts of **DAP** at 25 °C. The green and red signals correspond to the imide proton of the uracil core in the **Z-5AUP- and E-5AUP-containing** species, respectively.

Regarding the Z-5AUP·DAP complex, the titration experiments (Fig. 6) were performed in the dark at constant temperature (T = 298 K) with a freshly, light-isomerized solution of **5AUP** ($\delta_{NH} = 7.5$ ppm for the imide proton in the absence of **DAP**, no presence of **E-5AUP** was detected at the beginning of the titration). By adding increasing amounts of DAP, two different NH proton resonances started appearing, reaching the saturation chemical shift values of 12.1 and 11.7 ppm after the addition of 3.4 equivalents of DAP (Fig. 6). Considering the chemical shifts observed during the titration experiments performed for *E-5AUP*·DAP, the resonance at lower fields are attributed to E-5AUP, while that at higher fields to the Z isomer. Upon increasing additions of **DAP**, the intensity of the imide proton signal for the E isomer increases at the expenses of that pertaining to the Z isomer, which suggests $Z \rightarrow E$ interconversion (Fig. 6). As one can notice, this gives rise to defined fractions of the two isomers depending on the DAP concentration. For instance at [DAP]/[5AUP] ratios of 0.2, 0.5, and 2 the molar fractions (χ_E) of the *E* isomer (free and complexed) is 0.21, 0.34 and 0.66, respectively (Fig. 6). Because of this thermal $Z \rightarrow E$ interconversion, the estimation of the association constant (K_Z) for the Z-5AUP-DAP complex revealed to be not straightforward as its concentration is interconnected with the progressive increase of the other complex, namely *E-5AUP-DAP*. The fitting model (applied with Dynafit software) employed for estimating the association constant need to use as independent variables the concentration of DAP and that of the two isomers (measured from the integration of the imidic proton resonances). Using the K_E value estimated from the titration experiments of *E*-5AUP, we could estimate the K_Z value for the *Z*-5AUP·DAP complex to be 4040 ± 355 M⁻¹. Notably, this is almost half of that measured for its isomer *E-5AUP*·DAP.

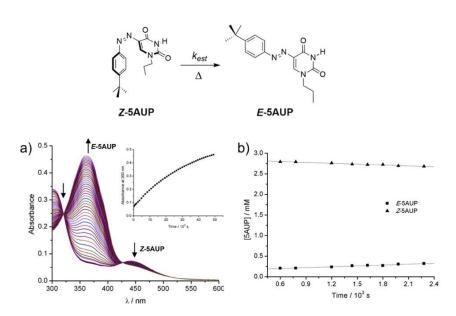


Figure 7. a) UV-Vis measurements of the thermal $Z\rightarrow E$ -5AUP isomerization at 25 °C, after sample irradiation at 360 nm (3.0·10⁻⁵ M toluene solution); b) Time-dependent variation of the concentration of the isomers estimated from the integrals of the NCH₂ protons peaks area as measured by ¹H-NMR after sample irradiation at 360 nm (3.0·10⁻³ M toluene- d_8 solution).

The rate constant of thermal $Z \rightarrow E$ isomerization for **5AUP** was investigated in the dark by both UV-Vis and ¹H-NMR measurements. The UV-Vis experiment shows an increasing of the absorbance at 360 nm over the time, corresponding to the increasing of the concentration for the E isomer (Fig. 7a). The process follows a

first order kinetic equation with a rate constant (k_{est}) value of 2.7 ± 0.01 · 10⁻⁵ s⁻¹ ($t_{1/2}$ = 7.1 h, see SI). The ¹H-NMR analysis was restricted to the first instants of the reaction according to the initial rate method (Fig. 7b). The k_{est} value obtained from ¹H-NMR measurements corresponds to 2.5 ± 0.2 · 10⁻⁵ s⁻¹, which is in good agreement with the value obtained from the UV-Vis measurements. As a control experiment, we have studied the thermal $Z\rightarrow E$ isomerization of the reference dialkylated uracil derivative (**5AUPP**), where also the N(3) position bears a propyl chain. The k_{est} value obtained from UV-Vis measurements corresponds to 3.06 ± 0.03 · 10⁻⁵ s⁻¹ (see SI), which is consistent with that calculated for the $Z\rightarrow E$ conversion of **5AUP** in the absence of **DAP**.

