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1 Contrasting evolutionary history, anthropogenic declines and genetic contact in the

2 northern and southern white rhinoceros (Ceratotherium simum)

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Abstract

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The white rhinoceros (Ceratotherium simum) has a discontinuous African distribution, which is limited by the extent of sub-Saharan grasslands. The southern population (SWR) declined to its lowest number around the turn of the 19th century, but recovered to become the world's most numerous rhinoceros. In contrast, the northern population (NWR) was common during much of the 20th century, declining rapidly since the 1970s, and now only two post-reproductive individuals remain. Despite this species' conservation status, it lacks a genetic assessment of its demographic history. We therefore sampled 232 individuals from extant and museum sources and analysed ten microsatellite loci and the mtDNA control region. Both marker types reliably partitioned the species into SWR and NWR, with moderate nuclear genetic diversity and only three mtDNA haplotypes for the species, including historic samples. We detected ancient interglacial demographic declines in both populations. Both populations may also have been affected by recent declines associated with the colonial expansion for the SWR, and with the much earlier Bantu migrations for the NWR. Finally, we detected post-divergence secondary contact between NWR and SWR, possibly occurring as recently as the last glacial maximum. These results suggest the species was subjected to regular periods of fragmentation and low genetic diversity, which may have been replenished upon secondary contact during glacial periods. The species' current situation thus reflects prehistoric declines that were exacerbated by anthropogenic decline beginning with the rise of late Holocene technological advancement in Africa. Importantly, secondary contact suggests a potentially positive outcome for a hybrid rescue conservation strategy, although further genome wide data are desirable to corroborate these results.

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50 51 **Keywords**: white rhinoceros, anthropogenic declines, demographic history, secondary contact, conservation

Introduction

The white rhinoceros (*Ceratotherium simum*) is the most common of the world's five remaining rhinoceros species. It has borne the brunt of rhinoceros losses during the global acceleration in illegal hunting, which began in 2008 because of increasing demand for horn products in South-East and East Asia. The species is an obligate grazer, thriving historically in two geographically separated grassland areas in sub-Saharan Africa, and has consequently been divided by taxonomists. The southern white rhinoceros (SWR) is endemic to southern Africa, historically occurring in much of the sub-region, south of the Zambezi river, including Namibia, Botswana, Zimbabwe and South Africa (Figure 1A & B, after Rookmaaker & Antoine 2012). The northern white rhinoceros (NWR) was endemic to a narrow belt of grassland from west of the Nile River and Albertine Rift, comprising parts of Uganda, South Sudan, the Democratic Republic of the Congo (DRC), Chad and the Central African Republic (Figure 1A & B). The recent histories of both populations are well known, independent, and contrastingly reflect events occurring in Africa and the Middle East since the colonial period from the 18th century (Figure 1C).

In southern Africa, the northwards spread of colonialism from the Cape of Good Hope resulted in the extermination of the SWR across most of the sub-region (Player and Feely 1960). Even before the turn of the 19th century, the SWR had undergone a population decline so severe that only 100 – 200 individuals remained, restricted to around the confluence of the Black and White Umfolozi Rivers in Zululand (Vaughan-Kirby 1920). However, in 1895 colonial authorities declared the white rhinoceros royal game and proclaimed the area the Umfolozi Junction Reserve (Brooks 2006). With the dedicated conservation action of wildlife authorities in South Africa, this small population increased steadily throughout the 20th century (Figure 1C) to become a conservation success story. The current severe poaching epidemic is threatening to undo these gains, and it is predicted that if present trends continue, the SWR population will start to decline again in 2018 (Ferreira et al. 2015). Efforts to curb recent losses are ineffective with only marginal decreases in poaching rates in 2015 and 2016, with more than 1,000 African rhinoceros killed every year since 2013. Such a population contraction, in the absence of gene flow from other sources, could negatively affect the genetic diversity and evolutionary potential of the SWR through genetic drift.

 The demographic recovery of the SWR is all the more remarkable because the 20th century also brought the near eradication of all other rhinoceros populations across the world. The NWR was still common throughout most of its range at the turn of the 20th century (Roosevelt and Heller 1914, Pitman 1931) and numbers were still relatively high until the 1960s (Emslie and Brooks 1999), when demand for rhino horn, mainly on the Arabian peninsula, precipitated the penultimate poaching epidemic. Political instability and ineffective conservation measures during the ensuing period saw the rapid decline of NWR numbers in the wild (Figure 1C), with the last wild individuals extirpated in Uganda by 1980 (Edroma 1982), in Sudan by 1984 (Emslie and Brooks 1999) and finally the population in Garamba National Park, Democratic Republic

of the Congo (Hillman-Smith et al. 1986), declared extinct in 2008. The NWR now survives only in captivity, and with two post-reproductive individuals (two females) remaining, its chances of survival look bleak. The imminent extinction of the NWR has sparked several conservation efforts to prevent the loss of what little remains of the population's genetic diversity.

