Structure-Guided Engineering of Immunotherapies Targeting TRBC1 and

**TRBC2 in T Cell Malignancies** 

## **Supplementary Materials**

#### **Supplementary Figures**

#### Supplementary Fig. 1: TRBC1/TRBC2 homology



Overlay of TCR proteins containing the  $\alpha\beta1$  (blue) or  $\alpha\beta2$  (orange) chains showing almost complete overlap with the exception of the Asn119/Lys120 and Lys119/Asn120 for TRBC1 and TRBC2, respectively (dashed circle). Central and right panels show surface analysis of TCR $\alpha\beta1$  and TCR $\alpha\beta2$ , showing very comparable surface charge (blue=positive and red=negative) and hydrophobic patch (green) distribution.

### Supplementary Fig. 2: SPR humanized Jovi-1



Affinity determination of murine (Mu)Jovi-1 and HuJovi-1 binding to TRBC1 (top) and TRBC2 (bottom) recombinant TCR HA1.7. Jovi-1/TRBC1 KD = 1.60 nM, HuJovi-1/TRBC1 KD = 2.57 nM. Source data are provided as a Source Data file.



### Supplementary Fig. 3 Humanized Jovi-1 CAR expression profile

a, Surface plasmon resonance affinity kinetics of anti-idiotype antibody for MuJovi-1 (left), HuJovi-1 (center) or non-related antibody (right). b, ELISA binding titration of anti-idiotype antibody for MuJovi-1, HuJovi-1 or non-related control antibody in human IgG1 format. Antiidiotype detected with Anti-rabbit IgG HRP conjugated secondary antibody. EC50 value obtained with Graphpad Prism v9. n=2 technical replicate, data shown as mean  $\pm$  SD. c, Overview of PBMC sorting in TRBC1<sup>+</sup> and TRBC2<sup>+</sup> populations (top). (Bottom) Human PBMCs are first sorted using biotinylated muJovi-1 antibody via magnetic biotin selection kit. Unbound fraction corresponding to  $\text{TRBC2}^+$  cells. Bound fraction is eluted from beads and corresponds to TRBC1<sup>+</sup> population. Staining obtained with muJovi-1 PE conjugated on CD3<sup>+</sup> gated population. (Right) % of contaminating TRBC1<sup>+</sup> (orange) or TRBC2<sup>+</sup> (blue) cells from 27 individual processed donors. Data shown as box and whiskers min to max. d, Example of flow cytometry analysis of PBMC transduction efficiency for MuJovi-1 CAR (red) and HuJovi-1 CAR (blue). Transduction and expression of CAR detected with anti-CD34 Ab (for RQR8 marker) and anti-idiotype (for CAR). Similar degree of transduction efficiency was determined for both CAR constructs. e, Ratio of RQR8 (via anti-CD34) and CAR (via antiidiotype) MFI signal for MuJovi-1 (orange) and HuJovi-1 (blue) CAR constructs. Significantly higher expression detected for HuJovi-1 CAR. Unpaired t test, \*\*\* p < 0.001, n=4 individual PBMC donors. Data shown as mean  $\pm$  SD. Source data are provided as a Source Data file.



**a**, TCR expression levels on endogenously expressing cell lines Jurkat TRBC1 (n=3 technical replicate), H9 TRBC1 (n=3 technical replicate), HPB-ALL TRBC2 (n=3 technical replicate), T-ALL1 TRBC2 (n=3 technical replicate), HD-MAR2 TRBC2 (n=2 technical replicate), engineered cell lines (n=3 biologically independent samples) and healthy human PBMC CD4+ and CD8+ (n=4 biologically independent samples). TCR detected with antibody clone WT31.

Data shown as mean  $\pm$  SD. **b**, TRBC chain profiling on T cell lines. TCR expression detected with anti-CD3 $\epsilon$ , TRBC1 expression detected with MuJovi-1 and TRBC2 expression detected with modified KFN aTRBC2 antibody (biotin). **c**, TCR expression levels on primary T-PLL tumor samples TRBC1<sup>+</sup> (blue, n=3 biologically independent samples) and TRBC2<sup>+</sup> (orange, n=3 biologically independent samples). TCR detected with antibody clone WT31. Data shown as mean  $\pm$  SD. Range of TCR<sup>+</sup> cells within samples was 79.8-96.3% and 95.6-97.9% for T-PLL TRBC1<sup>+</sup> and T-PLL TRBC2<sup>+</sup>, respectively. Source data are provided as a Source Data file.



#### Supplementary Fig. 5. Characterization of HuJovi-1 CAR

Cytotoxicity assay showing efficacy of MuJovi-1 CAR (orange) compared with HuJovi-1 CAR (blue) on WT and engineered Jurkat cell lines (n=8 biologically independent samples) (**a**), on H9 TRBC1 (n=6 biologically independent samples), and HD-MAR2 TRBC2 (n=4 biologically independent samples). Data shown as mean  $\pm$  SD. (**b**). Non-targeting aCD19 CAR (green) was used as a negative control CAR construct (1:4 E:T ratio, 72h). No killing was observed above background for TCR KO or TRBC2<sup>+</sup> cells. Efficacy of HuJovi-1 CAR was equivalent to the murine CAR. \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001. Two-way ANOVA with Dunnett's post