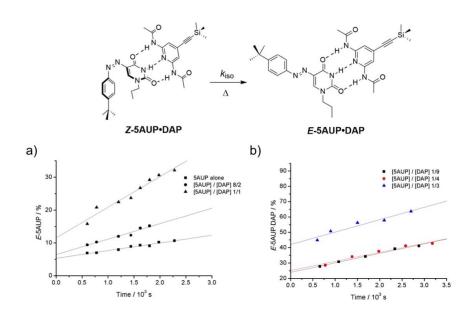


Figure 8. a) Kinetic profiles of the thermal $Z \rightarrow E$ isomerization of **5AUP** in toluene- d_8 in the absence and in the presence of different **DAP** concentrations measured by ¹H-NMR at 25 °C after sample irradiation at 360 nm; b) Kinetic profiles of the thermal $Z \rightarrow E$ isomerization of **5AUP**·**DAP** complex in the presence of exceeding quantities of **DAP** measured by ¹H-NMR at 25 °C in toluene- d_8 after samples irradiation at 360 nm; initial sample concentrations: (■) [**5AUP**]₀ = 1.0·10⁻³ M, [**DAP**]₀ = 9.4·10⁻³ M; (●) [**5AUP**]₀ = 2.2·10⁻³ M, [**DAP**]₀ = 9.6·10⁻³ M, and (▲) [**5AUP**]₀ = 3.2·10⁻³ M, [**DAP**]₀ = 10.2·10⁻³ M.

To understand if the supramolecular interaction has an influence on the thermal $Z\rightarrow E$ isomerization rate, further ¹H-NMR investigations have been made using Z-5AUP solutions in the presence of variable DAP concentrations (Fig. 8). In the presence of a large excess of DAP, the maximum concentration of complex (about 96%) is achieved (Fig. 8b). This allows calculating the rate constant of the thermal $Z\rightarrow E$ isomerization process within the complex 5AUP·DAP, which resulted to be 1.0 \pm 0.3 \cdot 10⁻⁴ s⁻¹ (see SI). It is now apparent that the $Z\rightarrow E$ isomerization process within the complex is about 4 times faster than that occurring for 5AUP alone (2.5 \pm 0.2 \cdot 10⁻⁵ s⁻¹).

Scheme 3 summarizes the relevant equilibria and kinetic processes involving the *Z*-5AUP and *E*-5AUP isomers along with their complexes with **DAP**. The isomer fractioning concept implies the photogeneration of *Z*-5AUP in the presence of a given concentration of **DAP**. The fast isomerization within the H-bonded complex will produce a *Z/E* mixture with an isomeric composition determined by the amount of *Z*-5AUP·DAP complex and subjected to a drift toward the thermodynamic equilibrium composition due to the slower background thermal isomerization of free *Z*-5AUP. As a consequence, the possibility to produce a

Z/E mixture with a defined isomeric composition and stable in time depends on the k_{iso}/k_{est} ratio. On the other hand, being the equilibration of **DAP** between the two **5AUP** isomers fast in the isomerization time scale, this process should not influence the system which experiences a time averaged composition ruled by the association constants. In our case the k_{iso}/k_{est} ratio is about 4. Although this acceleration does not to allow a complete separation between the two kinetic processes, a simulation shows that, for example, starting from a 1:1 mixture of **Z-5AUP/DAP** after about 10 hours the complex is fully isomerized but the χ_E (free and complexed) is 0.78 with the amount exceeding the 50% deriving from the interconversion of the free **Z-5AUP**. However, the isomeric composition obtained is relatively stable in time. For instance, after 5 hours the χ_E value has changed only by 0.09, going from 0.78 to 0.87.