The plight of the NWR has also precipitated a debate on whether the evolutionary relationship between the two populations could allow for interbreeding and genetic rescue as a conservation strategy (Saragusty et al. 2016), enabling the retention of at least some of the NWR's genetic diversity. The only known NWR-SWR hybrid was a female (Nasi), born in captivity in 1977. Although she survived 30 years in captivity, she never bred, and this has raised questions about the level of reproductive isolation between the two white rhinoceros populations. Although studies have revealed both morphological, behavioural and genetic differences between the SWR and NWR (Groves et al. 2010, Cinková and Policht 2014, Harley et al. 2016), the evolutionary processes giving rise to this differentiation have not been discussed. Several authors have attempted to compare these fossils with extant SWR and/or NWR (Groves 1975, Geraads 2005, 2010), but with limited success due to the scarcity of well-preserved fossil material and difficulty in delimiting species/populations from fossil remains.

However the fossil record demonstrates clearly that the prehistoric distribution of the white rhinoceros was wider than its recognised historical range. The presence of anatomically modern white rhinoceros in Pleistocene Tanzania, Ethiopia, Libya, Eritrea, and Kenya (Geraads 2011) suggests a demographic history of population contraction and expansion. Fluctuation between cold and arid glacial periods with wet and warm interglacials would have respectively expanded and contracted the grassland biomes on which the white rhinoceros is dependent (Figure 1D). The evolutionary consequences of such climatic fluctuations, especially with regard to demographic isolation, depends on whether climatically driven range expansions allowed NWR and SWR populations to come into demographic secondary contact. The deep divergence between NWR and SWR implied by analysis of mtDNA (0.46 – 0.97 Mya, Harley et al. 2016) is indicative of a prolonged period of demographic isolation between NWR and SWR maternal lineages. However, due to the maternal inheritance of mtDNA, it has a lower effective population size (Ne) than nuclear DNA and its lineages assort more quickly into monophyletic clades. Nuclear markers, especially those that evolve rapidly (such as microsatellites) would be expected to perform reliably in an analysis of demography and isolation by quantifying prehistoric levels of differentiation and gene-flow between populations (Kuhner et al. 1998).

Here we analysed genetic variation in the white rhinoceros with the aim of more appropriately informing conservation management. We use both nuclear microsatellites and mtDNA to determine levels of genetic variation across a sample of both NWR and SWR populations, and from both wild and captive populations. To estimate the losses in genetic diversity resulting from 20th century population declines, we also measured

the genetic diversity of historic (pre-bottleneck) museum material for comparison. Additionally, we also tested the hypothesis that both populations underwent prehistoric demographic size changes, and determined whether the NWR and SWR came into secondary genetic contact after their initial divergence.

Methods

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- 132 Samples and loci
- Samples were collected from wild (ESM1, Table S1) and captive (ESM1, Table S2) animals for both SWR
- and NWR. A total of 217 SWR samples (174 wild, 42 captive) and 15 NWR samples (8 wild, 7 captive) were
- obtained from extant and historic (museum) material representing the entire species range (full details are
- provided in ESM1, and permit information in ESM1, Table S3). The 5' end of the control region using primers
- 137 mt15996L (5'-TCCACCATCAGCACCCAAAGC-3') and mt16502H (5'-
- 138 TTTGATGGCCCTGAAGTAAGAACCA-3') were used to amplify a 477 bp fragment of the control region.
- Samples were also amplified for 10 microsatellite loci (ESM1, Table S4). The number of markers used in
- this study is comparable both with the number and identity of markers used in other publication on
- rhinoceros (Harley et al. 2005; Kotze et al. 2014; Moodley et al. 2017). Markers were selected at random
- and were developed from a variety of target species (black rhinoceros, SWR and pig). For detailed
- molecular and quality control methods, see ESM2.
- 145 Genetic diversity
- We included four previously published mtDNA control region sequences (Genbank accessions AF187836,
- AF187837, AF187838 and AF187839; Brown and Houlden 2000), as well as seven mitochondrial genomes
- from wild individuals, three of which were from wild SWR and four from wild NWR prior to that population's
- extirpation (Harley et al. 2016). Diversity was estimated for all populations separately. Since captive animals
- were from a variety of zoos and animal parks, we pooled all captive individuals into SWR and NWR groups.
- For microsatellites, we calculated the mean number of alleles, observed (H₀) and unbiased expected
- heterozygosity (H_E) using GENETIX (Belkhir et al. 2004). Allelic richness (AR) was computed by resampling
- to correct for sample size differences among populations. Both AR and and inbreeding coefficients (F_{IS})
- were calculated in FSTAT (Goudet 1995). Mitochondrial DNA diversity for both control region and whole
- genomes was assessed for levels of polymorphism and haplotype diversity, as well as nucleotide diversity
- 156 (π), in Arlequin v3.5 (Excoffier and Lischer 2010). Tajima's D (Tajima 1989) and Fu's Fs (Fu 1997) statistics
- (13) (13) In a local to the local and a local to the local and the local
- were also calculated in Arlequin to determine whether sequences showed evidence for population size
- 158 changes.

