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# Pre-transplant *FLT3*-ITD MRD assessed by high-sensitivity PCR-NGS determines post-transplant clinical outcome

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#### Abstract:

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## Short title: Pre-transplant FLT3-ITD MRD and clinical outcome

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FMS-like tyrosine kinase-3 internal tandem duplication (FLT3-ITD) is identified in ~25% of patients with acute myeloid leukemia (AML), making it one of the most common variants identified in this disease.<sup>1</sup> Although frontline incorporation of the FLT3 inhibitor midostaurin has been shown to improve survival of patients with FLT3 mutant AML, it is broadly accepted that allogeneic hematopoietic cell transplant (allo-HCT), performed in approximately 25-30% of patients in first remission, is a key contributor to enhanced outcomes.<sup>2</sup> Two studies have shown that detection of FLT3-ITD prior to transplant in morphologic remission using conventional fluorescence-based polymerase chain reaction (PCR) techniques with 1-5% sensitivity, was associated with higher post-HCT relapse risk (39-59%), compared to patients "negative" for FLT3-ITD (relapse risk 23-41%).<sup>3,4</sup> Assessment of FLT3-ITD by capillary electrophoresis (CE)-based approaches, however, has low sensitivity (~1%), in contrast to high coverage PCR and next-generation sequencing (NGS), which enables detection of *FLT3*-ITD with 100-1000-fold greater sensitivity.<sup>5-7</sup> Using NGS to assess FLT3-ITD with a limit of detection (LOD) of 10<sup>-4</sup> to 10<sup>-5</sup>, the proportion of patients measurable residual disease (MRD) negative after 2 cycles of intensive chemotherapy combined with midostaurin was 67%.8 Currently, it is not known whether detection of FLT3-ITD clones below the sensitivity of CE-based methods has clinical relevance in forecasting post-HCT relapse risk, especially if myeloablative conditioning (MAC) is administered. One study incorporating peripheral blood (PB) detection of FLT3-ITD prior to MAC or reduced-intensity conditioned (RIC) HCT found that 9/10 patients positive for FLT3-ITD MRD [variant allele frequency (VAF), 0.03%-3.97%] relapsed, compared to 1/7 relapsing if FLT3-ITD MRD was negative pre-HCT.9

We therefore sought to evaluate the prognostic impact of MRD detection by PCR-NGS to detect *FLT3*-ITD with high sensitivity prior to allo-HCT and to determine the added value of this approach compared to CE. The study was approved by Alfred Health, Peter MacCallum Cancer Centre, Royal Melbourne Hospital (181/21), and All-Wales (08/MRE09/29) ethics committees<sub>a</sub> and allAll subjects gaveprovided informed consent and the study was conducted according to the principles established by the Declaration of Helsinki. Patients with *FLT3*-ITD AML at diagnosis undergoing their first allo-HCT in morphologic remission (bone marrow (BM) blasts <5%) and with pre-HCT DNA available (92 BM and 12 PB) were included in this retrospective cohort (Supplemental Figure S1). Strong concordance between BM and PB sampling was noted (Supplemental methods). None of the patients were exposed to FLT3 inhibitor maintenance post-HCT. Amplicon-based PCR-NGS of exons 14-15 detected *FLT3*-ITDs of at least 6 base pairs (bp) to a minimum depth of 500K reads,

resulting in an LOD of 0.001% (Supplemental Methods).<sup>7</sup> *NPM1* MRD by RT-qPCR and NGS were determined as previously described.<sup>10,11</sup> Kaplan-Meier survival estimates were calculated from HCT date to the date of death or last follow-up (overall survival [OS]); and/or morphologic relapse (relapse-free survival [RFS]). Cumulative incidence of relapse (CIR) was estimated considering transplant-related mortality (TRM) as a competing risk. Cox proportional hazards model was used for uni- and multivariate analyses.

