

OPEN ACCESS Check for updates

Updates on radiotherapy-immunotherapy combinations: Proceedings of 8th Annual ImmunoRad Conference

Fereshteh Talebi^a, Fabiana Gregucci ^{ba}, Jalal Ahmed^b, Nir Ben Chetrit^a, Brian D. Brown^b, Timothy A. Chan ^{bc,d}, Dhan Chand^e, Julie Constanzo^f, Sandra Demaria^a, Dmitry I. Gabrilovich^g, Encouse Golden^a, Andrew Godkin^h, Chandan Guhaⁱ, Gaorav P. Gupta^j, Aisha Hasan^k, Fernanda G. Herrera^{Lm,n}, Howard Kaufman^o, Donna Li^p, Alan A. Melcher^q, Sierra McDonald^r, Taha Merghoub^{s,t}, Arta M. Monjazeb^u, Sébastien Paris^v, Sean Pitroda^w, Anguraj Sadanandam[×], Dörthe Schaue^y, Laura Santambrogio^a, Phillippe Szapary^z, Julien Sage^{aa}, James W. Welsh^{bb}, Anna Wilkins^q, Kristina H. Young^{cc,dd}, Eric Wennerberg^q, Laurence Zitvogel^{ee}, Lorenzo Galluzzi^{ff}, Eric Deutsch⁹⁹, and Silvia C. Formenti^a

^aDepartment of Radiation Oncology, Weill Cornell Medicine, New York, NY, USA; ^bIcahn Genomics Institute, Department of Immunology and Immunotherapy, Icahn School of Medicine at Mount Sinai, New York, NY, USA; Department of Cancer Sciences, Global Center for Immunotherapy and Precision Immuno-Oncology, Cleveland Clinic, Cleveland, OH, USA; ^dCase Western University School of Medicine, Cleveland, OH, USA; ^eAgenus Inc, Lexingston, MA, USA; ^fInstitut de Recherche en Cancérologie de Montpellier (IRCM), INSERM U1194, Université de Montpellier, Institut régional du Cancer de Montpellier (ICM), Montpellier, France; ⁹AstraZeneca, Gaithersburg, MD, USA; ^hDivision of Infection and Immunity/Systems Immunity University Research Institute, School of Medicine, Cardiff University, Cardiff, UK; ⁱDepartments of Radiation Oncology and Pathology, Albert Einstein College of Medicine, New York, NY, USA; ^jDepartment of Radiation Oncology, Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, NC, USA; * Johnson & Johnson, New Brunswick, NJ, USA; AGORA Cancer Research Center, Swiss Cancer Center Leman, Lausanne, Switzerland; Services of Radiation Oncology and Immuno-Oncology, Centre Hospitalier Universitaire Vaudois, Lausanne, Switzerland; "Department of Oncology, Ludwig Institute of Cancer Research, University of Lausanne, Lausanne, Switzerland; ^oAnkyra Therapeutics, Cambridge, MA, USA; ^pUniversity of Wisconsin, Madison, WI, USA; ^qDivision of Radiotherapy and Imaging, The Institute of Cancer Research, London, UK; UNC Lineberger Comprehensive Cancer Center, University of North Carolina, Chapel Hill, NC, USA; ^sSwim Across America and Ludwig Collaborative Laboratory, Department of Pharmacology, Weill Cornell Medicine, New York, NY, USA; 'Sandra and Edward Meyer Cancer Center and Parker Institute for Cancer Immunotherapy, Weill Cornell Medicine, New York, NY, USA; "Department of Radiation Oncology, University of California, San Diego, CA, USA; "Nanobiotix, Paris, France; "Department of Radiation and Cellular Oncology and Ludwig Center for Metastasis Research, University of Chicago, Chicago, IL, USA; *Division of Molecular Pathology, The Institute of Cancer Research, London, UK; *Department of Radiation Oncology, David Geffen School of Medicine at UCLA, Los Angeles, CA, USA; ^zInterventional Oncology, Johnson & Johnson, New Brunswick, NJ; ^{aa}Departments of Genetics and Pediatrics, Stanford University, Stanford, California; bbDepartment of Radiation Oncology, MD Anderson Cancer Center, University of Texas, Houston, TX, USA; "Division of Radiation Oncology, The Oregon Clinic, Portland, OR, USA; de Earle A. Chiles Research Institute, Providence Cancer Institute, Portland, OR, USA; eeGustave Roussy, INSERM U1015, Division of Medicine, Paris-Saclay University, Center of Clinical Investigations BIOTHERIS, Villejuif, France; "Cancer Signaling and Microenvironment Program, Fox Chase Cancer Center, Philadelphia, PA, USA; 99Department of Radiation Oncology, Gustave Roussy, INSERM U1030, Division of Medicine, Paris-Saclay University, RHU LySAIRI "Lymphocyte-Sparing Artificial Intelligence-quided Radio-Immunotherapy", Villejuif, France