E-5AUP

$$K_{\text{est}} =$$
 $K_{\text{est}} =$
 $K_{\text{est}} =$

Scheme 3. Summary of the relevant complexation equilibria and isomerization kinetics involving 5AUP and DAP.

Theoretical rationale. To shed further light on the causes affecting the acceleration of the switching process, we have performed density functional theory (DFT) calculations, using the PWSCF code of the Quantum ESPRESSO distribution, 64 and the Perdew-Burke-Ernzehof (PBE) functional 65 which was used before in careful studies of similar systems. 66,67 First, we optimized the gas-phase structures of **DAP**, **Z-5AUP**, **E-5AUP** molecules and those of their **Z**- and **E-5AUP·DAP** complexes (see SI). The calculations confirm the higher stability of the *E* isomers with respect to that of the *Z* both in both free and complex species. In particular, the calculated energy of the metastable *Z* isomer is 0.65 and 0.61 eV higher than the *E* level for **5AUP** and **5AUP·DAP**, respectively. As expected, these energy differences between this configurational isomers are comparable with the values previously computed for azobenzene by DFT techniques (0.66 eV) as well as a broad range of techniques including highly sophisticated coupled-cluster and diffusion Monte Carlo techniques (0.52-0.73 eV) 67 . Analysis of our DFT results suggests a peculiar redistribution of the electron density, which occurs around the N=N bond upon formation of the complex of **Z-5UAP** molecule. This is visible in Figure 9, where the electron density difference ρ_{tot} - $\Sigma \rho_{\text{mol}}$ is shown,

portraying the electron density readjustment following the "switching on" of the interaction between the two molecules in the complex (ρ_{tot} is the electron density of the complex and $\Sigma \rho_{mol}$ is the sum of the densities of the two isolated single molecules with same atomic position, in the gas phase). The alternated lobes of opposite signs (Figure 9, inset) fingerprints the onset of hybridization of electronic states, mixing states with π_g^* antibonding orbital components with bonding molecular states that do not have a nodal plane between the two N atoms of the C-N=N-C moiety. This is expected to lower the torsional barrier associated with the C-N=N-C group upon formation of the **Z-5UAP·DAP** complex, consistent with the experimental observations.

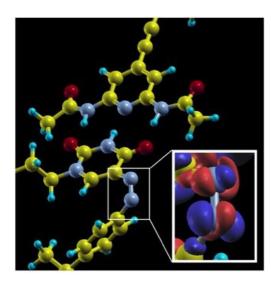


Figure 9. Side-view of the optimized structure of **Z-5AUP·DAP**. The zoomed region displays the change of the calculated electron density in the N=N bond region of **Z-5AUP** upon complexation with **DAP** (electron density accumulation is depicted in red and depletion in blue).

Finally, in an effort to estimate the effect of complexation on the $Z\rightarrow E$ isomerization barrier, we investigated the Z-5AUP-DAP $\rightarrow E$ -5AUP-DAP and the Z-5AUP $\rightarrow E$ -5AUP conversion, using the nudged elastic band (NEB) technique 68,69,70 to model the transition barrier. We note that solvent effects, as well as temperature effects should be fully included to properly account for the enthalpic and entropic contributions that define the free energy barrier relevant to the experimental interconversion conditions. While some techniques exist that could capture complex electron-phonon dynamics and level crossing occurring in system like the present ones during the isomerization transition⁷¹, a simple estimate of the barrier lowering upon DAP complexation relying on error cancellation can be simply obtained from our DFT static potential energy results. These suggest that the Z to E barrier gets smaller upon Z-5AUP complexation with DAP. Our computed 60 meV energy barrier reduction value (which would correspond to a speed-up factor of about 10 at 300K) is furthermore comparable with the 34meV free energy barrier reduction value (corresponding to the experimentally observed speedup of about 4) obtained from the ratio of our experimental thermal conversion rates further assuming the same attempt rate for the $Z\rightarrow E$ conversion in free Z-5AUP and H-bonded complex Z-5AUP-DAP.

