

Supplementary Material

3D Variability Analysis Reveals a Hidden Conformational Change Controlling Ammonia Transport in Human Asparagine Synthetase

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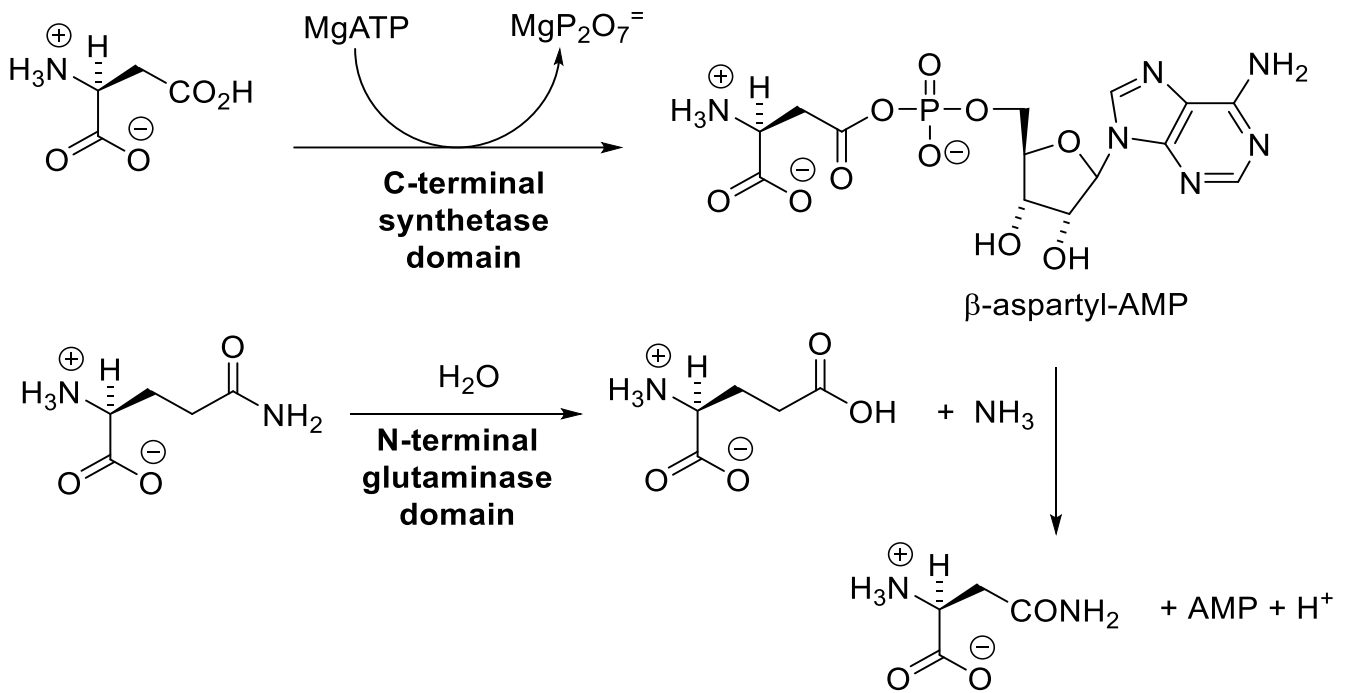
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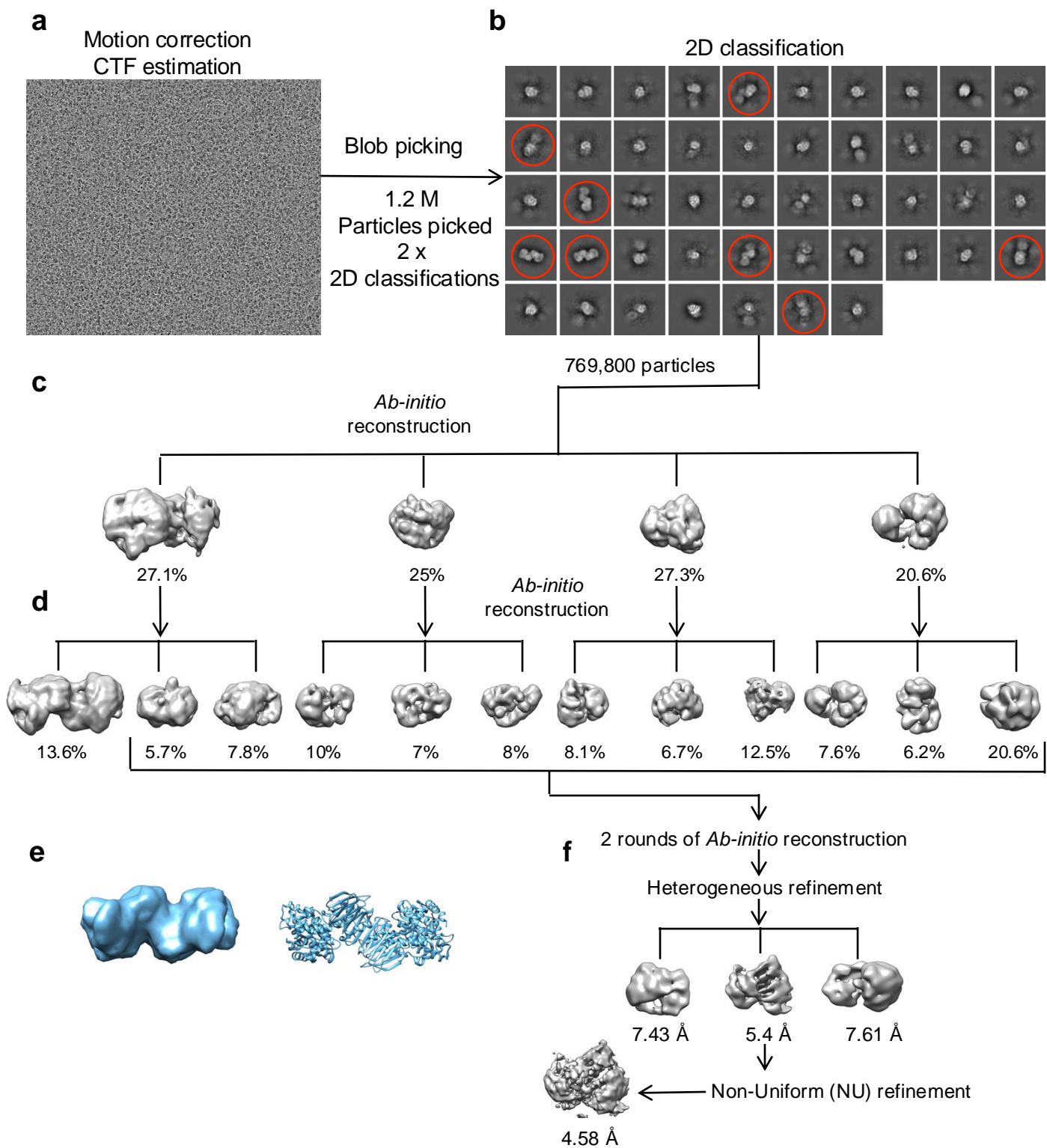
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Supplementary Figure 1. Reactions catalyzed by the active sites in the N- and C-terminal domains of asparagine synthetase. Ammonia released in the glutaminase site is channeled to the synthetase site through an intramolecular tunnel linking the two active sites.



Supplementary Figure 2. Unbiased initial image analysis to assess the mode of ASNS dimerization.

a Representative motion corrected micrograph.

b 2D class average: 2D projections resembling to the head-to-head dimer are indicated (red circle).

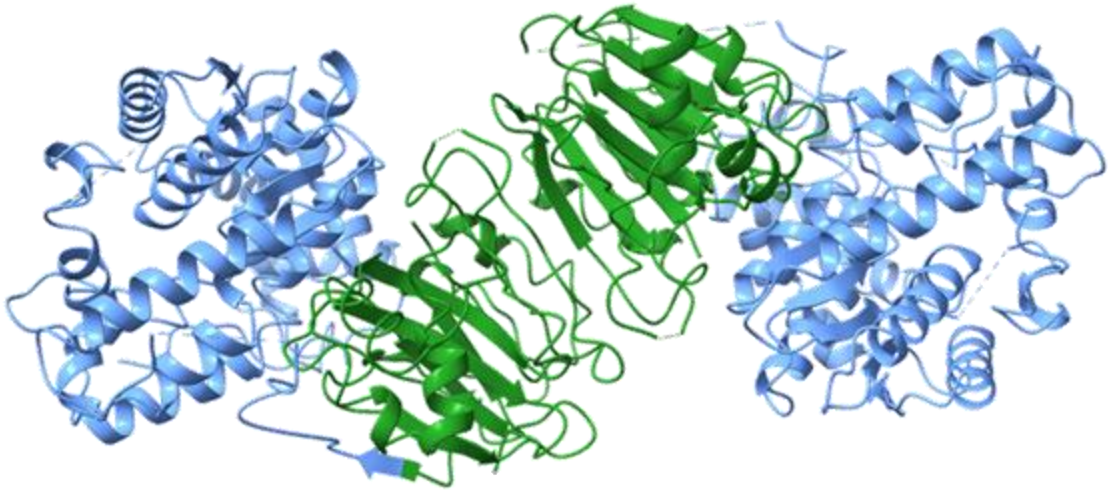
c The first round of *Ab-initio* reconstruction, yielding 4 different reconstructions.

d 2nd round of *Ab-initio* reconstruction, yielding a total of 12 different reconstructions.

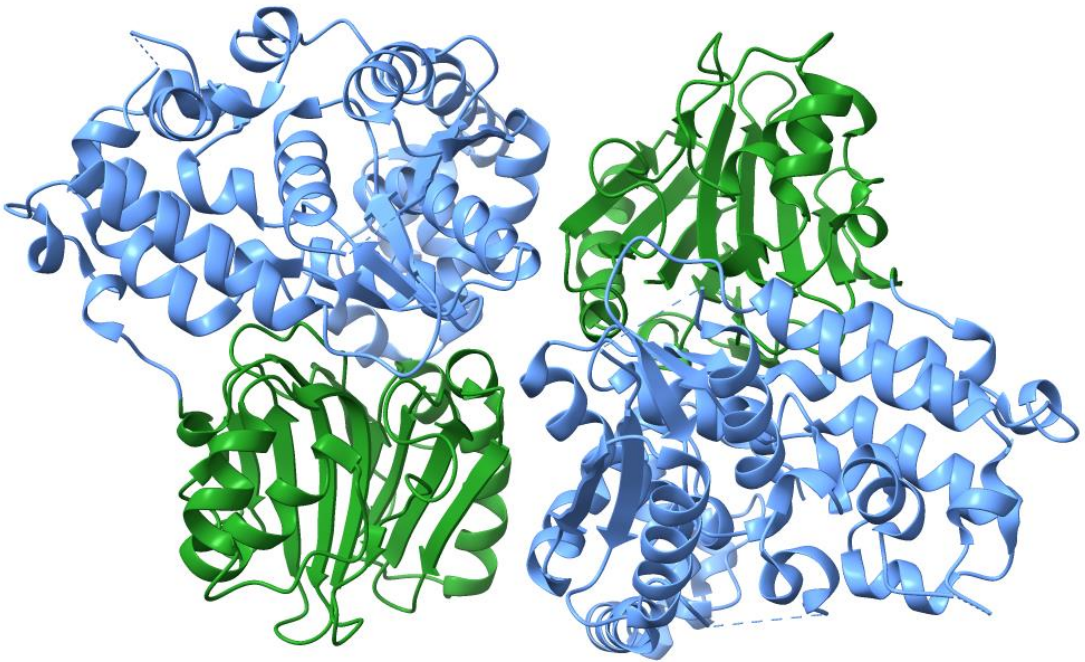
e Stimulated EM density at 12 Å (on the left) derived from the crystal structure (PDB: 6GQ3) on the right.

f Heterogeneous refinement resulting in 3 reconstructions followed by NU refinement.

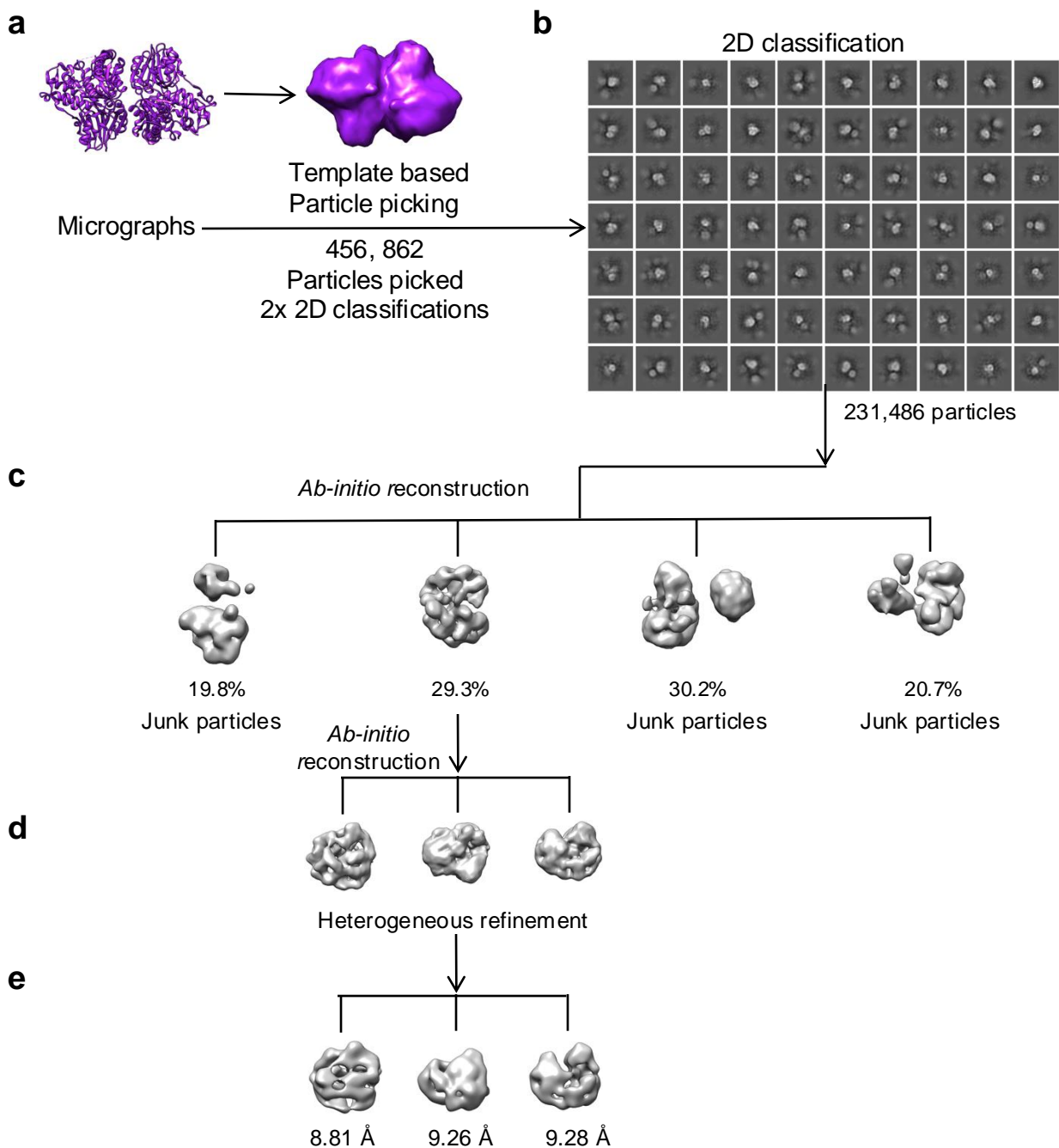
a



b



Supplementary Figure 3. Cartoon representations of the human and bacterial ASNS dimer. a Ribbon diagram of the "head-to-head" dimer formed by human ASNS. **b.** Ribbon diagram of the "head-to-tail" dimer observed in the X-ray crystal structure of *Escherichia coli* AS-B. The N- and C-terminal domains of both structures are colored green and blue, respectively.



Supplementary Figure 4. Image analysis using AS-B as a template to assess the mode of ASNS dimerization.

a Stimulated EM density at 12 Å (on the right) derived from the crystal structure (PDB: 1CT9) on the left.

b Selected 2D class averages of particles picked by templated-based approach.

c *Ab-initio* reconstruction, yielding 4 different reconstructions, 3 of which were considered to be junk.

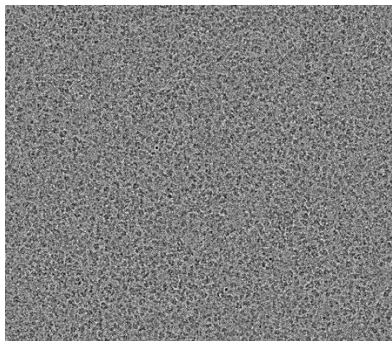
d 2nd round of *Ab-initio* reconstruction, yielding a total of 3 reconstructions.

e Heterogeneous refinement resulting in a total of 3 3D reconstructions.

Supplementary Table 1. Cryo-EM data collection, refinement and validation statistics

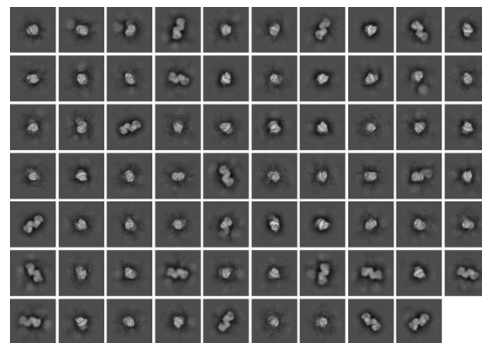
Data collection and processing		
	WT apo-ASNS (EMDB-40764) (PDB 8SUE)	R142I ASNS Variant (EMDB-44253) (PDB 9B6C)
Magnification	X 150,000	X 150,000
Voltage (kV)	200	200
Electron exposure (e/Å ²)	40	40
Defocus range (µm)	-0.8 to -2.0	-0.8 to -2.0
Pixel size (Å)	0.93	0.93
Symmetry imposed	C2	C2
Initial particle images (no.)	2,464,389	3,123,932
Final particle images (no.)	47,929	145,711
Map resolution (Å)	3.53	3.35
FSC threshold	0.143	0.143
Map resolution range	2.8-4.0	2.5-3.5
Refinement		
Initial model used	6GQ3	8SUE
Model resolution (Å)	2.3	2.2
FSC threshold	0.143	0.143
Map resolution range (Å)	2.8-4.0	2.5-4.0
Map sharpening B factor (Å ²)	119.8	118.5
Model composition		
Non-hydrogen atoms	8049	8124
Protein residues	1019	1031
Ligand (Mg)	0	0
<i>B</i> -factors (min/max/mean)		
Protein	36.67/74.22/51.65	10.29/103.04/36.04
Ligand	---	---
R.m.s. deviations		
Bond length (Å)	0.003	0.004
Bond angles (°)	0.638	0.571
Validation		
MolProbability score	2.05	1.91
Clashscore	12.53	7.78
Poor Rotamers (%)	0.36	1.65
Ramachandran plot		
Disallowed (%)	0	0
Allowed (%)	6.83	4.66
Favored (%)	93.17	95.34

Motion correction
CTF estimation



Template based
Particle picking
2, 464,387
Particles picked
3 x 2D classifications

2D classification

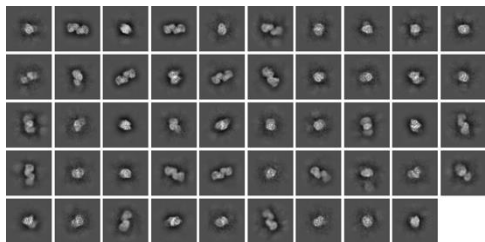


201, 704 particles

Generate template based on
Ab-initio reconstruction
Template based particle picking

860,742 particles

2D classification



423,294 particles

Combined particles
Remove duplicate

2D classification

278,210 particles

2 rounds of *Ab-initio* reconstructions
to remove small particles

102,546 particles

3rd *Ab-initio*
reconstruction

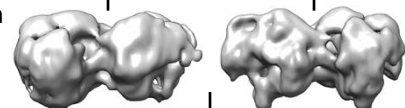


59.6%

40.4%

77,956 particles

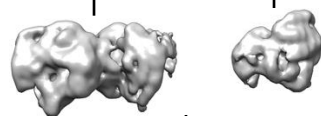
4th *Ab-initio*
reconstruction



39.5%

60.5%

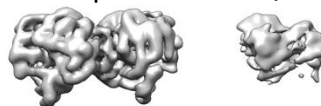
Ab-initio
reconstruction



59.6%

40.4%

Heterogeneous
refinement



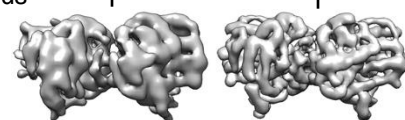
5.75 Å

6.09 Å

186,770 particles

100,440 particles

Heterogeneous
refinement



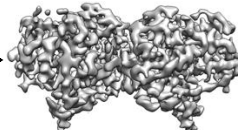
6.48 Å

4.78 Å

30,027 particles

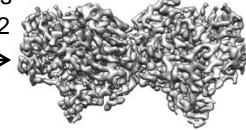
47,929 particles

Non-uniform
refinement C1
Beam tilt
correction



4.17 Å

Non-uniform
refinement
CTF & defocus
corrections C2

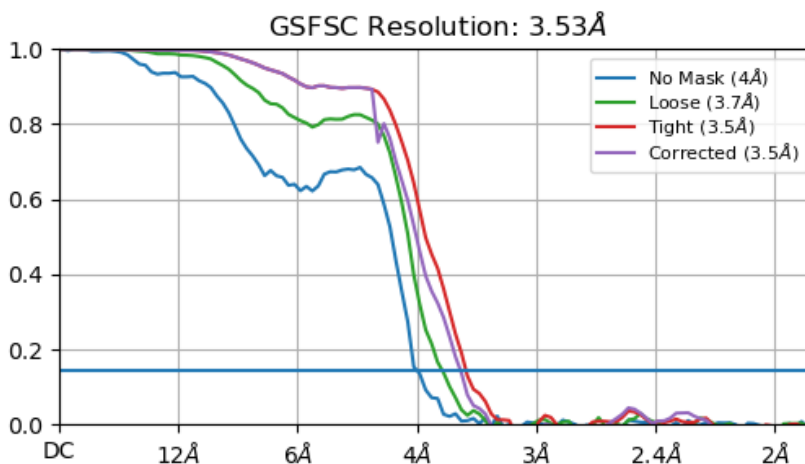


3.53 Å

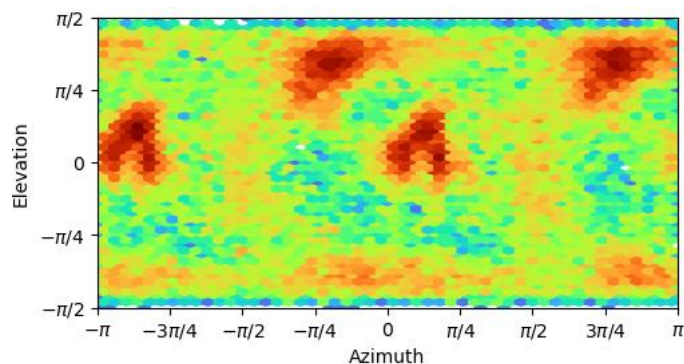
47,929 particles

Supplementary Figure 5. Summary of the cryo-EM data processing workflow using cryoSPARC v3.2.2.

