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# Influence of temperature for the azide displacement in benzodiazepine derivatives: Experimental and DFT study of competing S<sub>N</sub>1, S<sub>N</sub>2 and double S<sub>N</sub>2 reaction pathways

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article info

abstract

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Molecules carrying the azido functionality are widely used as intermediates in medicinal chemistry. Pyrolobenzodiazepines are molecules with a broad range of interesting pharmacological properties. Due to the wide importance of azido-PBD as a key intermediate in many synthetic pathways, in this work we have investigated the effect of a variety of experimental parameters on the mechanism of nucleophilic substitution reaction for introducing an azide group into the PBD scaffold. This study was carried out by combined experimental and DFT approaches, showing that S<sub>N</sub>2 and double S<sub>N</sub>2 pathways are possible and highly dependent on the reaction temperature.

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## Introduction

Pyrolo[2,1-c][1,4]benzodiazepines (PBD) (1) are a class of naturally occurring molecules that can be isolated from *Streptomyces* bacteria and that exhibit a broad spectrum of biological activities, featuring its antitumor properties as DNA minor groove binding agents. The first PBD molecule isolated was anthramycin (2) [1]. After that, a number of PBD molecules have been isolated from natural sources and efforts have been applied to synthesize new PBD derivatives that also presents pharmacological and biological properties, including anticancer, binding to GABA<sub>A</sub> receptor and formation of antibody-drug conjugates (ADCs) [2–6].

PBD molecules consist of a tricyclic structure, including an aromatic A-ring, a diazepine B-ring and a pyrrolidine C-ring (3), differing from one another specially by substitutions on the A and C rings, and by the degree of unsaturation of the pyrrolidine ring (Fig. 1). All naturally occurring PBD have S configuration at C-

11a, which provides a three-dimensional conformation that allows the molecules to fit perfectly in the minor groove of DNA [7].

Concerning the structure activity relationships regarding PBD derivatives as anticancer agents, studies have shown that modifications at the substituents of the C-2 position can significantly alter the DNA-binding ability and the cytotoxicity of the molecules. For example, investigations involving anthramycin showed that the removal of its C-2 side chain led to a strong decline of its DNA binding capacity [8,9]. This effect is due to the location along the minor groove of the extended side chains at C-2. Once accommodated along the minor groove, the groups of the side chain can interact with the minor groove floor through hydrogen bonds and Van der Waals interactions [1]. Those interactions stabilize the adduct and enhance the DNA binding affinity leading to higher cytotoxic potency. The comprehension of these interactions has helped researchers to design novel PBD analogues including monomers and dimers with modifications at the C-2 position, with enhanced DNA binding ability and cytotoxicity (Fig. 1) [2,3].

Therefore, we hypothesized that the addition of a second pharmacophore nucleus as a substituent to the C-2 position in the PBD scaffold, would entail increased DNA binding affinity and therefore