Conclusions

In conclusion, in this work we have synthesized photoswitchable uracil-azobenzene conjugates presenting direct anchoring of the phenyldiazenyl group to the uracil nucleobase, with the 5-functionalized derivatives depicting a slow thermal switching process and a nearly quantitate two-way isomerization. In the presence of a 2,6-diamidopyridine, triply H-bonded complexes were formed both in solution and at the solid state. UV-Vis and 'H-NMR investigations of the photochemical and thermal isomerization kinetics in toluene solutions showed that the thermal $Z \rightarrow E$ interconversion is four-fold accelerated upon formation of the Hbonded complex when compared to that of the uracil derivative alone. DFT calculations suggest that the formation of the triply H-bonded complex trigger weakening and elongation of the N=N double bond, through increase of its π_g^* antibonding character. This is expected to reduce the N=N torsional barrier and thus rationalize the acceleration of the thermal $Z \rightarrow E$ isomerization. This binding-induced acceleration of the isomerization process revealed to give access to tailored isomers fractioning (χ) with limited variation of the mixture over a few hours, although in our system the acceleration factor is rather small to ensure a complete time separation of the two kinetic processes. Structural modifications of the aza-derivative are currently under investigation to increase the gap between the isomerization rates within and outside the complex. Such types of systems are indeed of great appeal as they could give access to responsive architectures, which upon exposition to an external stimulus undergo a structural perturbation (e.g., unfolding, unzipping and denaturation).72-75 This reactive state could eventually develop into a new minimum in the presence of the relevant binder (e.g., drugs, nucleic acid strand, enzyme active sites and metabolites), the concentration of which can control both structure and its fraction. For instance, this modified system could be used to design photoresponsive nucleic acids that could undergo controlled unfolding and folding of DNA helixes,76 possibly opening new horizons into light-triggered gene manipulation.

ASSOCIATED CONTENT

Supporting Information

Synthetic protocols and spectroscopic characterisations. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

bonifazid@cardiff.ac.uk

Notes

The authors declare no competing financial interest.

ACKNOWLEDGEMENT

D.B. gratefully acknowledges the Science Policy Office of the Belgian Federal Government (BELSPO-IAP 7/05 project) and the EU through the ERC COLORLAND project. P.T. and D.B. thank the HORIZON2020 Marie Sklodowska Curie Actions through the INFUSION project. R.V. thanks UNamur for her doctoral and teaching assistant position. The authors thank the physical chemistry and characterization (PC²) at the UNamur and Bernadette Norberg for the X-ray measurements and data interpretation.