The cohort included 104 patients with a median age of 49 years (17-68). One guarter had prior exposure to FLT3-inhibitors, 43% had a baseline FLT3-ITD allelic ratio ≥0.5, 75% had co-existing NPM1 mutation, 86% transplanted in first remission (CR1) and 49% received MAC (Table 1). The higher-than-expected proportion of patients with NPM1 co-mutation is explained by the NCRI AML17 cohort being enriched for patients undergoing NPM1 MRD monitoring. Twelve patients received MRD directed therapy prior to HCT to treat molecular NPM1mut relapse (Supplemental Figure S2). Among the 104 patients in morphologic remission pre-HCT, 37% had FLT3-ITD MRD detected by PCR-NGS (median VAF 0.041% [range, 0.0011-9.352]). Of the cases who were FLT3-ITD positive by PCR-NGS, only 7/36 (19%) were positive by CE (Figure 1A). The median time from the date pre-HCT FLT3-ITD MRD was assessed to the date of HCT was 27.5 days (range, 1-87). The 2-year RFS was 78%, 32%, 40% and 0% for *FLT3*-ITD MRD levels <0.001 (negative), ≥0.001-<0.1%, ≥0.1-<1%, and ≥1%, respectively (Figure 1B), with the corresponding CIR and OS according to the level of *FLT3*-ITD MRD shown in Supplemental Figure S3. Therefore, post-HCT relapse was substantial even for patients with FLT3-ITD MRD levels <1%, below the detection threshold of CE. For patients positive for FLT3-ITD by both PCR-NGS and CE, CIR was 100% within the first 6 months post-HCT (Figure 1C for CIR and Supplemental Figure S4 for RFS). For patients FLT3-ITD MRD positive by PCR-NGS but not by CE, the CIR post-HCT was 67% (Figure 1C). In contrast, patients with FLT3-ITD MRD levels <0.001% (negative) by PCR-NGS had a post-HCT relapse risk of 16%. OS post-HCT was also negatively impacted by the presence of FLT3-ITD MRD pre-HCT, with ≤26% alive at 4 years, compared to 74% if *FLT3*-ITD MRD was negative (Figure 1D). Similar to previously published results,<sup>9</sup> MAC was not able to attenuate the high relapse risk associated with pre-HCT FLT3-ITD MRD, with RFS and OS comparable to that observed with RIC (Supplemental Figure S5). Multivariate analysis considering factors listed in Supplemental Table S1 demonstrated that pre-HCT FLT3-ITD MRD (hazard ratio [HR] 4.94, p=<0.0001), transplant in CR2 vs CR1 (HR 2.39, p=0.05) and T-cell depletion (HR 2.80, p=0.006) were the most important determinants of post-HCT relapse risk and survival.

We next sought to determine whether *FLT3*-ITD MRD provided additional prognostic value among patients also known to be *NPM1* mutant. Of 71 patients with both *FLT3*-ITD and

*NPM1* mutation at AML diagnosis with available pre-HCT MRD for both markers, RFS and OS were most favorable for those negative for both *NPM1* and *FLT3*-ITD MRD pre-HCT (Figures 1E and 1F). In contrast, outcomes were dismal for patients double positive for both *NPM1* and *FLT3*-ITD MRD. Interestingly, intermediate RFS and OS were observed for patients positive for either *NPM1* or *FLT3*-ITD MRD prior to HCT, suggesting MRD assessment for *NPM1* and *FLT3*-ITD had complementary clinical value (Figures 1E and 1F). In this *FLT3*-ITD cohort, low-level *NPM1* MRD (<2%) was associated with inferior prognosis, despite allo-HCT (Supplemental Figure S6).<sup>12</sup>

Our results highlight the dismal prognosis associated with pre-HCT detection of FLT3-ITD MRD using high-sensitivity PCR-NGS-based approaches. The poor prognosis associated with FLT3-ITD MRD was relevant even at very low MRD levels and was not ameliorated by MAC HCT. FLT3-ITD MRD detection pre-HCT may therefore be an indication for future MRD directed therapeutic strategies, in either the pre- or post-HCT setting. Clinical studies in *FLT3*mut relapsed/refractory AML indicate that gilteritinib may reduce *FLT3*-ITD VAF to  $\leq 10^{-4}$ in 29.2% of patients achieving complete remission or complete remission with partial hematologic recovery.<sup>13</sup> Gilteritinib combined with the BCL-2 inhibitor venetoclax reduced *FLT3*-ITD levels to  $\leq 10^{-2}$  in 56.7% patients achieving morphologic remission.<sup>14</sup> Although literature supports the clinical efficacy of post-transplant FLT3 inhibitor maintenance in reducing relapse risk and death,<sup>15</sup> the efficacy of FLT3 inhibitors in suppressing *FLT3*-ITD MRD levels in the post-HCT setting has not been formally demonstrated. This is likely to be addressed by the BMT-CTN1506 study, which randomized patients post-HCT to either gilteritinib or placebo maintenance with PCR-NGS MRD assessments planned at pre- and post-HCT timepoints. An intriguing future question is whether FLT3-ITD MRD assessment by high-coverage PCR-NGS could identify patients most likely to benefit from FLT3-directed therapy, either before or after transplant. In conclusion, FLT3-ITD assessment by PCR-NGS has important clinical value and warrants further investigation as a guide for precision-based therapeutic strategies aimed at improving the natural history of patients identified to be MRD positive pre-HCT.

The data that support the findings of this study are available from the corresponding author, AHW, upon reasonable request.