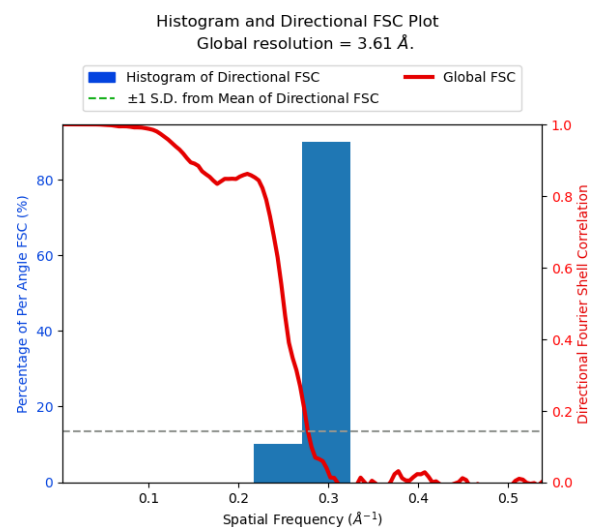
a Gold standard FSC curve for WT human ASNS



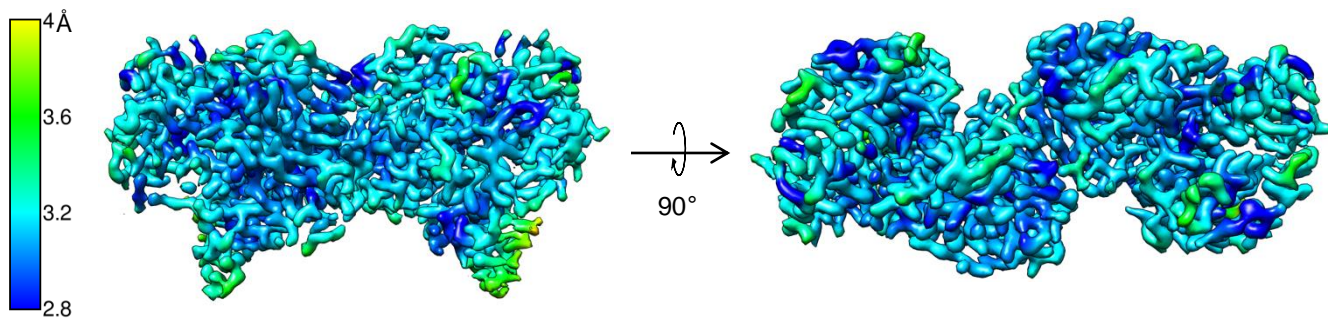
b Viewing distribution plots



c 3D FSC



d Local resolution



Supplementary Figure 6. Assessment of the quality of cryo-EM map.

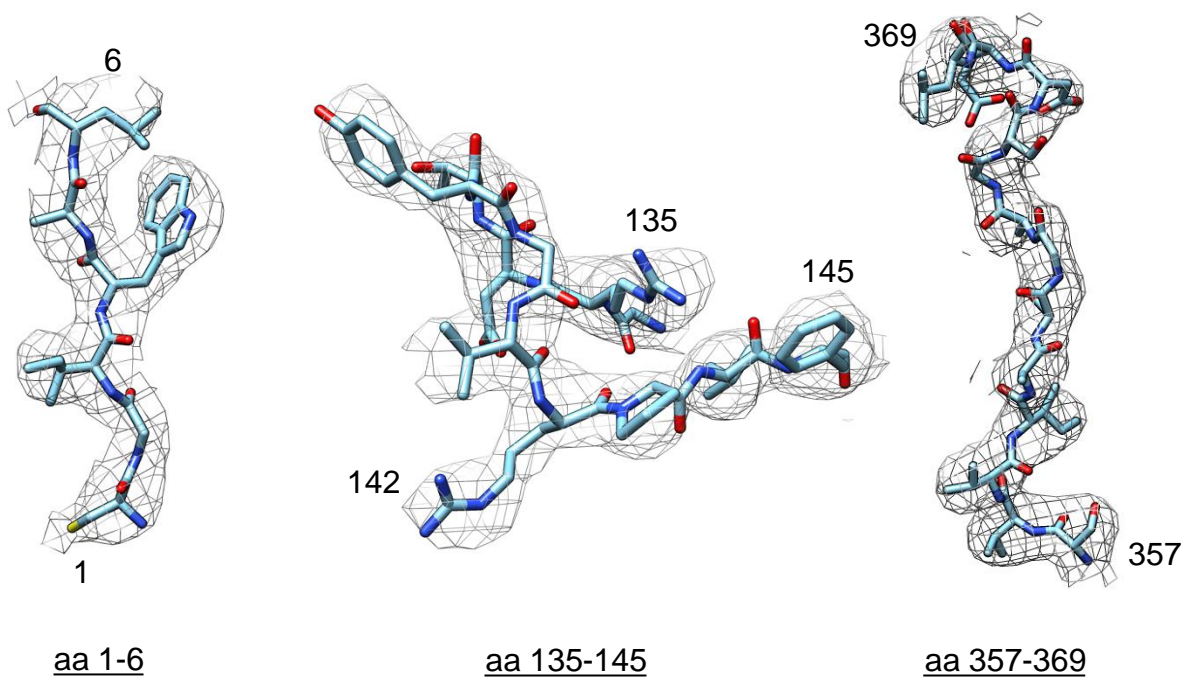
a Gold standard FSC curve for the map of hASNS generated in cryoSPARC v3.2.

b Viewing distribution plots for the map generated in cryoSPARC v3.2.

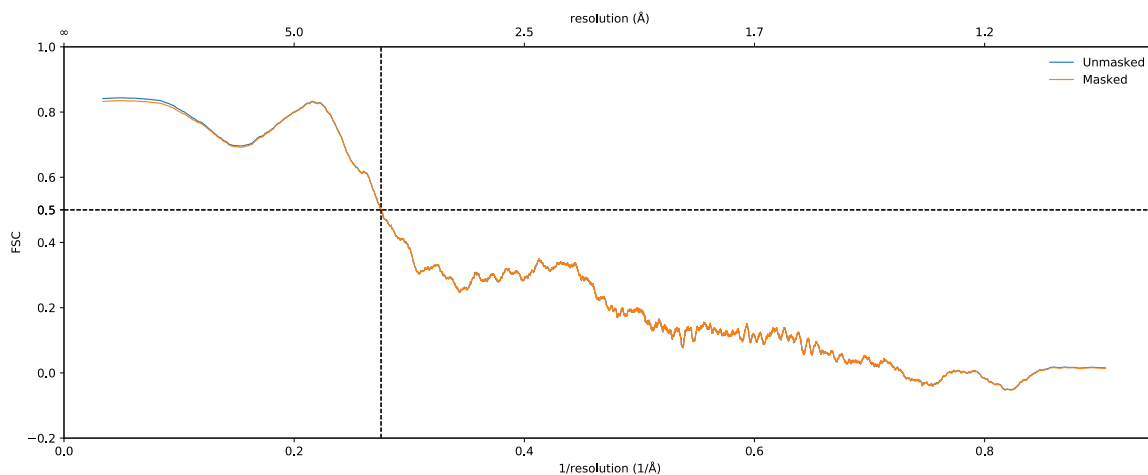
c 3D FSC: histogram and directional FSC plot for map global resolution are shown.

d Local resolution of the map (color scale shown on the left).

a Representative densities and corresponding models for WT human ASNS



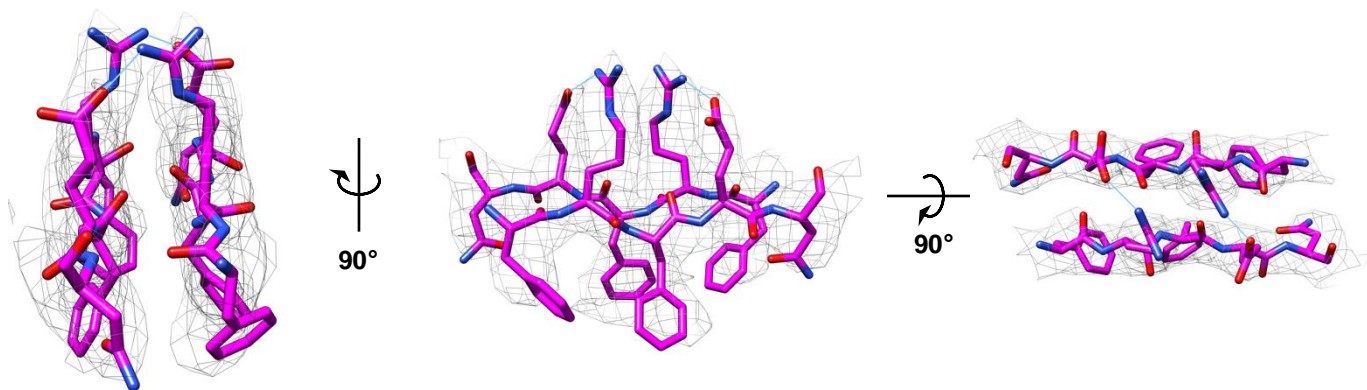
b Map-model FSC curve



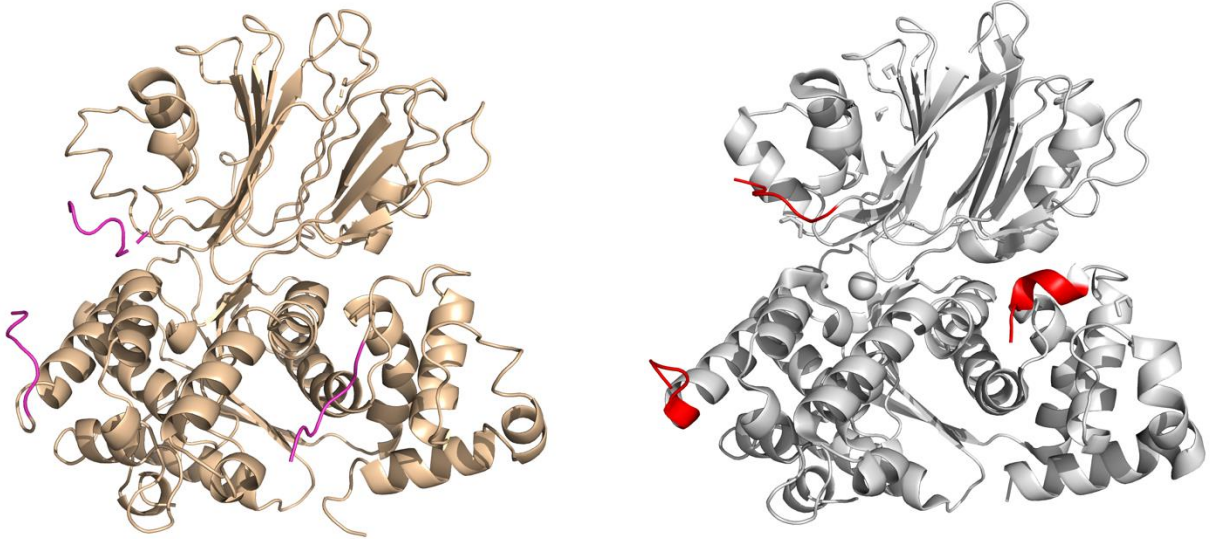
Supplementary Figure 7. Validating the cryo-EM structure: map and its model.

a Representative density of the map for residues in the N-terminal catalytic region (aa 1-6), the ammonia tunnel region (aa 132-145), and the C-terminal active site (aa 357-369)

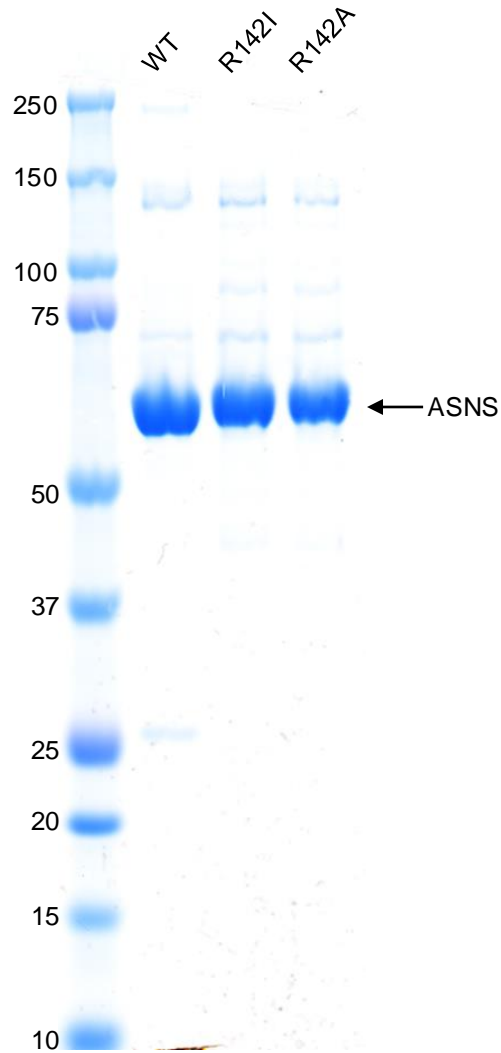
b Model-to-map FSC curves generated in Phenix.



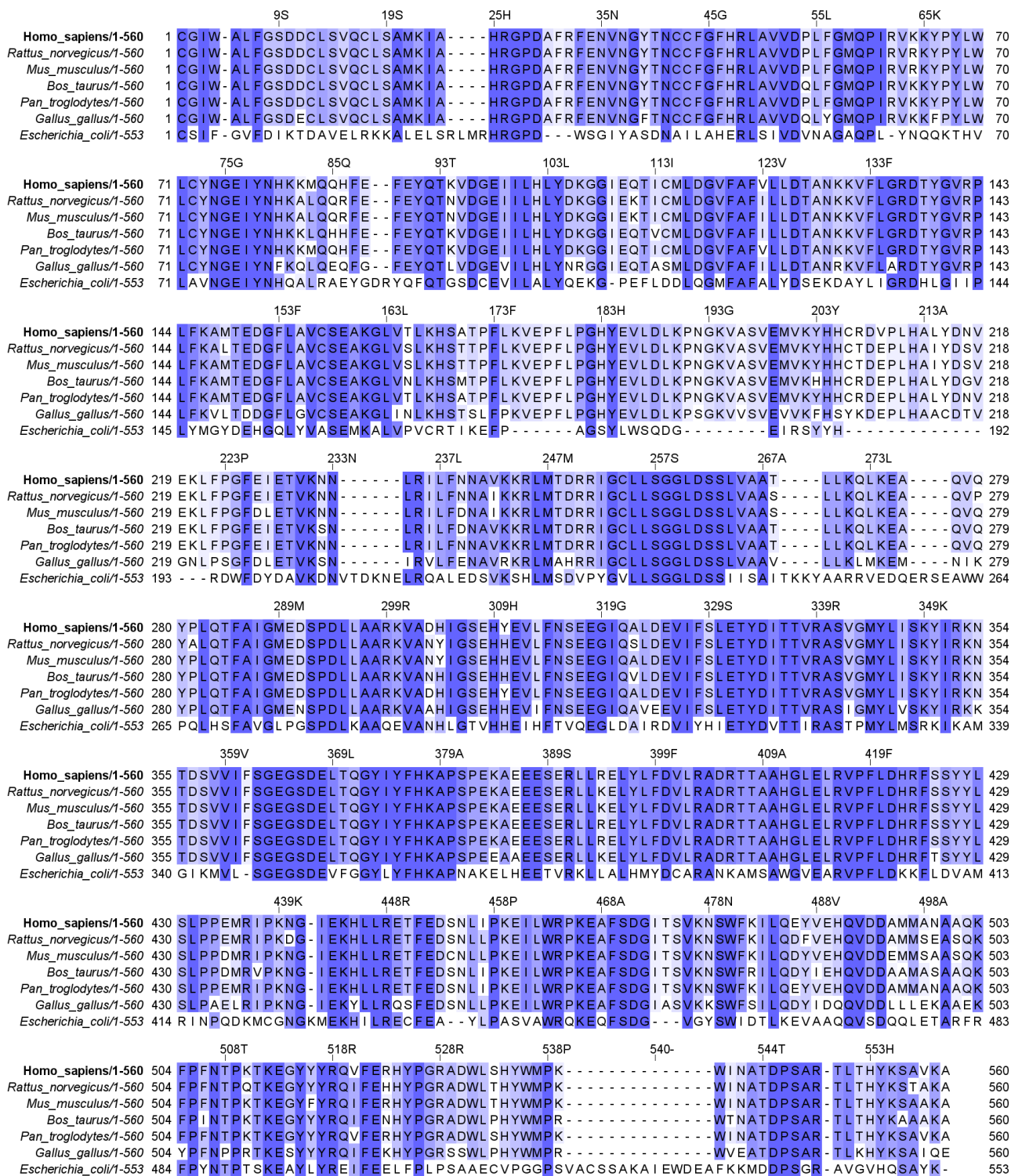
Supplementary Figure 8. Dimerization region with the map and model. Density from the EM map is shown for residues located in the dimerization region (aa 30-35) is displayed with the model. Salt bridges in between residues Arg-32 or Glu-34 (monomer A) and Glu-34 or Arg-32 (monomer B) are indicated by lines colored in cyan. Color scheme: C, magenta; N, blue; O, red.



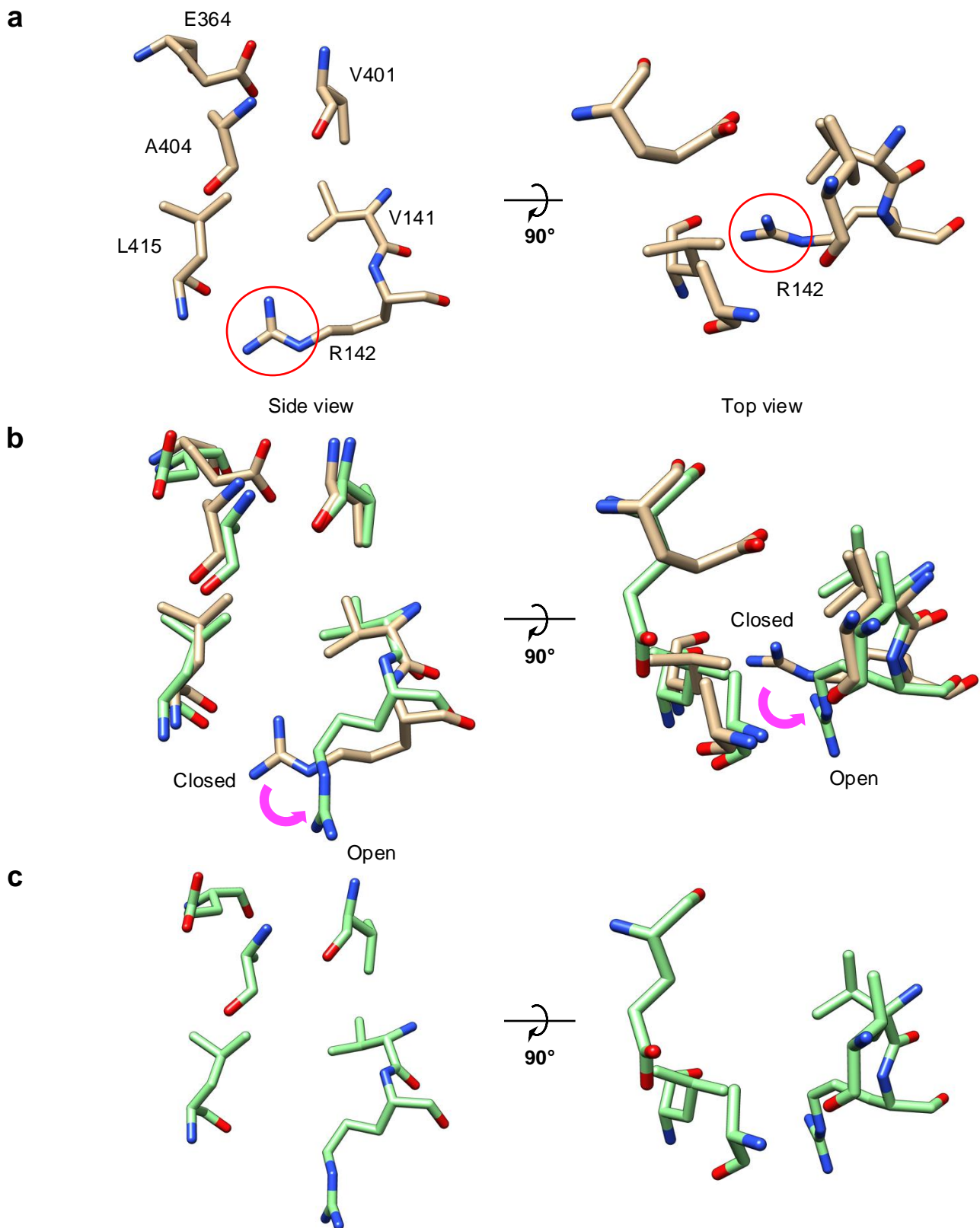
Supplementary Figure 9. The EM-derived (left) and X-ray crystal (right) structures of the human asparagine synthetase monomer. Ten residues adjacent to the experimentally undefined regions are colored in magenta and red for the EM-derived and X-ray structures, respectively.