- Genetic structure
- Population structure using microsatellite variation was assessed using Bayesian k-means clustering in
- STRUCTURE (Pritchard et al. 2000). We assumed an admixture model and analysis was run ten times for
- k = 1-7 with each randomly started run consisting of 500,000 Markov Chain Monte Carlo (MCMC) iterations,
- assuming correlated allele frequencies, discarding the first 100,000 iterations. The optimal k for the
- microsatellite data was determined as the highest value that was biologically interpretable. MtDNA structure
- was deduced by constructing a phylogenetic network of control region sequences. We used the median-

joining method in Network v5.0.0.1 (Bandelt et al. 1999) with equal weighting on all nodes and using a correction cost algorithm.

Evolutionary timeframe

In order to obtain a time frame for the evolutionary history of the species, we reconstructed a species level maternal phylogeny from the seven mitochondrial genomes sequenced by Harley et al. (2016). We conducted Bayesian phylogenetic dating using BEAST v2.4.3 (Drummond and Rambaut 2007) as this allowed us to parameterise splits in the tree with soft-bounded priors based on known fossil information, using the same mammalian mtDNA genomes and priors described by Harley et al. (2016), except that all five calibration times were used simultaneously in a single analysis. We used the Tamura-Nei model for nucleotide substitution with gamma correction, as deduced by jModelTest v2 (Darriba et al. 2012), placing a relaxed, lognormal prior on the clock rate to account for potential differences in the molecular clock. The analysis was facilitated by a heuristic 100 million-step exploration of the likelihood surface using a Markov chain Monte Carlo simulation, sampling the chain every 100,000 steps and discarding the first 10%.

Ancient and recent changes in effective population size

Ancient demographic change in both SWR (n = 20, excluding zoo individuals) and NWR (n = 15) populations was inferred using MSVar v.1.3 (Beaumont 1999; Storz & Beaumont 2002), with Ne being the size of a model population that has the same rate of genetic drift as the rhinoceros population of interest. Wide priors were set for all parameter estimates to allow for uncertainties in the data. Three potential scenarios were performed separately for SWR and NWR, assuming different ancestral (N1) and current (N0) effective population sizes. These were (i) a stable population (N1=N0), (ii) a population decline (N1>N0) and (iii) a population expansion (N1<N0). Further details of priors and MCMC runs are given in ESM1, Table S5.

The more recent demographic history of the white rhinoceros, during which humans may have driven population size changes, was investigated through Approximate Bayesian Computation (ABC) simulations (Beaumont et al. 2002). This approach is unlike the likelihood calculations of the data performed by MSVar, but instead simulates a finite set of potential demographic scenarios, which are then compared to the observed data using sets of summary statistics. Demographic histories for SWR and NWR were thus tested independently by exploratory simulations of six scenarios in ABCTOOLBOX v1.1 (Wegmann et al. 2010): a null model, two expansion models, two bottleneck models and a model with two bottlenecks (ESM, Figure S2). Under expansion and bottleneck scenarios we tested whether the timing of the demographic event coincided with sub-Saharan Africa's two most important anthropogenic events - the migration of iron-age, agriculturalist Niger-Congo language speakers (Bantu) into eastern and southern Africa 400-2,000 years ago (ya; Grollemund et al. 2015) and the expansion of colonial-era European influence into the region (present to 400 ya).