test. Data shown as mean  $\pm$  SD. Ns, not significant; NT, Non-transduced CAR T cells. **c**, CAR T cell expansion following 5 days incubation with Jurkat target cells at 1:4 E:T ratio (n=3 biologically independent samples). Data shown as mean  $\pm$  SD. Two-way ANOVA with Dunnett's post-test against aCD19 CAR cohort. \* p < 0.05. **d**, Cytotoxicity assay for MuJovi-1 and HuJovi-1 CARs against healthy T cells (TRBC1<sup>+</sup>) at 72h with 1:4, 1:1 and 4:1 E:T ratios (n=4). \*\*: p < 0.01, \*: p < 0.05. Two way ANOVA with Dunnett's post test. Data shown as mean  $\pm$  SD. Ns, not significant; NT, Non-transduced CAR T cells. **e**, CAR T cell count at 72h post incubation with healthy donor T cells (in **d**) at 4:1, 1:1 and 1:4 E:T ratios (n=4 biologically independent samples). No reduction in cell count for HuJovi-1 CAR compared to aCD19 CAR (i.e. no reverse killing). Data shown as mean  $\pm$  SD. \* p < 0.05, \*\* p < 0.01. Two-way ANOVA with Turkey's post test. Ns = not significant. Source data and exact p-values are provided as a Source Data file.

# Supplementary Fig. 6. In vivo characterization of HuJovi-1 CAR



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**a**, Schematic of NSG model with 5e6 Jurkat TRBC1<sup>+</sup> cells/animal, treated with 1e6 CAR T cells/mouse (n = 6/group, aCD19 CAR n = 5). **b**, Total radiance bioluminescent imaging (BLI) of treated mice.



**a**, Median total radiance BLI of Jurkat TRBC1<sup>+</sup> tumor burden post CAR-T cell injection (n = 6, aCD19 CAR n = 5) (left) (Data shown as median with 95% CI) and Kaplan-Meier survival curve (cut-off 1e9 p/s/cm<sup>2</sup>/sr) (right). Two-way ANOVA with Sidak's post test ns = non-significant. Log-rank test \* p < 0.05, \*\* p < 0.01. **b**, Individual mouse total flux radiance BLI of Jurkat TRBC1<sup>+</sup> tumor burden (in **a**) post CAR-T cell injection (n = 6, aCD19 CAR n = 5). Source data and exact p-values are provided as a Source Data file.

#### Supplementary Fig. 8: Affinity profile of HuJovi-1 derived clones



Affinity (KD = M) of *in silico*-derived HuJovi-1 mutants towards TRBC1 and TRBC2. Orange = KFN (Mut3); blue = HuJovi-1; grey = HuJovi-1 mutants with mutations on positions 26, 27, 28, 31, 32, 94, 96, 97 and 98 of the heavy chain (Kabat numbering). When kinetic could not be measured, value was fixed at 1e-4. Source data provided as a Source Data file.

Supplementary Fig. 9: Identification of HuJovi-1 variants with increased TRBC2 binding affinity using phage display.



**a**, Detailed selection strategy for TRBC2 enrichment from structural design phage library. For each panning strategy, the selection antigen is shown in green and the de-selection antigen is shown in red.  $\Delta$  indicates depletion. Round one input titers are shown while output titers are shown for round two. **b**, Representative phage ELISA output from two panning strategies. Left: 1425 was selected for TRBC2 with no pressure for TRBC1, while 1427 was selected under selection pressure from TRBC1 for both rounds one and two.

Supplementary Fig. 10: Molecular interface between KFN and TRBC1.



**a**, Close-up view of the interactions between the TRBC1 and the CDRs of the KFN Fab (PDB ID 7AMR). The TCR  $\beta$ -chain is colored in pink, the heavy chain is colored in light green. Residues Asn119 and Lys120, of the TCR  $\beta$ -chain, and Lys28, Phe32 and Asn96 (Kabat) are shown as stick representations. **b**, Electron density of the CDRs around the epitope of the TCR. The TCR  $\beta$ -chain is colored in pink while the heavy and light chains are colored in green and grey, respectively.



### Supplementary Fig. 11: PBMC TRBC1/TRBC2 staining.

**a**, Gating strategy for human PBMC CD3<sup>+</sup>. **b**, Dot plot of three independent stainings of healthy donor PBMCs with HuJovi-1 IgG (anti-human AF647) and biotinylated KFN IgG muFc (Streptavidin-PE). % of TRBC1<sup>+</sup> relative to TRBC2<sup>+</sup> populations in gated area depicted in graphs.

Supplementary Fig. 12: Kinetic binding profiles of Jovi-1 and KFN antibodies to TRBC1 and TRBC2.



**a**, Kinetic profiles of HuJovi-1 and KFN for TRBC1 and TRBC2 measured by SPR according to a 1:1 Langmuir model. **b**, Reverse kinetic profiles of chip immobilized TRBC1 and TRBC2 against huJovi-1 and KFN IgG, to determine avidity impact on the interaction (Bivalent analyte fit). Source data provided as a Source Data file.



Supplementary Fig. 13: Solubility and stability of TRBC1 and 2 antibodies.

Solubility (**a**) and stability (**b**) of HuJovi-1 and KFN antibodies in IgG format. **c**, Solubility of HuJovi-1 and KFN antibodies in scFv-Fc format. Summary of stability parameters in the two antibody formats. Data expressed as mean  $\pm$  SD. Source data provided as a Source Data file.