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## Authorship

Contribution: A.H.W. and S.L. designed the research; A.I. designed and validated the PCR-NGS assay; A.I., N.S.A., N.P., J.J., M.R., M.M. and A.G. sequenced and analyzed the samples or provided other molecular analysis tools and interpretation; J.O. and S.L. performed clinical data collection; S.L. performed the statistical analysis; S.L., R.D., J.O., I.S.T., C.C.C., A.B., D.R., Z.H.Y., K.G., I.T., S.J., N.H.R. and A.H.W. contributed patients or analyzed and interpreted data; S.L. and A.H.W. wrote the manuscript; all authors read and approved the manuscript.

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## References

- 1. Papaemmanuil E, Gerstung M, Bullinger L, et al. Genomic classification and prognosis in acute myeloid leukemia. *N Engl J Med.* 2016;374(23):2209-2221.
- 2. Stone RM, Mandrekar SJ, Sanford BL, et al. Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. *N Engl J Med.* 2017;377(5):454-464.
- Gaballa S, Saliba R, Oran B, et al. Relapse risk and survival in patients with FLT3 mutated acute myeloid leukemia undergoing stem cell transplantation. *Am J Hematol.* 2017;92(4):331-337.
- 4. Helbig G, Koclega A, Wieczorkiewicz-Kabut A, et al. Pre-transplant FLT3/ITD status predicts outcome in FLT3-mutated acute myeloid leukemia following allogeneic stem cell transplantation. *Ann Hematol.* 2020;99(8):1845-1853.
- 5. Thol F, Kölking B, Damm F, et al. Next-generation sequencing for minimal residual disease monitoring in acute myeloid leukemia patients with FLT3-ITD or NPM1 mutations. *Genes, Chromosomes and Cancer.* 2012;51(7):689-695.
- Bibault JE, Figeac M, Helevaut N, et al. Next-generation sequencing of FLT3 internal tandem duplications for minimal residual disease monitoring in acute myeloid leukemia. *Oncotarget*. 2015;6(26):22812-22821.
- 7. Blätte TJ, Schmalbrock LK, Skambraks S, et al. getITD for FLT3-ITD-based MRD monitoring in AML. *Leukemia*. 2019:1.
- Herzig JK, Rucker FG, Schmalbrock LK, et al. Next-generation sequencing (NGS)-based measurable residual disease (MRD) monitoring in acute myeloid leukemia with FLT3 internal tandem duplication (*FLT3*-ITD+ AML) treated with additional midostaurin (abstract). *Blood*. 2020;136:21-22.

- 9. Hourigan CS, Dillon LW, Gui G, et al. Impact of conditioning intensity of allogeneic transplantation for acute myeloid leukemia with genomic evidence of residual disease. *J Clin Oncol.* 2020;38(12):1273-1283.
- 10. Ivey A, Hills RK, Simpson MA, et al. Assessment of minimal residual disease in standard-risk AML. *N Engl J Med.* 2016;374(5):422-433.
- 11. Blombery P, Jones K, Doig K, et al. Sensitive NPM1 mutation quantitation in acute myeloid leukemia using ultradeep next-generation sequencing in the diagnostic laboratory. *Arch Pathol Lab Med.* 2018;142(5):606-612.
- 12. Dillon R, Hills R, Freeman S, et al. Molecular MRD status and outcome after transplantation in NPM1-mutated AML. *Blood.* 2020;135(9):680-688.
- 13. Altman JK, Perl AE, Hill JE, Rosales M, Bahceci E, Levis MJ. The impact of FLT3 mutation clearance and treatment response after gilteritinib therapy on overall survival in patients with FLT3 mutation-positive relapsed/refractory acute myeloid leukemia. *Cancer Med.* 2021;10(3):797-805.
- 14. Daver N, Perl AE, Maly J, et al. Venetoclax in combination with gilteritinib demonstrates molecular clearance of FLT3 mutation in relapsed/refractory FLT3-mutated acute myeloid leukemia (abstract). *Blood.* 2021;138:691.
- 15. Burchert A, Bug G, Fritz LV, et al. Sorafenib maintenance after allogeneic hematopoietic stem cell transplantation for acute myeloid leukemia with FLT3-internal tandem duplication mutation (SORMAIN). *J Clin Oncol.* 2020;38(26):2993-3002.