ABSTRACT

The annual ImmunoRad Conference has established itself as a recurrent occasion to explore the possibility of combining radiation therapy (RT) and immunotherapy (IT) for clinical cancer management. Bringing together a number of preclinical and clinical leaders in the fields of radiation oncology, immuno-oncology and IT, this annual event fosters indeed essential conversations and fruitful exchanges on how to address existing challenges to expand the therapeutic value of RT-IT combinations. The 8th edition of the ImmunoRad Conference, which has been held in October 2024 at the Weill Cornell Medical College of New York City, highlighted exciting preclinical and clinical advances at the interface between RT and IT, setting the stage for extra progress toward extended benefits for patients with an increasing variety of tumor types. Here, we critically summarize the lines of investigation that have been discussed at the occasion of the 8th Annual ImmunoRad Conference.

Introduction

Over the past decade, considerable efforts have been dedicated to the development of safe and effective combinatorial regimens involving radiation therapy (RT) and immunotherapy (IT) for clinical cancer management, building on the notion that – at least when employed according to specific dose and fractionation schedules, and when delivered focally to limited target volumes that do not involve tumor-draining lymph nodes (TDLNs) and/or considerable amounts of circulating lymphocytes – RT can mediate robust immunostimulatory effects.^{1–4} Importantly, while a few randomized clinical trials demonstrated that RT indeed can be safely and effectively combined with IT in specific oncological settings, *e.g.*, patients with stage III, unresectable non-small-cell lung carcinoma

CONTACT Eric Deutsch eric.deutsch@gustaveroussy.fr Department of Radiation Oncology, Gustave Roussy, INSERM U1030, Division of Medicine, Paris-Saclay University, RHU LySAIRI "Lymphocyte-Sparing Artificial Intelligence-guided Radio Immunotherapy", 114 Rue Édouard Vaillant, Villejuif 94805, France; Silvia C. Formenti formenti@med.cornell.edu Department of Radiation Oncology, Weill Cornell Medicine, 525 East 68th Street, New York, NY 10065, USA

© 2025 The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The terms on which this article has been published allow the posting of the Accepted Manuscript in a repository by the author(s) or with their consent.

ARTICLE HISTORY

Received 23 April 2025 Revised 12 May 2025 Accepted 13 May 2025

KEYWORDS

Abscopal effect; CAR T cells; CD8⁺ cytotoxic T lymphocytes; low-dose radiation therapy; T_{REG} cells; tumor-associated macrophages (NSCLC) receiving an immune checkpoint inhibitor (ICI) specific for CD274 (best known as PD-L1) after chemoradiotherapy,⁵ many others failed to document any clinical benefit from the addition of ICIs to RT employed according to standard-of-care (SOC),^{6–8} pointing to the existence of numerous obstacles against the widespread applicability of RT-IT combinations in the clinic.^{1,9,10}

The ImmunoRad conference has been jointly established by Dr. Silvia Formenti from the Weill Cornell Medical College (New York, US) and Dr. Eric Deutsch from Gustave Roussy (Paris, France) in 2016, with the specific aim to understand the nature of – and hence ultimately circumvent – such obstacles, de facto fostering progress at the interface between RT and IT to achieve superior clinical outcome for an ever increasing number of patients with cancer. Since then, with the only exception of year 2020 as imposed by the COVID-19 pandemic,^{11,12} the conference has been held annually, alternating between New York and Paris, attracting an ever wider and more diverse group of attendants.^{13,14}

