Supplementary Information Available

Highly luminescent metallacages featuring bispyridyl ligands functionalised with BODIPY for imaging in cells

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Experimental section

General

Chemicals. All reagents and solvents were obtained from commercial suppliers and used without further purification, unless otherwise stated. Triethylamine was distilled under nitrogen before use. $^1$H NMR, $^{13}$C($^1$H) NMR, $^{11}$B NMR and $^{19}$F NMR spectra were recorded on a 500 MHz DMX (Bruker) or 400 MHz AV spectrometer (Bruker). Chemical shifts are given in parts per million (ppm). Abbreviations for NMR multiplicities are: singlet (s), doublet (d), triplet (t), quartet (q), doublet of doublets (dd), doublet of triplets (dt) and multiplet (m). Coupling constants $J$ are given in Hz. The following solvents were used as internal standards: DMSO-$d_6$: 2.50 ppm ($^1$H NMR) and 39.52 ppm ($^{13}$C NMR); CDCl$_3$: 7.26 ppm ($^1$H NMR) and 77.16 ppm ($^{13}$C NMR); acetone-$d_6$: 2.05 ppm ($^1$H NMR) and 29.84 ppm ($^{13}$C NMR); DMF-$d_7$: 8.03 ppm ($^1$H NMR) and 163.15 ppm ($^{13}$C NMR). High resolution ESI-MS spectra were recorded on a Walter Synapt G2SI QTOF.

Synthesis of ligands

BODIPY B1,[1] B2[2] and ligand L1[3] have been synthesised adapting previously reported procedures and the analytical data is in accordance with the literature.

- 3,3’-((5-azido-1,3-phenylene)bis(ethyne-2,1-diyl))dipyridine (L2)

\[
\begin{align*}
\text{L1} & \xrightarrow{1) \text{NaNO}_2 (6 \text{M HCl/H}_2\text{O}, 0 \degree \text{C}, 30 \text{ min}} \quad 2) \text{NaN}_3 (\text{H}_2\text{O}), \text{ r.t., 2 h} \\
\text{N}_3 & \quad \text{N}_3
\end{align*}
\]

Scheme S1. Synthesis of ligand L2.

L1 (136 mg, 459 $\mu$mol, 1.00 eq.) was dissolved in 6M HCl (4.5 mL) and cooled down to 0 °C (water/ice). NaNO$_2$ (39.7 mg, 575 $\mu$mol, 1.20 eq.) was dissolved in water (3 mL) and added dropwise to the reaction. After 30 min, the reaction was allowed to warm to r.t. and NaN$_3$ (59.3 mg, 912 $\mu$mol, 2.00 eq.) dissolved in water (3 mL) was added dropwise and stirred for 2 h. The pH-value was adjusted to 8 with 2M NaOH, the
formed precipitate filtered over a glass-fritted funnel (porosity 3) and washed with a small amount of water, EtOH and Et₂O to give L2 as a pale brown solid (96.3 mg, 300 μmol, 65%).

**^1H NMR** (400 MHz, CDCl₃): δ [ppm] = 8.78 (d, J = 1.3 Hz, 2H, Hₐ), 8.59 (dd, J = 1.7, 4.9 Hz, 2H, Hₖ), 7.82 (dt, J = 1.9, 7.9 Hz, 2H, Hₜ), 7.51 (t, J = 1.4 Hz, 1H, Hₑ), 7.31 (dd, J = 4.9, 7.9 Hz, 2H, Hₑ), 7.19 (d, J = 1.4 Hz, 2H, Hᵦ).