## Table 1. Baseline characteristics

	All patients	Pre-HCT: FLT3-ITD positive <sup>1</sup>		Pre-HCT: FLT3-ITD
Variables at AML				negative
diagnosis, n (%)	(n=104)	PCR-NGS pos	PCR-NGS pos,	PCR-NGS neg
		& CE pos (n=7)	CE neg (n=29)	(n=66)
Median age; years (range)	49 (17-68)	44 (25-68)	51 (26-68)	47 (17-68)
Males	47 (45)	1 (14)	18 (62)	27 (42)
AML subtype	X = /	· · · · ·	- (- )	
- De novo	97/102 (95)	7 (100)	26 (90)	62/64 (97)
- Secondary	5/102 (5)	-	3 (10)	1/64 (3)
Prior therapy				
- No FLT3 inhibitor	76 (73)	6 (86)	18 (62)	50 (76)
<ul> <li>FLT3 inhibitor</li> </ul>	28 (27)	1 (14)	11 (38)	16 (24)
Karyotype risk <sup>2</sup>				
- Favorable	-	-	-	-
- Intermediate	95/96 (99)	7/7 (100)	28/28 (100)	59/59 (100)
- Adverse	1 /96 (1)	-	-	-
<i>FLT3</i> -ITD allelic ratio				
(1 NA)				
- <0.5	59/103 (57)	3 (43)	15/28 (54)	40 (61)
- ≥0.5	44/103 (43)	4 (57)	13/28 (46)	26 (39)
NPM1 mutant	78 (75)	4 (57)	21 (72)	52 (79)
	26 (25)	3 (43)	8 (28)	14 (21)
FLI3-IKD mut	4/96 (4)		2/28 (7)	2/59 (3) 57/50 (07)
FL13-1KD wild-type	92/96 (96)		20/28 (93)	57/59 (97) 22/40 (57)
DNMT3A mutant	33/39 (30)	2/2 (100)	9/10 (30) 7/16 (44)	23/40 (37)
Driver lines of thorapy	20/59 (44)	2/2 (100)	7/10 (44)	17/40 (43)
pre-HCT				
- 1	75 (72)	2 (29)	19 (66)	52 (79)
- 2	29 (28)	5 (71)	10 (34)	14 (21)
 Conditioning <sup>4</sup>	20 (20)	0 (7 1)		()
- Mveloablative	40/82 (49)	3/6 (50)	12/22 (55)	24/53 (44)
- Reduced intensity	42/82 (51)	3/6 (50)	10/22 (45)	29/52 (56)
Transplant status				
CR1	89 (86)	3 (43)	25 (86)	59 (89)
CR2	15 (14)́	4 (57)	4 (14)	7 (11)
T-cell depletion <sup>5</sup>	41/81 (51)	4/5 (80)	12/22 (54)	25/53 (47)
Unrelated donor <sup>6</sup>	52/100 (52)	3/6 (50)	9/28 (32)	40/65 (62)
Median time from last	42.5 (12-			
treatment to day 0 of	147)			
HCT; days (range)				

<sup>1</sup>2 without available sample for CE correlation; <sup>2</sup>Determined according to UK MRC criteria. 8 patients with unknown karyotype; <sup>3</sup>*DNMT3A* mutation testing in 59 patients only; <sup>4</sup>Conditioning intensity information available in 82 patients. Myeloablative conditioning regimes were busulfan and cyclophosphamide (Bu/Cy), cyclophosphamide and total body

irradiation (Cy/TBI) and fludarabine and 4 days of busulfan (Flu/Bu4); <sup>5</sup>T-cell depletion information available in 81 patients; <sup>6</sup>Donor source information available in 100 patients. Abbreviations: HCT, hematopoietic cell transplant; CE, capillary electrophoresis; PCR-NGS, polymerase chain reaction-next-generation sequencing; mut, mutant; CR1, first remission; CR2, second remission.

## **Figure Legends**

## Figure 1.

(A) The majority of *FLT3*-ITD MRD detected by PCR-NGS pre-HCT is below the threshold of conventional capillary electrophoresis (CE). The limit of detection (LOD) for *FLT3*-ITD as assessed by CE and PCR-NGS are indicated. Blue boxes indicate 2 patients positive for *FLT3*-ITD by PCR-NGS but without available CE data.

# (B) Relapse risk post-HCT is associated with *FLT3*-ITD PCR-NGS VAF ≥0.001%.

Kaplan-Meier estimates of relapse-free survival according to pre-HCT *FLT3*-ITD MRD levels showing highest relapse risk for *FLT3*-ITD MRD  $\geq$ 1%, lowest risk for *FLT3*-ITD MRD <0.001% and intermediate risk for levels between 0.001-<1%.

# (C-D) Pre-HCT FLT3-ITD MRD is associated with inferior clinical outcomes. Kaplan-

Meier estimates of (C) Cumulative incidence of relapse (with transplant-related mortality as a competing risk in 14 patients not included in this curve) and (D) Overall survival according to pre-HCT *FLT3*-ITD PCR-NGS and CE status. Two patients with positive *FLT3*-ITD by PCR-NGS lacking CE data were excluded.

# (E-F) Pre-HCT FLT3-ITD and NPM1 MRD are prognostic for clinical outcome post-HCT.

Kaplan-Meier estimates of (E) Relapse-free and (F) Overall survival according to pre-HCT *FLT3*-ITD PCR-NGS MRD and *NPM1* MRD in 71 of 78 co-mutated for both *FLT3*-ITD and *NPM1*. Seven patients with insufficient material for *NPM1* MRD assessment were excluded.