#### а Hinge-TyrpTM-41BBz CD8Stk-TyrpTM-CD28z CD28Stk-CD28TM-CD28z aCD19 CAR Non-transduced 02 02 0.44 RQR8 Anti-idiotype / aMurine IgG2a b \*\*\*\* г \*\* 10 Transduction efficiency % (RQR8) CAR/RQR8 MFI ratio 8 80 60 6 4 40 4-20 2 С TRBC1 CAR-T HuJovi-1-Hinge-TyrpTM-41BBz HuJovi-1-CD8Stk-TyrpTM-28z 600 • . HuJovi-1-CD28Stk-CD28TM-CD28z • . aCD19-CD8Stk-41BBz A T.ALL THECA HOMARATIRECA TCRYO d TNFα IL2 IFNγ 10000

## Supplementary Fig. 14. Functional characterization of HuJovi-1 CAR



a, Representative dot plot for HuJovi-1 CAR T cells stained with anti-CD34 (for RQR8 marker protein) and anti-idiotype (for CAR). b, % transduction efficiency based on RQR8<sup>+</sup> CAR T cell population by flow cytometry (left, n = 10 biologically independent samples) and surface CAR expression as CAR (anti-idiotype)/RQR8 (anti-CD34) ratio (right, n= 4 biologically independent samples). Data shown as mean ± SD. One-way ANOVA with Tukey's post test, \* p < 0.05, \*\* p < 0.01, \*\*\*\* p < 0.0001. c, FACS-based killing of HPB-ALL TRBC1 (n=9) biologically independent samples), HPB-ALL TRBC2 (n=9 biologically independent samples), HPB-ALL TCR KO (n=6 biologically independent samples), H9 TRBC1 (n=6 biologically independent samples), T-ALL1 TRBC2 (n=9 biologically independent samples) and HD-MAR2 TRBC2 (n=4 biologically independent samples) cell lines by HuJovi-1 CAR T cells at 1:8 E:T ratio, 72h. Data shown as mean  $\pm$  SD. \*p < 0.05, \*\* p <0.01, \*\*\* p>0.001 by two-way ANOVA and Dunnett's test for multiple comparisons versus aCD19 CAR. d, Cytometric bead array assay measurement for cytokine and cytolytic mediators by HuJovi-1 CAR-T cells against target cell lines. H9 values for Granzyme A, Granzyme B and Perforin were plotted separately due to high constitutive expression. Data presented as geometric mean. Source data and exact p-values are provided as a Source Data file.





a, Representative dot plot for KFN CAR-T cells stained with anti-CD34 (for RQR8 marker protein) and anti-idiotype (for CAR). b, % transduction efficiency based on RQR8<sup>+</sup> CAR-T cell population by flow cytometry (left, n = 11 biologically independent samples) and surface CAR expression as CAR (anti-idiotype)/RQR8 (anti-CD34) ratio (right, n = 4 biologically independent samples). Data shown as mean ± SD. One-way ANOVA with Tukey's post test, \*\*\* p < 0.001, \*\*\*\* p < 0.0001. c, Surface plasmon resonance (SPR) affinity kinetics of antiidiotype antibody for KFN antibody. **d**, Flow cytometry-based killing of HPB-ALL TRBC1 (n=9 biologically independent samples), HPB-ALL TRBC2 (n=9 biologically independent samples), HPB-ALL TCR KO (n=6 biologically independent samples), H9 TRBC1 (n=6 biologically independent samples), T-ALL1 TRBC2 (n=9 biologically independent samples) and HD-MAR2 TRBC2 (n=4 biologically independent samples) cell lines by HuJovi-1 CAR-T cells at 1:8 E:T ratio, 72h. Data shown as mean  $\pm$  SD. \*p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001 by two-way ANOVA and Dunnett's test for multiple comparisons versus aCD19 CAR. e, Cytometric bead array assay measurement for cytokine and cytolytic mediators by HuJovi-1 CAR-T cells against target cell lines. H9 values for Granzyme A, Granzyme B and Perforin were plotted separately due to high constitutive expression. Data presented as geometric mean. Source data and exact p-values are provided as a Source Data file.



Supplementary Fig. 16. Gating strategy for flow cytometry-based killing assay

Representative gating strategy for flow-cytometry-based killing assay against target cell lines and normal T cells (**a**) or primary tumor samples (**b**).