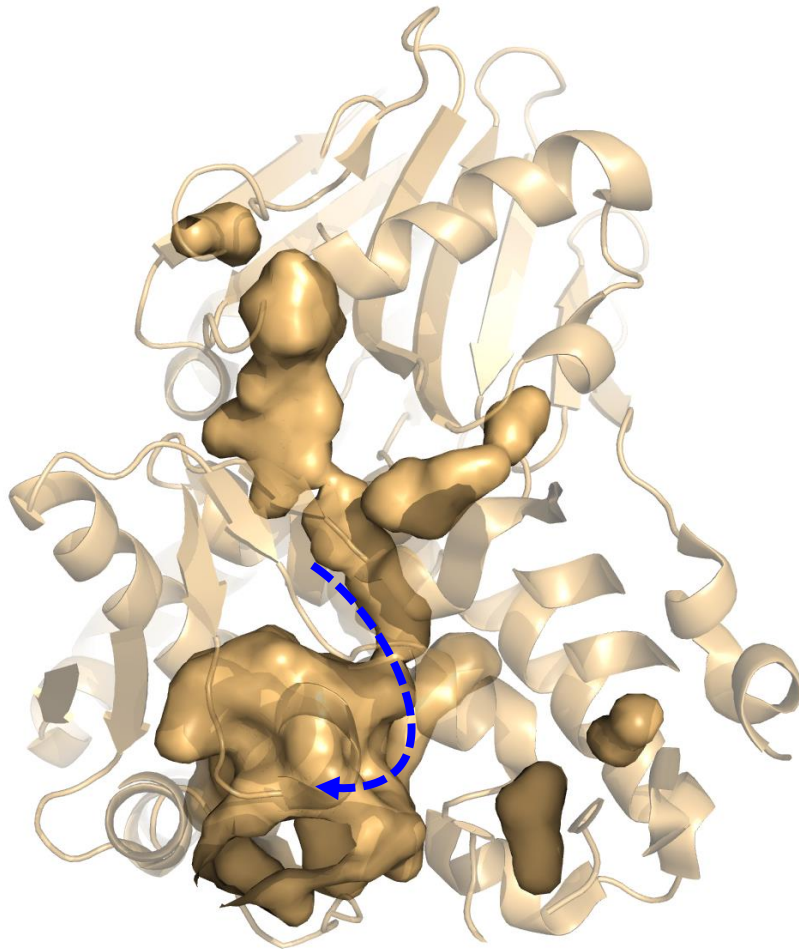
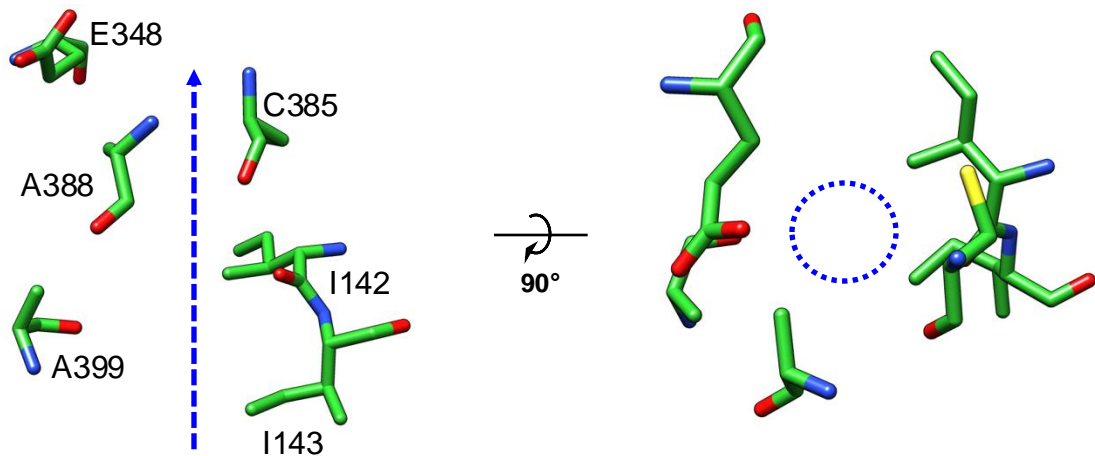
Supplementary Figure 10. SDS-PAGE of recombinant WT ASNS, and the recombinant R142I and R142A ASNS variants used in this study. Approximately 6 μ g samples of each recombinant protein were subjected to SDS-PAGE and stained with Coomassie Brilliant Blue. Each ASNS band is indicated by arrow. Left hand column is composed of molecular weight markers (numbers indicate the marker size in kDa). M: molecular weight marker; WT: wild-type ASNS; R142I; R142I variant; R142A; R142A variant.



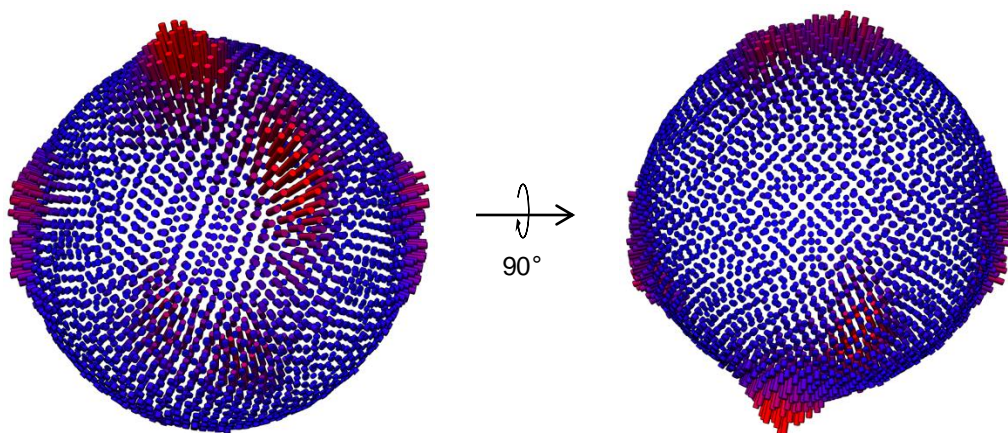
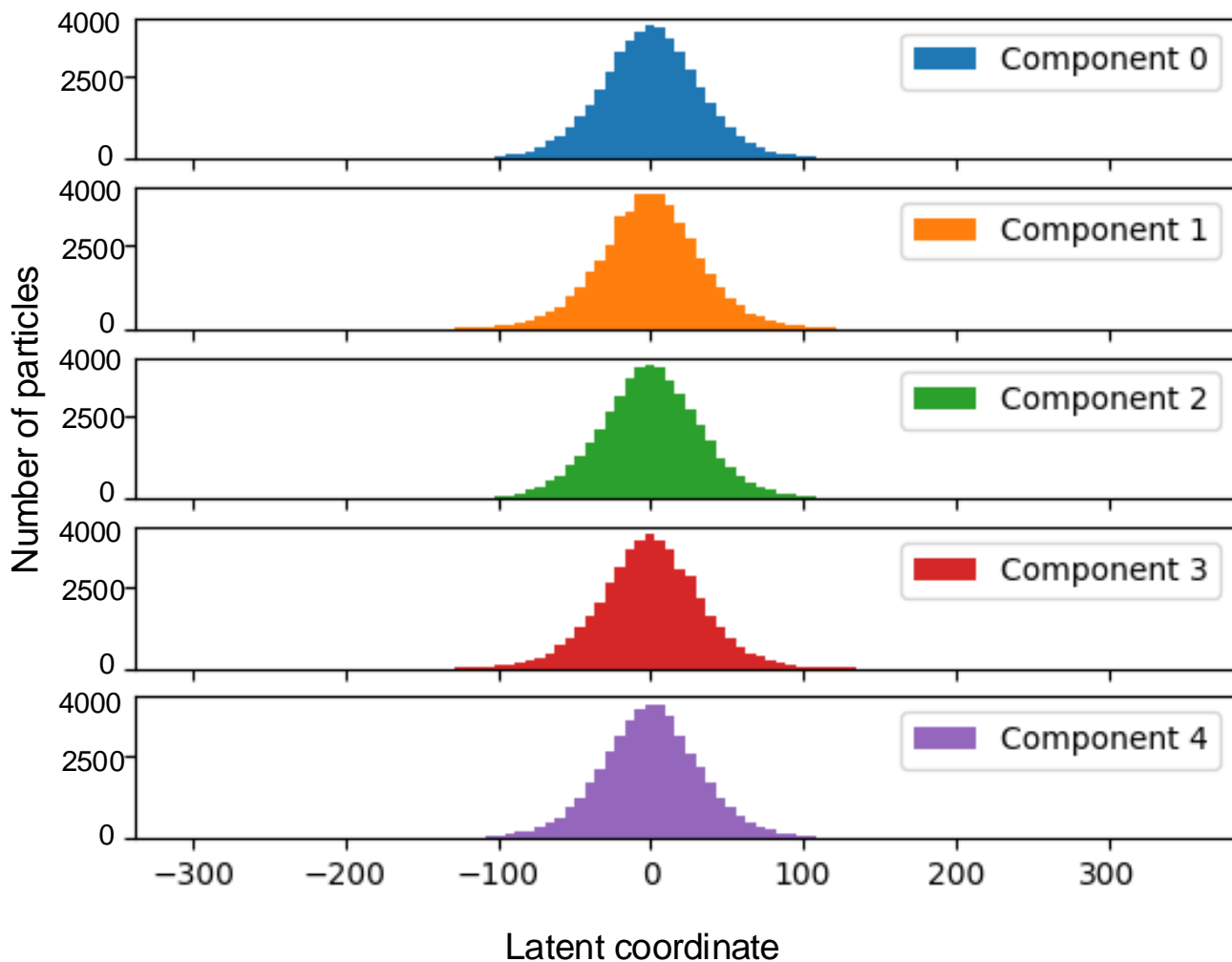
Supplementary Figure 11. Sequence alignment of the deduced sequences for glutamine-dependent asparagine synthetase in mammals, chickens and *Escherichia coli*. Shading indicates conserved (dark) and highly conserved (light) residues



Supplementary Figure 12. Example of conformational changes in the ammonia tunnel interconvert the “open and closed” forms in WT ASNS, as detected by 3D variability analysis.
a Critical residues constituting the ammonia tunnel showing how the Arg-142 side-chain (R142) blocks the tunnel in the EM structure
b Superimposition of tunnel residues in 3DVA component1 superimposed with those shown in a, indicating how a conformational change of R142 interconverts the open and closed forms of the tunnel
c Open conformation detected in the 3DVA component1

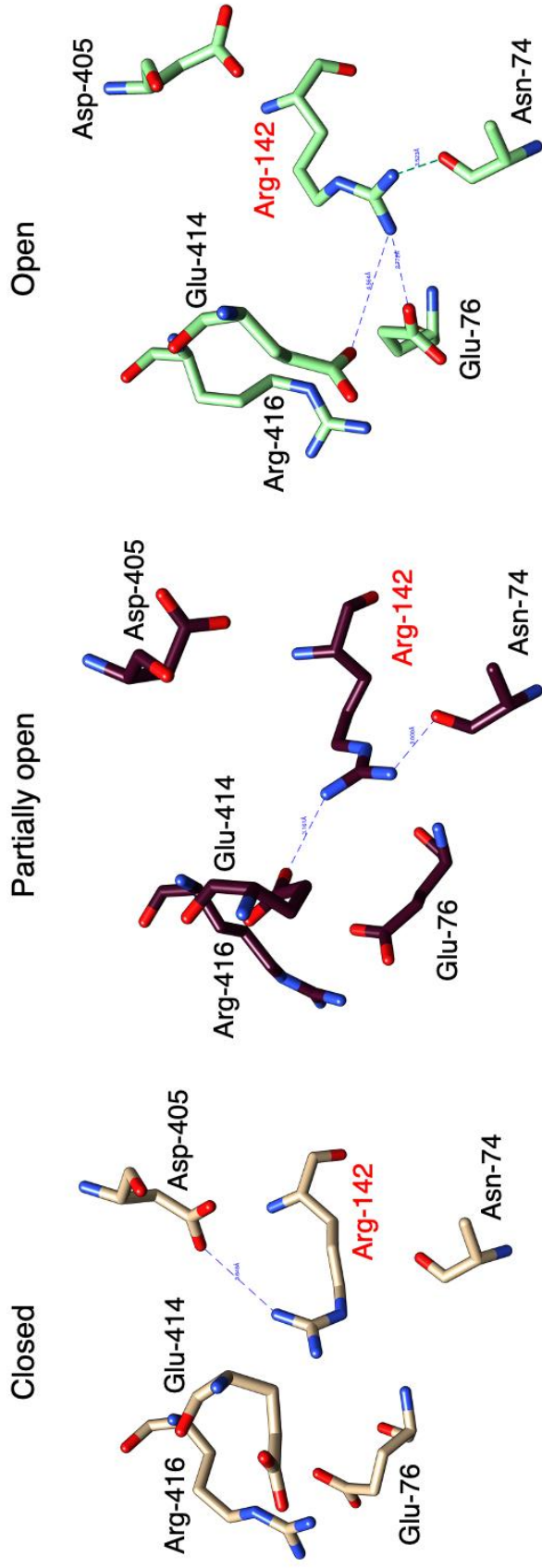
a**b**

Supplementary Figure 13. The open tunnel observed in the X-ray crystal structure of the C1A variant of *Escherichia coli* AS-B complexed with glutamine and AMP. **a The blue dashes represent the location of the open tunnel for NH_3 translocation from the N-terminal glutaminase site to the C-terminal synthetase site. **b** Critical residues constituting the tunnel: side view (left) and top view (right). The dashed blue arrow and circle illustrate an open tunnel configuration.**

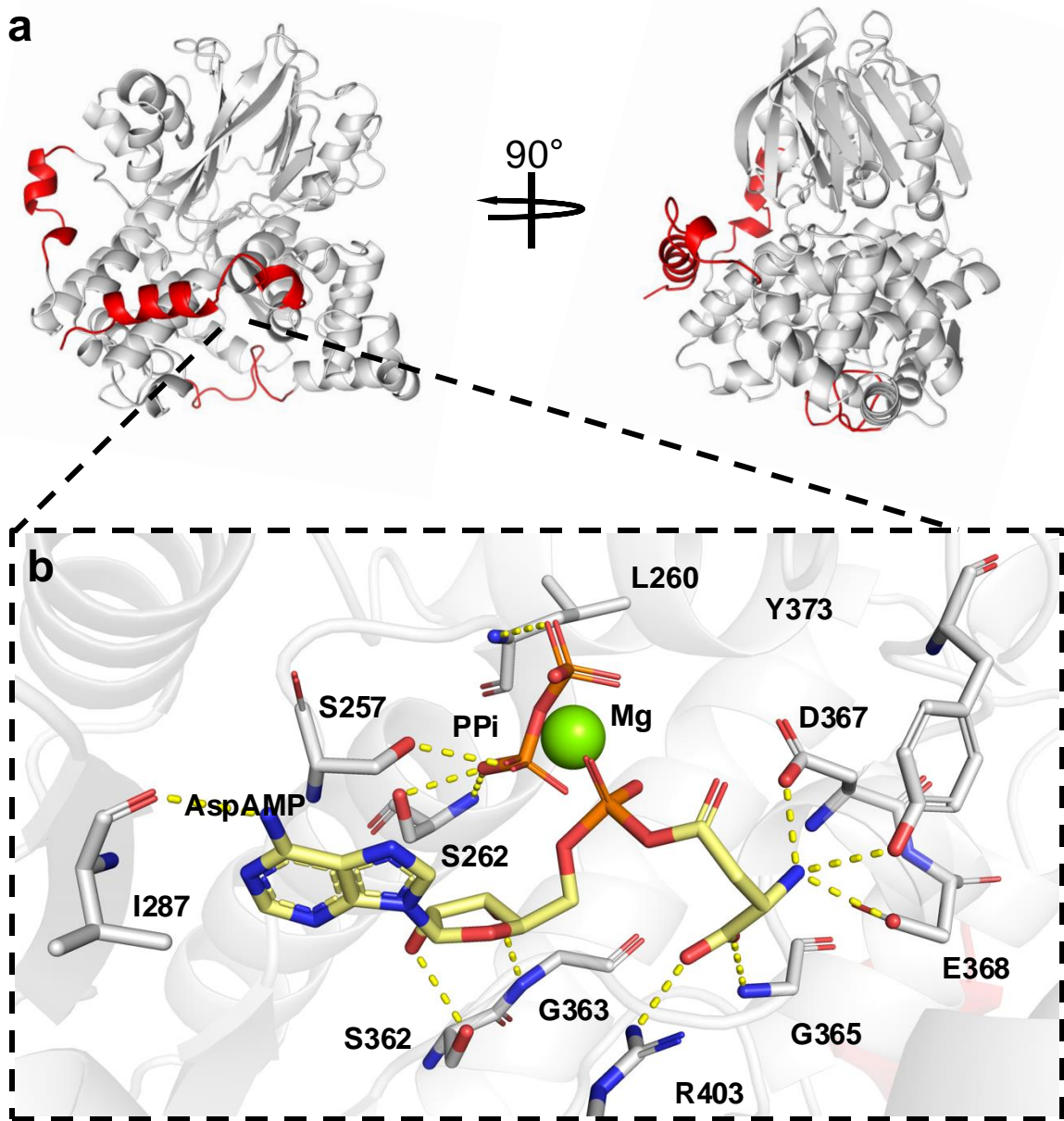
a**b****Supplementary Figure 14. PCA and particle distributions.**

a 3D representation of particle distribution used in the reconstruction; 2 views at 90° are represented. Red cylinders indicate the amount of particle in each rotation angle for the reconstruction.

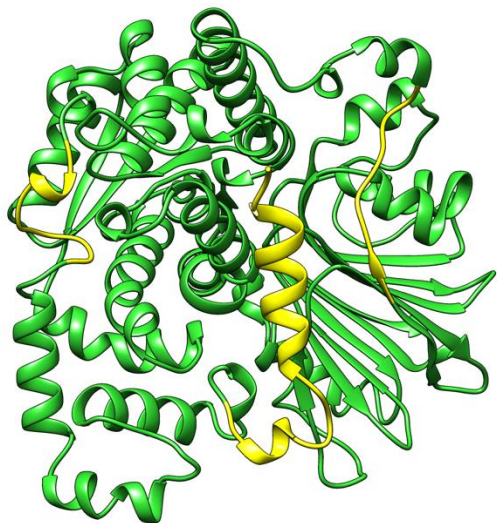
b Particle distribution in latent space, represented per principal component as a function of latent coordinate.



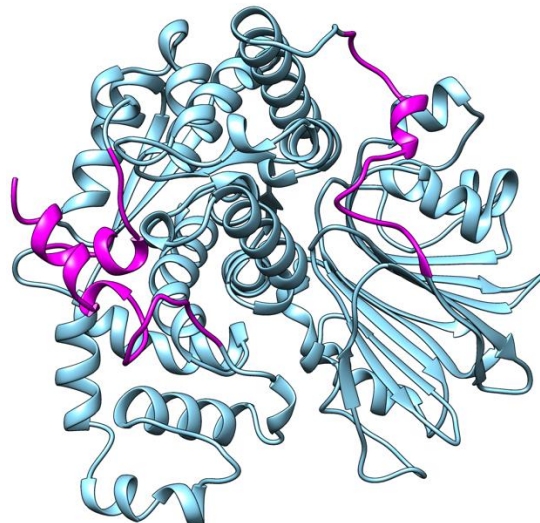
Supplementary Figure 15. Distance-based definitions of closed, open, and partially open Arg-142 conformations seen in the 3DVA analysis. (Left) In the closed conformation, blocking ammonia access to the tunnel, Arg-142 only makes a direct electrostatic interaction with the carboxylate of Asp-405. (Middle) In the partially open conformation, the Arg-142/Asp-405 interaction is disrupted, and the side chains move apart. As a result, the guanidinium group becomes closer to the side chains of Asn-74 and Glu-414. (Right) In the fully open conformation, which permits ammonia access to the tunnel, the guanidinium moiety of Arg-142 forms an additional interaction with the carboxylate of Glu-76. The Arg-142/Asp-405 and Arg-142/Glu-414 distances can therefore be used as a measure of whether the side chain is in the closed conformation (shorter/longer, respectively) or open conformation (shorter/longer, respectively).