The 8th Annual ImmunoRad Conference has been held on October 3rd to 5th, 2024, at the Weill Cornell Medical College in New York City, continuing such a tradition of excellence. This year, the conference welcomed no less than 430 participants from over 19 different countries worldwide and featured more than 30 thought-provoking presentations from established and emergent scientists working on RT-IT combinations, encompassing an opening Keynote lecture from Dr. Laurence Zitvogel (Gustave Roussy), 4 lectures delivered as part of a CME-accredited primer on the ability of RT to alter the immunological tumor microenvironment (TME) plus 21 main talks, 4 short talks selected from abstracts, as well as a closing panel discussion with 4 panelists from industry. Alongside, the event hosted more than 25 poster presentations by trainees and young researchers cultivating an interest in the possibility to achieve superior therapeutic outcomes by combining RT and IT. Following up on previous editions of the event, outstanding contributions from young investigators in the fields were recognized with travel awards and poster prizes. The conference program incorporated discussions on tumorhost interactions, predictive biomarkers for patient stratification, and innovative clinical trial design, overall providing an interactive platform for sharing new knowledge and fostering promising research and clinical collaborations. For how the ImmunoRad Conference is built, all attendees enjoyed the chance to directly engage with key opinion leaders, facilitating in-depth discussions on the most pressing challenges and opportunities in RT-IT combinations.

Here, we report on the main topics that have been discussed at the occasion of the 8th Annual ImmunoRad Conference in New York City as we set the stage for the 9th installment of this exciting event, which will be held in Paris, on September 17th to 19th, 2025.

The dirty secrets of radiation oncology

As part of the opening Keynote lecture, Prof. Laurence Zitvogel (Gustave Roussy) provided critical insights into the influence of the gut microbiota – i.e., the bacterial, viral and fungal communities populating the intestinal tract¹⁵ – on RT and IT

outcomes in patients with cancer.¹⁶⁻¹⁸ Over the past decade, pivotal work from Dr. Zitvogel and others underscored indeed the critical impact of the gut microbiome on the systemic immune set point, affecting the sensitivity of various tumor types to various therapeutic interventions.¹⁹⁻²² In this context, microbiota-targeting strategies such as fecal microbiota transplantation (FMT) and probiotics were highlighted as effective approaches to overcome treatment resistance and improve disease outcome.²³⁻²⁶ Moreover, Dr. Zitvogel presented compelling evidence supporting the value of intestinal low-dose radiotherapy (ILDR) as a combinatorial partner for stereotactic body radiotherapy (SBRT) and ICIs in metastatic pancreatic ductal adenocarcinoma (PDAC).²⁷ Mechanistically, ILDR appears indeed to elicit immunogenic changes in the gut microbiome that result in a reduced development of gutderived regulatory T (T_{REG}) cells and improved tumor infiltration by non-exhausted CD8⁺ T cells.²⁷ These findings point to ILDR as to safe and effective strategy to modulate the gut microbiome in support of superior disease outcome in patients with cancer receiving RT-IT combinations.

Radiation therapy-induced modulation of the tumor immune microenvironment

The first main session of the 8th annual Immunorad Conference was CME-accredited Primer on the capacity of RT to influence the TME.

