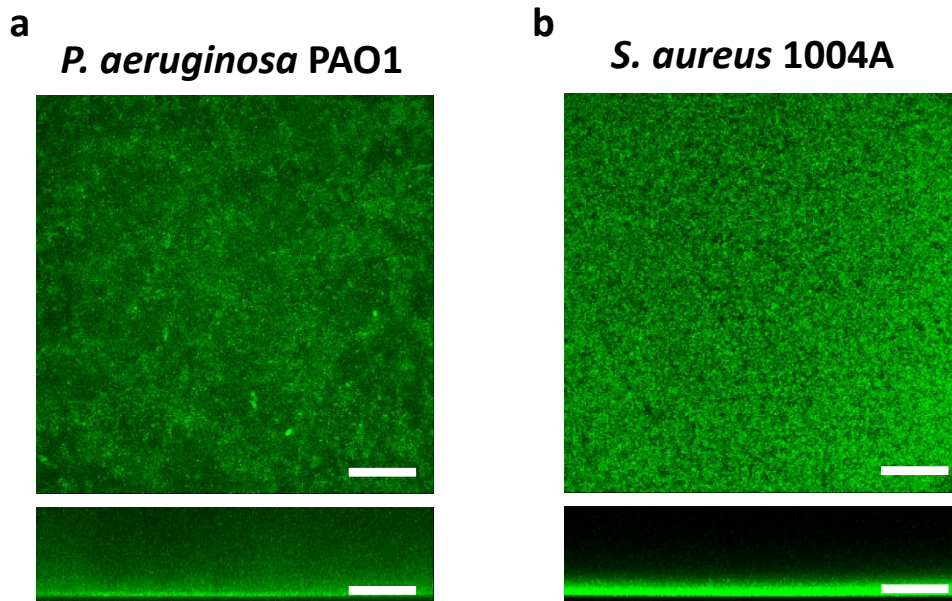


## **SUPPLEMENTARY MATERIALS:**

**Quantifying the effects of antibiotic treatment on the extracellular polymer network of antimicrobial resistant and sensitive biofilms using multiple particle tracking.**

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## SUPPLEMENTARY FIGURE



**Supplementary Fig. 1. Comparison of *P. aeruginosa* and *S. aureus* biofilm structures.**

CLSM 3D imaging of (a) *P. aeruginosa* PAO1 and (b) *S. aureus* 1004A (MRSA) biofilms grown for 72 h at 37°C in MH broth, using Syto9<sup>®</sup> staining (Scale bar, 40 μm; n=3).

## SUPPLEMENTARY MATERIALS AND METHODS

*P. aeruginosa* bacterial strain.

*P. aeruginosa* PAO1 used in this study closely resembles PAO1\_Orsay and PAO1\_ATCC 15692 strains, as demonstrated in high-throughput genome resequencing in an unrelated study (European Nucleotide Archive[ENA] project number PRJEB36146 and accession number for the draft genome sequence GCA\_902860215).

Calculation of mean square displacement  $\langle \text{MSD} \rangle$ , effective diffusion coefficient  $\langle \text{Deff} \rangle$ , and heterogeneity of particle diffusion

The mean square displacement  $\langle \text{MSD} \rangle$  was determined as follows:

$$\text{MSD}_{(n)} = (X_{\Delta t})^2 + (Y_{\Delta t})^2 \quad (1)$$

where the distance the nanoparticle (n) moved over a selected time frame (t) in the X-Y trajectory was expressed as a squared displacement (SD).

The effective diffusion coefficient  $\langle \text{Deff} \rangle$ , of the nanoparticles determined by the following equation:

$$\langle \text{Deff} \rangle = \langle \text{MSD} \rangle / (4 * \Delta t) \quad (2)$$

where 4 is a constant relating to the 2-dimensional mode of video capture and  $\Delta t$  is the selected time interval.

Nanoparticle diffusion in water ( $D^\circ$ ) was calculated by the Stokes–Einstein equation at 37 °C:

$$D^\circ = k_B T / 6\pi\eta r \quad (3)$$

where  $k_B$  is the Boltzmann constant, T is absolute temperature,  $\eta$  is water viscosity, and r is

radius of the nanoparticle.

The diffusion of the nanoparticles was also expressed as the parameter, % ratio  $[D_{eff}]/[D^{\circ}]$ .

The heterogeneity of particle diffusion was measured by profiling the diffusion coefficients ( $\Delta t = 2$  sec) of all individual particles within the entire population (360 particles) from the highest (90th) to the lowest (10th) percentiles in  $\langle D_{eff} \rangle$  values.

### Error arising from experimental noise

The error arising from experimental noise (tracking resolution  $[\sigma]$ ) was measured for each of the *FluoSphere* particles individually, by fixing the particles onto a glass-bottomed imaging dish (MatTek life sciences) with cyanoacrylate-based glue and tracking their movements. Using this set-up, 20 videos were analysed using ImageJ software with Mosaic plugin to independently measure  $\sigma^2$  by determining the X- and Y-directional displacement of the particles at the lowest temporal resolution (0.033 frame per second). The value of  $\sigma^2$  was subtracted from the MSD measurement at the lowest frame rate to achieve final measurements of MSD and  $\langle D_{eff} \rangle$ . The calculated values of  $\sigma$  ranged between 3.45 to 3.96 nm for each of the *Fluospheres* (Supplementary Table 1).

**Supplementary Table 1. Tracking resolution ( $\sigma$ ) of the *FluoSphere*<sup>®</sup> particles**

<i>FluoSpheres</i> <sup>®</sup>	Particle size (nm)	$\sigma$ (nm)
-ve carboxylate	40	3.96
-ve carboxylate	100	3.82
-ve carboxylate	200	3.45
-ve carboxylate	500	3.56
+ve amine	200	3.69