**^13C NMR** (101 MHz, DMSO-d₆): δ [ppm] 151.8, 149.5, 141.0, 138.8, 130.7, 123.9, 123.7, 122.5, 118.8, 90.3, 87.8.

**HRMS** (ESI, MeCN): calcd. for C₂₀H₁₂N₅ [M+H]⁺: m/z = 322.1094; found: 322.1112, δ = 5.9 ppm.

- **N-(3,5-bis(pyridin-3-ylethynyl)phenyl)-4-(2,8-diethyl-5,5-difluoro-1,3,7,9-tetramethyl-5H-4λ₄,5λ₄-dipyrrolo[1,2-c:2’,1’-f][1,3,2]diazaborinin-10-yl)benzamide (LB1)**

![Scheme S2. Synthesis of LB1.](image_url)

A mixture of B1 (152 mg, 357 μmol, 1.00 eq.), L1 (105 mg, 356 μmol, 1.00 eq.), CMPI (360 mg, 1.41 mmol, 4.00 eq.) and DMAP (433 mg, 3.54 mmol, 10.0 eq.) was dissolved in anhydrous DMF (18 mL) and stirred under a nitrogen atmosphere at 130 °C. After 18 h, the solvent was removed under reduced pressure, the residue dissolved in DCM (50 mL) and washed with water (3x 40 mL) and brine (1x 40 mL). The organic layer was dried over MgSO₄ and filtered over a glass-fritted funnel (porosity 3). The solvent was removed
under reduced pressure and the crude compound purified via silica column chromatography (EtOAc:n-hexane = 1:1 → 3:1, Rf = 0.21) to give product LB1 as a red/pink solid (108 mg, 154 μmol, 43%).

\(^1\)H NMR (400 MHz, acetone-\(d_6\)): \(\delta [\text{ppm}] = 9.97\) (s, 1H, NH), 8.80 (d, \(J = 1.3\) Hz, 2H, H\(_a\)), 8.61 (dd, \(J = 1.7, 4.9\) Hz, 2H, H\(_b\)), 8.30 (d, \(J = 8.3\) Hz, 2H, H\(_c\)), 8.21 (d, \(J = 1.5\) Hz, 2H, H\(_d\)), 7.99 (dt, \(J = 1.9, 7.9\) Hz, 2H, H\(_e\)), 7.61 (d, \(J = 8.2\) Hz, 2H, H\(_f\)), 7.57 (t, \(J = 1.4\) Hz, 1H, H\(_g\)), 7.47 (dd, \(J = 4.9, 7.9\) Hz, 2H, H\(_h\)), 2.51 (s, 6H, NCC\(\text{H}_3\)), 2.36 (q, \(J = 7.5\) Hz, 4H, C\(\text{H}_2\)CH\(_3\)), 1.36 (s, 6H, CC\(\text{H}_3\)), 0.99 (t, \(J = 7.5\) Hz, 6H, CH\(_2\)CH\(_3\)).

\(^{13}\)C NMR (101 MHz, acetone-\(d_6\)): \(\delta [\text{ppm}] = 165.4, 156.0, 154.8, 152.9, 150.1, 148.3, 141.0, 140.2, 139.3, 139.0, 133.8, 130.5, 129.7, 129.4, 124.4, 124.3, 124.2, 120.6, 96.1, 91.8, 87.5, 17.5, 14.9, 12.7, 12.1.

\(^{11}\)B NMR (128 MHz, acetone-\(d_6\)): \(\delta [\text{ppm}] = 0.76\).

\(^{19}\)F NMR (376 MHz, acetone-\(d_6\)): \(\delta [\text{ppm}] = -145.1\).

HRMS (ESI, MeCN): calcld. for C\(_{44}\)H\(_{38}\)BFN\(_5\)O [M-F] \^+ : m/z = 682.3125; found: 682.3139, \(\delta = -2.1\) ppm.