#### Supplementary Fig. 17. CAR T cell trogocytosis

**a**, Co-culture of CD19 CAR (green), KFN CAR (orange) and NT PBMC (purple) on TRBC1 effector cells (left) or CD19 CAR (green), HuJovi-1 CAR (Blue) and non-transduced (NT) PBMC (purple) on TRBC2 effector cells (right) with CellTrace Violet (CTV) labelled TRBC1<sup>+</sup>, TRBC2<sup>+</sup> PBMC or in the absence of target cells. Graphs show CTV uptake by effector cells following 30 minutes, 1h, 4h or 24h co-culture with target cells (1:1 E:T ratio, n = 3 biologically independent samples). Data shown as mean  $\pm$  SD. **b**, Co-culture of CD19 CAR (green), KFN CAR (orange), HuJovi-1 CAR (blue) or NT PBMC (purple) with TRBC1<sup>+</sup> (left) or TRBC2<sup>+</sup> (right) target cells. Graphs show TRBC1 (left) or TRBC2 (right) downregulation from TRBC1<sup>+</sup> or TRBC2<sup>+</sup> target cells following 30 minutes, 1h, 4h or 24h co-culture with HuJovi-1 CAR or KFN CAR, respectively (1:1 E:T ratio, n = 3 biologically independent samples). Data shown as mean  $\pm$  SD. Duta shown as mean  $\pm$  SD. Two-way ANOVA with Dunnett's post-test against NT effector cells, \* p < 0.05, \*\* p < 0.01. **c**, Co-culture of CD19 CAR (green), KFN CAR (orange), HuJovi-1 CAR (blue) or NT PBMC (purple) with TRBC1<sup>+</sup> (right) target cells. Graphs uptake by target cells following 30 minutes, 1h, 4h or 24h co-culture with effector cells, \* p < 0.05, \*\* p < 0.01. **c**, Co-culture of CD19 CAR (green), KFN CAR (orange), HuJovi-1 CAR (blue) or NT PBMC (purple) with TRBC1<sup>+</sup> (left) or TRBC2<sup>+</sup> (right) target cells. Graphs show transduction marker RQR8 uptake by target cells following 30 minutes, 1h, 4h or 24h co-culture with effector cells (1:1 E:T ratio, n = 3 biologically independent samples).

Data shown as mean  $\pm$  SD. Two-way ANOVA with Dunnett's post-test against NT effector cells, \* p < 0.05, \*\* p < 0.01. Source data and exact p-values are provided as a Source Data file.



Supplementary Fig. 18. In vitro CAR-T profiling

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**a**, Differentiation profile of CAR-T cells for sorted TRBC1<sup>+</sup> PBMC (KFN CAR and aCD19 CAR) and TRBC2<sup>+</sup> PBMC (HuJovi-1 CAR and aCD19 CAR), incubated for 6 days with 0, 1, 5 and 10% contaminating TRBC2<sup>+</sup> PBMC (for KFN CAR and aCD19 CAR) or TRBC1<sup>+</sup> PBMC (for HuJovi-1 CAR and aCD19 CAR). Data shown as mean  $\pm$  SD. Tcm = T central memory, Tem = T effector memory, Temra = terminally differentiated effector memory cells, Tn = naïve T cell. n = 3 biologically independent samples. **b**, Exhaustion profile of CAR T

cells for sorted TRBC1<sup>+</sup> PBMC (KFN CAR and aCD19 CAR) and TRBC2<sup>+</sup> PBMC (HuJovi-1 CAR and aCD19 CAR) based on the expression of 0, 1, 2 or 3 antigens between PD-1, TIM3 and CXCR5. Cells were incubated for 6 days with 0, 1, 5 or 10% contaminating TRBC2<sup>+</sup> PBMC (for KFN CAR and aCD19 CAR) or TRBC1<sup>+</sup> PBMC (for HuJovi-1 CAR and aCD19 CAR). n = 3 biologically independent samples. Data shown as mean  $\pm$  SD. Source data provided as a Source Data file.



# Supplementary Fig. 19. Bioluminescent imaging for NSG HPB-ALL in vivo model

Total radiance bioluminescent imaging (BLI) of NSG mice engrafted with HPB-ALL TRBC2 cell line and treated with KFN-CD28Stk-CD28TM-CD28z, HuJovi1-CD8Stk-TyrpTM-CD28z, aCD19 CAR T cells or non-transduced PBMC, at 5e6 CAR T cells/mouse.



### Supplementary Fig. 20. Analysis of NSG HPB-ALL in vivo model

Individual mouse total radiance BLI of HPB-ALL TRBC2<sup>+</sup> tumor burden post CAR-T cells (n = 6). Source data provided as a Source Data file.

# Supplementary Fig. 21. Bioluminescent imaging for NSG Jurkat TRBC1/TRBC2 *in vivo* model

		Jurkat I	RBC1		Jurkat TRBC2			
Days after CAR T cells injection	Non transduced	aCD19 CAR	HuJovi-1 CD8Stk- TyrpTM-CD28z	KFN CD28Stk- CD28TM-CD28z	Non transduced	aCD19 CAR	HuJovi-1 CD8Stk- TyrpTM-CD28z	KFN CD28Stk- CD28TM-CD28z
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66	- - - 1.0				- x10 <sup>6</sup> - 3.0			are dae
	- x10 <sup>6</sup> - - - 0.5				- 2.0			
	Radiance				Radiance			
	(p/sec/cm <sup>2</sup> /sr) Color scale Min = 8.30e5 Max = 6.80e6				Color scale Min = 8.30e5 Max = 6.80e6			

Jurkat TRBC1

Jurkat TRBC2

Total radiance BLI of NSG mice engrafted with Jurkat TRBC1 (left) or Jurkat TRBC2 (right) cell lines and treated with KFN-CD28Stk-CD28TM-CD28z, HuJovi1-CD8stk-TyrpTM-CD28z, aCD19 CAR-T cells or non-transduced PBMC, at 5e6 CAR T cells/mouse.

NSG - Jurkat TRBC1

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**a**, Individual mouse total radiance BLI of Jurkat TRBC1<sup>+</sup> tumor burden post CAR-T cells (n = 6). # xeno-GvHD event. **b**, Individual mouse total radiance BLI of Jurkat TRBC2<sup>+</sup> tumor burden post CAR-T cells (n = 6). Source data are provided as a Source Data file.