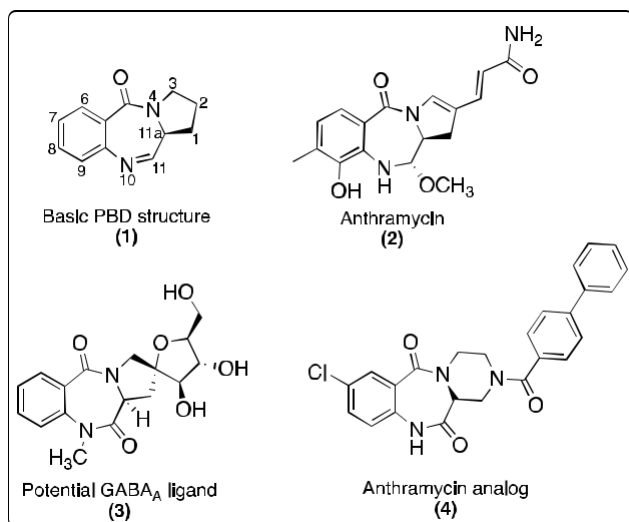


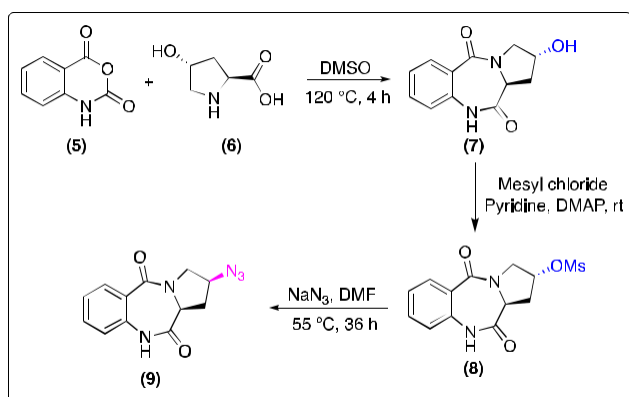
Fig. 1. The pyrolobenzodiazepine (PBD) scaffold and the structure of pharmacologically active PBDs.

pharmacological activity. Triazoles [10], tetrazoles [11], amides [12,13] and its derivatives have shown anticancer activity when linked with another pharmacophore function. These hybrid molecules, containing two or more pharmacophore units, can potentialize the anticancer effect of the PBD core, reduce side effects and overcome other drawbacks associated with PBD derivatives.

## Results and discussion

To obtain the triazole and tetrazole heterocycles mentioned above, a suitable key intermediate is the azido group. The azido functionality has been successfully applied in carbohydrate chemistry [14], synthesis of heterocycles and peptide chemistry [15]. The azido group can be obtained via different reactions, such as substitution or addition via the insertion of the  $N_3$  group, diazotization, diazo transfer, cleavage of triazines and azide rearrangement. Among those, the most common method applied for the synthesis of alkyl azides is the classic nucleophilic substitution reaction [16].

In the present work, the azido-PBD derivative was synthesized according to Scheme 1, starting with a condensation reaction between isatoic anhydride and trans-hydroxy-L-proline, followed by the conversion of the hydroxyl functionality into a better leaving group. After that, the desired  $S_N2$  reaction with  $NaN_3$  would



Scheme 1. Synthesis of the azido-PBD from isatoic anhydride and trans-hydroxy-L-proline.

afford the azido-PBD derivative with inversion of configuration at the nucleophilic center.

As presented in Scheme 1, in the first attempt to obtain the azido-PBD derivative, we followed the procedure published by Schultz and collaborators [17] where the reaction was carried out at 55 LC in DMF for 36 h. The product obtained was recrystallized from acetone, affording colorless crystals. The azide displacement went exclusively with inversion of configuration at C-2 leading to one diastereomer as confirmed by NMR elucidation and crystal structure. In order to optimize the reaction, we decided to apply different conditions to the azide displacement, as described by Rault and collaborators [18] that performed the reaction at 100 LC in DMF for 5 h. However, it was not observed a 100% inversion of configuration at C-2 under these conditions, leading to a mixture of the two possible azido-PBD diastereomers. Such a result could imply that a change in mechanism is occurring at different temperatures.

In the interest of understanding this phenomenon, different reaction conditions were investigated, including the effect of solvents, leaving groups, temperature and reaction times, as presented in Table 1. The reactions were performed using the same ratio between the reactants (1 equiv.) and  $NaN_3$  (5 equiv.) accompanied by TLC (eluent: ethyl acetate) until all the reactant was consumed. The results obtained for the different conditions are presented in Table 1, including the yield and the diastereomeric ratios obtained in the cases where the reactions led to the mixture. The ratio was assigned by  $^1H$  NMR analysis, as shown in Fig. 2.