- 10-(4-(1-(3,5-bis(pyridin-3-ylethynyl)phenyl)-1H-1,2,3-triazol-4-yl)phenyl)-2,8-diethyl-5,5-difluoro-1,3,7,9-tetramethyl-5H-4\(\lambda^4\),5\(\lambda^4\)-dipyrrolo[1,2-c:2',1'-f][1,3,2]diazaborinine (LB2)

\[\text{L2} + \text{B2} \rightarrow \text{LB2}\]

**L2** (15.8 mg, 49.2 μmol, 1.00 eq.) was dissolved in DCM (0.5 mL) and diluted with MeOH:water (3:1, 6 mL).

**B2** (54.5 mg, 135 μmol, 3.00 eq.), DCM (1 mL) and a solution of L-(+)-ascorbic acid (11.6 mg, 65.9 μmol, 1.35 eq.), NaOH (3.10 mg, 77.5 μmol, 1.60 eq.) and CuSO₄·5 H₂O (1.10 mg, 4.41 μmol, 0.10 eq.) in water (2 mL) was added to the reaction mixture and stirred at room temperature. After 5 h, the reaction was quenched with a saturated NH₄OAc solution (20 mL) and stirred for 1 h. The reaction solution was washed with DCM (3x 20 mL) and the combined organic layers with brine (1x 30 mL). The organic layer was dried over Na₂SO₄, filtered over a glass-fritted funnel (porosity 3) and the solvent removed under reduced pressure. The crude compound was purified via silica column chromatography (DCM:MeOH = 100:5, Rᵣ = 0.34) to give product **LB2** as a purple solid (19.7 mg, 27.1 μmol, 55%).

**¹H NMR** (400 MHz, acetone-ᴅ₆): δ [ppm] = 9.34 (s, 1H, Ḣ_i), 8.84 (d, J = 1.2 Hz, 2H, Ḣ_a), 8.64 (dd, J = 1.6, 4.9 Hz, 2H, Ḣ_c), 8.35 – 8.20 (m, 4H, Ḣ_f, Ḣ_g), 8.04 (dt, J = 1.9, 8.0 Hz, 2H, Ḣ_d), 7.92 (t, J = 1.4 Hz, 1H, Ḣ_e), 7.55 (d, J = 8.3 Hz, 2H, Ḣ_h), 7.50 (dd, J = 4.9, 7.9 Hz, 2H, Ḣ_k), 2.52 (s, 6H, NCC₃H₃), 2.37 (q, J = 7.5 Hz, 4H, CH₂CH₃), 1.44 (s, 6H, CCH₃), 1.00 (t, J = 7.5 Hz, 6H, CH₂CH₃).

**¹³C NMR** (126 MHz, acetone-ᴅ₆): δ [ppm] = 154.5, 153.0, 150.5, 148.4, 139.4, 139.1, 138.7, 136.5, 134.9, 132.2, 130.0, 127.2, 126.5, 125.7, 125.7, 124.4, 123.8, 123.6, 120.5, 120.2, 90.7, 89.1, 17.5, 15.0, 12.6, 12.1.

**¹¹B NMR** (160 MHz, acetone-ᴅ₆): δ [ppm] = 0.83.


**HRMS** (ESI, MeCN): calcd. for C₄₅H₃₈BFN₇ [M+]: m/z = 706.3281; found: 706.3214, δ = -7.7 ppm.
Synthesis of metallacages

A solution of Pd precursor (2 eq.) and ligand (4 eq.) in DMSO was stirred at r.t. for 1 h (Scheme S4). Afterwards, precipitation by addition of acetone and diethyl ether and consecutive filtration gave the respective cages C1.BF\textsubscript{4}, C1.NO\textsubscript{3} and C2.NO\textsubscript{3}.

Scheme S4. General synthesis scheme for C1.BF\textsubscript{4}, C1.NO\textsubscript{3} and C2.NO\textsubscript{3}.