Dr. James W. Welsh (MD Anderson Cancer Center) discussed strategies to overcome resistance to IT in patients with cancer through the so-called "radscopal effect", i.e., the ability of low-dose RT (LDRT) to reprogram the TME in favor of tumor-targeting immune responses.^{28,29} In this context, Dr. Welsh emphasized the importance of optimizing radiation doses to activate specific immune cell populations, notable natural killer (NK) cells with low-dose radiation and CD8⁺ cytotoxic T lymphocytes (CTLs) with high-dose RT (HDRT). Moreover, he presented data supporting the notion that a beneficial increase in the intratumoral CTL/T_{REG} cell ratio requires the early administration of pembrolizumab, an ICI specific for programmed cell death 1 (PDCD1, best known as PD-1), coupled with stereotactic body radiotherapy (SBRT) optionally along with functionalized hafnium oxide nanoparticles or C-C motif chemokine receptor 8 (CCR8)-targeting strategies.30,31

Dr. Fernanda G. Herrera (University of Lausanne) followed up on Dr. Welsh presentation by providing additional data on the promise of using LDRT as a strategy to recruit immune effector cells, notably $CD8^+$ CTLs, to the TME in support of superior therapeutic responses to ICIs.^{32,33} Her team found that such an beneficial effect can be elicited by RT doses as low as one fraction of 1 Gy in preclinical models of ovarian cancer, resulting in improved intratumoral CTL/T_{REG} cell ratio and the local expression of pro-inflammatory cytokines and costimulatory molecules.³⁴ Alongside, Dr. Herrera presented results from the RACIM trial, which tested a combination of LDRT plus ICIs, cyclophosphamide and prostaglandin E synthase 2 (PTGES2, best known as COX2) inhibitors in patients with solid tumors exhibiting poor lymphocytic infiltration. In this clinical study, responders experienced an increase in intratumoral CD8⁺ CTLs, whereas the TME of nonresponders appeared to promote the accumulation of macrophages, which are generally associated with local immunosuppression.³⁵ These findings highlight the clinical potential of LDRT as a strategy to optimize IT outcomes.

Dr. Eric Deutsch (Gustave Roussy) further explored the importance of RT dose in the context of RT-IT combinations. Indeed, while SBRT can effectively control tumor growth in a variety of settings, and (at least in some cases) elicit immunological alterations that can be harnessed with ICIs,³⁶ it cannot always be employed in a safe manner (for instance when target volumes are in the close proximity to an organ at risk). In the case of metastatic disease, LDRT may hence offer a complementary strategy to HDRT for lesions that are ineligible to the latter, resulting in superior systemic disease control in the context of IT with ICIs. Dr. Deutsch also emphasized the notion that spatially varied RT doses create considerable degrees of immune heterogeneity in the TME, with areas enriched in CD8⁺ CTLs alongside regions abundantly infiltrated by with monocytes and neutrophils.³⁷ Importantly, targeting these immunosuppressive myeloid cell populations was shown to improve tumor control in preclinical models of colorectal and breast cancer, de facto compensating for the regional detrimental effects of heterogeneous RT doses.³⁷ In conclusion, the presentation by Dr. Deutsch reinstated the need for further research into optimal dosing, treatment intervals, and irradiation areas for improved therapeutic outcomes in patients receiving RT plus IT.38

Dr. Lorenzo Galluzzi (Weill Cornell Medical College) summarized an abundant preclinical and clinical literature demonstrating that - depending on a number of RT-related, cancerrelated and host-related variables - RT can have both beneficial and detrimental effects on lymphocytes and their interactions with malignant cells, which has major implications for the development of effective RT-IT combination. To exemplify this concept, Dr. Galluzzi alluded to the fact that while circulating lymphocytes including T and B cells are highly sensitive to the cytotoxic effects of RT, intratumoral CD8⁺ CTLs, especially tissue-resident CD8⁺ T cells, persist upon RT in support of tumor-targeting immunity.³⁹ Moreover, he commented on the ability of low-dose total body irradiation (TBI) delivered before CAR T cell infusion to improve the therapeutic effects of the latter in preclinical models of leukemia and pancreatic cancer,^{40,41} and presented unpublished results from his team demonstrating that hypofractionated RT in 3 doses of 10 Gy each prevents CDK4/6 inhibitors from eliciting the secretion of C-C motif chemokine ligand 2 (CCL2) in a uniquely translational model of HR^+ breast cancer,^{42–45} thus impeding the recruitment of immunosuppressive $\gamma\delta$ T cells to the TME. These data exemplify the highly context-dependent effects of RT on lymphocytes.