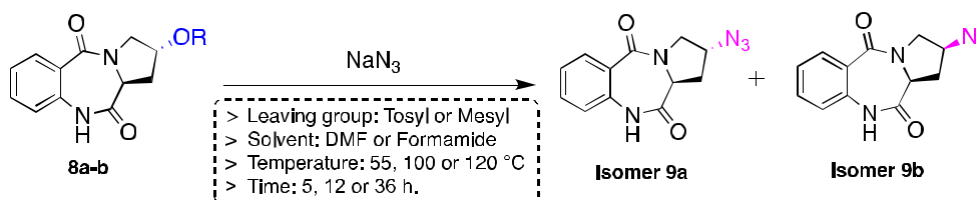
Concerning the solvent choices, the reactions were carried out in DMF and formamide. DMF is a polar aprotic solvent, which can improve the efficiency of the nucleophile in an  $S_N2$ -type reaction while formamide, besides being a polar solvent, also shows the ability to make hydrogen bonds, which is not usually ideal in an  $S_N2$ -type reaction. However, it was not observed significant changes concerning the diastereomeric ratio obtained for the different reactions, indicating that the solvent is not important in changing the mechanistic pathway in the case of azido-PBD formation. The only difference we could observe between the two solvents is the isolated yield, which is higher when using DMF for all reaction conditions (1–3 and 7–9). The reaction time also does not seem to have an impact on the mechanism of the reaction, although as expected longer reaction times were observed for lower temperatures.

The reaction was performed at different temperatures based on previously reported works [16,17]. As discussed before, when the reaction was carried out at 55 LC, just one product was observed, and it was the one corresponding to a 100% inversion of configuration at C-2. The same result was observed when applying different leaving groups and solvents, confirming a substitution by direct displacement ( $S_N2$ ) mechanism (Entries 3, 6, 9 and 12 in Table 1). However, once the temperature was increased to 100 LC, the formation of a mixture of diastereomers started to be observed, where the main diastereomer formed was still 9b. The same results were obtained for all the reactions performed at 100 LC, regardless of which leaving group or solvent was used, with diastereomeric ratios similar to one another (Entries 2, 5, 8 and 11). When the reactions were performed at 120 LC, the mixture of diastereomers was observed for all the entries. However, the diastereomeric ratio of the products changed

drastically, leading to the isomer 9a being the major one for entries 1, 7 and 10, except for entry 4, where the ratio is close to 50:50.

X-ray crystallography was employed to unequivocally characterize stereochemistry and conformation of one of the obtained azido-PBD diastereomers. A single crystal of compound 9b was obtained by slow evaporation of acetone from the solution of pure diastereomer. 9b crystallizes in the  $P2_1$  space group containing one

Table 1  
Evaluation of different reaction conditions for the azide displacement reaction.



Entry	LG <sup>a</sup>	Temperature	RT <sup>b</sup>	Solvent	<sup>1</sup> H NMR ratio Isomer 9a: 9b	Yield (%)
1	Tosyl	120 LC	12 h	DMF	Mixture (59:41)	94
2	Tosyl	100 LC	5 h	DMF	Mixture (9:91)	85
3	Tosyl	55 LC	36 h	DMF	(0:100)	85
4	Tosyl	120 LC	12 h	Formamide	Mixture (49:51)	29
5	Tosyl	100 LC	5 h	Formamide	Mixture (15:85)	45
6	Tosyl	55 LC	36 h	Formamide	(0:100)	55
7	Mesyl	120 LC	12 h	DMF	Mixture (63:37)	82
8	Mesyl	100 LC	5 h	DMF	Mixture (36:64)	62
9	Mesyl	55 LC	36 h	DMF	(0:100)	47
10	Mesyl	120 LC	12 h	Formamide	Mixture (65:35)	50
11	Mesyl	100 LC	5 h	Formamide	Mixture (9:91)	46
12	Mesyl	55 LC	36 h	Formamide	1 diastereoisomer	50

<sup>a</sup> Leaving Group.

<sup>b</sup> Reaction time.

independent molecule per unit cell. The asymmetric unit is shown in Fig. 3, and the stereochemistry of C2 confirms that the reaction occurred through S<sub>N</sub>2 mechanism. The crystal packing is stabilized by intermolecular hydrogen bonds between oxygen atom O1 and amine group (N2-H7). Each molecule shows intermolecular inter-actions with two other adjacent molecules generating a supramolecular chain along the crystallographic direction b (see Fig. S1 for details).