REFERENCES

- (1) Dugave, C.; Demange, L. *Chem. Rev.* **2003**, *103*, 2475.
- (2) Hassam, M.; Taher, A.; Arnott, G. E.; Green, I. R.; van Otterlo, W. A. L. *Chem. Rev.* 2015, 115, 5462.
- (3) Bléger, D.; Hecht, S. Angew. Chem. Int. Ed. 2015, 54, 11338.
- (4) Mukhopadhyay, R. D.; Praveen, V. K.; Ajayaghosh, A. Mater. Horiz. 2014, 1, 572.
- (5) Kathan, M.; Hecht, S. Chem. Soc. Rev. 2017, 46, 5536.
- (6) Klajn, R. Chem. Soc. Rev. 2014, 43, 148.
- (7) Hartley, G. S. *Nature* **1937**, *140*, **28**1.
- (8) Moreno, J.; Gerecke, M.; Grubert, L.; Kovalenko, S. A.; Hecht, S. Angew. Chem. Int. Ed. 2016, 55, 1544.
- (9) Yagai, S.; Karatsu, T.; Kitamura, A. *Chem. Eur. J.* **2005**, *11*, 4054.
- (10) Goulet-Hanssens, A.; Utecht, M.; Mutruc, D.; Titov, E.; Schwarz, J.; Grubert, L.; Bléger, D.; Saalfrank, P.; Hecht, S. J. Am. Chem. Soc. 2017, 139, 335.
- (11) Bléger, D.; Schwarz, J.; Brouwer, A. M.; Hecht, S. J. Am. Chem. Soc. 2012, 134, 20597.
- (12) Klajn, R. Pure Appl. Chem. 2010, 82, 2247.
- (13) Zeitouny, J.; Aurisicchio, C.; Bonifazi, D.; De Zorzi, R.; Geremia, S.; Bonini, M.; Palma, C.-A.; Samorì, P.; Listorti, A.; Belbakra, A.; Armaroli, N. *J. Mater. Chem.* **2009**, *19*, 4715.
- (14) Liaros, N.; Couris, S.; Maggini, L.; De Leo, F.; Cattaruzza, F.; Aurisicchio, C.; Bonifazi, D. *ChemPhysChem* **2013**, *14*, 2961.
- (15) Maggini, L.; Marangoni, T.; Georges, B.; Malicka, J. M.; Yoosaf, K.; Minoia, A.; Lazzaroni, R.; Armaroli, N.; Bonifazi, D. *Nanoscale* **2013**, *5*, 634.
- (16) Weber, C.; Liebig, T.; Gensler, M.; Zykov, A.; Pithan, L.; Rabe, J. P.; Hecht, S.; Bléger, D.; Kowarik, S. *Sci. Rep.* **2016**, *6*, 25605.
- (17) Bléger, D.; Ciesielski, A.; Samorì, P.; Hecht, S. Chem. Eur. J. 2010, 16, 14256.
- (18) Lee, J.; Klajn, R. Chem. Commun. 2015, 51, 2036.
- (19) Kundu, P. K.; Klajn, R. ACS Nano 2014, 8, 11913.
- (20) Klajn, R.; Browne, K. P.; Soh, S.; Grzybowski, B. A. Small 2010, 6, 1385.
- (21) Mahesh, S.; Gopal, A.; Thirumalai, R.; Ajayaghosh, A. J. Am. Chem. Soc. 2012, 134, 7227.