- **C1.BF\textsubscript{4}**

A solution of Pd(MeCN\textsubscript{4})(BF\textsubscript{4})\textsubscript{2} (6.00 mg, 14.3 µmol, 2.00 eq.) and LB1 (20.0 mg, 28.5 µmol, 4.00 eq.) in DMSO (1.5 mL) was stirred for 1 h at r.t.. Following precipitation by addition of acetone (3 ml) and diethyl ether (excess), the solid was filtered over a glass-fritted funnel (porosity 4) to give cage C1.BF\textsubscript{4} as a red solid (19.0 mg, 5.56 µmol, 78%).
\(^1\)H NMR (400 MHz, acetone-\(d_6\)): \(\delta\) [ppm] = 9.95 (s, 1H, NH), 9.91 (s, 2H, \(H_b\)), 9.55 (d, \(J = 4.9\) Hz, 2H, \(H_b\)), 8.39 – 8.13 (m, 6H, \(H_a, H_g, H_i\)), 7.96 (s, 1H, \(H_b\)), 7.88 – 7.73 (m, 2H, \(H_b\)), 7.57 (s, 2H, \(H_i\)), 2.52 (s, 6H, NCC\(H_3\)), 2.48 (s, 4H, CH\(_2\)CH\(_3\)), 1.30 (s, 6H, CC\(H_3\)), 0.96 (t, \(J = 6.94\) Hz, 6H, CH\(_2\)C\(H_3\)).

\(^{13}\)C NMR (101 MHz, acetone-\(d_6\)): \(\delta\) [ppm] = 165.1, 161.8, 153.9, 153.5, 150.6, 143.0, 139.5, 138.0, 134.9, 132.9, 130.2, 129.3, 128.9, 128.5, 127.4, 124.5, 123.4, 122.7, 116.3, 93.9, 84.5, 16.5, 14.0, 11.7, 11.2.

\(^{11}\)B NMR (128 MHz, acetone-\(d_6\)): \(\delta\) [ppm] = 0.66 (\(BF_2\)), -0.38 (\(BF_4\)).

\(^{19}\)F NMR (376 MHz, acetone-\(d_6\)): \(\delta\) [ppm] = -145.1 (d, \(J = 33.2\) Hz, \(BF_2\)), -145.3 (d, \(J = 33.2\) Hz, \(BF_2\)), -149.9 (s, \(BF_4\)).

HRMS (ESI, MeCN): calcd. for C\(_{161}\)H\(_{130}\)B\(_3\)F\(_6\)N\(_18\)O\(_4\)Pd\(_2\)[M-4BF\(_4\)-(C\(_{17}\)H\(_{22}\)BF\(_2\)N\(_2\))+2H\(^2+\)]: \(m/z = 1367.9508\); found: 1367.6577.

- **C1.NO\(_3\)**

A solution of Pd(NO\(_3\))\(_2\) \(\cdot\) 2 H\(_2\)O (4.60 mg, 17.3 \(\mu\)mol, 2.00 eq.) and LB1 (23.1 mg, 32.9 \(\mu\)mol, 4.00 eq.) in DMSO (1.5 mL) was stirred for 2 h at r.t. Following precipitation by addition of acetone (2 mL) and diethyl ether (excess), the solid was filtered over a glass-fritted funnel (porosity 4) to give cage C1.NO\(_3\) as a red solid (24.6 mg, 7.53 \(\mu\)mol, 91%).

\(^1\)H NMR (500 MHz, DMSO-\(d_6\)): \(\delta\) [ppm] = 10.69 (s, 1H, NH), 9.69 (s, 2H, \(H_b\)), 9.42 (s, 2H, \(H_b\)), 8.30 (d, \(J = 7.9\) Hz, 2H, \(H_b\)), 8.24 (s, 2H, \(H_b\)), 8.16 (d, \(J = 7.4\) Hz, 2H, \(H_b\)), 7.84 (s, 2H, \(H_b\)), 7.72 (s, 1H, \(H_b\)), 7.59 (d, \(J = 7.8\) Hz, 2H, \(H_b\)), 2.44 (s, 6H, NCC\(H_3\)), 2.28 (s, 4H, CH\(_2\)CH\(_3\)), 1.25 (s, 6H, CC\(H_3\)), 0.93 (s, 6H, CH\(_2\)CH\(_3\)).