Radiation therapy and antigen presentation

Dr. Laura Santambrogio (Weill Cornell Medical College) discussed the critical role of the brain lymphatic system in maintaining homeostasis through meningeal lymphatics and deep cervical lymph nodes.^{46,47} More specifically, she

presented unpublished data demonstrating that disruptions in this drainage, whether through lymphatic ligation, cauterization, or ablation, result in metabolic stress and oxidadamage. Using state-of-the-art metabolomic tive approaches,⁴⁸ her team identified a shift from the NAD⁺ salvage pathway to de novo synthesis, coupled with an increased flux through the pentose phosphate pathway and consequent glutathione neosynthesis, as major consequences of disruptions in the brain lymphatic system, culminating with markers of neurodegeneration including synaptic loss, disrupted neurogenesis, and protein aggregation. As emphasized by Dr. Santambrogio, these findings are highly relevant for patients with head and neck squamous cell carcinoma (HNSCC) receiving RT as the potential of the latter to impair lymphatic clearance may contribute to the development of neurodegenerative conditions like Alzheimer's disease. This research underscores the critical importance of preserving brain lymphatic function during cancer therapy to limit long-term neurological side effects.

Dr. Dörthe Schaue (David Geffen School of Medicine) discussed the connection between RT and antigen processing and presentation. RT is indeed known to promote antigen presentation by malignant cells via a number of mechanisms, including the upregulation of multiple components of the antigen presentation machinery (APM), generally culminating in increased cancer cell immunogenicity and visibility to CD8⁺ CTLs.^{49,50} However, malignant cells often acquire APM defects, most likely as a strategy to evade anticancer immunity,⁵¹⁻⁵³ which also prevents RT from stimulating CD8⁺ CTL responses. Dr. Schaue noted that increased expression levels of APM components such as proteasome 20S subunit beta 8 (PSMB8, best known as LMP7) is critical for immunological tumor control, correlating with improved tumor infiltration by immune cells and superior overall survival across various cancer types,⁵⁴⁻⁵⁷ pointing to LMP7 and the associated APM as potential therapeutic targets for the development of more effective RT-IT combinations.

Dr. Andrew Godkin (Cardiff University) went on to first discuss the ability of cyclophosphamide to modulate the immune response of solid tumors, based on findings from animal models of colorectal carcinoma (CRC) as well as human CRC samples, proposing cyclophosphamide as a potential combinatorial partner for RT. Specifically, he highlighted that low-dose cyclophosphamide effectively depletes intratumoral T_{REG} cells, as demonstrated in preclinical tumors models,^{58,59} as well as in a clinical trial testing cyclophosphamide optionally in combination with a cancer vaccine in patients with metastatic CRC.⁵⁸ Previous findings support the notion that cyclophosphamide may also improve the interaction between RT and IT by depleting.⁶⁰ He also showed that cyclophosphamide can modulate the gut microbiome, resulting in further neoantigen recognition.⁵⁸

Finally, he noted that high biologically effective doses (BEDs >100 Gy) of RT have been associated with immunosuppressive effects as illustrated by an elevated neutrophil-to-lymphocyte ratio (NLR) as a biomarker of worsened progression-free survival (PFS), and concluded by commenting on how RT can alter the nature of peptides presented by MHC molecules.

Immune inhibitory effects of radiation therapy

In the last session of Day 1, Dr. Nir Ben Chetrit (Weill Cornell Medical College) discussed the potential of reprogramming tumor-associated macrophages (TAMs) in combination with RT to improve disease outcome in cancer patients.⁶¹ Using single-cell RNA sequencing (scRNAseq), he demonstrated that TAMs commonly found in breast cancer can either support immune effector cells (tumor-niche macrophages, TNMs) or suppress anticancer immunity (stromal-associated macrophages, SAMs), the latter contributing to poor disease outcomes and hence representing promising targets for novel immunotherapeutic interventions.⁶² Dr. Ben Chetrit also presented results from macrophage reprogramming CRISPR screens in TAMs suggesting that inhibiting notch receptor 4 (NOTCH4) may constitute a valid approach to reprogram SAMs toward an immunostimulatory TNM-like phenotype, at least in preclinical models of breast and ovarian cancer.