- Gopal, A.; Hifsudheen, M.; Furumi, S.; Takeuchi, M.; Ajayaghosh, A. *Angew. Chem. Int. Ed.* **2012**, *51*, 10505.
- (23) Rajaganesh, R.; Gopal, A.; Mohan Das, T.; Ajayaghosh, A. Org. Lett. 2012, 14, 748.
- (24) Das, S.; Ranjan, P.; Maiti, P. S.; Singh, G.; Leitus, G.; Klajn, R. Adv. Mater. 2013, 25, 422.
- (25) Mativetsky, J. M.; Pace, G.; Elbing, M.; Rampi, M. A.; Mayor, M.; Samorì, P. J. Am. Chem. Soc. **2008**, 130, 9192.
- (26) Castellanos, S.; Goulet-Hanssens, A.; Zhao, F.; Dikhtiarenko, A.; Pustovarenko, A.; Hecht, S.; Gascon, J.; Kapteijn, F.; Bléger, D. *Chem. Eur. J.* **2016**, 22, 746.
- (27) Ferri, V.; Elbing, M.; Pace, G.; Dickey, M. D.; Zharnikov, M.; Samorì, P.; Mayor, M.; Rampi, M. A. *Angew. Chem. Int. Ed.* **2008**, *47*, 3407.
- Döbbelin, M.; Ciesielski, A.; Haar, S.; Osella, S.; Bruna, M.; Minoia, A.; Grisanti, L.; Mosciatti, T.; Richard, F.; Prasetyanto, E. A.; De Cola, L.; Palermo, V.; Mazzaro, R.; Morandi, V.; Lazzaroni, R.; Ferrari, A. C.; Beljonne, D.; Samorì, P. *Nat. Commun.* 2016, 7, 11090.
- (29) Mukhopadhyay, R. D.; Praveen, V. K.; Hazra, A.; Maji, T. K.; Ajayaghosh, A. Chem. Sci. 2015, 6, 6583.
- (30) Babu, S. S.; Praveen, V. K.; Ajayaghosh, A. Chem. Rev. 2014, 114, 1973.
- (31) Velema, W. A.; Szymanski, W.; Feringa, B. L. J. Am. Chem. Soc. 2014, 136, 2178.
- (32) Broichhagen, J.; Frank, J. A.; Trauner, D. Acc. Chem. Res. 2015, 48, 1947.
- (33) Woolley, G. A. Acc. Chem. Res. 2005, 38, 486.
- (34) Schütt, M.; Krupka, S. S.; Milbradt, A. G.; Deindl, S.; Sinner, E.-K.; Oesterhelt, D.; Renner, C.; Moroder, L. *Chem. Biol.* **2003**, *10*, 487.
- (35) Stein, M.; Middendorp, S. J.; Carta, V.; Pejo, E.; Raines, D. E.; Forman, S. A.; Sigel, E.; Trauner, D. *Angew. Chem. Int. Ed.* **2012**, *51*, 10500.
- (36) Chen, W. P. Process Biochem. 1980, 15, 36.
- (37) Graham Solomons, J. T.; Zimmerly, E. M.; Burns, S.; Krishnamurthy, N.; Swan, M. K.; Krings, S.; Muirhead, H.; Chirgwin, J.; Davies, C. *J. Mol. Biol.* **2004**, 342, 847.
- (38) Terada, T.; Mukae, H.; Ohashi, K.; Hosomi, S.; Mizoguchi, T.; Uehara, K. Eur. J. Biochem. 1985, 148, 345.
- (39) Fischer, G.; Schmid, F. X. Biochemistry 1990, 29, 2205.
- (40) Balbach, J.; Schmid, F. X. In *Mechanisms of protein folding*; Pain, R. H., Ed.; Oxford University Press: Oxford, 2000; pp 212–249.
- (41) Herder, M.; Pätzel, M.; Grubert, L.; Hecht, S. Chem. Commun. 2011, 47, 460.
- (42) Goswami, S.; Ghosh, K.; Halder, M. *Tetrahedron Lett.* **1999**, 40, 1735.
- (43) Goodman, A.; Breinlinger, E.; Ober, M.; Rotello, V. M. J. Am. Chem. Soc. 2001, 123, 6213.
- (44) Vollmer, M. S.; Clark, T. D.; Steinem, C.; Ghadiri, M. R. Angew. Chem. Int. Ed. 1999, 38, 1598.
- (45) Lohse, M.; Nowosinski, K.; Traulsen, N. L.; Achazi, A. J.; von Krbek, L. K. S.; Paulus, B.; Schalley, C. A.; Hecht, S. *Chem. Commun.* **2015**, *51*, 9777.
- (46) Arroyave, F. A.; Ballester, P. J. Org. Chem. 2015, 80, 10866.