\(^{13}\)C NMR (126 MHz, DMSO-\(d_6\)): \(\delta\) [ppm] = 183.5, 165.1, 157.9, 153.6, 153.0, 150.6, 143.0, 140.2, 139.3, 138.5, 137.9, 132.8, 129.6, 128.6, 127.4, 122.2, 122.1, 93.7, 86.8, 85.9, 85.1, 16.4, 14.5, 12.3, 11.5.

\(^{11}\)B NMR (128 MHz, DMSO-\(d_6\)): \(\delta\) [ppm] = 0.65.

\(^{19}\)F NMR (376 MHz, DMSO-\(d_6\)): \(\delta\) [ppm] = -142.9.

HRMS (ESI, MeOH): calcd. for C\(_{178}\)H\(_{157}\)B\(_4\)F\(_7\)N\(_{20}\)O\(_6\)Pd\(_2\)Na\(_2\)[M-4NO\(_3\)-3H-\(F\)+2MeOH+2Na\(^2+\)]: \(m/z = 1552.0486\); found: 1552.0068.

- **C2.NO\(_3\)**
A solution of Pd(NO$_3$)$_2$·2H$_2$O (4.10 mg, 15.4 µmol, 2.00 eq.) and LB2 (22.3 mg, 30.7 µmol, 4.00 eq.) in DMSO (1.5 mL) was stirred for 1 h at r.t.. Following precipitation by addition of acetone (3 mL) and diethyl ether (excess), the solid was filtered over a glass-fritted funnel (porosity 4) to give cage C2.NO$_3$ as a dark purple solid (24.0 mg, 7.14 µmol, 93%).

$^1$H NMR (500 MHz, DMSO-$d_6$): $\delta$ [ppm] = 9.83 (s, 2H, $H_a$), 9.59 (s, 1H, CH), 9.45 (s, 2H, $H_b$), 8.38 (s, 2H, $H_c$), 8.31 (d, $J = 8.3$ Hz, 2H, $H_d$), 8.30 (d, $J = 7.9$ Hz, 2H, $H_e$), 8.05 (s, 1H, $H_f$), 7.89 (s, 2H, $H_i$), 7.51 (d, $J = 8.1$ Hz, 2H, $H_j$), 2.44 (s, 6H, NCC$_3$H$_3$), 2.22 – 2.32 (m, 4H, CH$_2$CH$_3$), 1.31 (s, 6H, CCH$_3$), 0.93 (t, $J = 7.4$ Hz, 6H, CH$_2$CH$_3$).

$^{11}$B NMR (128 MHz, DMSO-$d_6$): $\delta$ [ppm] = 0.77.

$^{19}$F NMR (376 MHz, DMSO-$d_6$): $\delta$ [ppm] = -142.9.

HRMS (ESI, MeOH): calcd. For C$_{140}$H$_{103}$B$_2$F$_4$N$_{24}$Pd$_2$ [M-4NO$_3$-(C$_{17}$H$_{22}$BF$_2$N$_2$)-(C$_{23}$H$_{26}$BF$_2$N$_2$)$_2$]$^{2+}$: m/z = 1214.8926; found: 1214.3605.
Quantum Yield Determination
Quantum yield of fluorescence was calculated by comparison to a reference standard (Rhodamine 6G in degassed ethanol, $\phi = 94\%$ at room temperature). UV-Visible absorption spectra were recorded on a Cary 60 UV-Vis spectrometer from Agilent Technologies. Emission spectra were recorded on a Cary Eclipse Fluorescence Spectrophotometer from Agilent Technologies. The selected fluorophore was dissolved in degassed DMSO to a concentration corresponding to UV-Visible absorbance 0.8 A.U. ($\lambda_{(\text{max})} = 523 - 535$ nm; 25 °C). The solution was transferred to a fluorescence spectrophotometer and an emission spectrum was recorded (excitation wavelength 595 nm).