Dr. Taha Merghoub (Weill Cornell Medical College) discussed the critical impact of intratumoral neutrophils on the efficacy of multiple (immuno)therapeutic anticancer agents, including RT. He explained that co-stimulatory molecules and ICIs not only promote the recruitment of neutrophils to the TME, but also activate effector mechanisms including the release of so-called "neutrophil extracellular traps" (NETs) and nitric oxide synthase 2 (NOS2)-dependent cytotoxicity.⁶³ This may be particularly relevant for tumors that evade immune recognition by adoptively transferred T cells by losing the expression of the antigenic target of the latter.⁶³ Dr. Merghoub also highlighted the considerable therapeutic challenges posed by tumor heterogeneity, which characterizes tumors,⁶⁴ most (if not all) solid and local immunosuppression,⁶⁵ calling for the development of combinatorial strategies targeting both malignant and immune TME components. Finally, he explored the potential of combining RT with immunostimulatory antibodies and ICIs, which (at least in some settings) rely on neutrophil function to maximize T cell responses. Such an approach stands out as a promising strategy to eradicate highly heterogeneous tumors that are resistant to conventional (immuno)therapeutics.

Dr. Dmitry I. Gabrilovich (AstraZeneca) closed Day 1 by discussing the dual role of myeloid cells in cancer sensitivity to RT and IT, emphasizing their two main functional states: classical (pro-inflammatory) and pathological (immunosuppressive). ^{61,66} He commented on the mechanisms through which TAMs and MDSCs contribute to tumor progression, metastasis, and resistance to (immuno)therapy, including the activation of transforming growth factor beta (TGF- β) and vascular endothelial growth factor (VEGF) signaling, the release of immunosuppressive factors such as interleukin 10 (IL10), as well as the remodeling of the tumor stroma via matrix metalloproteases (MMPs). Dr. Gabrilovich also discussed on the possibility of activating ferroptosis, a regulated form of cell death driven by oxidative stress and consequence lipid peroxidation, as a therapeutic strategy against cancer.^{67,68} RT has been shown to elicit ferroptosis in malignant cells,⁶⁹ but whether ferroptosis potentiates or suppresses anticancer immunity remains a matter of debate.^{70–72} Moreover, the ferroptotic death of intratumoral MDSCs and neutrophils potently suppresses anticancer

immunity by CD8⁺ CTLs,⁷³ suggesting that caution should be employed when developing ferroptosis-promoting strategies for cancer (immuno)therapy.

Oral presentations (day 2)

Day 2 started with two oral presentations that were selected from the pool of abstracts submitted to this edition of ImmunoRad.

Following up on a first oral communication by Anne-Gaëlle Goubet (University of Geneva), Donna Li (University of Wisconsin) presented her research on the role of NF- κ B activation by RT-elicited DNA damage on anticancer immune responses.^{74–76} Her unpublished results – originating from mice lacking a critical component of the canonical machinery for NF- κ B signaling,⁷⁷ which results in impaired NF- κ B responses to genotoxic stress – demonstrate that RT-driven NF- κ B activation supports tumor infiltration by CD8⁺ CTLs. Interestingly, such a beneficial reconfiguration of the TME was accompanied by the repolarization of TAMs toward an immunostimulatory state, as demonstrated by reduced colony stimulating factor 1 receptor (CSF1R) expression. These findings suggest that targeting the immunosuppressive receptor CSF1R may improve the susceptibility of some tumors to RT.