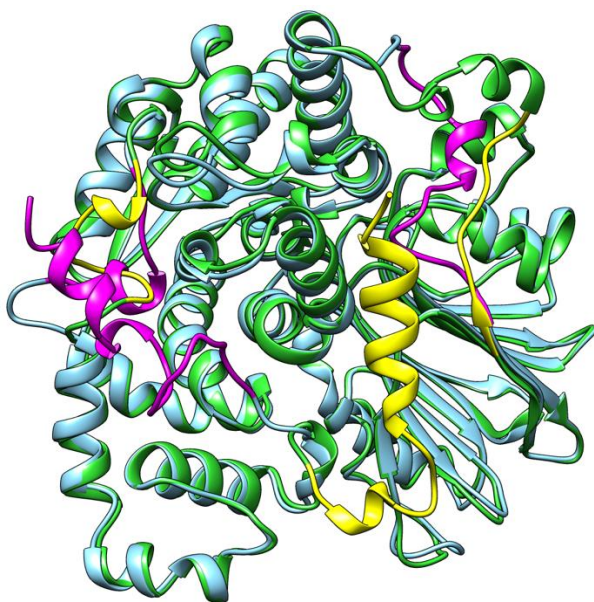
Supplementary Figure 16. Computational models constructed from the Robetta structure of human ASNS. **a** Cartoon representation of the full-length model of human apo-ASNS used in the computational studies, showing the reconstructed loops (red). **b** Close-up of the synthetase active site in the computational model of the human ASNS/ β -aspartyl-AMP/PP_i ternary complex. Key protein residues (C, grey) and the ligands (C, yellow) are rendered as sticks and Mg²⁺ is shown by the green sphere. Intermolecular hydrogen bonds are indicated by the dashed yellow lines. (Note that these models were also used to generate initial structures for the apo-R142I variant and its β -aspartyl-AMP/PP_i ternary complex).

a

ROBETTA ASNS model

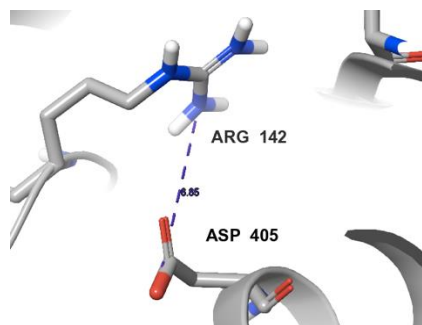
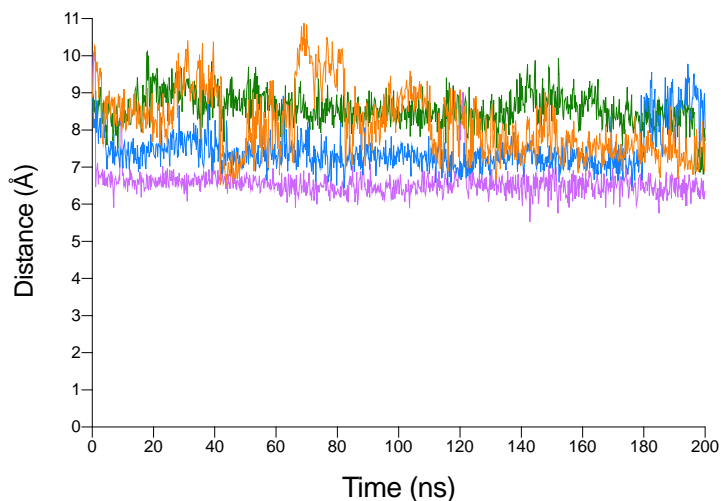
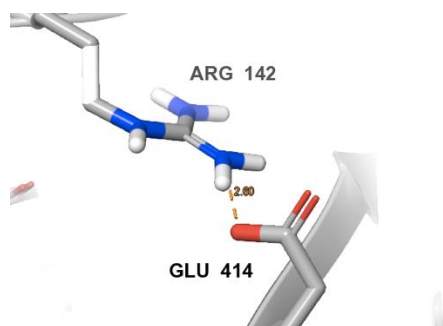
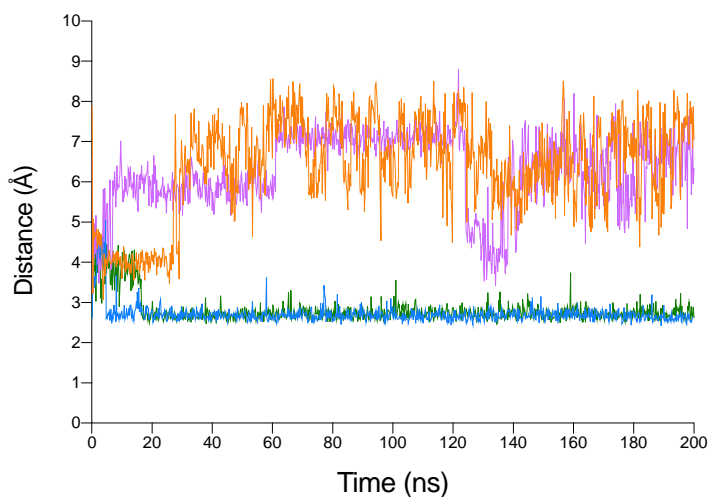


AF3 ASNS model

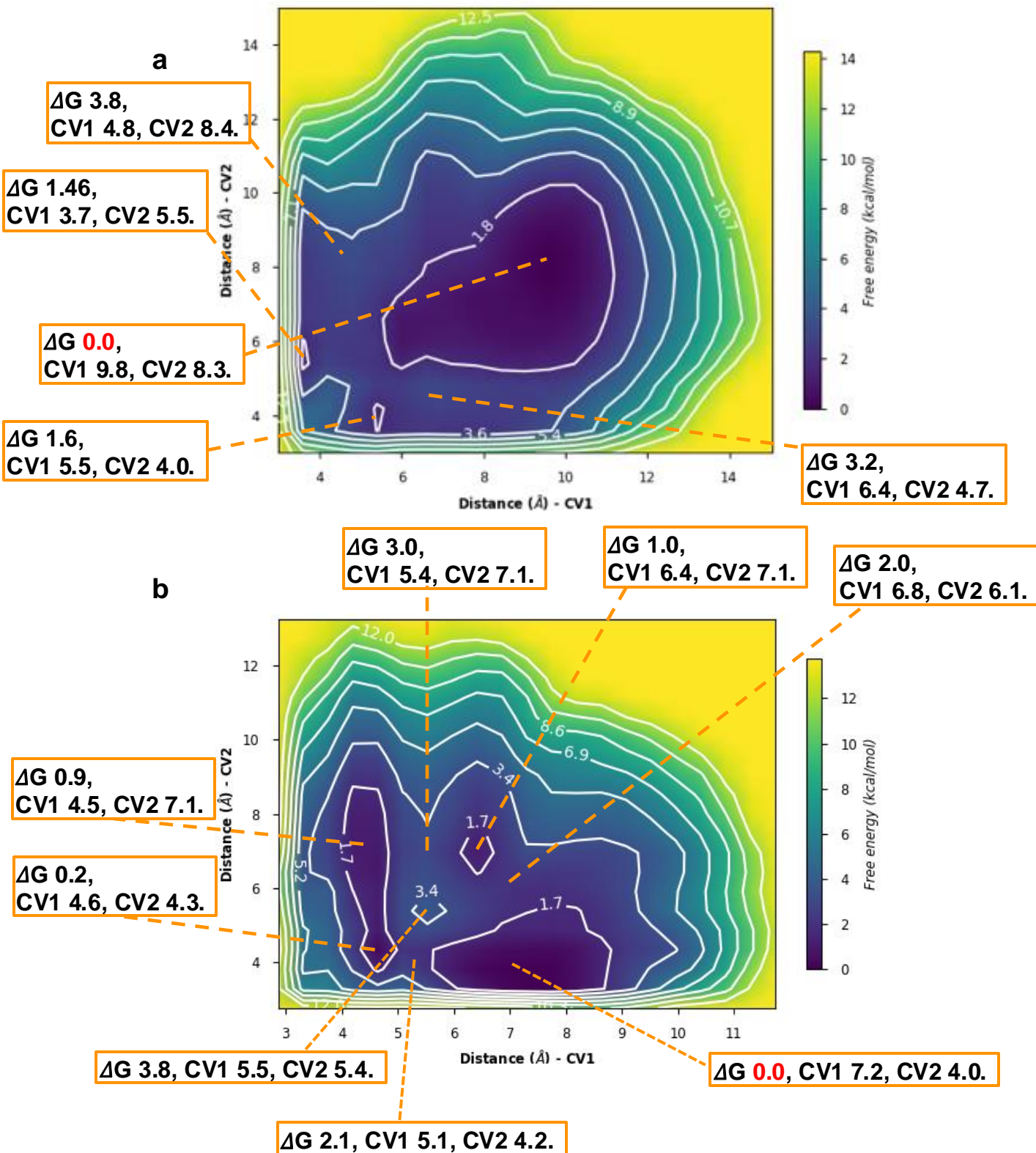
b

Superimposed

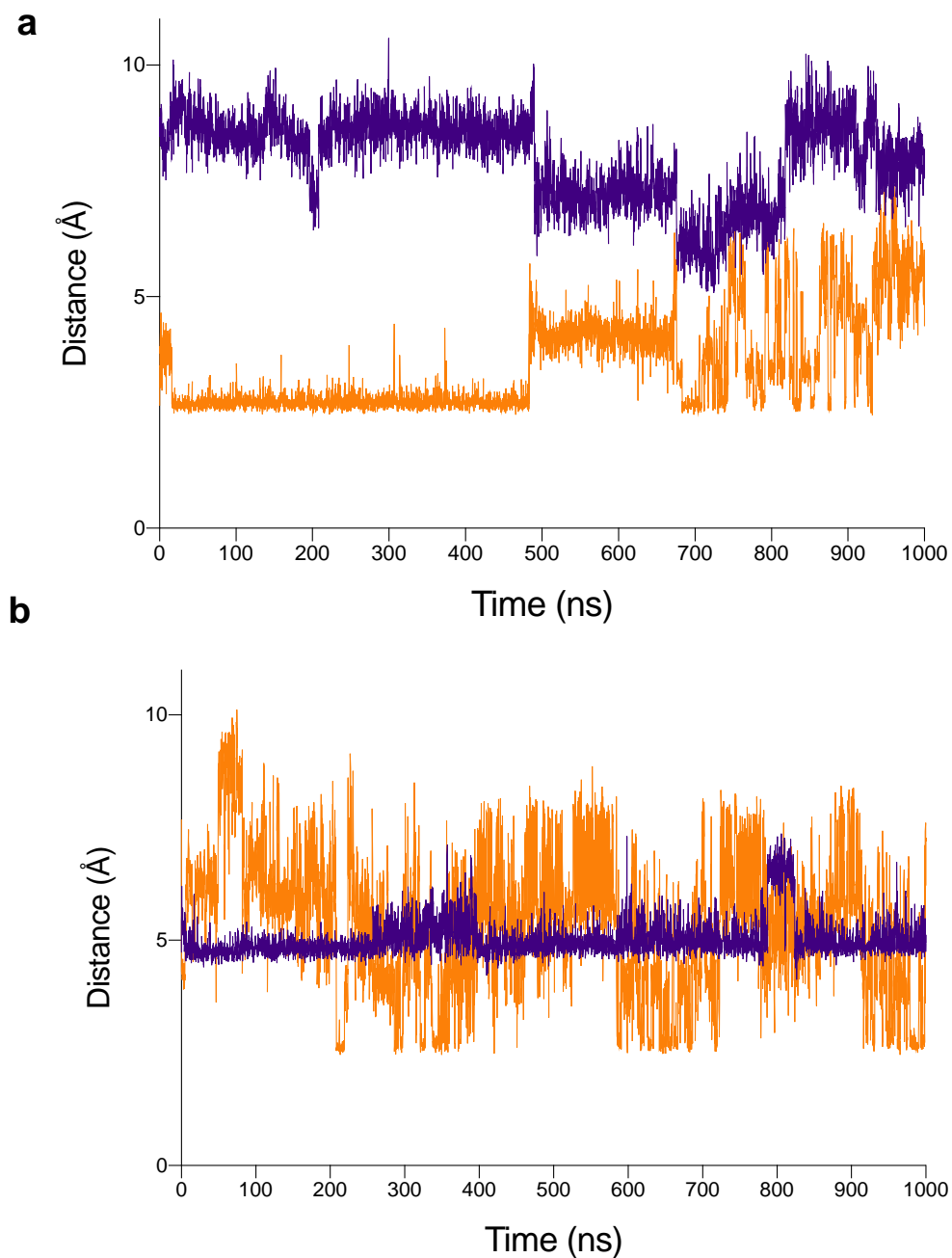
Supplementary Figure 17. Comparison of ROBETTA-generated and Alphafold3 (AF3)-generated models for the WT human apo-ASNS monomer. **a** Ribbon representation of the ROBETTA model (lime green), and the AF3-generated model (sky blue). Loop regions (residues 201-220, 465-475) as well as the C-terminal tail (residues 536-560), which are not seen in the X-ray or cryo-EM structures are colored in yellow for the ROBETTA model and magenta for the AF3 model. **b** Superimposed models of ROBETTA-generated and AF3-generated models (RMSD: 0.629 Å). The major difference is associated with the position of the C-terminal region in the two models.

a**Arg142-Asp405****b****Arg142-Glu414**

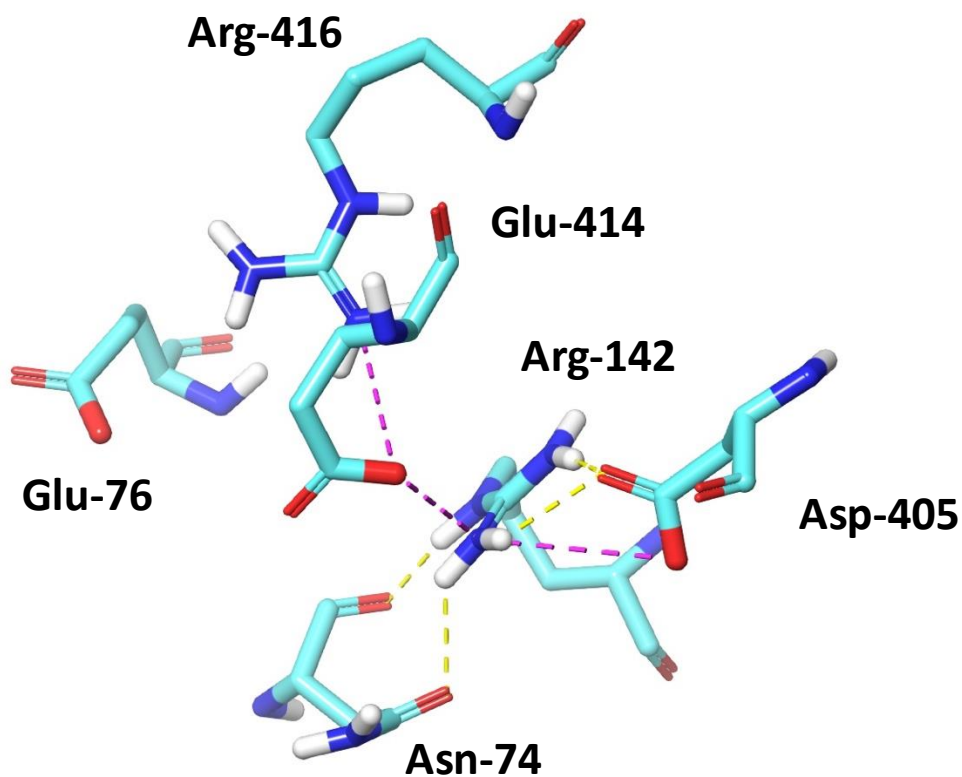
Supplementary Figure 18. Replicate 200 ns MD simulations of the apo-ASNS monomer. **a** Plots of the distance between Arg-142 and Asp-405 (shown on the left) in the four replicates of the 200 ns MD simulations of the apo-ASNS monomer. **b** Plots of the distance between Arg-142 and Glu-414 (shown on the left) in the four replicates of the 200 ns MD simulations of the apo-ASNS monomer. Distances are calculated between the NH₂ nitrogen atoms of Arg-142 and carboxylate oxygen atoms of Asp-405 or of Glu-414, respectively, as indicated by the dashed lines. Shorter Arg-142/Asp-405 (longer Arg-142/Glu-414) distances correspond to Arg-142 adopting the “closed” conformation in the cryo-EM structure that blocks ammonia access. Similarly, longer Arg-142/Asp-405 (shorter Arg-142/Glu-414) distances correspond to Arg-142 adopting the “open” conformation in the cryo-EM structure that permits ammonia access. Source data are provided as a Source Data file.



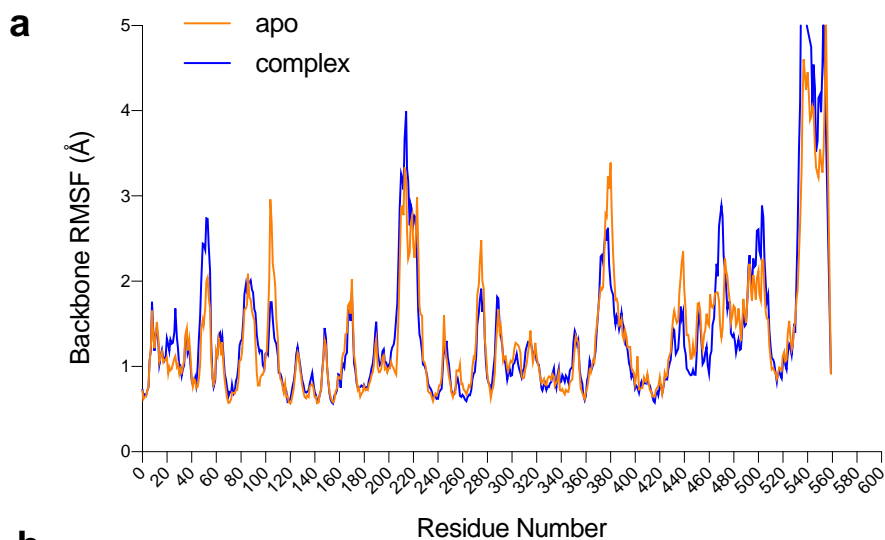
Supplementary Figure 19. Metadynamics-based free energy surfaces for the Arg-142 side chain in (a) apo-ASNS and (b) the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex. Two distance-based collective variables were defined for these simulations (Supplementary Figs. S15), which correspond to the distances between the center-of-mass (COM) of the guanidinium group in Arg-142 and the COM of the carboxylate moieties in either Asp-405 (CV1) or Glu-414 (CV2).



Supplementary Figure 20. Microsecond simulations of apo-ASNS and the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex in their monomeric form. **a Plots of the distance between Arg-142 and Glu-414 (orange) and between Arg-142 and Asp-405 (purple) in the microsecond MD simulation of the full-length apo-ASNS monomer. **b** Plots of the distance between Arg-142 and Glu-414 (orange) and between Arg-142 and Asp-405 (purple) in the microsecond MD simulation of the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex. All distances are defined as described in the legend to Fig. S17. Source data are provided as a Source Data file.**

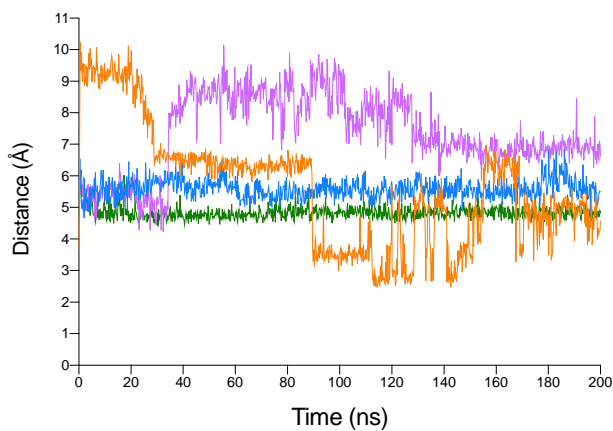


Supplementary Figure 21. Residue network showing the Arg-142/Asn-74 interaction. Structure is taken from a snapshot ($t = 200$ ns) in the MD trajectory of apo-ASNS. Hydrogen bonds and salt bridges are indicated by yellow and violet dashed lines, respectively. Color scheme: C, cyan; H, white; N, blue; O, red.