At the final stage of this investigation, DFT calculations have been applied to provide deeper insight into the effect of temperature over the alternative reaction mechanisms for OMs displacement by azide in the investigated benzodiazepine derivatives. Using 8b as substrate for this investigation, the energy profile of three competing reaction channels were computed, S<sub>N</sub>1, S<sub>N</sub>2 and

“double” S<sub>N</sub>2, as shown in Fig. 4. As observed during the experimental investigations, the S<sub>N</sub>2 pathway (Fig. 4) presented the low-est free energy barrier from 8b (27.2 kcal mol<sup>-1</sup>), which clearly justify the exclusive formation of product 9b at low reaction temperatures. However, with increasing temperature, a mixture of products was observed which raised the hypothesis of a possible change in the observed mechanisms, either via a S<sub>N</sub>1 mechanism or via the so-called “double” S<sub>N</sub>2 as proposed by Szabó and Czakó [18,19].

Firstly, we investigated the energy cost to form a carbocation species in the reaction mixture by elimination of the substituent group at C-2 (either OMs or N<sub>3</sub>). As described in previous works, computing the energy barriers for formation of such intermediate is, in general, a good approximation to estimate the rate constant

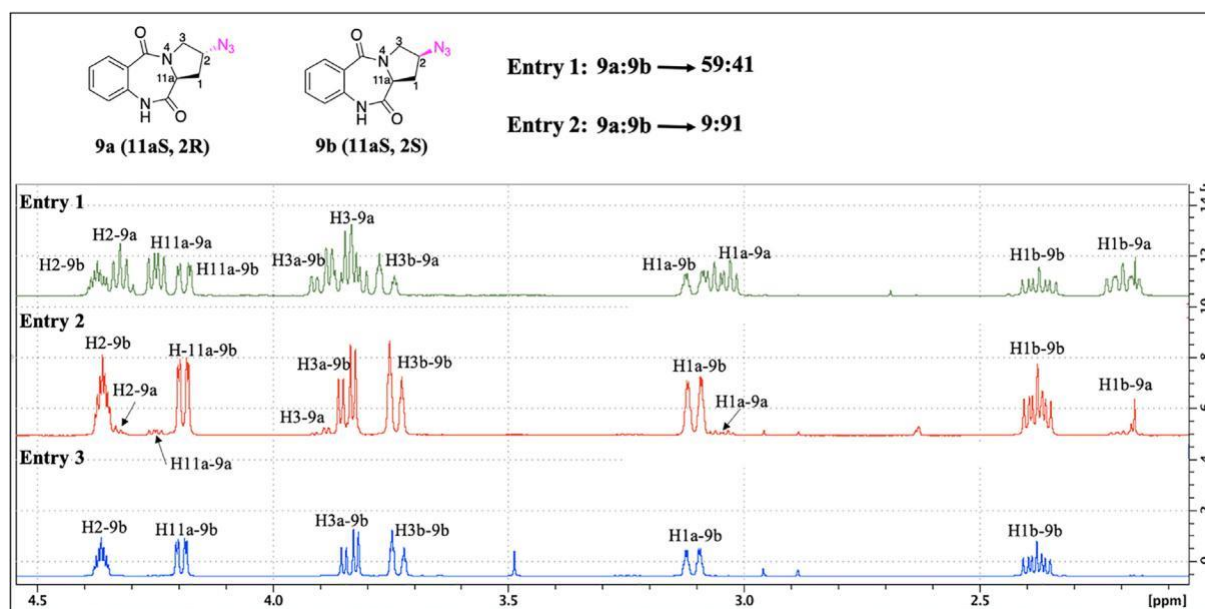


Fig. 2. <sup>1</sup>H NMR spectra from aliphatic region used for the determination of the diastereomeric ratio. (CDCl<sub>3</sub>, 400.13 MHz).