Stability studies by UV-Visible Spectroscopy
UV-visible absorption spectra to investigate the stability of the metallacages in solution were recorded on a Cary 60 UV-Vis spectrometer from Agilent Technologies. For each compound, stock solutions at a concentration of $3 \cdot 10^{-3}$ M were prepared. An aliquot was diluted either with 1x PBS (pH 7.4) or deionised water and the UV-Vis spectra measured at different times immediately after dilution at room temperature over 24 h. The cuvette was then shaken, and another spectrum recorded, to determine if the compound was altered during the 24 h or if the reduction in absorption was only due to precipitation.

Metallacage stability in the presence of L-glutathione
C1.BF$_4$ was dissolved in a 9:1 ratio of DMSO-$d_6$:D$_2$O (0.5 mL) and a $^1$H NMR spectrum was recorded. L-glutathione was added to the solution to achieve a final concentration of 2 mM L-glutathione (0.3 mg) and the first $^1$H NMR spectrum immediately recorded. Afterwards, spectra were recorded every 5 min for the first hour, followed by every hour for the following 17 hours. Finally, another 0.3 mg of L-glutathione were added after 18 hours to ensure that the reaction was complete. The ratio of the metallacage: ligand in solution was calculated by comparing the integral value of peak H$_b$ of the metallacage to the peak of H$_b$ of the ligand.

Cisplatin encapsulation studies
$^1$H NMR spectroscopy
Each metallacage (4.4 µM, 1.00 eq.) was dissolved in 1 mL DMF-$d_7$ and a $^1$H NMR spectrum was recorded. Afterwards, Cisplatin (8.8 µM, 2.00 eq.) was added to the NMR tube and the deuterated solution was stirred for 10 min before $^1$H NMR spectrum was recorded. Finally, NMR spectra were compared to evaluate any chemical shifts due to the encapsulation of cisplatin. Both spectra were calibrated to the residual solvent signal of the carbonyl proton of DMF (8.03 ppm).

$^{195}$Pt NMR spectroscopy
Cisplatin (2 mg, 6.7 µM, 1 eq.) was dissolved in DMF (0.5 mL) and transferred to an NMR tube with a capillary insert filled with DMF-$d_7$. Thus, the $^{195}$Pt NMR spectra was recorded. To this sample, the selected
metallacage (C1.BF₄) (6.7 µM, 1 eq.) was added and the solution stirred for 10 min before another ¹⁹⁵Pt NMR spectrum of 1 equiv. of metallacage and 1 equiv. of cisplatin was recorded. Afterward, a second equivalent of cisplatin (2 mg, 6.7 µM, 1 eq.) was added to the solution to achieve a cisplatin:cage ratio of 2:1, and a final ¹⁹⁵Pt NMR spectrum was recorded. The resulting spectra were compared to observe any chemical shifts between the free cisplatin and metallacage complex.

**Cell culture maintenance**

Human malignant melanoma cell line A375 was obtained from ATCC and maintained in culture according provider instructions. Cells were cultured in a humidified atmosphere at 37 °C and 5% CO₂ using DMEM Dulbecco’s Modified Eagle Medium (DMEM, 4.5 g/L glucose, Corning, Thermo Fischer Scientific) supplemented with 10% fetal bovine serum (FBS, Eu-approved South American Origin, Thermo Fisher Scientific) and 1% penicillin/streptomycin (Gibco) and passaged when reaching confluence.