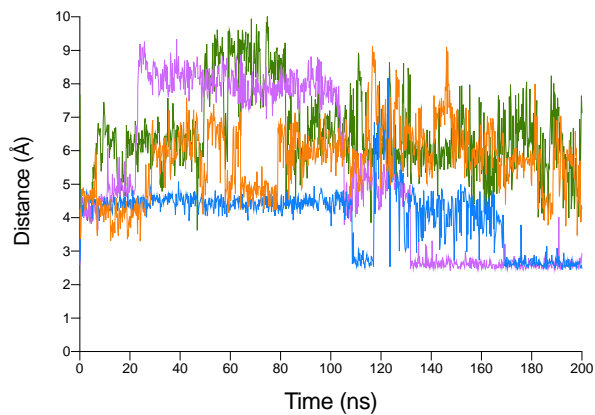


b

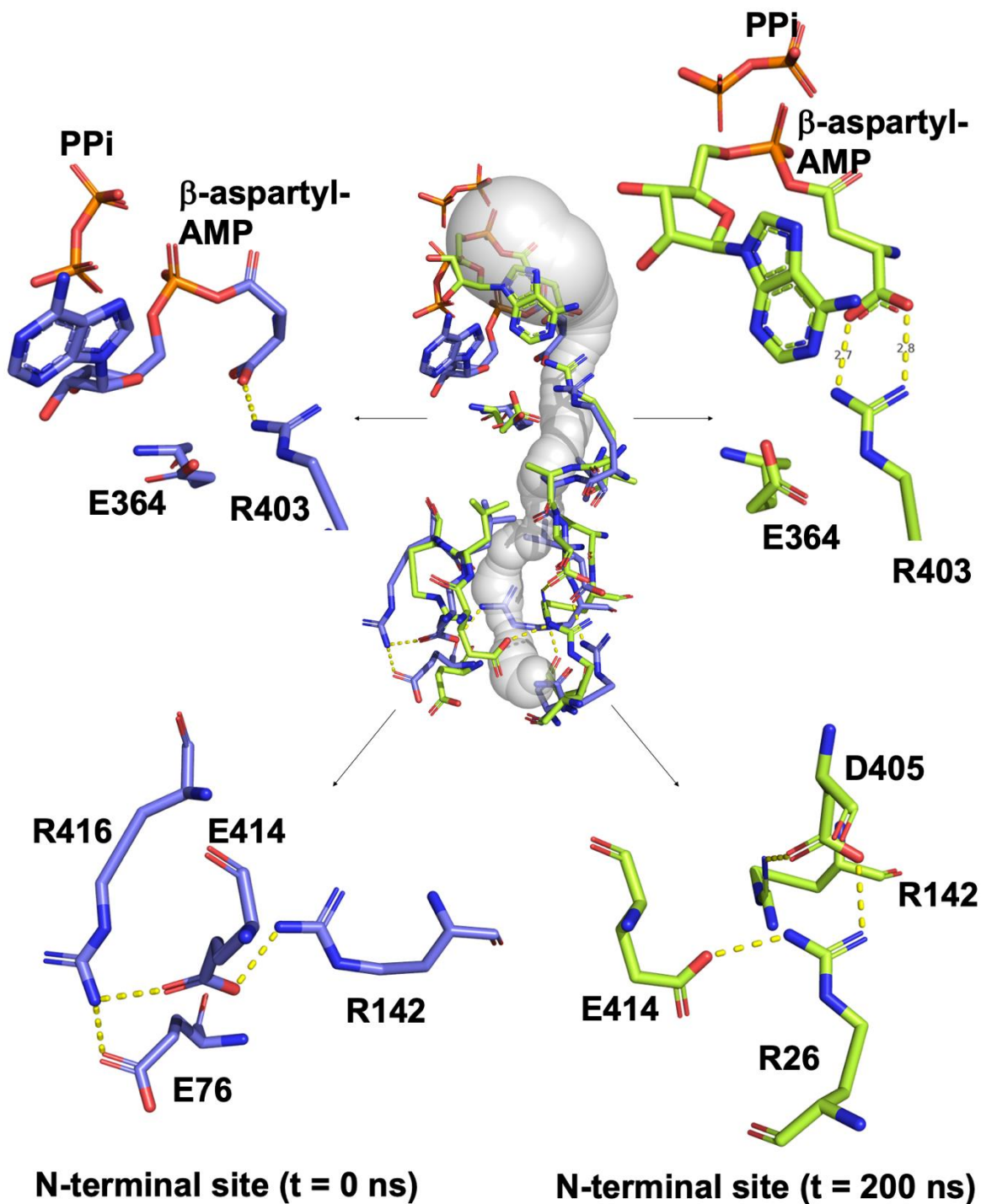
Arg142-Asp405



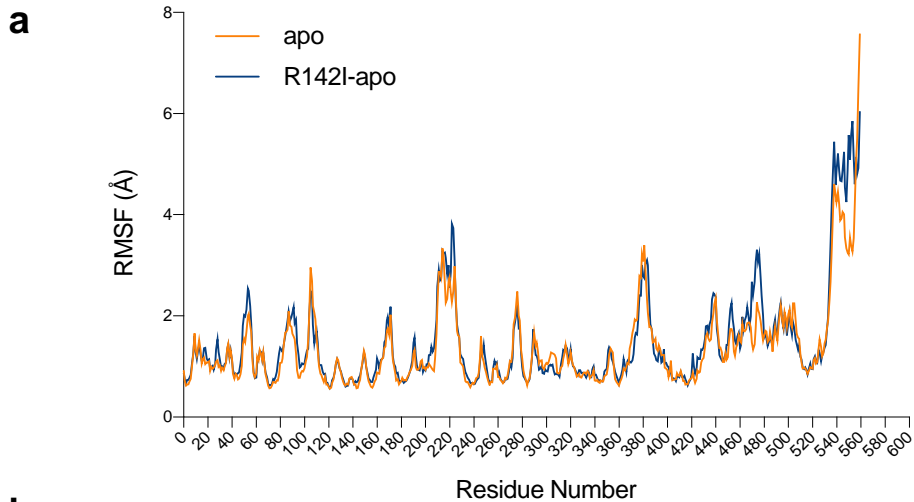
Arg142-Glu414



Supplementary Figure 22. 200 ns MD simulations of the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex. **a** Comparison of backbone RMSF values computed for the apo-ASNS monomer (orange) and the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex (blue). **b** Replicate 200 ns MD simulations of the WT ASNS/ β -aspartyl-AMP/ PP_i ternary complex. (left) Plots of the distance between Arg-142 and Asp-405 and (right) between Arg-142 and Glu-414 in the four replicates. All distances are defined as described in the legend to Fig. S17. Source data are provided as a Source Data file.



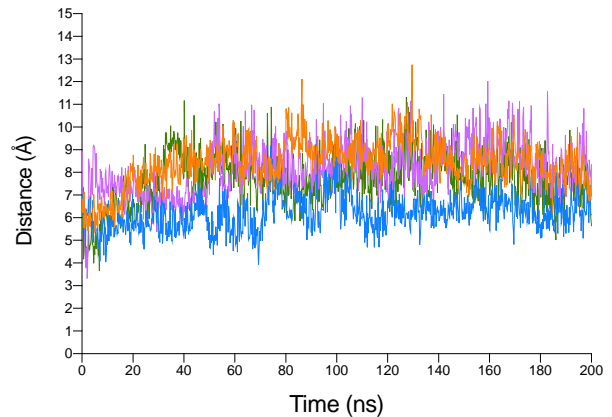
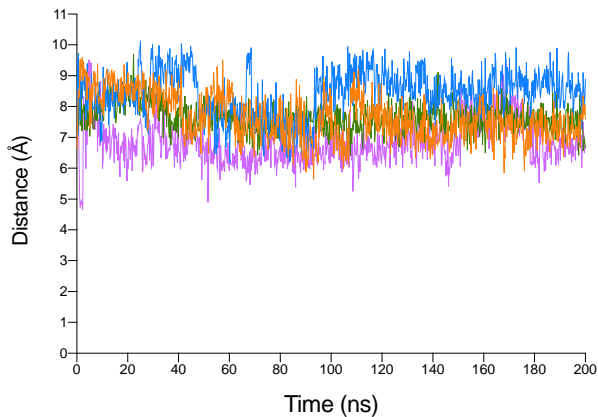
Supplementary Figure 23. Communication network between the N-terminal active site and the C-terminal active site as seen in the MD trajectory of the WT ASNS/ β -aspartyl-AMP/ PPi ternary complex. Residues are shown in sticks; carbon atoms are shown in purple and lime for the $t = 0$ ns and $t = 200$ ns snapshots, respectively. Oxygen atoms are shown in red, and nitrogen atoms are shown in blue. Yellow dashed lines show residue interactions within 2.8 Å. The tunnel present in the final snapshot of the 200 ns simulation is shown in gray.



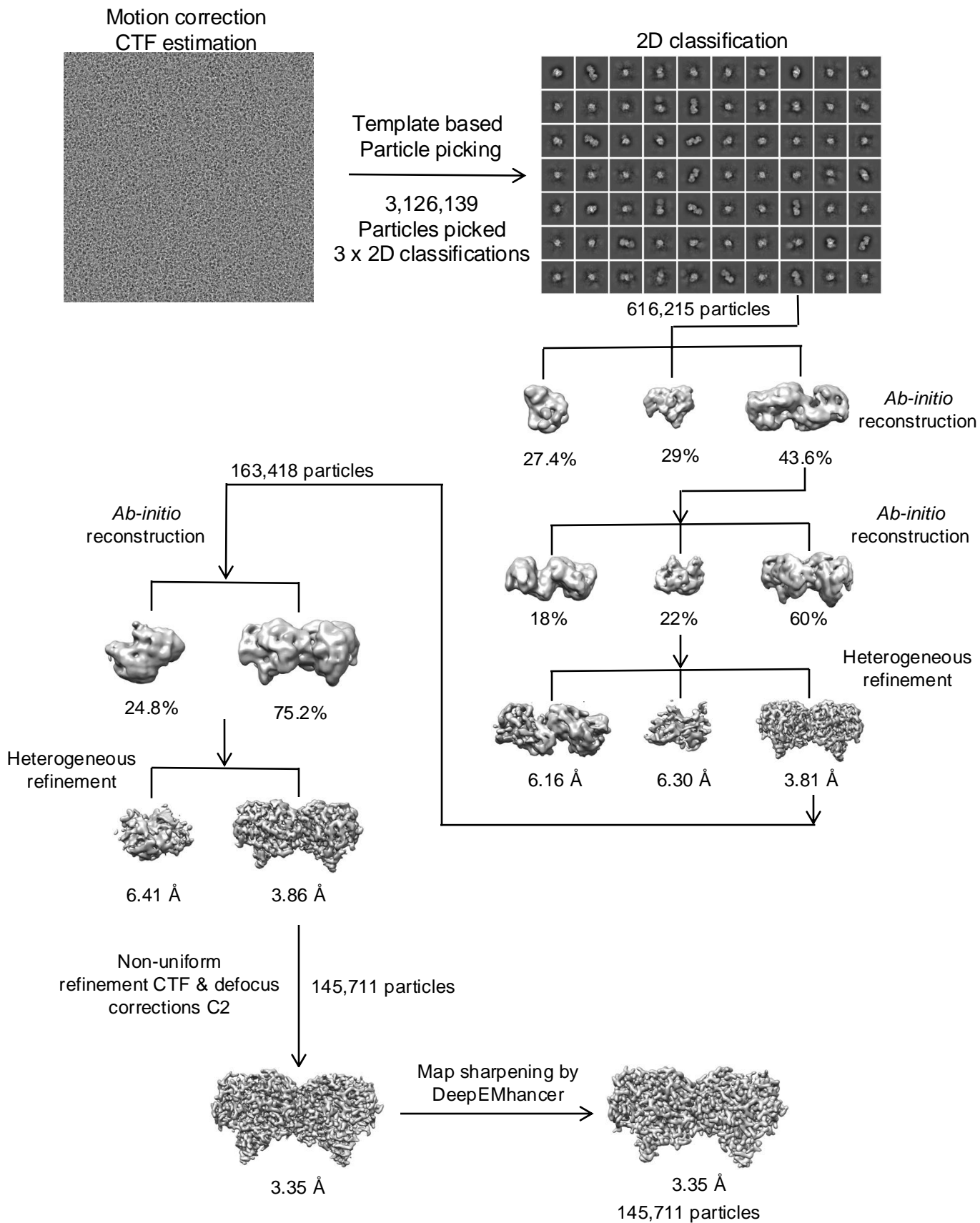
b

Ile142-Asp405

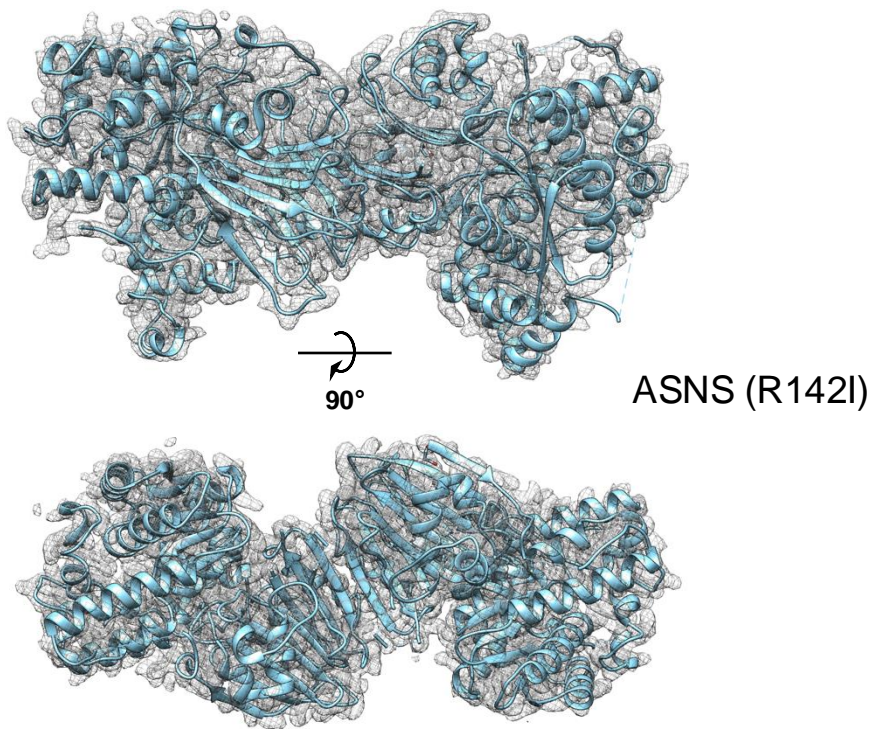
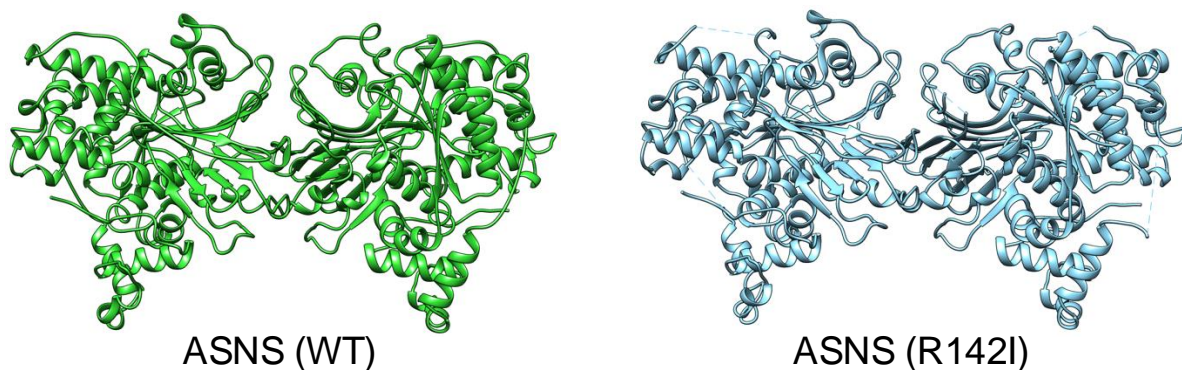
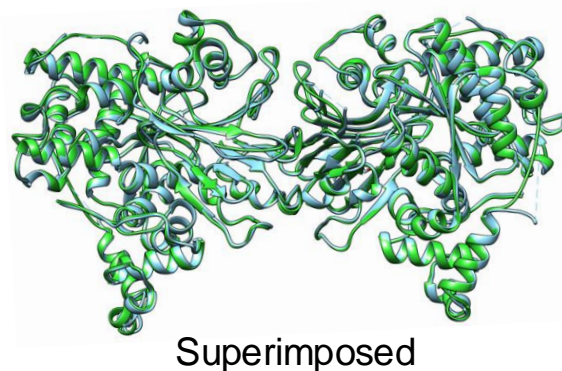
Ile142-Glu414



Supplementary Figure 24. 200 ns MD simulations of the apo-R142I variant. **a** Comparison of backbone RMSF values computed for the apo-ASNS monomer (orange) and the apo-R142I variant (blue). **b** Replicate 200 ns MD simulations of the apo-R142I variant. (left) Plots of the distance between Ile-142 and Asp-405 and (right) between Ile-142 and Glu-414 in the four MD simulation replicates. Distances were calculated between the delta carbon atom of Ile-142 and one of the carboxylate oxygen atoms of Asp-405 or of Glu-414, respectively. See Fig. S17 for the location of these oxygen atoms. Source data are provided as a Source Data file.

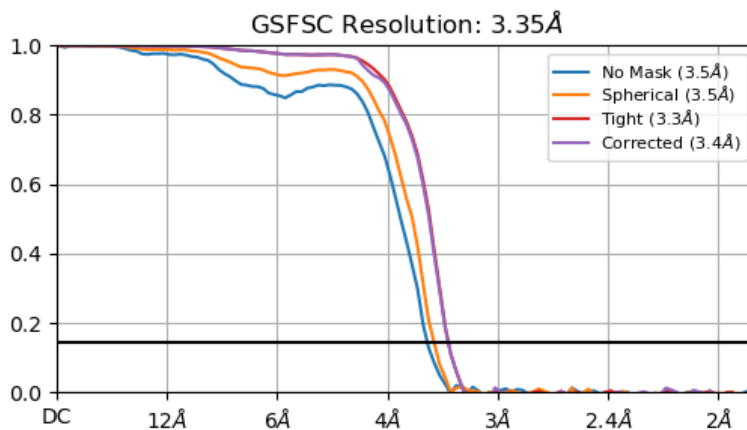


Supplementary Figure 25. Summary of the cryo-EM data processing workflow of ASNS R1421 variant using cryoSPARC v4.4.1.

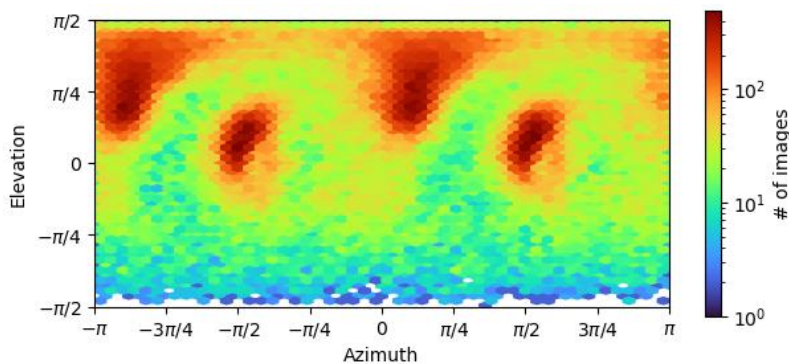
a**b****c**

Supplementary Figure 26. Structure of the R142 ASNS variant and its comparison to WT ASNS
a Ribbon representation of the structure of the R142I ASNS variant (PDB: 9B6C) colored in sky blue with its EM map (EMD-44253) in grey. **b** Comparison between WT vs. R142I variant structures: side view of the WT model (PDB: 8SUE) colored in lime green on the left, and the R142I variant model (PDB: 9B6C) in sky blue on the right. **c** Superimposed models of WT and the R142I variant (RMSD: 0.483 Å)

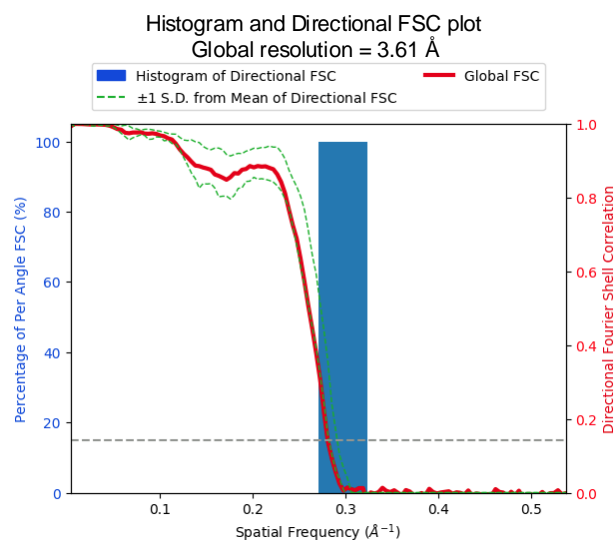
a Gold standard FSC curve



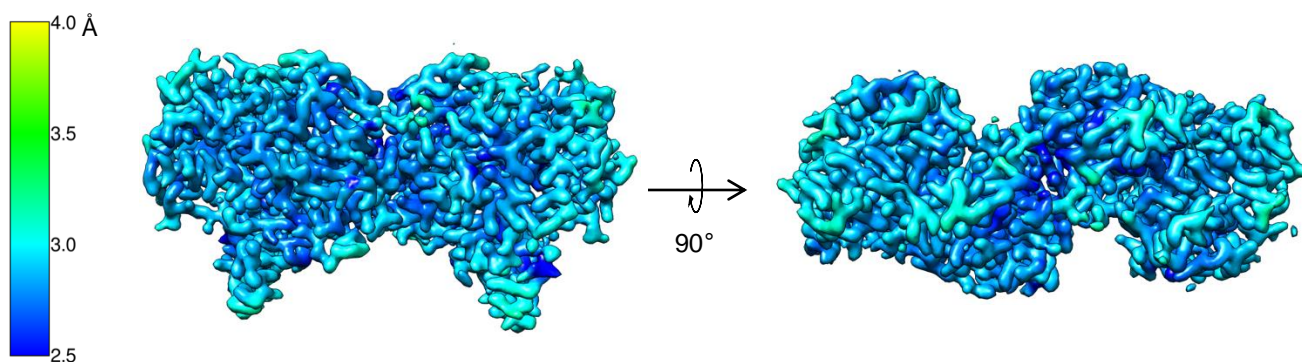
b Viewing distribution plots



c 3D FSC



d Local resolution



Supplementary Figure 27. Assessment of the quality of cryo-EM map of R142I ASNS variant.