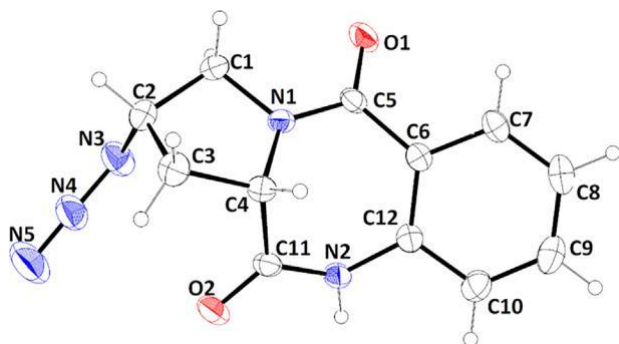
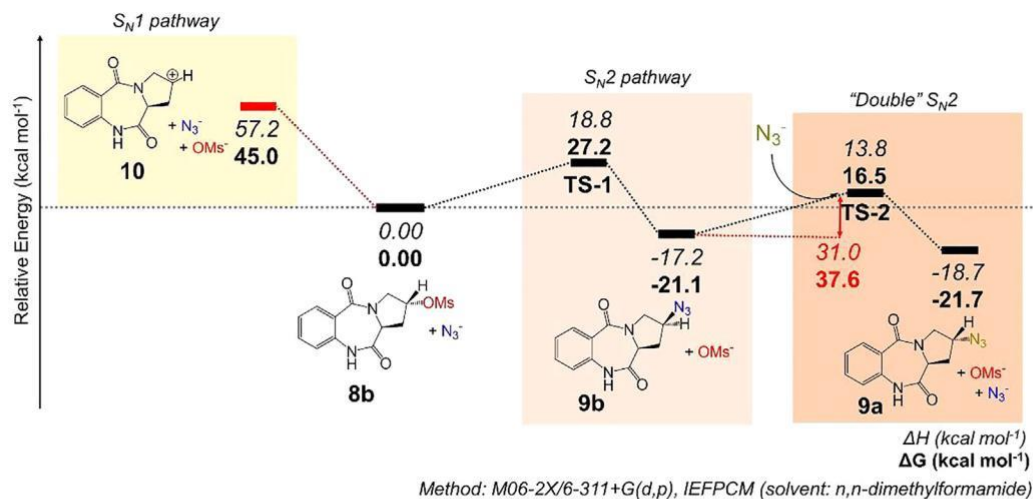


Fig. 3. ORTEP view of the asymmetric unit of 9b with thermal ellipsoids drawn at 30% of probability.

for conventional  $S_N1$  reactions [20]. As it can be seen in Fig. 4, such carbocation would be a highly energetic intermediate, presenting relative free energy higher than 45 kcal mol<sup>-1</sup> in comparison to the isolated reactants. The possible stabilization of 10 by interactions with nucleophile and leaving group was also tested by the construction of a [OMs 10 N<sub>3</sub>] complex, however the resulting decrease in energy was smaller than 2 kcal mol<sup>-1</sup> when the anions were kept 3.5 Å away from the carbocationic center.

On the other hand, the alternative “double”  $S_N2$  process presents much lower free energy barrier than that to form the above-mentioned carbocation intermediate. Starting from intermediate 9b, which has relative free energy around 21.0 kcal mol<sup>-1</sup> compared to 8b, the activation Gibbs free energy to form the product with a second inversion via a second  $S_N2$  reaction is 37.6 kcal mol<sup>-1</sup>. Although this is a relatively high energy, requiring heat to be surmounted, it is lower than the activation Gibbs free energy for the  $S_N1$  process. As a result, the most favorable process identified here for OMs substitution in 8b is an initial  $S_N2$  reaction, with a Gibbs free energy barrier of 27.2 kcal mol<sup>-1</sup>, followed by a “double”  $S_N2$  step presenting a free energy barrier of 37.6 kcal mol<sup>-1</sup>. These results are consistent with the experimental observation of enantioselective formation of 9a and 9b at distinct temperatures. Formation of 9b occurs with the lowest energy barrier. As this is a stable intermediate, its lifetime is enough to dissipate energy. At low temperature this is the main product formed. Increasing the temperature 9b can be converted into 9a by reaction with excess N<sub>3</sub> in the reaction mixture. Furthermore, the slightly higher stability of 9a regarding 9b is also consistent with the observation of increased amount of 9a when longer reaction time is used. This observation was investigated experimentally by treating diastereomerically pure 9b with sodium azide in DMF at 120 °C overnight. This resulted in 63:37 9a:9b ratio, confirming the preference for a “double”  $S_N2$  pathway at higher temperatures.