Secondary contact between northern and southern white rhinoceros

We also tested for the possibility that NWR and SWR could have come into secondary genetic contact since they diverged from each other. This may have occurred during the late Pleistocene during which the grassland biome would have periodically been continuous between eastern and southern Africa. We therefore built a two-population model that included uni-and bi-directional migration. First, we tested for migration (uni- and bidirectional) at any time during the last glacial period (LGP) of the late Pleistocene (14,000 – 106,000 ya, scenarios 2-4) which followed the end of the Eemian interglacial. We then subdivided the LGP to attempt to differentiate between recent migration during the last glacial maximum (LGM, 14,000 – 26,000 ya, scenarios 5-7) and earlier migration during the LGP (26,000 – 106,000 ya, scenarios 8-10). Last, we tested the null hypothesis against a model of ancient (pre-Eemian) migration (130,000 – 500,000 ya, scenarios 11-13). For details of model parameterisation see ESM2.

We were concerned that individuals in our data set could be closely related and we therefore removed all individuals with a relatedness values (r) of 0.3 or higher and reran all single- and two-population ABC simulations.

Results

A total of 232 white rhinoceros were genotyped at 10 microsatellite loci (ESM3) and 419 bp of the mitochondrial control region (ESM4) was sequenced in 63 individuals. The level of missing data is given as the number and percentage (%) of failed genotypes for the historical and modern data (ESM1, Table S6). Three loci (RHI32A, RH17B and RH17C) for the historic NWR sample showed a high proportion of missing data (> 50%). To determine if missing data at these loci affected the overall observed structure between populations, we reran the Structure analyses for k = 1-7 without these loci, and found that there was no change in the overall result.

Genetic diversity

The effect of ascertainment bias was limited in this study as our results were consistent with previous studies using different types of markers in rhinoceros (Merenlender et al. 1989). All populations were found to be in Hardy-Weinberg equilibrium and individual loci were randomly associated (in linkage equilibrium). Nuclear microsatellite genetic diversity was moderate to low, with an average of eight alleles per locus and heterozygosity ranging from 0.48-0.56 (ESM, Table S7). SWR ($H_0 = 0.48$) were more diverse than NWR ($H_0 = 0.46$), but both populations had lower observed than expected heterozygosity and positive, but not significant, inbreeding coefficients (F_{IS} SWR = 0.09, F_{IS} NWR = 0.33). The extant wild SWR population (six subpopulations) had slightly lower diversity ($H_0 = 0.47$) than captive SWR individuals (($H_0 = 0.51$). Within the captive SWR there was no difference in genetic diversity between founders and their offspring (zooborn). In contrast, the wild (museum sampled) NWR population was more diverse than our sample of captive NWR animals, returning a higher mean number of alleles (3.4 vs 2.4) and heterozygosity (0.48 vs 0.43). Yet, despite comprising only seven individuals, the captive NWR sample had similar allelic diversity to all sampled wild SWR subpopulations from reserves in South Africa today.

Mitochondrial genetic diversity was low for both SWR and NWR (ESM2, Table S8), comprising three haplotypes in total, with SWR comprising two haplotypes and NWR just one. Captive SWR contained both haplotypes but had lower nucleotide diversity than wild SWR (0.003 vs 0.005). In both populations, genetic diversity of 19 and 20th century haplotypes were the same as extant levels. We amplified the control region for one ancient SWR individual (shot in South Africa in 1869) which possessed one of the two haplotypes detected in extant SWR populations. Surprisingly, our entire historic NWR sample, containing early 20th century individuals from the three range states in which the NWR was most common, all harboured the same haplotype as NWR population in captivity.

Genetic structure

- Both nuclear and mitochondrial markers structured the species into two distinct populations/clades (ESM2,
- Figure S1), corresponding to SWR and NWR. For microsatellite data, k = 2 returned the highest likelihood,
- with no recent admixture detected between populations (ESM2, Figure S1A). When the data were analysed

at higher, less likely, *k* models, substructure within SWR was revealed with the separation of mainly captive individuals (K4), Mthethomusha Game Reserve (K5) and Origstad and Nkomazi Game Reserves (K6, ESM2, Fig. S4,). However, since we could not assume that our captive sample was taken from a naturally breeding population, only wild SWR were used for subsequent ABC analyses. However, the multilocus profile of the only known SWR-NWR hybrid individual Nasi clearly shows an admixed profile. The mitochondrial control region showed two monophyletic clades, separated by 30 mutational steps (ESM2, Figure S3B). Haplotypes within the SWR were separated by four mutations.