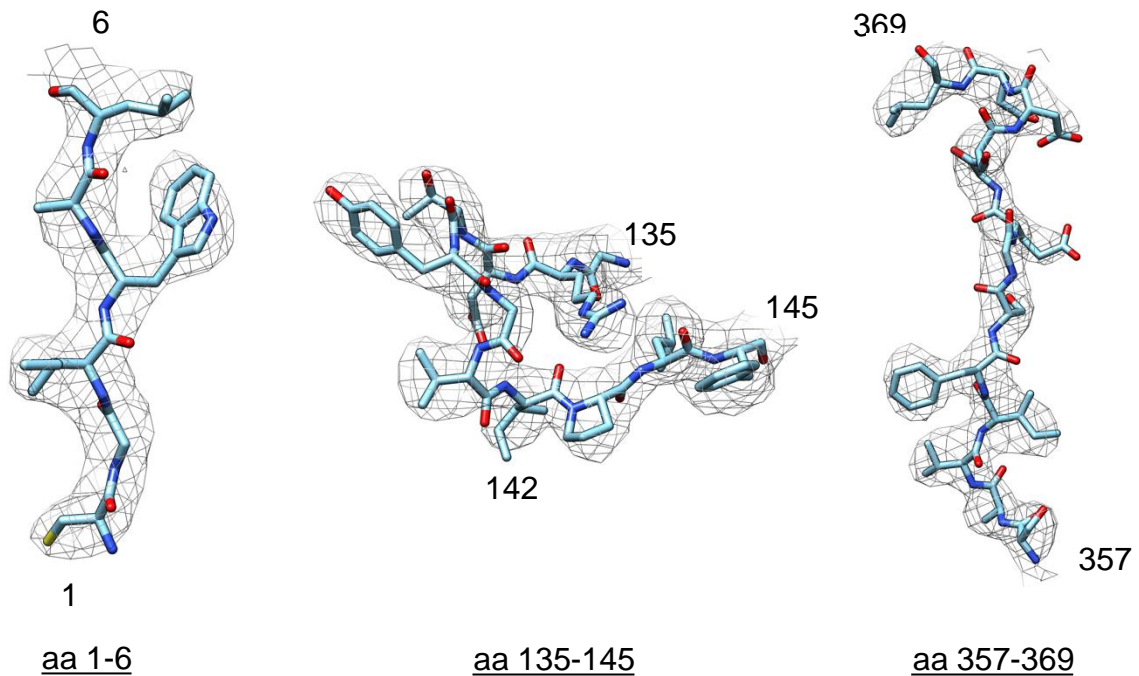
a Gold standard FSC curve for the map of R142I variant generated in cryoSPARC v4.4.1.

b Viewing distribution plots for the map generated in cryoSPARC v4.4.1.

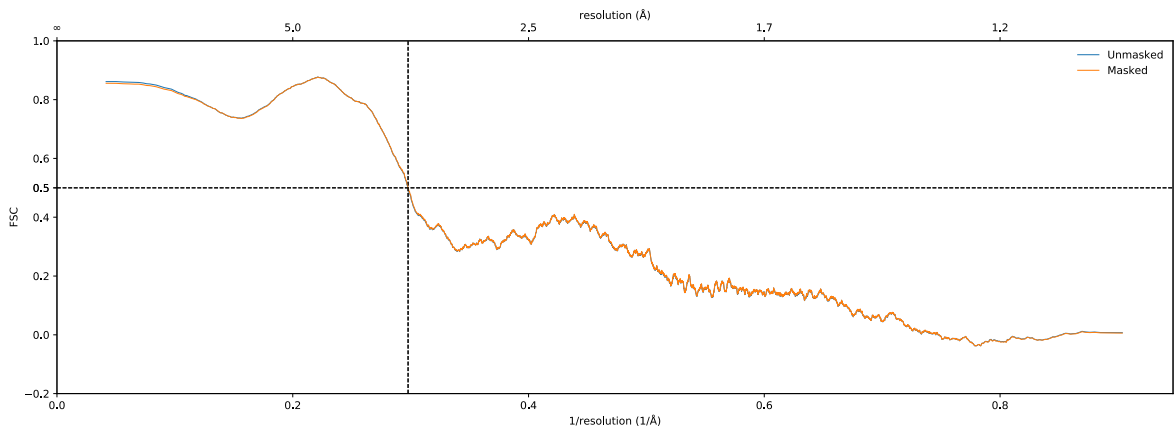
c 3D FSC: histogram and directional FSC plot for map global resolution are shown.

d Local resolution of the map (color scale shown on the left).

a Representative densities and corresponding models



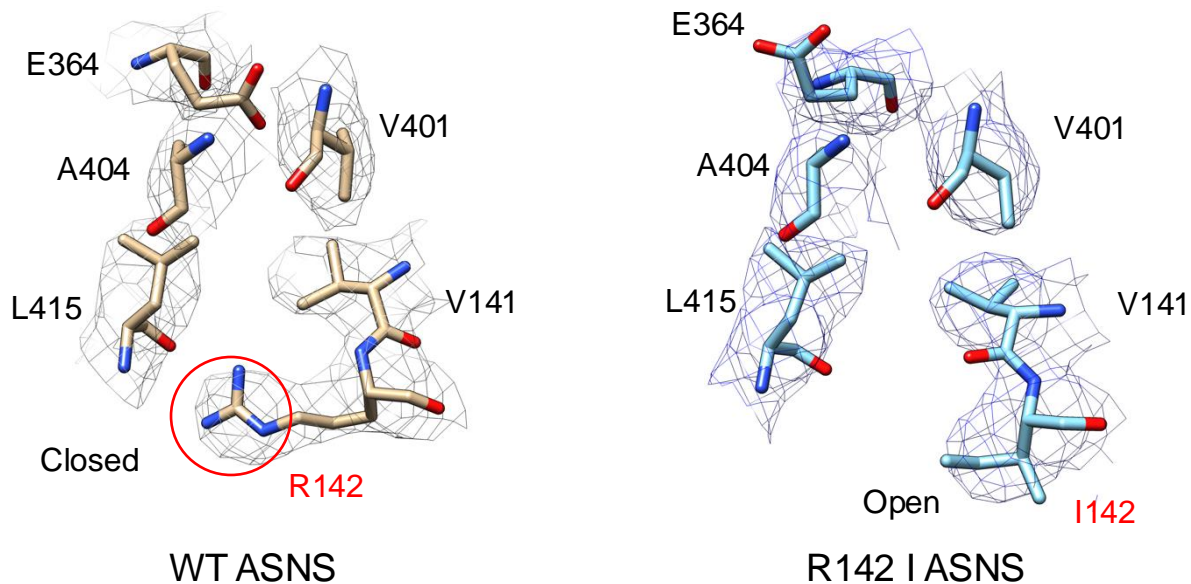
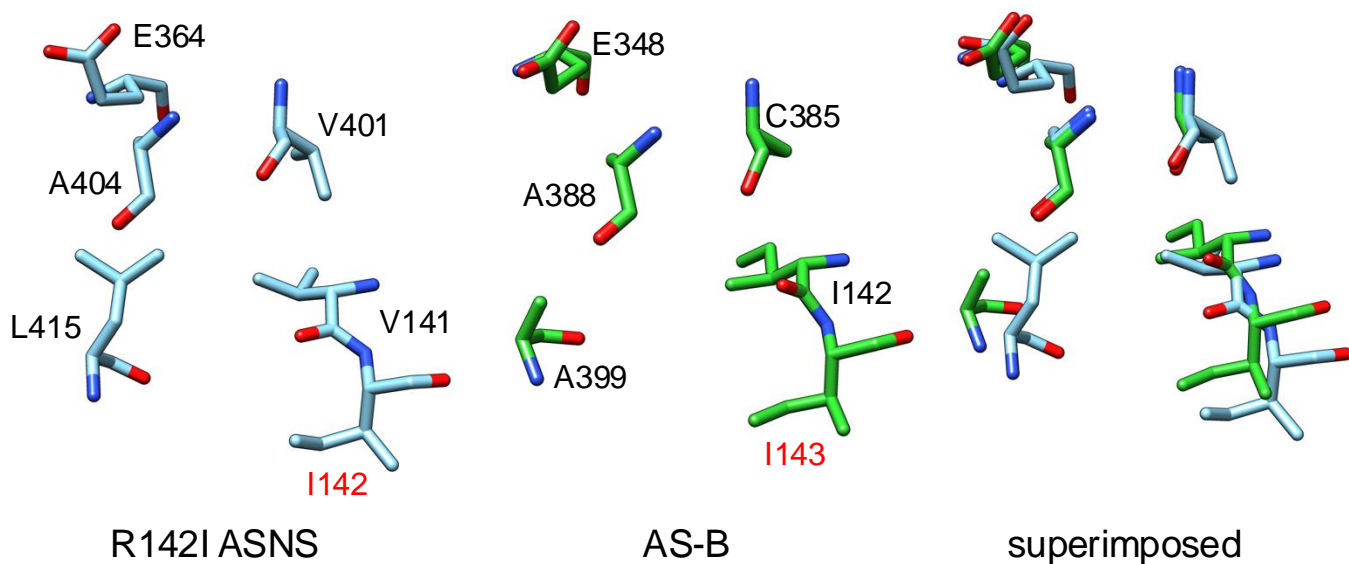
b Map-model FSC curve



Supplementary Figure 28. Validating the cryo-EM structure of ASNS R142I variant : map and its model.

a Representative density of the map for residues in the N-terminal catalytic region (aa 1-6), the ammonia tunnel region (aa 135-145), and the C-terminal active site (aa 357-369).

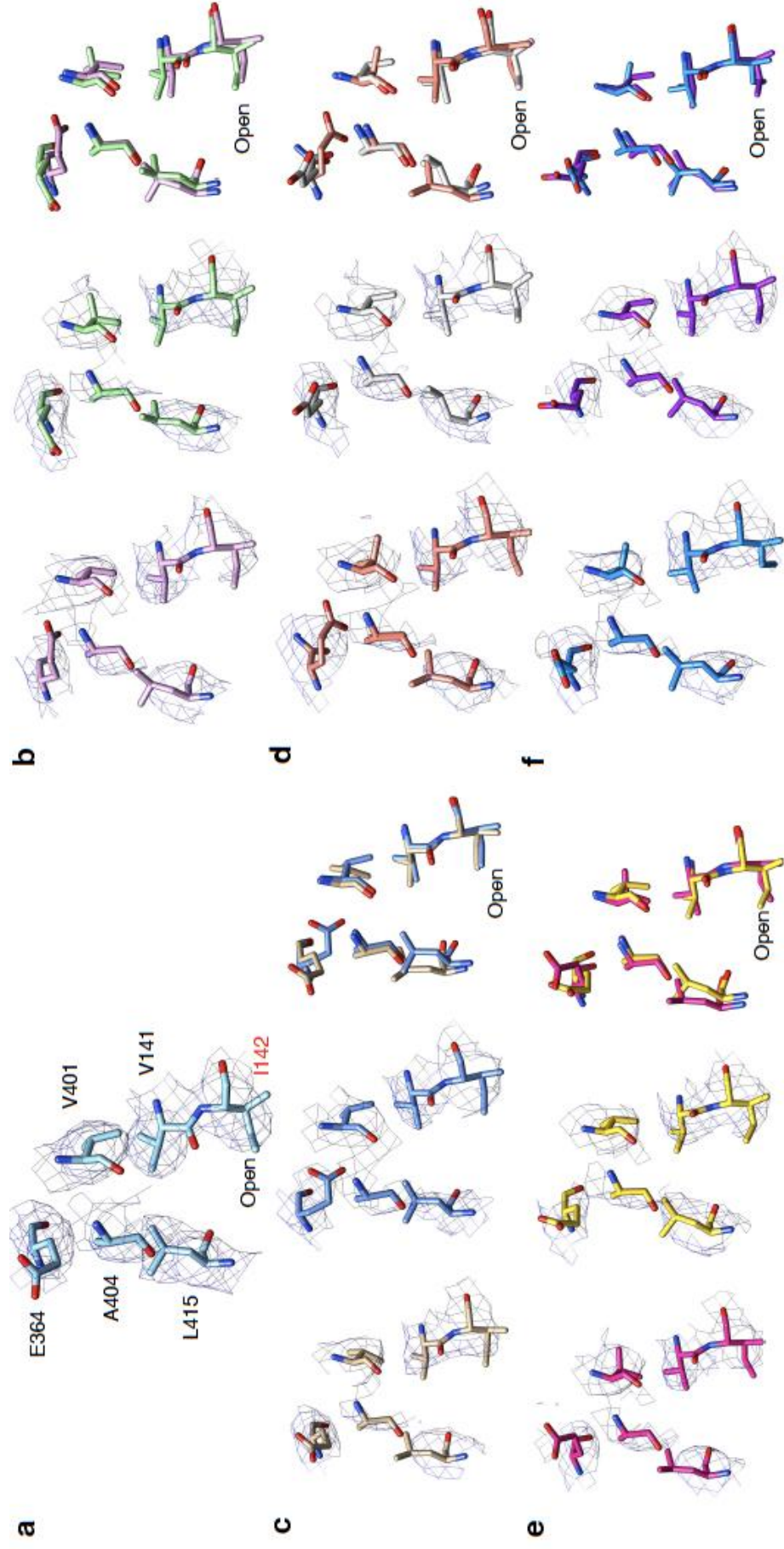
b Model-to-map FSC curves generated in Phenix.

a**b**

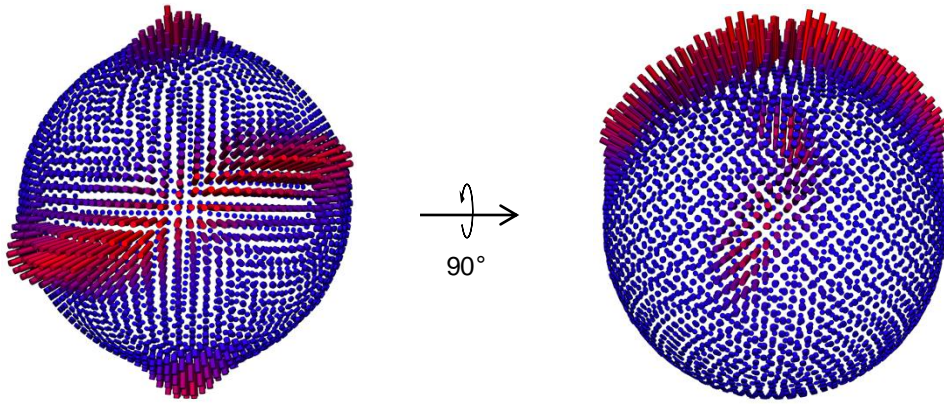
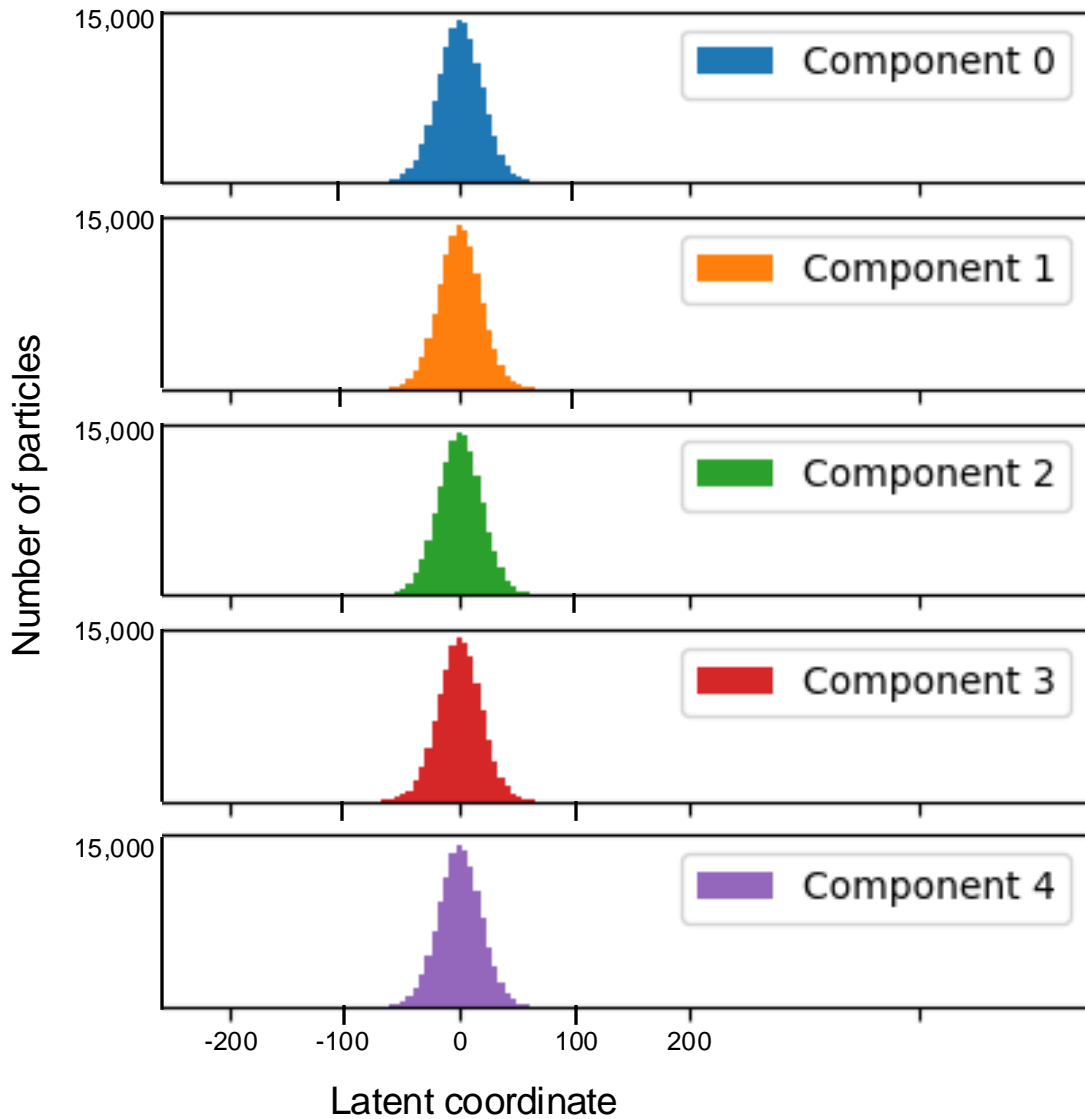
Supplementary Figure 29. Structural comparison in tunnel region between the R142I variant and AS-B.

a Comparison of tunnel residues between WT ASNS (left) and the R142I variant (right).

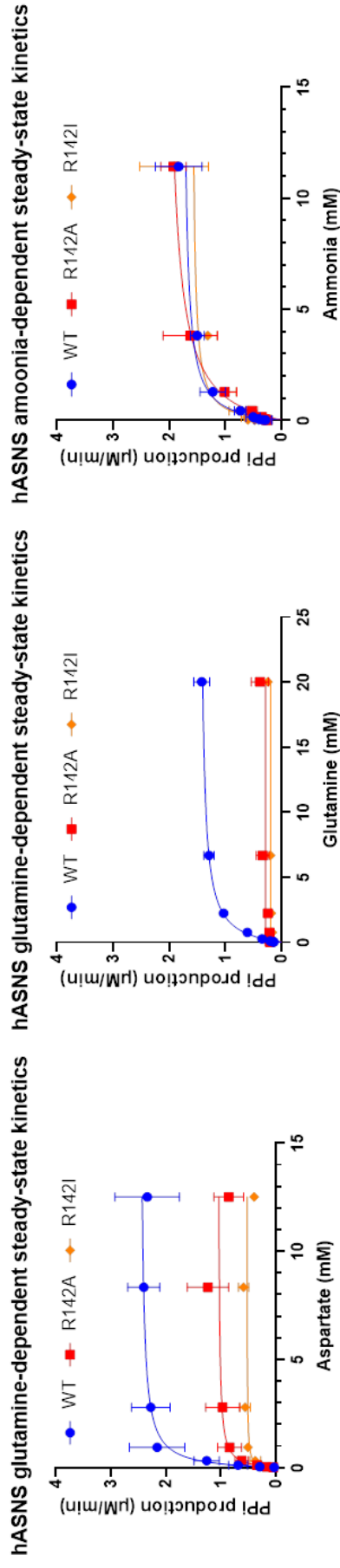
b Comparison of tunnel residues between the R142I variant (left) and AS-B (middle), superimposed (right). R142I variant residues, V141, I142, E364, V401, A404, L415 correspond to the residues of bacterial enzyme AS-B, I142, I143, E352, C385, A388, and A399. Residues of WT ASNS are colored in brown, the R142I variant in sky blue, and AS-B in lime green.