(a) Energy profile for competing  $S_N1$ ,  $S_N2$ , and “double”  $S_N2$  pathways



(b) Optimized carbocation and transition state structures

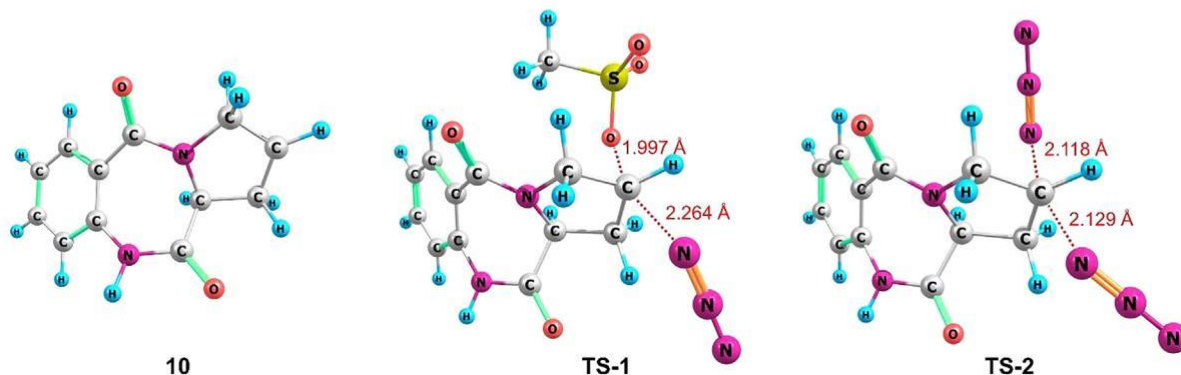


Fig. 4. Computed energy profile for alternative  $S_N1$ ,  $S_N2$ , and “double”  $S_N2$  reaction pathways for OMs displacement by azide in 8b.

## Conclusions

This study showed that small changes in temperature led to retention of configuration in the azide displacement of PBD derivatives. This is in contrast to previously reported literature, which describes 100% inversion of configuration at the substitution center. 1D and 2D-NMR techniques were used to identify products and follow the changes in the reaction over time. Furthermore, X-ray crystallography was employed to unequivocally characterize stereochemistry and conformation of one of the obtained azido-PBD diastereomers. While products with inversion of configuration are the commonly expected outcome for this reaction, we have shown that small changes in the reaction temperature can lead to the diastereomer with retention of configuration. Results of DFT calculations have provided a theoretical insight into the effect of the temperature over the reaction mechanism, showing that a “double” S<sub>N</sub>2 mechanism takes place at higher temperatures, which was confirmed experimentally. Our results can be useful when evaluating the pharmacological profile of molecules prepared from key azido-PBD intermediates, where molecules with high enantiomeric purity are required. In addition, the ability to obtain both diastereomers is also an advantage, once it allows for evaluating their pharmacological properties, which can vary considerably.

## Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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## Appendix A. Supplementary data

Supplementary data (detailed experimental procedures, characterization data (<sup>1</sup>H, <sup>13</sup>C NMR and HRMS), copies of spectra and further information concerning DFT studies and X-ray) are provided. CCDC 2041317 contains the supplementary crystallographic data as CIF file for this paper. This data can be obtained free of charge from The Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/structures](http://www.ccdc.cam.ac.uk/structures)) to this article can be found online at <https://doi.org/10.1016/j.tetlet.2021.152937>.