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- Evolutionary timeframe
- Using complete mitochondrial genomes (ESM5), which contained greater levels of within-population 267 polymorphism relative the control region, BEAST analysis inferred a divergence time between the two white 268 269 rhino lineages at approximately 0.97 million years (Myr), but with a large 95% highest posterior density 270 (HPD) of 0.5-1.5 Myr (ESM2, Figure S1C), with African rhinoceros species (black and white) identified as sister taxa with a most recent common ancestor (MRCA) of 11 Myr (HPD95: 6.4 - 16.0 Myr). Low 271 272 mitochondrial DNA diversity within each white rhinoceros population resulted in very shallow but similar MRCA times. SWR lineages coalesce to 38,000 years ago (va. HPD95: 7,400 - 77,400 va) and NWR 273 274 mitochondrial genomes shared a common ancestor 34,000 ya (HPD95: 8,800 – 67,000 ya).

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- 276 Ancient changes in effective population size
- 277 While analyses of mtDNA neutrality indicated a history of population contraction for the species, with the 278 majority yielding positive Fu's Fs values, these were not significant for the control region or for mitochondrial 279 genomes (ESM2 Table S8). However, microsatellite data analyses using MSVar revealed that both SWR 280 and NWR have undergone an ancient reduction in effective population size (EMS1, Table S9; ESM2, Figure S5), Independent runs invoking stable, expansion and contraction models all converged to the same 281 282 posterior values placing current population size (N_0) consistently lower than ancestral population size (N_1) EMS1, Table S9; ESM2, Figure S5A & C). The timing of these population contraction events could be dated 283 to the mid-Holocene, between 3,400 – 5,800 ya for SWR and the early Holocene-late Pleistocene (7,000 – 284 285 29,000 ya) for NWR (EMS2, EMS1, Table S9; ESM2, Figure S5B & D).

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- Recent demographic change
- The null ABC model (Scenario 1; ESM2, Figure S2) of no recent change in population size could be rejected for both SWR and NWR. Instead, highest model support for both populations was for a single decline or bottleneck (Table 1; BF > 3). For SWR, the best model selected was that of a population bottleneck during the colonial period (scenario 4 SWR; Table 1) with a modal time for the beginning of the decline of 264 years (HPD90 138 394 years). In contrast, the best model for NWR was a demographic bottleneck during the time of the Bantu expansion into East Africa (scenario 5 NWR; Table 1), occurring about 1,370 ya (HPD90 518 1,869 years).

Secondary contact and gene flow

Combining both SWR and NWR data, parameterised according to the two best single-population scenarios above, we found the marginal densities for all migration models to be higher than the null model of no post-divergence migration (Table 1). Among migration scenarios, bidirectional migration was more likely than any equivalent unidirectional scenario). The highest marginal densities and Bayes factors among bidirectional models were for scenarios set within the LGP (Table 1). Within the LGP, recent LGM secondary contact was the most likely of all tested scenarios, but could not be significantly differentiated from later LGP migration.

 All ABC simulations were also run without closely related individuals (r > 0.3). This reduced the sample size of SWR and NWR to 11 and 10 respectively. Nevertheless, all runs returned similar results to those above, with the exception that colonial and Bantu period population bottleneck scenarios could no longer be distinguished from each other for NWR. The results of these additional simulations are provided in ESM1, Tables S10 & S11.

Discussion

We generated molecular data from a sample that included the recent recorded white rhinoceros range, with samples from extant and historic specimens back to the 19^{th} century. We observed much higher allelic variation at microsatellites than for mtDNA, likely reflecting the differences in effective population size between the two markers (approximately 4:1). Levels of microsatellite heterozygosity in white rhinoceros are lower than East, West and southern African black rhinoceros populations (H_E : 0.71-0.74, Moodley et al. 2017) but higher than the relatively unmanaged south-western black rhinoceros of Namibia or Angola (H_E : 0.42-0.49). We also found that historic levels of NWR diversity were greater than extant levels, demonstrating the negative genetic consequences of the NWR's colonial-era history of hunting and habitat destruction during the latter part of the 20^{th} century. However, for mtDNA, we found that even in colonial times maternal genetic diversity was already as low as it is in extant populations. Therefore, the evolutionary process which reduced maternal variation in both white rhinoceros populations is very likely to have occurred prior the time of sampling in the late 1800's.

The lower diversity of SWR individuals born in captivity relative to wild-born founders may also indicate a loss of diversity, even in the space of one or two generations. Owing to the increased erosive power of genetic drift in small populations, this effect may increase as time progresses. We therefore suggest an active management plan for captive bred individuals, where multilocus genetic profiles can be used to maintain genetic diversity. Similarly, genetic drift in isolation has differentiated some SWR populations, and more active management between reserves is encouraged to help ameliorate these effects.