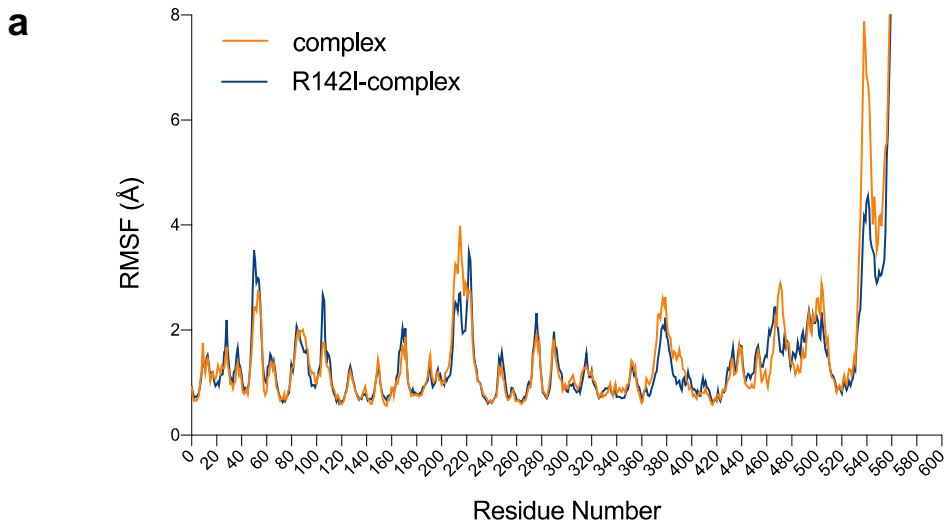
Supplementary Figure 30. PCA-derived structures of the ammonia tunnel in the human R142I variant obtained from 3D variability analysis and 3D refinement. a Side view of critical residues (Val-141, Ile-142, Glu-364, Val-401, Ala-404, Val-414, Leu-415) constituting the ammonia tunnel derived from the consensus EM map. Mutated residue Ile-142 is colored in red. Side view of critical residues constituting ammonia tunnel derived from the variable EM maps derived from 3DVA. Critical residues constituting ammonia tunnel derived from variable maps at frame 1 (left, one end) and frame 20 (middle, the other end) are superimposed (right): b PCA component 0, c PCA component 1, d PCA component 2, e PCA component 3, f PCA component 4.

a**b****Supplementary Figure 31. PCA and particle distributions .**

a 3D representation of particle distribution for the reconstruction of ASNS R142I variant; 2 views at 90° are represented. Red cylinders indicate the amount of particle in each rotation angle for the reconstruction. **b** Particle distribution in latent space, represented per principal component as a function of latent coordinate.

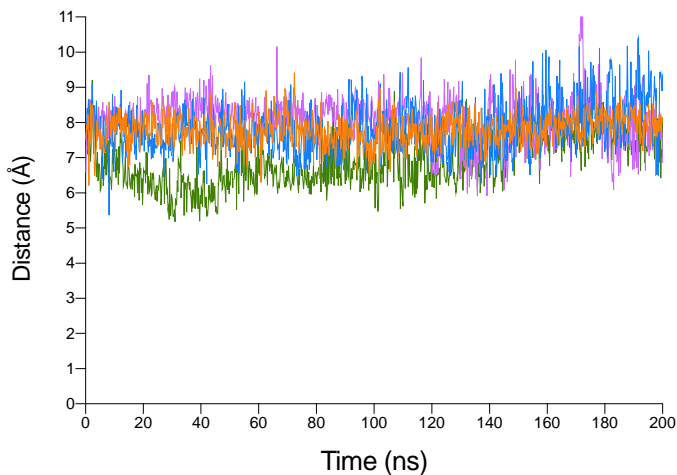


Supplementary Figure 32. Kinetic plots for the steady-state experiments. (Left) Pyrophosphate production vs. L-aspartate concentration (0-12.5 mM), at fixed concentrations of L-glutamine (20 mM), 10 mM MgCl₂, 2 mM dithiothreitol and 5 mM ATP in 100 mM EPPS, pH 8 for WT ASNS and the R142I and R142A ASNS variants. (Middle) Pyrophosphate production vs. L-glutamine concentration (0-20 mM), at fixed concentrations of aspartate (10 mM), 10 mM MgCl₂, 2 mM dithiothreitol and 5 mM ATP in 100 mM EPPS, pH 8 for WT ASNS and the R142I and R142A ASNS variants. (Right) Pyrophosphate production vs. ammonia concentration (0-11.4 mM), at fixed concentrations of L-aspartate (10 mM), 10 mM MgCl₂, 2 mM dithiothreitol and 5 mM ATP in 100 mM EPPS, pH 8 for WT ASNS and the R142I and R142A ASNS variants. Data analysis was performed using GraphPad Prism. Source data are provided as a Source Data file.

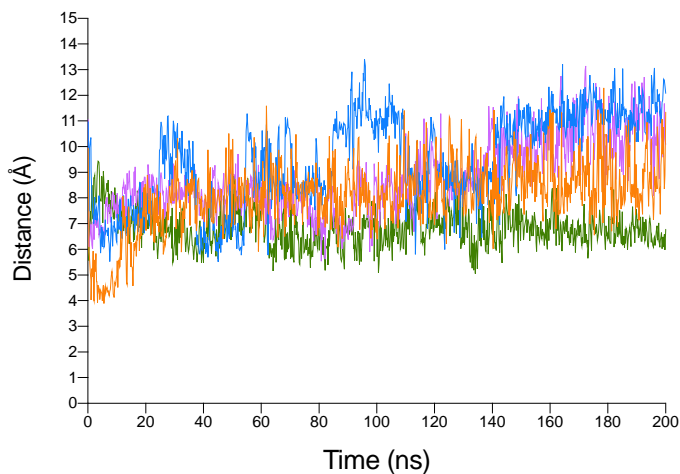


b

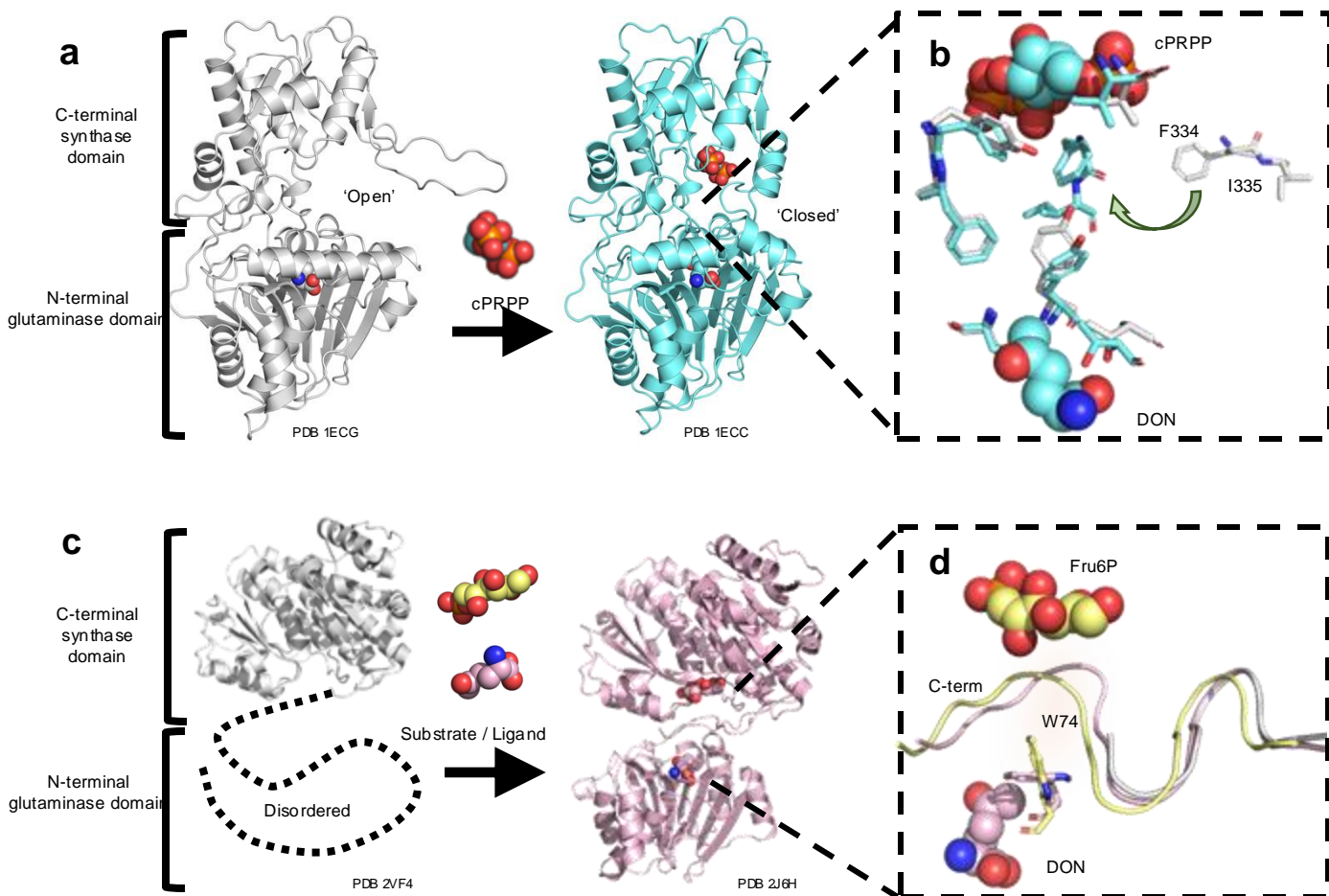
Ile142-Asp405



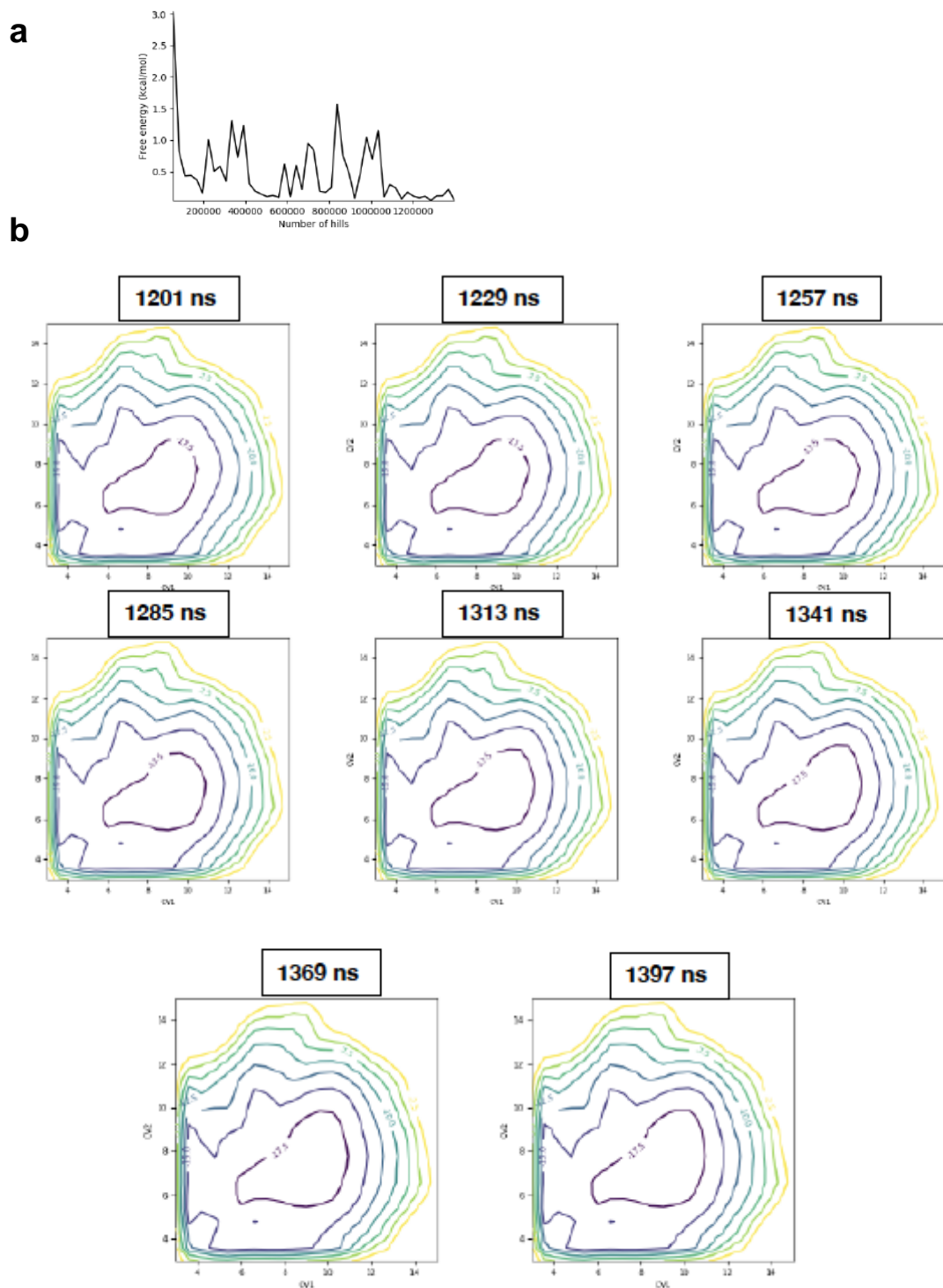
Ile142-Glu414



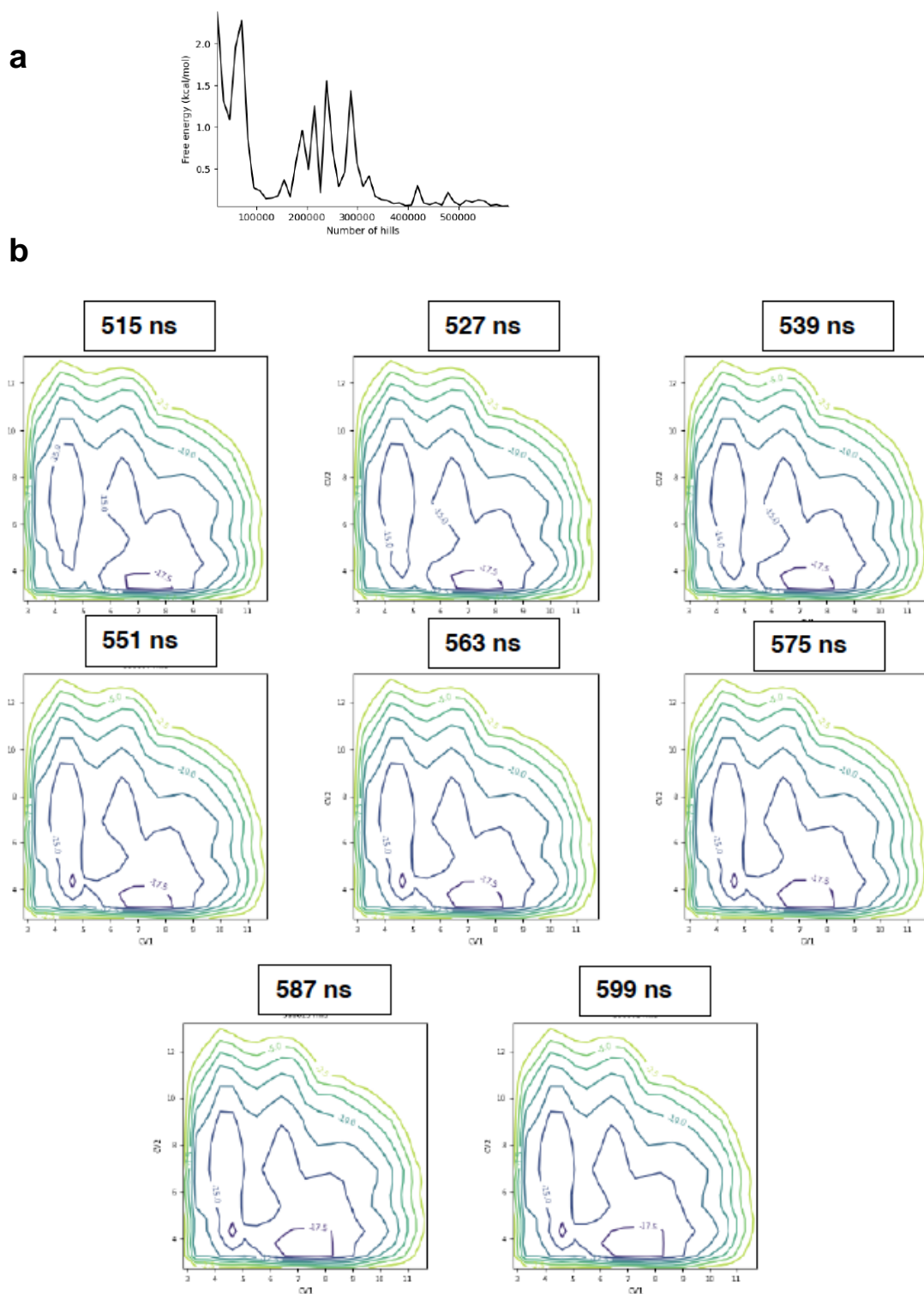
Supplementary Figure 33. 200 ns MD simulations of the R142I/ β -aspartyl-AMP/PP_i ternary complex. **a** Comparison of backbone RMSF values computed for the WT ASNS/ β -aspartyl-AMP/PP_i ternary complex (orange) and the R142I/ β -aspartyl-AMP/PP_i ternary complex (blue). **b** Replicate 200 ns MD simulations of the R142I/ β -aspartyl-AMP/PP_i ternary complex. (left) Plots of the distance between Ile-142 and Asp-405 and (right) between Ile-142 and Glu-414 in the four MD simulation replicates. Distances were calculated between the delta carbon atom of Ile-142 and one of the carboxylate oxygen atoms of Asp-405 or of Glu-414, respectively. See Fig. S17 for the location of these oxygen atoms. Source data are provided as a Source Data file.



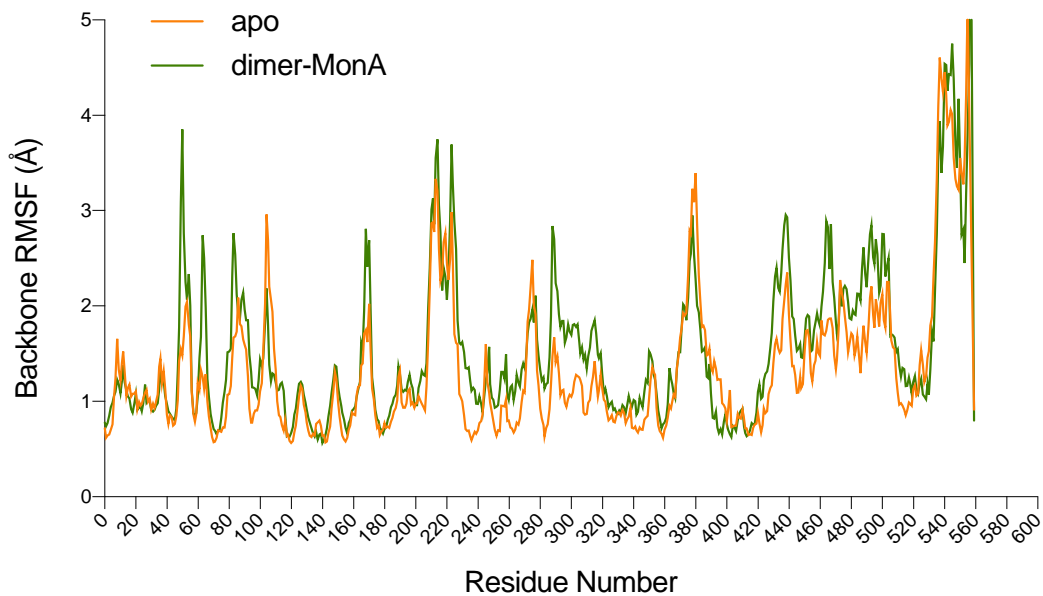
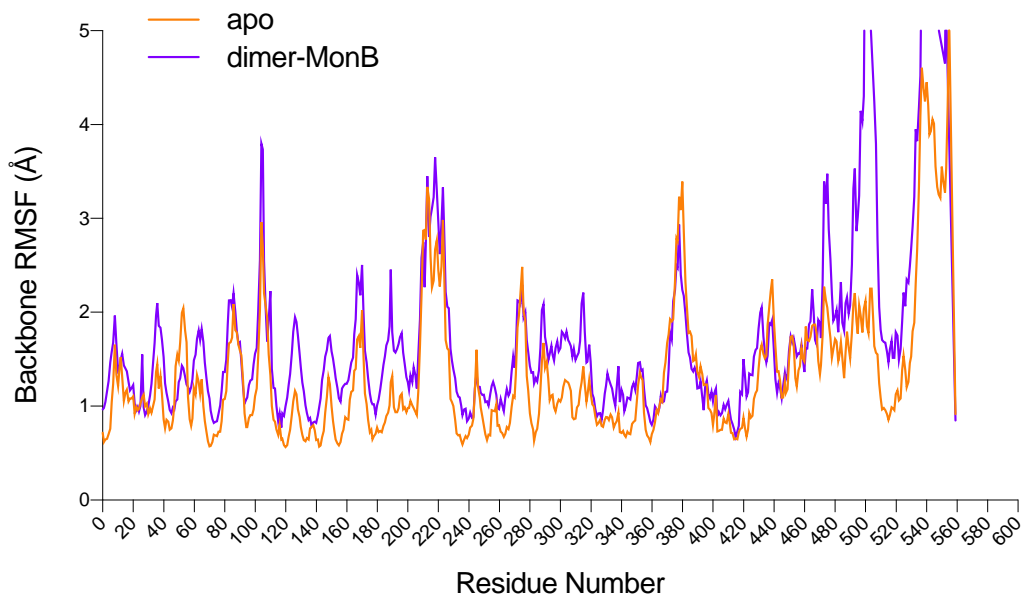
Supplementary Figure 34. **a** cPRPP binding induces a conformational change in an active site loop of the synthase domain of glutamine PRPP amidotransferase (GPA). Cartoon representations of the apo-enzyme (PDB 1ECG) and the complex between cPRPP and DON-modified GPA (PDB 1ECC) are shown in gray and cyan, respectively. **b** When a stable PRPP analog (cPRPP) binds to the DON-modified GPA, the loop residues Phe-334 and Ile-335 reorient (indicated by the green arrow) to create a tunnel connecting the N- and C-terminal active sites (green shading). Residues in the apo-enzyme (PDB 1ECG) and the cPRPP/DON-modified GPA complex (PDB 1ECC) are shown as gray and cyan sticks, respectively. **c** Substrate/ligand binding is needed to stabilize the N-terminal domain of GFAT, which is not resolved in the X-ray crystal structure of the free enzyme (gray) (PDB 2VF4)²¹. When glucose-6-phosphate is present in DON-modified GFAT (light pink), however, both domains of the enzyme are observed (PDB 2J6H). Glucose-6-phosphate and DON are rendered as yellow and light pink spheres, respectively. **d** Close-up of the open intramolecular tunnel in the fructose-6-phosphate/DON-modified GFAT complex (yellow) (PDB 4AMV) formed by rotation of Trp-74. The C-terminal tail is disordered in the free enzyme (gray) but becomes ordered in the two complexes (pink, yellow). Fructose-6-phosphate and DON are rendered as yellow and light pink spheres, respectively.