## References

- [1] J. Mantaj, P.J.M. Jackson, K.M. Rahman, D.E. Thurston, *Angew. Chem. Int. Ed.* 56 (2017) 462–488.
- [2] M.S.K. Sutherland, R.B. Walter, S.C. Jeffrey, P.J. Burke, C. Yu, H. Kostner, I. Stone, M.C. Ryan, D. Sussman, R.P. Lyon, W. Zeng, K.H. Harrington, K. Klusman, L. Westendorf, D. Meyer, I.D. Bernstein, P.D. Senter, R.B. Dennis, J.G. Drachman, *A. McEarchem, Blood* 122 (2013) 1455–1463.
- [3] A. Mieczkowski, M. Psurski, M. Baginski, B. Bieszczad, M. Mroczkowska, M. Wilczek, J. Czajkowska, D. Trzybinski, K. Wozniak, J. Wietrzyk, *Bioorg. Med. Chem. Lett.* 28 (2018) 618–625.
- [4] A.C. Araújo, A.P. Rauter, F. Nicotra, C. Airoldi, B. Costa, L. Cipolla, *J. Med. Chem.* 54 (2011) 1266–1275.
- [5] G.C. Kemp, A.C. Tiberghien, N.V. Patel, F. D’Hooge, S.M. Nilapar, L.R. Adams, S. Corbett, D.G. Williams, J.A. Hartley, P.W. Howard, *Bioorg. Med. Chem. Lett.* 27 (2017) 1154–1158.
- [6] A.C. Tiberghien, J.N. Levy, L.A. Masterson, N.V. Patel, L.R. Adams, S. Cobert, D.G. Williams, J.A. Hartley, P.W. Howard, *ACS Med. Chem. Lett.* 7 (2016) 983–987.
- [7] J.A. Hartley, *Expert. Opin. Investig. Drugs* 20 (2011) 733–744.
- [8] M.L. Kopka, D.S. Goodsell, I. Baikalov, K. Grzeskowiak, D. Cascio, R.E. Dickerson, *Biochemistry* 33 (1994) 13593–13610.
- [9] S.B. Hoewitz, S.C. Chang, A.P. Grollman, A.B. Borkovec, *Science* 174 (1971) 159–161.
- [10] Z. Xu, S.-J. Khao, Y. Liu, *Eur. J. Med. Chem.* (2019) 111700.
- [11] L. Lauko, P.H.J. Kouwer, A.E. Rowan, *Anti-Cancer Agents Med. Chem.* 17 (2017) 1856–1868.
- [12] Y. Lu, Z. Wang, C.-M. Lib, J. Chen, J.T. Dalton, W. Li, D.D. Miller, *Bioorg. Med. Chem.* 18 (2010) 477–495.
- [13] T. Sreelatha, S. Kandhasamy, R. Dinesh, S. Shruthy, S. Shweta, D. Mukesh, D. Karunakaran, R. Balaji, N. Mathivanan, *Bioorg. Med. Chem. Lett.* 24 (2014) 3647–3651.
- [14] S.B. Ferreira, A.C.R. Sodero, M.F.C. Cardoso, E.S. Lima, C.R. Kaiser, F.P. Silva-Jr, V. F. Ferreira, *J. Med. Chem.* 53 (2010) 2364–2375.
- [15] I.F. Silva, P.R.C. Martins, E.G. Silva, S.B. Ferreira, V.F. Ferreira, K.R.C. Costa, M.C. Vasconcellos, E.S. Lima, F.C. Da Silva, *Med. Chem.* 9 (2013) 1085–1090.
- [16] Y. Bi, P.G. Schultz, *Bioorg. Med. Chem. Lett.* 6 (1996) 2299–2300.
- [17] A.C. Gillard, M. Alkhalid, S. Rault, *Heterocycl. Commun.* 2 (1996) 409–414.
- [18] I. Szabo, G. Czako, *Nature Commun.* 6 (2015) 5972.
- [19] J. Xie, W.L. Hase, *Science* 352 (2016) 32–33.
- [20] N. Streidl, B. Denegri, O. Kronja, H.A. Mayr, *Acc. Chem. Res.* 43 (2010) 1537–1549.