**Antiproliferative assay**

To evaluate the inhibition of cell growth, 96-well tissue culture-treated plates (Corning) were seeded in a concentration of 15000 cells/well with 200 µL full medium. Working solutions of ligand and cage samples were prepared in the required concentration by diluting fresh stock solutions (5 x 10⁻³ M in DMSO) of the corresponding compound in aqueous complete DMEM medium accordingly. Dilutions (1 mM stock) of reference compound cisplatin (Sigma-Aldrich) were freshly prepared in aqueous solution and mixed with the metallacages prior each experiment. Cage formation and cisplatin encapsulation were confirmed by ¹H NMR spectroscopy as previously reported.[4] Following the initial 24 h incubation required for cell adhesion, cells were incubated for an additional 24 h with 200 µL of the compounds’ dilution per well. Afterwards, 20 µL/well of CellTiter-Blue® reagent was added to the assay plate, shaken 10 sec and incubated for 4 hours at 37 °C and 5% CO₂. Fluorescent intensity (531ₐₑ/595ₑₘ nm) from each well was quantified in quadruplicates for each experiment using a multi-well plate reader (VICTOR X5, Perking Elmer). The percentage of surviving cells was calculated, using GraphPad Prism software, from the ratio of fluorescence intensity of treated to untreated cells, corrected for the interfering fluorescence of the BODIPY. The EC₅₀ value for each compound was calculated as the concentration showing 50% decrease in cell growth, when compared to controls, using a nonlinear fitting of [concentration] vs response. Data is presented as mean ± SEM of at least three independent experiments.

**Fluorescence microscopy assays**

Round glass coverslips (Ø 13mm, VWR) sterilised by UV-light were inserted in 24-well tissue culture-treated plates (Corning). Cells were seeded at a concentration of 50.000 cells/well and incubated at 37 °C under humidified atmosphere with 5% CO₂ for 48 h. The medium was discarded and fresh medium containing 5 µM of either cage or ligand was added. Following 2 h of incubation at either 37 °C under tissue culture
conditions or 4 °C in the fridge, the glass coverslips were removed from the wells, washed 4x with 1x Phosphate Buffered Saline (PBS, Corning) and fixed with 4% formaldehyde (Alfa-Aesar) for 20 min at room temperature (r.t.). The coverslips were washed 3x with 1x PBS and incubated for 1 min with 40 µL 1:1000 from a 1 mg/mL stock solution of 4',6-Diamidino-2-phenylindole dihydrochloride (DAPI, Sigma-Aldrich/ MERCK) at r.t.. After washing the coverslips thrice with 1x PBS they were mounted on glass microscope slides (VWR) using Mowiol® 4-88 (Sigma-Aldrich). Fluorescence images were obtained using either a Zeiss Axio Vert.A1 epifluorescent microscope or a Leica SP5 confocal laser-scanning microscope. For confocal imaging, two laser lines were used: 405 Blue Diode (excitation wavelength 405 nm, laser intensity 30%) for DAPI and Argon (excitation wavelength 514 nm, laser intensity 30%) for the complexes, captured sequentially to avoid bleedthrough. A HCX PL APO 63x 1.4 NA oil immersion objective was used for all images with Leica Type F immersion oil. Captured “XYZ” images from the Leica SP5 and “XY” images from the Zeiss were analysed using ImageJ. All images were captured under the same settings within each experiment and treated equally following acquisition.
Figures

NMR spectra

Figure S1. $^1$H NMR (400 MHz, CDCl$_3$) spectrum of L2.

Figure S2. $^{13}$C NMR (101 MHz, DMSO-$d_6$) spectrum of L2.
Figure S3. $^1$H NMR (400 MHz, acetone-$d_6$) spectrum of LB1.

Figure S4. $^{13}$C NMR (101 MHz, acetone-$d_6$) spectrum of LB1.
Figure S5. $^{11}$B NMR (128 MHz, acetone-$d_6$) spectrum of LB1.

Figure S6. $^{19}$F NMR (376 MHz, acetone-$d_6$) spectrum of LB1.
Figure S7. $^1$H NMR (400 MHz, acetone-$d_6$) spectrum of LB2.

Figure S8. $^{13}$C NMR (126 MHz, acetone-$d_6$) spectrum of LB2.
Figure S9. $^{11}$B NMR (160 MHz, acetone-$d_6$) spectrum of LB2.

Figure S10. $^{19}$F NMR (471 MHz, acetone-$d_6$) spectrum of LB2.
Figure S11. $^1$H NMR (400 MHz, acetone-$d_6$) spectrum of C1.BF$_4$.