Structure and evolutionary timeframe

Microsatellite clustering clearly differentiated the white rhinoceros into two distinct populations (ESM2, Figure S1), an observation already made using mtDNA (Groves et al. 2010, Harley et al. 2016) and for the nuclear amelogenin gene (Groves et al. 2010). Both these studies, however, made use of more limited datasets comprising two and seven individuals, respectively. The present study therefore is the first to use large sample sizes and microsatellite markers, and in the case of the NWR the historic sample covered much of the population's range. For maternally inherited mtDNA, dated using five mammalian calibration points, we estimated the divergence of mtDNA lineages at just under a million years, but with wide confidence limits (± 500,000 years). ABC simulations were not able to narrow this estimate. These wide limits underscore model uncertainty and the wide prior distributions on fossil calibration points. Nevertheless, these divergence estimates provide a general time frame for the initial split between NWR and SWR populations. Genomic analysis will likely be needed to date the divergence of the two white rhinoceros populations more precisely.

A coalescent analysis of prehistoric effective population size changes using our microsatellite data allowed us to infer late Pleistocene to mid-Holocene population contractions for both SWR and NWR (ESM2, Figure S4). Since the effective population size inferred through coalescent simulation is a measure of the effect of genetic drift on the genealogical process, reported numbers reflect the minimum number of effective breeders required by the studied population to ameliorate the loss of further diversity through drift. The inferred timeframe largely overlaps with the confidence limits for the coalescence of intra-population mtDNA lineages (ESM2, Figure S1C), which could potentially have occurred as recently as 6,000 – 7,000 ya and as early as 77,000 ya. Although confidence limits on the posterior distribution of these times were large, for SWR, the inferred decline suggests that this population was adversely affected by the grassland contraction that occurred after the LGM. In the NWR, we obtained a signal both a post-LGM (7,000 ya) and a pre-LGM (26,000 – 29,000 ya) population decline, also possibly in response to grassland contraction, highlighting the white rhinoceros' dependence on suitable grassland habitats. These prehistoric population contractions may have been partly responsible for low mtDNA genetic variation detected among colonial-era NWR and SWR samples.

Recent human-associated population declines

Microsatellite analysis also allowed us to infer very recent population declines associated with human movements in Africa. We stress that although the timeframes for the recent NWR and SWR bottlenecks were defined to test for an association with known human historical events, our ABC simulations do not provide a causal link between the human activity and white rhinoceros demography. NWR precolonial population decline, may be coincident with the arrival of Bantu speakers from West Africa. Recent reconstructions have inferred that the Bantu expansion proceeded first in a south-easterly direction from Cameroon, avoiding rainforest and taking advantage of a savannah corridor that started to open ~4,000 ya (e.g. Vincens et al. 1998), accelerating ~2,500 ya (Bayon et al. 2012) and leading to colonisation of east Africa and the Great Lakes region around 2,000 ya (Grollemund et al. 2015). While the Bantu were predominantly agriculturalists, utilising grassland habitats on which to grow newly domesticated strains of millet and sorghum, they were also in possession of iron age smelting technology, and thus capable of hunting larger game animals, either directly, or through interactions with and spread of iron-age technology through local hunter-gatherers (e.g. Patin et al. 2014). It is also possible that Bantu speakers associated with people from further afield, either with Arab and South Asian traders via the eastern coast of Africa or with Romans via the Nile Valley. In either case, demand for rhinoceros products, and potentially even live animals, may have helped intensify the decline in effective population size in the NWR observed during this period.

In comparison, we recovered a clear signal for a more recent human-induced population decline in the SWR, during the occupation of southern Africa by Europeans. This population decline is historically well documented, with the SWR reaching its lowest number of approximately 100 animals over a hundred years

after the median time of decline (Figure 2), although the actual time at which the SWR was at its lowest number falls well within the confidence limits of our posterior distribution. Interestingly, although both populations were reduced to low numbers by humans, current effective population size confidence limits did not overlap, showing that the SWR was reduced to significantly lower effective numbers. Although the effective numbers of NWR destroyed by humans was greater, significantly lower effective size for SWR could reflect the more efficient destruction of white rhinoceros by mechanised hunting during the colonial times.