Supplementary Figure 35. Convergence of the metadynamics-based free energy surface for the Arg-142 side chain in the human apo-ASNS monomer. Two distance-based collective variables were defined for these simulations (Supplementary Fig. S17), which correspond to the distances between the center-of-mass (COM) of the guanidinium group in Arg-142 and the COM of the carboxylate moieties in either Asp-405 (CV1) or Glu-414 (CV2). **a** Distance of free energies as a function of time, **b** Convergence of the free energy surface in the final stages of the MTD simulation.



Supplementary Figure 36. Convergence of the metadynamics-based free energy surface for the Arg-142 side chain in the WT ASNS/ β -aspartyl-AMP/PP_i ternary complex. Two distance-based collective variables were defined for these simulations (Supplementary Fig. S17), which correspond to the distances between the center-of-mass (COM) of the guanidinium group in Arg-142 and the COM of the carboxylate moieties in either Asp-405 (CV1) or Glu-414 (CV2). **a** Distance of free energies as a function of time, **b** Convergence of the free energy surface in the final stages of the MTD simulation.

a**b**

Supplementary Figure 37. Comparison of backbone RMSF values computed from 200 ns MD trajectories of the full-length apo-ASNS monomer and the apo-ASNS dimer. a Values for the apo-ASNS monomer and monomer A in the dimer are shown in orange and green, respectively. **b** Values for the apo-ASNS monomer and monomer B in the dimer are shown in orange and purple, respectively. The deviations in the C-terminal region are not important for the tunnel structure and dynamics. Source data are provided as a Source Data file.

Supplementary Table 2. Average values and standard deviation of nitrogen-oxygen distances (Å) between the Arg-142 side chain and selected residues, as calculated from four 200 ns replicate MD simulations of the human apo-ASNS monomer. Atom names correspond to the PDB naming convention for arginine, aspartate and glutamate residues. The molecular snapshots shown in Fig. 5a correspond to structures sampled from the trajectory of replica MD simulation 1.

WT ASNS	Replica 1	Replica 2	Replica 3	Replica 4
Asn-74				
OD1-NE	4.5 ± 1.1	5.8 ± 1.1	6.1 ± 0.9	6.1 ± 2.3
OD1-NH1	6.2 ± 1.0	6.1 ± 1.1	7.9 ± 0.9	8.0 ± 2.1
OD1-NH2	6.2 ± 1.6	7.3 ± 1.1	8.0 ± 1.0	7.8 ± 2.3
Glu-76				
OE1-NE	9.4 ± 1.0	7.7 ± 0.8	7.5 ± 1.6	6.5 ± 0.9
OE2-NE	9.5 ± 1.2	7.5 ± 0.8	8.1 ± 1.5	6.5 ± 1.0
OE1-NH1	8.4 ± 0.9	7.7 ± 1.0	6.6 ± 1.8	7.4 ± 1.2
OE2-NH1	8.5 ± 1.1	7.5 ± 1.0	7.6 ± 1.3	7.4 ± 1.4
OE1-NH2	9.2 ± 1.3	6.2 ± 0.8	7.7 ± 1.5	5.9 ± 1.1
OE2-NH2	9.3 ± 1.4	6.0 ± 0.8	8.6 ± 1.2	5.9 ± 1.1
Asp-405				
OD1-NE	7.2 ± 0.9	6.5 ± 0.4	4.0 ± 0.3	7.2 ± 0.9
OD2-NE	6.5 ± 1.1	8.2 ± 0.4	5.1 ± 0.4	7.7 ± 0.9
OD1-NH1	8.9 ± 1.3	5.4 ± 0.5	5.0 ± 0.3	7.6 ± 1.0
OD2-NH1	8.2 ± 0.9	7.5 ± 0.5	6.6 ± 0.4	8.0 ± 0.9
OD1-NH2	7.4 ± 1.1	7.4 ± 0.4	2.8 ± 0.4	8.5 ± 1.0
OD2-NH2	6.6 ± 1.0	9.5 ± 0.4	4.5 ± 0.5	9.0 ± 1.0
Glu-414				
OE1-NE	6.3 ± 1.0	4.8 ± 0.2	6.0 ± 1.0	4.6 ± 1.0
OE2-NE	6.7 ± 0.8	4.8 ± 0.2	6.7 ± 0.8	4.6 ± 1.0
OE1-NH1	6.7 ± 1.4	2.8 ± 0.3	6.2 ± 1.1	5.3 ± 1.1
OE2-NH1	7.1 ± 1.0	3.5 ± 0.4	7.3 ± 0.9	5.0 ± 1.1
OE1-NH2	6.0 ± 1.6	3.5 ± 0.2	5.2 ± 1.1	3.9 ± 0.9
OE2-NH2	6.4 ± 1.2	2.7 ± 0.2	6.3 ± 0.9	3.5 ± 0.1

Supplementary Table 3. Shortest nitrogen-oxygen distances (Å) between the Arg-142 side chain and selected residues in the snapshots (Fig. 5a) obtained from the 200 ns MD simulation of the human apo-ASNS monomer. Atom names correspond to the PDB naming convention for arginine, aspartate and glutamate residues.

WT ASNS	20 ns	80 ns	140 ns	200 ns
Asn-74	2.7 (OD1-NH2)	4.4 (OD1-NE)	5.4 (OD1-NE)	7.2 (OD1-NE)
Glu-76	7.0 (OE1-NH2)	7.1 (OE1-NH1)	8.7 (OE1-NH1)	7.1 (OE1-NH1)
Asp-405	5.7 (OD1-NH2)	6.9 (OD1-NH2)	5.7 (OD2-NE)	5.4 (OD2-NE)
Glu-414	2.5 (OE1-NH2)	7.2 (OE1-NH2)	5.5 (OE1-NH2)	5.0 (OE1-NE)

Supplementary Table 4. Average values and standard deviation of nitrogen-oxygen distances (Å) between the Arg-142 side chain and selected residues, as calculated from four 200 ns replicate MD simulations of the the human ASNS/ β -aspartyl-AMP/MgPP_i ternary complex. Atom names correspond to the PDB naming convention for arginine, aspartate and glutamate residues. The molecular snapshots shown in Fig. 5b correspond to structures sampled from the trajectory of replica MD simulation 1.

Complex	Replica 1	Replica 2	Replica 3	Replica 4
Asn-74				
OD1-NE	4.8 ± 1.7	5.9 ± 1.8	5.2 ± 0.9	5.1 ± 0.9
OD1-NH1	6.8 ± 1.5	8.0 ± 1.7	6.9 ± 1.1	6.8 ± 1.1
OD1-NH2	6.5 ± 2.0	7.8 ± 2.0	6.5 ± 1.2	7.0 ± 0.9
Glu-76				
OE1-NE	10.4 ± 1.4	8.2 ± 1.0	5.4 ± 1.5	6.9 ± 1.9
OE2-NE	10.4 ± 1.3	8.9 ± 1.3	5.5 ± 1.6	6.8 ± 1.9
OE1-NH1	10.1 ± 1.4	10.0 ± 1.1	6.4 ± 1.5	8.0 ± 1.9
OE2-NH1	10.1 ± 1.2	10.6 ± 1.5	6.6 ± 1.7	7.9 ± 2.0
OE1-NH2	11.0 ± 2.0	8.3 ± 1.0	4.5 ± 1.8	6.1 ± 2.0
OE2-NH2	11.0 ± 1.8	8.9 ± 1.4	4.7 ± 2.0	6.0 ± 2.0
Asp-405				
OD1-NE	4.5 ± 0.7	4.7 ± 0.6	6.7 ± 1.1	4.1 ± 0.3
OD2-NE	5.3 ± 0.9	6.0 ± 0.4	8.2 ± 1.1	6.0 ± 0.3
OD1-NH1	4.6 ± 1.4	4.9 ± 0.6	6.0 ± 1.3	3.0 ± 0.5
OD2-NH1	5.6 ± 1.9	5.6 ± 0.3	7.4 ± 1.2	5.0 ± 0.4
OD1-NH2	3.5 ± 1.3	5.9 ± 0.6	8.0 ± 1.3	4.9 ± 0.4
OD2-NH2	4.3 ± 1.8	6.9 ± 0.5	9.3 ± 1.4	7.0 ± 0.3
Glu-414				
OE1-NE	5.7 ± 1.1	3.7 ± 0.8	5.3 ± 0.8	6.3 ± 1.5
OE2-NE	5.9 ± 1.0	4.3 ± 0.6	5.8 ± 1.3	6.2 ± 1.5
OE1-NH1	6.0 ± 1.1	5.0 ± 0.7	5.1 ± 1.1	6.6 ± 1.4
OE2-NH1	6.3 ± 0.8	5.9 ± 0.7	5.7 ± 2.1	6.6 ± 1.3
OE1-NH2	5.3 ± 1.4	3.1 ± 0.8	5.0 ± 1.4	5.3 ± 1.6
OE2-NH2	5.7 ± 1.0	4.1 ± 0.9	5.4 ± 2.4	5.3 ± 1.6

Supplementary Table 5. Shortest nitrogen-oxygen distances (Å) between the Arg-142 side chain and selected residues in the snapshots (Fig. 5b) obtained from the 200 ns MD simulation of the ASNS/ β -aspartyl-AMP/MgPP_i ternary complex. Atom names correspond to the PDB naming convention for arginine, aspartate and glutamate residues.

Complex	20 ns	80 ns	140 ns	200 ns
Asn-74	7.4 (OD1-NE)	6.2 (OD1-NE)	4.4 (OD1-NE)	3.3 (OD1-NH2)
Glu-76	9.1 (OE2-NH1)	10.1 (OE2-NH1)	9.0 (OE1-NH1)	7.9 (OE2-NH2)
Asp-405	5.0 (OD1-NE)	2.7 (OD1-NH2)	2.7 (OD2-NH2)	2.6 (OD1-NH1)
Glu-414	2.7 (OE1-NH2)	4.6 (OE1-NE)	5.2 (OE2-NH2)	3.9 (OE2-NH2)

Supplementary Table 6. Supplementary Movies and Associated DOIs.

Movie	Content	DOI
1	Conformational changes seen in the 3DVA-generated structures for WT apo-ASNS along principal component 0	10.5281/zenodo.14026196
2.	Conformational changes seen in the 3DVA-generated structures for WT apo-ASNS along principal component 1	10.5281/zenodo.14026244
3	Conformational changes seen in the 3DVA-generated structures for WT apo-ASNS along principal component 2	10.5281/zenodo.14026249
4	Conformational changes seen in the 3DVA-generated structures for WT apo-ASNS along principal component 3	10.5281/zenodo.14026276
5	Conformational changes seen in the 3DVA-generated structures for WT apo-ASNS along principal component 4	10.5281/zenodo.14026295
6	Conformational changes seen in the 3DVA-generated structures for the R142I ASNS along principal component 0	10.5281/zenodo.14028720
7	Conformational changes seen in the 3DVA-generated structures for the R142I ASNS along principal component 1	10.5281/zenodo.14028728
8	Conformational changes seen in the 3DVA-generated structures for the R142I ASNS along principal component 2	10.5281/zenodo.14028732
9	Conformational changes seen in the 3DVA-generated structures for the R142I ASNS along principal component 3	10.5281/zenodo.14028736
10	Conformational changes seen in the 3DVA-generated structures for the R142I ASNS along principal component 4	10.5281/zenodo.14028740

Supplementary Table 6 (continued). Supplementary Movies and Associated DOIs.

Movie	Content	DOI
11	Movements of tunnel residues in WT apo-ASNS for the replicate MD simulation used to generate Fig. 5A	10.5281/zenodo.14028743
12	Movements of tunnel residues in the WT ASNS/ β -aspartyl-AMP/MgPP _i ternary complex for the replicate MD simulation used to generate Fig. 5B	10.5281/zenodo.14028749
13	Movements of tunnel residues in the R142I ASNS variant for the replicate MD simulation used to generate Fig. 6A	10.5281/zenodo.14028757
14	Movements of tunnel residues in the R142I/ β -aspartyl-AMP/MgPP _i ternary complex for the replicate MD simulation used to generate Fig. 6B	10.5281/zenodo.14028763

Supplementary Table 7. PROPKA-based residue ionization states.

	WT	R142A	R142I
CYS_1	-1	-1	-1
ASP_10	-1	-1	-1
ASP_11	-1	-1	-1
LYS_22	1	1	1
HIS_25	0	0	0
ARG_26	1	1	1
ASP_29	-1	-1	-1
ARG_32	1	1	1
GLU_34	-1	-1	-1
HIS_47	0	0	0
ARG_48	1	1	1
ASP_53	-1	-1	-1
ARG_62	1	1	1
LYS_64	1	1	1
LYS_65	1	1	1
GLU_76	-1	-1	-1
HIS_80	0	0	0
LYS_81	1	1	1
LYS_82	1	1	1
HIS_86	0	0	0
GLU_88	-1	-1	-1
GLU_90	-1	-1	-1
LYS_94	1	1	1
ASP_96	-1	-1	-1
GLU_98	-1	-1	-1
HIS_102	0	0	0
ASP_105	-1	-1	-1
LYS_106	1	1	1
GLU_110	-1	-1	-1
ASP_117	-1	-1	-1
ASP_126	-1	-1	-1
LYS_130	1	1	1
LYS_131	1	1	1
ARG_136	1	1	1
ASP_137	-1	-1	-1
ARG_142	1	-	-
LYS_146	1	1	1

	WT	R142A	R142I
GLU_150	-1	-1	-1
ASP_151	-1	-1	-1
GLU_159	-1	-1	-1
LYS_161	1	1	1
LYS_167	1	1	1
HIS_168	0	0	0
LYS_175	1	1	1
GLU_177	-1	-1	-1
HIS_183	0	0	0
GLU_185	-1	-1	-1
ASP_188	-1	-1	-1
LYS_190	1	1	1
LYS_194	1	1	1
GLU_199	-1	-1	-1
LYS_202	1	1	1
HIS_204	0	0	0
HIS_205	0	0	0
ARG_207	1	1	1
ASP_208	-1	-1	-1
HIS_212	0	0	0
ASP_216	-1	-1	-1
GLU_219	-1	-1	-1
LYS_220	1	1	1
GLU_226	-1	-1	-1
GLU_228	-1	-1	-1
LYS_231	1	1	1
ARG_235	1	1	1
LYS_243	1	1	1
LYS_244	1	1	1
ARG_245	1	1	1
ASP_249	-1	-1	-1
ARG_250	1	1	1
ARG_251	1	1	1
ASP_261	-1	-1	-1
LYS_271	1	1	1
LYS_274	1	1	1
GLU_275	-1	-1	-1

Supplementary Table 7 (continued). PROPKA-based residue ionization states.

	WT	R142A	R142I
GLU_290	-1	-1	-1
ASP_291	-1	-1	-1
ASP_294	-1	-1	-1
ARG_299	1	1	1
LYS_300	1	1	1
ASP_303	-1	-1	-1
HIS_304	0	0	0
GLU_308	-1	-1	-1
HIS_309	0	0	0
GLU_311	-1	-1	-1
GLU_317	-1	-1	-1
GLU_318	-1	-1	-1
ASP_324	-1	-1	-1
GLU_325	-1	-1	-1
GLU_331	-1	-1	-1
ASP_334	-1	-1	-1
ARG_339	1	1	1
LYS_349	1	1	1
ARG_352	1	1	1
LYS_353	1	1	1
ASP_356	-1	-1	-1
GLU_364	-1	-1	-1
ASP_367	-1	-1	-1
GLU_368	-1	-1	-1
HIS_377	0	0	0
LYS_378	1	1	1
GLU_383	-1	-1	-1
LYS_384	1	1	1
GLU_386	-1	-1	-1
GLU_387	-1	-1	-1
GLU_388	-1	-1	-1
GLU_390	-1	-1	-1
ARG_391	1	1	1
ARG_394	1	1	1
GLU_395	-1	-1	-1
ASP_400	-1	-1	-1
ARG_403	1	1	1

	WT	R142A	R142I
ASP_405	-1	-1	-1
ARG_406	1	1	1
HIS_411	0	0	0
GLU_414	-1	-1	-1
ARG_416	1	1	1
ASP_421	-1	-1	-1
HIS_422	0	0	0
ARG_423	1	1	1
GLU_434	-1	-1	-1
ARG_436	1	1	1
LYS_439	1	1	1
GLU_443	-1	-1	-1
LYS_444	1	1	1
HIS_445	0	0	0
ARG_448	1	1	1
GLU_449	-1	-1	-1
GLU_452	-1	-1	-1
ASP_453	-1	-1	-1
LYS_459	1	1	1
GLU_460	-1	-1	-1
ARG_464	1	1	1
LYS_466	1	1	1
GLU_467	-1	-1	-1
ASP_471	-1	-1	-1
LYS_477	1	1	1
LYS_482	1	1	1
GLU_486	-1	-1	-1
GLU_489	-1	-1	-1
HIS_490	0	0	0
ASP_493	-1	-1	-1
ASP_494	-1	-1	-1
LYS_503	1	1	1
LYS_510	1	1	1
LYS_512	1	1	1
GLU_513	-1	-1	-1
ARG_518	1	1	1
GLU_522	-1	-1	-1

Supplementary Table 7 (continued). PROPKA-based residue ionization states.

	WT	R142A	R142I
ARG_523	1	1	1
HIS_524	0	0	0
ARG_528	1	1	1
ASP_530	-1	-1	-1
HIS_534	1	1	1
LYS_539	1	1	1
ASP_545	-1	-1	-1
ARG_549	1	1	1
HIS_553	0	0	0
LYS_555	1	1	1
LYS_559	1	1	1

Supplementary Table 8. Raw Data Files in the Zenodo Repository and Associated DOIs.

Entry	Content	DOI
1	All EM-associated data for the dimeric apo-form of WT ASNS used to generate Figs. 1 and 2, and Supplementary Figs. 6-8	10.5281/zenodo.14029288
2.	All EM-associated data for the dimeric apo-form of the R142I ASNS variant used to generate Supplementary Figs. 26-29	10.5281/zenodo.14029280
3	All 3DVA-associated data for the dimeric apo-form of WT ASNS	10.5281/zenodo.14029380
4	All 3DVA-associated data for the dimeric apo-form of the R142I ASNS variant	10.5281/zenodo.14031045
5	Coordinates for all MD trajectories of the monomeric apo-form of WT ASNS	10.5281/zenodo.14032047
6	Coordinates for the MD trajectory of the dimeric apo-form of WT ASNS	10.5281/zenodo.14029294
7	Coordinates for all MD trajectories of the monomeric WT ASNS/ β -aspartyl-AMP/MgPP _i ternary complex	10.5281/zenodo.14031986
8	Coordinates for all MD trajectories of the monomeric apo-form of the R142I ASNS variant and the monomeric R142I/ β -aspartyl-AMP/MgPP _i ternary complex	10.5281/zenodo.14029317
9	Coordinates for the metadynamics trajectory of the monomeric apo-form of WT ASNS used to generate Supplementary Fig. 35	10.5281/zenodo.14042469
10	Coordinates for the metadynamics trajectory of the monomeric WT ASNS/ β -aspartyl-AMP/MgPP _i ternary complex used to generate Supplementary Fig. 36	10.5281/zenodo.14042474