Figure S12. $^{13}$C NMR (126 MHz, acetone-$d_6$) spectrum of C1.BF$_4$. 
Figure S13. $^{11}$B NMR (160 MHz, acetone-$d_6$) spectrum of C1.BF$_4$.

Figure S14. $^{19}$F NMR (471 MHz, acetone-$d_6$) spectrum of C1.BF$_4$. 
Figure S15. $^1$H NMR (500 MHz, DMSO-$d_6$) spectrum of C1.NO$_3$.

Figure S16. $^{13}$C NMR (126 MHz, DMSO-$d_6$) spectrum of C1.NO$_3$. 
Figure S17. $^{11}$B NMR (160 MHz, DMSO-$d_6$) spectrum of C1.NO$_3$.

Figure S18. $^{19}$F NMR (471 MHz, DMSO-$d_6$) spectrum of C1.NO$_3$. 
**Figure S19.** $^1$H NMR (500 MHz, DMSO-$d_6$) spectrum of C2.NO$_3$.

**Figure S20.** $^{11}$B NMR (128 MHz, DMSO-$d_6$) spectrum of C2.NO$_3$. 
Figure S21: $^{19}$F NMR (376 MHz, DMSO-$d_6$) spectrum of C2.NO$_3$. 
Stability studies by UV-Visible Spectroscopy

**Figure S22.** UV-Visible spectra of C1.BF₄ [25 x 10⁻⁶ M] in 1x PBS (left); and in water (right) recorded over 24 h and shaken cuvette afterwards.

**Figure S23.** UV-Visible spectra of C1.NO₃ [55 x 10⁻⁶ M] in 1x PBS (left); and in water (right) recorded over 24 h and shaken cuvette afterwards.

**Figure S24.** UV-Visible spectra of C2.NO₃ [86 x 10⁻⁶ M] in 1x PBS (left); and in water (right) recorded over 24 h and shaken cuvette afterwards.
Cisplatin encapsulation studies

$^1$H NMR Spectra

**Figure S25.** Stack $^1$H NMR spectra in DMF-$d_7$ of the aromatic region (7-11 ppm) of the cage C1.BF$_4$ before (top) and after (bottom) addition of 2 eq. of cisplatin. Both the cavity facing proton peaks $H_a$ and $H_e$ undergo downfield shifts, as well as $H_b$. Broadening of $H_e$ peak upon encapsulation of cisplatin may also be indicative of intermolecular forces between the host and guest complex.
Figure S26. Stack $^1$H NMR spectra in DMF-$d_7$ of the aromatic region (7-11 ppm) of the cage C1.NO$_3$, before (top) and after (bottom) addition of 2 eq. of cisplatin. The cavity facing proton peaks $H_a$ and $H_b$ both undergo upfield shifts, consistent with replacement of the encapsulated negatively charged NO$_3^-$ counterion by cisplatin. $H_b$ signal undergoes a slight downfield shift upon cisplatin addition.
Cell Uptake Studies

A) C1.BF$_4$

![CLSM images of a representative fixed A375 melanoma cell incubated for 2 h with A) 5 µM C1.BF$_4$ or B) 5 µM LB1. Counterstaining with DAPI. Different z-slices from top to bottom are shown as well as the sum of stack/maximum projection of stack. Scale bar represents 10 µm.]

B) LB1

Figure S27. CLSM images of a representative fixed A375 melanoma cell incubated for 2 h with A) 5 µM C1.BF$_4$ or B) 5 µM LB1. Counterstaining with DAPI. Different z-slices from top to bottom are shown as well as the sum of stack/maximum projection of stack. Scale bar represents 10 µm.
Figure S28. Fluorescence widefield microscopy (Zeiss) images of fixed cells comparing control to samples incubated with 5 µM of C1.BF₄, LB1 or B1 for 2 h at 37 °C (top three rows) or 4 °C (bottom three rows). Scale bar represents 50 µm.
## References