Post-divergence gene flow between SWR and NWR

We used two-population ABC analyses to demonstrate that although the NWR may have diverged from the SWR over a million years ago, both populations came into post-divergence secondary contact more recently during the LGP, and potentially even as recently as the LGM. The implications of this finding may prove central in ongoing debates about the specific status of the two white rhinoceros populations, and how best to manage their remaining genetic diversity in the future. The inferred post-divergence gene-flow was likely facilitated by savanna grassland expansions after the Eemian interglacial (115,000 – 130,000 va), but has ceased completely since the Holocene when NWR and SWR populations declined as their grassland habitat diminished. A potentially continuous distribution of the white rhinoceros is also supported by evidence of its occurrence east of the Nile river from the middle Pleistocene (Clark and Brown 2001), the LGP (Hillman-Smith et al. 1986) and as recently as the Holocene (Gifford et al. 1980). Therefore, the present-day absence of the white rhinoceros east of the Nile River can only be explained by the local extirpation of an East African population during the Holocene contraction, and with repopulation of Uganda, Kenya and Tanzania subsequently attenuated by the flow of the Nile. Taken together, these results suggest that the white rhinoceros has been resilient to population size contractions, which would have subjected local populations to periods of low genetic diversity during interglacial periods, but with diversity being potentially replenished during glacial periods by secondary contact.

Conservation implications

The contrasting histories of the northern and southern white rhinoceros have substantial implications for their conservation. Low diversity at both mtDNA and microsatellite loci implies that maintenance of genetic diversity should be a core conservation action for the species. Although the African Rhino Specialist Group advocates a lower limit of 20 founding individuals (Emslie et al. 2009) for new populations, some wild SWR populations like Mthethomusha, Origstad and Nkomazi have already differentiated from the original SWR stock due to management in isolation. Our results suggest not only a minimum number of founders for new populations, but also that microsatellite profiles should be used to select founding individuals from more than one source population. Additionally, low diversity of some populations should be ameliorated by regular and targeted translocations.

With most endangered species intensive genetic management of populations would be prohibitively expensive and/or logistically challenging. However, population genetic analysis carried out in forensic context is increasingly being applied in large African mammals, for example in both the forest and savannah elephant to identify the origins of seized animal products (Wasser et al. 2015) and to identify demographic units for conservation management (e.g. Ishida et al. 2018). Forensic studies require large genetic reference databases, thus a large and growing number of white rhinoceros have been routinely genotyped for forensic purposes (Harper et al. 2018), and we advocate making use of this unique genetic resource to aid the management of genetic diversity. With this database, it should be possible to monitor population diversity levels in real time, and select the profiles of immigrant individuals that would maximise population genetic diversity. Since landowners in South Africa are legally obliged to genotype their rhinoceros, it would also be possible, to monitor the breeding success of immigrant individuals as the calves of the next generation are added to the expanding database.

The situation for the NWR is very different, and here we show that this population is the end-point of a long period of both prehistoric and anthropogenic decline. With only two female individuals remaining, the role of genetics is presently confined to an evaluation of the potential outcomes of hybrid rescue involving the use of SWR genomes. The recent LGP secondary contact is a key result in this context, as it increases the likelihood that hybrid rescue could be positive and that the recently reported NWR/SWR hybrid embryos may provide a viable strategy for conservation of the NWR (Hildebrandt et al. 2018). However drawing such inference could be premature using a handful of genetic markers alone, and for this reason whole resequencing genomic data could be extremely useful in documenting locally adapted regions of the white rhinoceros genome that may be a priority for genetic management in white rhinoceros occupying the northern edge of the species' historic distribution, regardless of the origin of the animals.

An additional conservation implication of this work is that managed translocation of SWR into some portion of the NWR's historic range might be a viable approach to restore the ecological functionality that this large grazing mammal previously contributed to the northern savannah ecosystem it once occupied (Griffiths et al. 2011), although any such introductions would need to be closely monitored for evidence of a lack of local adaptation, genetic drift and inbreeding.