# Supplementary Tables

Clone	Molecule	Chain	Sequence
	Protein	Heavy chain	EVRLQQSGPDLIKPGASVKMSCKASGYTFTGYVMHWVKQRPGQGLEWIGF INPYNDDIQSNERFRGKATLTSDKSSTTAYMELSSLTSEDSAVYYCARGAG YNFDGAYRFFDFWGQGTTLTVSSASTKGPSVFPLAPSSKSTSGGTAALGCL VKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQT YICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKD TLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNST YRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVY TLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
MuJovi-1		Light chain	DVVMTQSPLSLPVSLGDQASISCRSSQRLVHSNGNTYLHWYLQKPGQSPKL LIYRVSNRFPGVPDRFSGSGSGTDFTLKISRVEAEDLGIYFCSQSTHVPYTFG GGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKV DNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGL SSPVTKSFNRGEC
	DNA	Heavy chain	GAGGTGCGGCTGCAGCAGAGCGGCCCTGACCTGATCAAGCCCGGCGCC AGCGTGAAGATGAGCTGCAAGGCCAGCGGCTACACCTTCACCGGCTAC GTGATGCACTGGGTGAAGCAGCGGCCTGGCCAGGGCCTGGAGTGGATC GGCTTCATCAACCCTTACAACGACGACATCCAGAGCAACGAGCGGTTCC GCGGCAAGGCCACCCTGACCAGCGACAAGAGCAGCACCACCGCCTACA TGGAGCTGAGCAGCCTGACCAGCGAGGACAAGAGCAGCACCACCGCCTACA CCCGCGGGAGCCGGCTACAACTTCGACGGCGCCTACCGGTTCTTCGACTT CTGGGGCCAGGGCACCACCCTGGCACCGTGAGCTCAgcgTCGACCAAGGGC CCATCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGCA CAGCGGCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCTGTGAC GGTCTCGTGGAACTCAGGCGCCTGACCAGCGGCGTGCACACCTTCCGG GCTGTCCTACAGTCCTCAGGACCTCAGCGGCGTGCACACCTTCCG GCTGTCCTACAGTCCTCAGGACCTACTTCCCCTCAGCAGCGTGGTGACCG TGCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCA CAAGCCCAGCAACACCAAGGTGGACAAGAAGTTGAGCCCAAATCTTG

# Supplementary Table 1: Antibody sequences.

			TGACAAAACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCTGGGG
			GGACCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGA
			TCTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGA
			AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCAT
			AATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGT
			GTGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGG
			AGTACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAA
			AACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACAC
			CCTGCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACC
			TGCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGA
			GCAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGG
			ACTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAG
			CAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCT
			CTGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
			GACGTGGTGATGACCCAGAGCCCACTGAGCCTGCCCGTGAGCCTGGGCG
			ACCAGGCCAGCATCAGCTGCCGGAGCAGCCAGCGGCTGGTGCACAGCA
		Light chain	ACGGCAACACCTACCTGCACTGGTACCTGCAGAAGCCCGGCCAGAGCCC
			TAAGCTGCTGATCTACCGGGTGAGCAACCGGTTCCCTGGCGTGCCCGAC
			CGGTTCAGCGGCAGCGGCAGCGGCACCGACTTCACCCTGAAGATCAGCC
			GGGTGGAGGCCGAGGACCTGGGCATCTACTTCTGCAGCCAGAGCACCC
			ACGTGCCCTACACCTTCGGAGGCGGCACCAAGCTGGAGATCAAGCGGA
			CGGTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAGTTG
			AAATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAG
			AGAGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAA
			CTCCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAG
			CCTCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAA
			AGTCTACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACA
			AAGAGCTTCAACAGGGGAGAGTGT
			QVQLVQSGAEVKKPGASVKVSCKASGYTFTGYVMHWVRQAPGOGLEWM
			GFINPYNDDIQSNERFRGRVTMTRDTSISTAYMELSRLRSDDTAVYYCARG
			AGYNFDGAYRFFDFWGQGTMVTVSSASTKGPSVFPLAPSSKSTSGGTAALG
HuJovi-1	Protein	Heavy chain	CLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
			QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKP
			KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYN
			STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQV

			YTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDS
			DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
			DIVMTQSPLSLPVTPGEPASISCRSSQRLVHSNGNTYLHWYLQKPGQSPRLLI YRVSNRFPGVPDRFSGSGSGSTDFTLKISRVEAEDVGVYYCSQSTHVPYTFG
		Light chain	OCTVI EIKRTVA APSVEIEPPSDEOI KSGTASVVCI I NNEYPREAKVOWKV
		Light cham	VUINLEINNI VAAI SVIIITI SDEVENSTASVI CELANI TI NEAN VUIN
			DNALQSGNSQESVIEQDSKDSIYSLSSILILSKADIEKHKVIACEVIHQGL
			SSPVTKSFNRGEC
			CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAAACCAGGCGCC
			AGCGTGAAGGTGTCCTGCAAGGCCAGCGGCTACACCTTTACCGGCTACG
			TGATGCACTGGGTGCGCCAGGCTCCAGGCCAGGGACTGGAATGGATGG
			GCTTCATCAACCCCTACAACGACGACATCCAGAGCAACGAGCGGTTCCG
			GGGCAGAGTGACCATGACCAGAGACACCAGCATCAGCACCGCCTACAT
			GGAACTGAGCCGGCTGAGAAGCGACGACACCGCCGTGTACTACTGCGC
			CAGAGGCGCCGGATACAACTTCGACGGCGCCTACAGATTCTTCGACTTC
			TGGGGCCAGGGCACAATGGTCACCGTGTCCTCTGCGTCGACCAAGGGCC
I			CATCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGGCAC
			AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCTGTGACG
l			GTCTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGG
l			CTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGT
			GCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCAC
		TT hain	AAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT
	DNA	Heavy chain	GACAAAACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGG
			GACCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGAT
			CTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAA
			GACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATA
			ATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTG
I			TGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGA
I			GTACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAA
I			ACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACC
			CTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCT
			GCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAG
			CAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGA
			CTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAGC
			AGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTC
			TGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA

		Light chain	GACATCGTGATGACCCAGAGCCCCCTGAGCCTGCCTGTGACACCTGGCG AACCTGCCAGCATCAGCTGCCGGTCTAGCCAGAGACTGGTGCACAGCAA CGGCAACACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCCCCC AGACTGCTGATCTACCGGGTGTCCAACAGATTCCCCGGCGTGCCCGATA GATTCAGCGGCTCTGGCAGCGGCACCGACTTCACCCTGAAGATCTCCCG GGTGGAAGCCGAGGACGTGGGCGTGTACTACTGCAGCCAGAGCACCCA CGTGCCCTACACCTTTGGCCAGGGCACCAAGCTGGAAATCAAGCGTACG GTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAGTTGAA ATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAG AGGCCAAAGTACAGTGGAAGGTGGATAACGCCTCCAATCGGGTAACT CCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCC TCAGCAGCACCCTGACGCTGAGCAAGCAAGCAAGCAAGCA
	Protein	Heavy chain	QVQLVQSGAEVKKPGASVKVSCKASGYKFTGFVMHWVRQAPGQGLEWM GFINPYNDDIQSNERFRGRVTMTRDTSISTAYMELSRLRSDDTAVYYCARG NGYNFDGAYRFFDFWGQGTMVTVSSASTKGPSVFPLAPSSKSTSGGTAALG CLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKP KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYN STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQV YTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDS DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
KFN (Mut3)		Light chain	DIVMTQSPLSLPVTPGEPASISCRSSQRLVHSNGNTYLHWYLQKPGQSPRLLI YRVSNRFPGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCSQSTHVPYTFG QGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKV DNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGL SSPVTKSFNRGEC
	DNA	Heavy chain	CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAAACCAGGCGCC AGCGTGAAGGTGTCCTGCAAGGCCAGCGGCTACaagTTTACCGGCtttGTG ATGCACTGGGTGCGCCAGGCTCCAGGCCAGGGACTGGAATGGATGG

			TCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGGCACAG
			CGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCTGTGACGGT
			CTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGGCT
			GTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGTGC
			CCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCACAA
			GCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGTGA
			CAAAACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGGGGA
			CCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGATCTC
			CCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAAGA
			CCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATAAT
			GCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTGTG
			GTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGAGT
			ACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCATCGAGAAAA
			CCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACCC
			TGCCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCTG
			CCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGC
			AATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGAC
			TCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAGCA
			GGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCT
			GCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
			GACATCGTGATGACCCAGAGCCCCCTGAGCCTGCCTGTGACACCTGGCG
			AACCTGCCAGCATCAGCTGCCGGTCTAGCCAGAGACTGGTGCACAGCAA
			CGGCAACACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCCCCC
			AGACTGCTGATCTACCGGGTGTCCAACAGATTCCCCGGCGTGCCCGATA
			GATTCAGCGGCTCTGGCAGCGGCACCGACTTCACCCTGAAGATCTCCCG
			GGTGGAAGCCGAGGACGTGGGCGTGTACTACTGCAGCCAGAGCACCCA
			CGTGCCCTACACCTTTGGCCAGGGCACCAAGCTGGAAATCAAGCGTACG
		Light chain	GTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAGTTGAA
			ATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAG
			AGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAACT
			CCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCC
			TCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAG
			TCTACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAA
			GAGCTTCAACAGGGGAGAGTGT
			QVQLVQSGAEVKKPGASVKVSCKASGYRFTGFVMHWVROAPGOGLEWM
RFN (Mut4)	Protein	Heavy chain	GFINPYNDDIQSNERFRGRVTMTRDTSISTAYMELSRLRSDDTAVYYCARG
			NGYNFDGAYRFFDFWGQGTMVTVSSASTKGPSVFPLAPSSKSTSGGTAALG

			CLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGT
			QTYICNVNHKPSNTKVDKKVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKP
			KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYN
			STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQV
			YTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDS
			DGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK
			DIVMTQSPLSLPVTPGEPASISCRSSQRLVHSNGNTYLHWYLQKPGQSPRLLI
			YRVSNRFPGVPDRFSGSGSGTDFTLKISRVEAEDVGVYYCSQSTHVPYTFG
		Light chain	QGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKV
			${\tt DNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGL}$
			SSPVTKSFNRGEC
			CAGGTGCAGCTGGTGCAGTCTGGCGCCGAAGTGAAGAAACCAGGCGCC
			AGCGTGAAGGTGTCCTGCAAGGCCAGCGGCTACCGCTTTACCGGCTTTG
			TGATGCACTGGGTGCGCCAGGCTCCAGGCCAGGGACTGGAATGGATGG
			GCTTCATCAACCCCTACAACGACGACATCCAGAGCAACGAGCGGTTCCG
		Heavy chain	GGGCAGAGTGACCATGACCAGAGACACCAGCATCAGCACCGCCTACAT
			GGAACTGAGCCGGCTGAGAAGCGACGACACCGCCGTGTACTACTGCGC
			CAGAGGCAACGGATACAACTTCGACGGCGCCTACAGATTCTTCGACTTC
			TGGGGCCAGGGCACAATGGTCACCGTGTCCTCCGCGTCGACCAAGGGCC
			CATCGGTCTTCCCCCTGGCACCCTCCTCCAAGAGCACCTCTGGGGGGCAC
			AGCGGCCCTGGGCTGCCTGGTCAAGGACTACTTCCCCGAACCTGTGACG
			GTCTCGTGGAACTCAGGCGCCCTGACCAGCGGCGTGCACACCTTCCCGG
			CTGTCCTACAGTCCTCAGGACTCTACTCCCTCAGCAGCGTGGTGACCGT
	DNA		GCCCTCCAGCAGCTTGGGCACCCAGACCTACATCTGCAACGTGAATCAC
			AAGCCCAGCAACACCAAGGTGGACAAGAAAGTTGAGCCCAAATCTTGT
			GACAAAACTCACACATGCCCACCGTGCCCAGCACCTGAACTCCTGGGGG
			GACCGTCAGTCTTCCTCTTCCCCCCAAAACCCAAGGACACCCTCATGAT
			CTCCCGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTGAGCCACGAA
			GACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGGAGGTGCATA
			ATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGCACGTACCGTG
			TGGTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTGAATGGCAAGGA
			GTACAAGTGCAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAA
			ACCATCTCCAAAGCCAAAGGGCAGCCCCGAGAACCACAGGTGTACACC
			CTGCCCCATCCCGGGATGAGCTGACCAAGAACCAGGTCAGCCTGACCT
			GCCTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAG
			CAATGGGCAGCCGGAGAACAACTACAAGACCACGCCTCCCGTGCTGGA
			CTCCGACGGCTCCTTCTTCCTCTACAGCAAGCTCACCGTGGACAAGAGC

		AGGTGGCAGCAGGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTC
		TGCACAACCACTACACGCAGAAGAGCCTCTCCCTGTCTCCGGGTAAA
		GACATCGTGATGACCCAGAGCCCCCTGAGCCTGCCTGTGACACCTGGCG
		AACCTGCCAGCATCAGCTGCCGGTCTAGCCAGAGACTGGTGCACAGCAA
		CGGCAACACCTACCTGCACTGGTATCTGCAGAAGCCCGGCCAGTCCCCC
		AGACTGCTGATCTACCGGGTGTCCAACAGATTCCCCGGCGTGCCCGATA
		GATTCAGCGGCTCTGGCAGCGGCACCGACTTCACCCTGAAGATCTCCCG
	Light chain	GGTGGAAGCCGAGGACGTGGGCGTGTACTACTGCAGCCAGAGCACCCA
		CGTGCCCTACACCTTTGGCCAGGGCACCAAGCTGGAAATCAAGCGTACG
		GTGGCTGCACCATCTGTCTTCATCTTCCCGCCATCTGATGAGCAGTTGAA
		ATCTGGAACTGCCTCTGTTGTGTGCCTGCTGAATAACTTCTATCCCAGAG
		AGGCCAAAGTACAGTGGAAGGTGGATAACGCCCTCCAATCGGGTAACT
		CCCAGGAGAGTGTCACAGAGCAGGACAGCAAGGACAGCACCTACAGCC
		TCAGCAGCACCCTGACGCTGAGCAAAGCAGACTACGAGAAACACAAAG
		TCTACGCCTGCGAAGTCACCCATCAGGGCCTGAGCTCGCCCGTCACAAA
		GAGCTTCAACAGGGGAGAGTGT