- (47) Hallett-Tapley, G. L.; D'Alfonso, C.; Pacioni, N. L.; McTiernan, C. D.; González-Béjar, M.; Lanzalunga, O.; Alarcon, E. I.; Scaiano, J. C. *Chem. Commun.* **2013**, *49*, 10073.
- (48) Titov, E.; Lysyakova, L.; Lomadze, N.; Kabashin, A. V.; Saalfrank, P.; Santer, S. J. Phys. Chem. C 2015, 119, 17369.
- (49)Simoncelli, S.; Aramendía, P. F.; Klajn, R.; Stoddart, J. F.; Grzybowski, B. A.; Zheng, Y. B.; Yang, Y.-W.; Jensen, L.; Fang, L.; Juluri, B. K.; Flood, A. H.; Weiss, P. S.; Stoddart, J. F.; Huang, T. J.; Molen, S. J. van der; Liao, J.; Kudernac, T.; Agustsson, J. S.; Bernard, L.; Calame, M.; Wees, B. J. van; Feringa, B. L.; Schönenberger, C.; Yoon, J. H.; Yoon, S.; Klajn, R.; Bishop, K. J. M.; Grzybowski, B. A.; Sidhaye, D. S.; Kashyap, S.; Sastry, M.; Hotha, S.; Prasad, B. L. V.; Dulkeith, E.; Morteani, A. C.; Niedereichholz, T.; Klar, T. A.; Feldmann, J.; Levi, S. A.; Veggel, F. C. J. M. van; Reinhoudt, D. N.; Möller, M.; Gittins, D. I.; Zhang, J.; Whitesell, J. K.; Fox, M. A.; Shin, K.; Shin, E. J.; Manna, A.; Chen, P.-L.; Akiyama, H.; Wei, T.-X.; Tamada, K.; Knoll, W.; Hallett-Tapley, G. L.; D'Alfonso, C.; Pacioni, N. L.; McTiernan, C. D.; González-Béjar, M.; Lanzalunga, O.; Alarcon, E. I.; Scaiano, J. C.; Nishimura, N.; Sueyoshi, T.; Yamanaka, H.; Imai, E.; Yamamoto, S.; Hasegawa, S.; Birnbaum, P. P.; Linford, J. H.; Style, D. W. G.; Hartley, G. S.; Dunn, N. J.; Humphries, W. H.; Offenbacher, A. R.; King, T. L.; Gray, J. A.; Sanchez, A. M.; Rossi, R. H.; Ciccone, S.; Halpern, J.; Taylor, H. S.; Oxford, S. M.; Henao, J. D.; Yang, J. H.; Kung, M. C.; Kung, H. H.; Nigra, M. M.; Arslan, I.; Katz, A.; Sun, Y.; Mayers, B.; Herricks, T.; Xia, Y.; Hagen, S.; Kate, P.; Peters, M. V.; Hecht, S.; Wolf, M.; Tegeder, P.; Jung, U.; Filinova, O.; Kuhn, S.; Zargarani, D.; Bornholdt, C.; Herges, R.; Magnussen, O.; Solomun, T.; Christmann, K.; Baumgaertel, H.; McNellis, E. R.; Meyer, J.; Reuter, K.; Bartram, M. E.; Koel, B. E.; Dozier, W.; Drake, J.; Klafter, J.; Blaaderen, A. van; Vrij, A.; Turkevich, J.; Stevenson, P. C.; Hillier, J.; McGilvray, K. L.; Granger, J.; Correia, M.; Banks, J. T.; Scaiano, J. C.; Graf, C.; Vossen, D. L. J.; Imhof, A.; Blaaderen, A. van; Stöber, W.; Fink, A.; Bohn, E. Catal. Sci. Technol. 2015, 5, 2110.
- (50) Dorđević, L.; Marangoni, T.; Miletić, T.; Rubio-Magnieto, J.; Mohanraj, J.; Amenitsch, H.; Pasini, D.; Liaros, N.; Couris, S.; Armaroli, N.; Surin, M.; Bonifazi, D. *J. Am. Chem. Soc.* **2015**, *137*, 8150.
- (51) Marangoni, T.; Bonifazi, D. Nanoscale 2013, 5, 8837.
- (52) Llanes-Pallas, A.; Yoosaf, K.; Traboulsi, H.; Mohanraj, J.; Seldrum, T.; Dumont, J.; Minoia, A.; Lazzaroni, R.; Armaroli, N.; Bonifazi, D. *J. Am. Chem. Soc.* **2011**, *133*, 15412.
- (53) Yoosaf, K.; Llanes-Pallas, A.; Marangoni, T.; Belbakra, A.; Marega, R.; Botek, E.; Champagne, B.; Bonifazi, D.; Armaroli, N. *Chem. Eur. J.* **2011**, *17*, 3262.
- (54) Marangoni, T.; Mezzasalma, S. A.; Llanes-Pallas, A.; Yoosaf, K.; Armaroli, N.; Bonifazi, D. *Langmuir* **2011**, *27*, 1513.
- (55) Llanes-Pallas, A.; Palma, C.; Piot, L.; Belbakra, A.; Listorti, A.; Prato, M.; Samori, P.; Armaroli, N.; Bonifazi, D. *J. Am. Chem. Soc.* **2009**, *131*, 509.
- (56) Llanes-Pallas, A.; Matena, M.; Jung, T.; Prato, M.; Stöhr, M.; Bonifazi, D. *Angew. Chem. Int. Ed.* **2008**, 47, 7726.
- (57) Feibush, B.; Figueroa, A.; Charles, R.; Onan, K. D.; Feibush, P.; Karger, B. L. *J. Am. Chem. Soc.* **1986**, *108*, 3310.
- (58) Hamilton, A. D.; Van Engen, D. J. Am. Chem. Soc. 1987, 109, 5035.
- (59) Vögtle, F.; Gorka, M.; Hesse, R.; Ceroni, P.; Maestri, M.; Balzani, V. *Photochem. Photobiol. Sci.* **2002**, *1*, 45.
- (60) Conti, I.; Garavelli, M.; Orlandi, G. J. Am. Chem. Soc. 2008, 130, 5216.
- (61) Takamiya, N.; Tada, M.; Kishimoto, T.; Takano, T.; Sato, Y. Bull. Sci. Eng. Res. 1986, 114, 12.
- (62) Bandara, H. M. D.; Burdette, S. C. Chem. Soc. Rev. 2012, 41, 1809.