452	Ethics
453	Samples collected for this study received ethical approval from Cardiff University and were collected in
454	accordance with the protocols/guidelines of the National Zoological Gardens of South Africa (NZG).
455	Where relevant, animals were handled under the guidelines of the American Society of Mammalogists
456	(ASM; Animal Care and Use Committee, 2011). All required permits are listed in ESM1, Table S3. All
457	museum samples were collected in accordance with the relevant guidelines and regulations of each
458	museum.
459	
460	Electronic Supplementary material
461	Five files are provided:
462	ESM1, Supplementary Tables
463	ESM2, Methods and Figures
464	ESM3, White rhinoceros microsatellite data
465	ESM4, White rhinoceros control region data
466	ESM5, White rhinoceros complete mitochondrial genomes
467	
468	Data
469	All DNA sequences generated in this study were submitted to Genbank (accession numbers inserted on
470	final acceptance of paper). Microsatellite data were uploaded to the Dryad database (website given upor
471	final acceptance).
472	
473	Competing interests
474	The authors have no competing interests.
475	
476	Authors' contributions
477	YM, IMR, JR, AK, CW and MWB conceived the study. YM, JR, JS, OR, RH, CW performed the fieldwork
478	IMR, DLD and SS performed the laboratory work. YM, IMR, DLD, SS and MWB conducted the analyses.
479	YM, IMR and MWB wrote the initial draft which was subsequently critically revised and approved by co-
480	authors.
481	
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484	Melzer at Ol Pejeta, Kenya in 2017. Copyright C. Metzler. Pictures kindly provided by the photographer.
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Figure and table captions

Figure 1. Distribution, phenotype and population trends in the white rhinoceros. A. Historic distribution of northern (NWR) and southern (SWR) white rhinoceros in sub-Saharan Africa (after Rookmaaker & Antoine, 2012). B. Northern (left) and southern (right) white rhinoceros females at OI Pejeta, Kenya. Copyright C. Melzer 2017. C. Population trends of NWR (red) and SWR (blue) through the 20th century. D. Extent of the grassland biome (in red) in Africa during the last glacial maximum (left) when it was continuous from north to south, and at present (right) where it is fragmented.

Tables

Table 1. Posterior estimates for demographic scenarios within and between Northern and Southern white rhinoceros based on approximate Bayesian computation. The best model among single and two-population scenarios are indicated in bold.

	Scenario	Scenario Description	Migration prior (kyr)	Posterior Mode (kyr)	Posterior NeM	Marginal Density	P-value	Bayes Factor (BF)
Southern White	1	Null model (stable population size)	-	-	-	0.01	0.00	-
Rhinoceros	2	Expansion during the colonial period	-	-	-	7.49E-05	0.00	0.007
	3	Expansion during the Bantu migrations	-	-	-	2.59E-37	0.00	2.59E-35
	4	Bottleneck during the colonial period	-	-	-	1.68	0.99	168.00
	5	Bottleneck during the Bantu migrations	-	-	-	0.12	0.18	1.2
	6	Two bottlenecks colonial period/Bantu migrations	-	-	-	3.61E-50	0.00	3.61E-48
Northern White	1	Null model (stable population size)	-	-	-	0.003	0.00	-
Rhinoceros	2	Expansion during the colonial period	-	-	-	1.28E-07	0.00	4.27E-05
	3	Expansion during the Bantu migrations	-	-	-	6.45E-35	0.00	2.15E-32
	4	Bottleneck during the colonial period	-	-	-	2.68E-68	0.00	8.93E-66
	5	Bottleneck during the Bantu migrations	-	-	-	0.68	1.00	226.67
	6	Two bottlenecks colonial period/Bantu migrations	-	-	-	1.72E-18	0.00	5.73E-16
_	1	Null model; No migration	-	-	-	739.66	0.56	-
Two population	2	Unidirectional migration (S-N)	Last glacial	21.0	5	7,211.49	0.76	9.75
models	3	Unidirectional migration (N-S)	maximum	18.1	72	2,432.03	0.80	3.29
	4	Bidirectional migration	(14-26)	S-N: 18.1	S-N: 7	19,241.30	0.92	26.01
	5	Unidirectional migration (S-N)	Last glacial	N-S: 21.8	N-S: 112	4,937.74	0.70	6.68
	6	Unidirectional migration (N-S)	period	43.7	7	1,929.91	0.77	2.61
	7	Bidirectional migration	(LGP) (14-106)	40.0	87	10,643.00	0.86	14.39
	8	Unidirectional migration (S-N)	,	S-N: 43.7	S-N: 7	4,819.14	0.65	6.52
	9	Unidirectional migration (N-S)	Early LGP	N-S: 40.9	N-S: 95	2,099.64	0.83	2.84
	10	Bidirectional migration	(26-106)	72.9	7	9,950.02	0.79	13.45
	11	Unidirectional migration (S-N)	Pre-Eamian	49.4	85	2,377.90	0.65	3.21
	12	Unidirectional migration (N-S)	interglacial	S-N: 73.7	S-N: 9	1,691.94	0.69	2.29
	13	Bidirectional migration	(130-540)	N-S: 47.0	N-S: 98	5,124.45	0.91	6.93