# Supplementary Table 2: X-ray data collection and refinement statistics.

	HuJovi- 1/TRBC1 (7AMP)	HuJovi- 1/TRBC2 (7AMQ)	KFN/TRBC2 (7AMS)	KFN/TRBC1 (7AMR)
Resolution (Å)	54.44–2.64 (2.69–2.64)	47.8–2.35 (2.43–2.35)	43.4–2.42 (2.46–2.42)	45.7–1.95 (2.13–1.95)‡
Wavelength (Å)	0.9282	0.9795	0.9795	0.9762
Space group	C2	C2	C2	C2
Unit cell (Å)	a=117.4 b=91.1 c=125.3 β=93.8	a = 114.5 b = 91.8 c = 123.4 β=94.6	a = 114.8 b = 91.2 c = 123.3 $\beta = 94.1$	a = 115.7 b = 91.0 c = 123.3 $\beta$ =93.9
Completeness (%)	99.9 (99.8)	99.6 (96.8)	99.9 (97.8)	ellipsoidal: 93.8 (87.8) spherical: 53.7 (11.6)
Redundancy	6.1 (6.3)	5.1 (4.3)	5.0 (5.1)	5.7 (4.9)
No. of observations/unique reflections	237,232 (38,805)	267,467 (52,746)	242,685 (48,522)	287,266 (49,996)
<i σ(i)=""></i>	9.9 (0.93)	8.3 (1.0)	7.0 (1.1)	10.4 (1.5)
CC(1/2) (%)	99.8 (62.5)	99.6 (71.2)	99.0 (71.3)	99.6 (52.4)
R <sub>merge</sub> (I) (%)	11.8 (166.9)	13.2 (122.8)	17.5 (165.9)	12.5 (101.8)
$R_{model}(F) R_{free}(F) (\%)$	23.9/28.9 (47.4)/(46.9)	21.6/25.6 (34.9)/(35.2)	19.9/24.6 (31.4)/(44.3)	17.9/22.5 (28.6)/(21.7)
No. of non-hydrogen atoms	6,858	7,287	7,382	7,539
No. of water molecules	20	502	557	683
Rms deviations from ideal geometry: bond lengths (Å)	0.007	0.007	0.007	0.10
Bond angles (degrees)	1.16	1.00	0.98	1.19
Mean B-factor all atoms (Å <sup>2</sup> )*	85.1	56.3	47.6	32.7
Mean B-factor protein chains A,B,H,L (Å <sup>2</sup> )*	81.8, 85.6, 87.7, 85.1	60.3, 57.0, 54.7, 51.6	53.7, 49.9, 45.4, 43.0	37.5, 34.2, 30.2, 27.6
Mean B-factor (solvent $\text{\AA}^2$ )*	54.6	57.8	47.6	37.4
Mean B-factor others (Å <sup>2</sup> )	79.6	66.4	56.6	41.4
Ramachandran plot quality <sup>#</sup>				
Favored regions (%)	94.4	98.0	97.1	97.4
Allowed regions (%)	5.3	1.8	2.7	2.3
Outliers (%)	0.3	0.2	0.2	0.3

Figures in parentheses are for the highest resolution shell. Other relevant quality indicators can be easily extracted from the PDB file header. \*Calculated with Moleman<sup>1</sup>; #calculated using a local Molprobity server<sup>2</sup>; ‡processed anisotropically with autoPROC/STARANISO. The anisotropic resolution limits were 1.85 Å, 2.3 Å, and 3.0 Å in the three principal directions of the anisotropic ellipsoid. When processed isotropically, the data gave a resolution of 2.56 Å with 100% completeness.

Residue	Original	Mutated	δ Affinity TRBC2	δ Affinity TRBC1	Net <b>ð</b> Affinity
H:28	THR	LYS	-13.32	-3.35	-9.97
H:28	THR	ARG	-12.41	-2.3	-10.11
H:96	ALA	ASN	-11.73	-0.62	-11.11
H:99	ASN	ARG	-11.21	-2.41	-8.8
H:35	HIE	TRP	-11.2	-3.42	-7.78
H:28	THR	HIP	-10.93	-15.51	4.58
H:99	ASN	TYR	-7.98	0.09	-8.07
H:100F	PHE	ARG	-7.93	-19.61	11.68
H:100A	ASP	ASN	-6.61	-2.24	-4.37
H:98	TYR	ARG	-6.27	1.33	-7.6
H:60	ASN	HIP	-5.77	-1.1	-4.67
H:100F	PHE	LYS	-5.08	-7.18	2.1
H:58	GLN	LYS	-4.98	0.39	-5.37
H:96	ALA	GLN	-4.95	-2.42	-2.53
H:31	GLY	MET	-4.94	1.71	-6.65
H:100F	PHE	HIP	-4.89	-8.78	3.89
H:99	ASN	GLN	-4.87	0.23	-5.1
H:100A	ASP	LEU	-4.65	-1.32	-3.33
H:31	GLY	LYS	-4.64	-1.4	-3.24
H:100G	PHE	ARG	-4.53	-1.56	-2.97
H:28	THR	MET	-4.31	4.25	-8.56
H:101	ASP	MET	-4.18	-6.36	2.18
H:28	THR	ILE	-4.15	3.96	-8.11
H:95	GLY	HIP	-4.11	-1.54	-2.57
H:60	ASN	TRP	-4.1	-1.1	-3
H:98	TYR	HIP	-4.07	-5.07	1
H:100B	GLY	PHE	-4.04	-3.27	-0.77
H:100B	GLY	ARG	-4.02	-2.18	-1.84
H:99	ASN	MET	-3.77	-0.1	-3.67
H:31	GLY	HID	-3.54	0.97	-4.51
H:52	ASN	PHE	-3.49	-0.39	-3.1
H:96	ALA	THR	-3.42	-1.81	-1.61
H:28	THR	TRP	-3.36	-1.02	-2.34
H:101	ASP	ILE	-3.3	-4.42	1.12
H:28	THR	TYR	-3.23	-0.33	-2.9
H:100A	ASP	THR	-3.2	-2.86	-0.34
H:95	GLY	ARG	-3.08	12.27	-15.35
H:96	ALA	HID	-3.05	-3.33	0.28

Supplementary Table 3: In silico mutagenesis of HuJovi-1 binding to TRBC2.

*In silico* identification of Hu-Jovi-1 variants with highest affinity increase to TRBC2. Three color grading: Red = high  $\delta$  affinity to TRBC2 or low  $\delta$  affinity to TRBC1. Green = high  $\delta$ 

affinity to TRBC1 or low  $\delta$  affinity to TRBC2. Net  $\delta$  affinity = difference between  $\delta$  affinity to TRBC2 and  $\delta$  affinity to TRBC1. Kabat numbering.

# **Supplementary References**

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