- (63) Kuzmic, P. Anal. Biochem. 1996, 237, 260.
- (64) Giannozzi, P.; Baroni, S.; Bonini, N.; Calandra, M.; Car, R.; Cavazzoni, C.; Ceresoli, D.; Chiarotti, G. L.; Cococcioni, M.; Dabo, I.; Dal Corso, A.; de Gironcoli, S.; Fabris, S.; Fratesi, G.; Gebauer, R.; Gerstmann, U.; Gougoussis, C.; Kokalj, A.; Lazzeri, M.; Martin-Samos, L.; Marzari, N.; Mauri, F.; Mazzarello, R.; Paolini, S.; Pasquarello, A.; Paulatto, L.; Sbraccia, C.; Scandolo, S.; Sclauzero, G.; Seitsonen, A. P.; Smogunov, A.; Umari, P.; Wentzcovitch, R. M. J. Phys. Condens. Matter 2009, 21, 395502.
- (65) Perdew, J. P.; Burke, K.; Ernzerhof, M. Phys. Rev. Lett. 1996, 77, 3865.
- (66) Turanský, R.; Konôpka, M.; Doltsinis, N. L.; Štich, I.; Marx, D. Phys. Chem. Chem. Phys. 2010, 12, 13922.
- (67) Dubecký, M.; Derian, R.; Mitas, L.; Štich, I. J. Chem. Phys. 2010, 133, 244301.
- (68) Jonsson, H.; Mills, K. W. J. *Classical and Quantum Dynamics in Condensed Phase Simulations*, 1998th ed.; Berne, B. J., Ciccotti, G., Coker, D. F., Eds.; World Scientific: Singapore, 1998.
- (69) Henkelman, G.; Uberuaga, B. P.; Jónsson, H. J. Chem. Phys. 2000, 113, 9901.
- (70) Henkelman, G.; Jónsson, H. J. Chem. Phys. 2000, 113, 9978.
- (71) Neukirch, A. J.; Shamberger, L. C.; Abad, E.; Haycock, B. J.; Wang, H.; Ortega, J.; Prezhdo, O. V.; Lewis, J. P. *J. Chem. Theory Comput.* **2014**, *10*, 14.
- (72) Wang, R.; Jin, C.; Zhu, X.; Zhou, L.; Xuan, W.; Liu, Y.; Liu, Q.; Tan, W. J. Am. Chem. Soc. 2017, 139, 9104.
- (73) Stafforst, T.; Hilvert, D. Angew. Chem. Int. Ed. 2010, 49, 9998.
- (74) Goldau, T.; Murayama, K.; Brieke, C.; Steinwand, S.; Mondal, P.; Biswas, M.; Burghardt, I.; Wachtveitl, J.; Asanuma, H.; Heckel, A. *Chem. Eur. J.* **2015**, *21*, 2845.
- (75) Matsunaga, D.; Asanuma, H.; Komiyama, M. J. Am. Chem. Soc. 2004, 126, 11452.
- (76) Freeman, N. S.; Moore, C. E.; Wilhelmsson, L. M.; Tor, Y. J. Org. Chem. 2016, 81, 4530.

Figure to Table of Content.