Rectal cancer as a model for studying RT-induced immune modulation

Dr. Encouse Golden (Weill Cornell Medical College) presented adenosine signaling as a significant obstacles to RT-driven anticancer immune responses.⁷⁸⁻⁸¹ RT favors indeed the accumulation of adenosine in TME, which mediates considerable immunosuppressive effects on both myeloid and lymphoid cells.⁸² In line with this notion, blocking the extracellular enzyme that converts AMP into adenosine, namely 5'nucleotidase ecto (NT5E, best known as CD73), has been shown to improve both local and distant tumor control by RT in preclinical models of various cancer types including CRC, alongside markers of response indicating improved tumor-targeting immunity.^{82,83} These observations led to the initiation of clinical studies including PANTHER and ARC-9 testing potent dual adenosine receptor inhibitors (e.g., etrumadenant) in combination with RT and/or ICIs and chemotherapy in patients with rectal cancer or metastatic CRC, respectively.^{84,85} Preliminary results from these studies are promising. Two patients with rectal cancer who experienced a complete response to treatment were indeed presented by Dr. Golden, highlighting the potential of this specific RT-IT combination.

Dr. Alan A. Melcher (Institute of Cancer Research) next presented on the capacity of neoadjuvant RT for rectal cancer triggers immunological changes that resemble those elicited by viral infection, a beneficial situation commonly known as viral mimicry,^{86,87} specifically in patients with pronounced responses to therapy.⁸⁸ He also highlighted the promise of using oncolytic virotherapy with T-Vec, a genetically engineered variant of herpes simplex virus (HSV-1),^{89,90} as a strategy to kill malignant cells through immunogenic cell death (ICD),⁹¹ resulting in a reshaped intratumoral TCR repertoire in favor of highly active CD8⁺ CTLs over their dysfunctional counterparts.^{92,93} Finally, Dr. Melcher reported on yet another HSV-1-derived oncolytic virus, i.e., RP1. Similar to T-vec, RP1 could be favorably combined with PD-1 blockers in preclinical models of thyroid carcinoma and melanoma, resulting in superior tumor control along with a reconfiguration of the intratumoral T cell compartment toward the enrichment of ICI-sensitive pre-exhausted CD8⁺ T cells.^{94,95} Globally, these observations suggest that oncolytic viruses may inform strategies to improve the sensitivity of patients with cancer exhibiting limiting infiltration by CD8⁺ CTLs to RT-IT combinations.

Dr. Kristina H. Young (Providence Cancer Institute) dissected the dual role of TGF-ß signaling in rectal cancer development and sensitivity to treatment. Indeed, while TGF-β acts as a prominent oncosuppressive factor at early stage of rectal carcinogenesis,96-98 it potently promotes immunoevasion and tumor progression at later stages of the disease.⁹⁸⁻¹⁰⁰ Adding an extra layer of complexity, a number of nonmalignant components of the TME, notably cancer-associated fibroblasts (CAFs) can produce high levels of TGF- β , hence compensating for potential defects in TGF-B production by neoplastic cells.^{101,102} A number of preclinical studies demonstrated that blocking TGF-β considerably improves local and distant tumor control by RT-IT combinations, alongside eliciting signs of superior anticancer immunity including a favorable increase in the intratumoral CTL/ T_{REG} cell ratio.^{103–107} In this context, Dr. Young highlighted the results of a Phase 2 trial testing neoadjuvant galunisertib (a small molecule inhibiting TGF- β signaling) with chemotherapy and RT to improve pathological complete response (pCR) rates in patients with locally advanced rectal cancer, demonstrating good tolerability and an overall response rate (ORR) of 32%.^{108,109} That said, gene expression studies suggest that specific CRC subtypes including CMS3 and CMS4 might respond differently to RT plus TGF- β , raising the need for the development of personalized approaches to implement this specific RT-IT combination in patients with CRC.

Radiation therapy and immunotherapy in the management of solid tumors

Dr. Gaorav P. Gupta (University of North Carolina) discussed the potential of combining RT with ICIs for the treatment of early stage breast cancer. He emphasized that while RT can elicit immunostimulatory mechanisms including ICD activation and CD8⁺ CTL recruitment, it can also promote immunosuppressive pathways including PD-L1 upregulation on the surface of malignant cells and T_{REG} recruitment to the TME.¹ Clinical trials enrolling women with triple-negative breast cancer (TNBC), such as PEARL and P-RAD, have shown that combining PD-L1 or PD-1 blockers with appropriate RT doses improves pCR rates, with cancer-related factors like MHC Class I expression levels influencing disease outcome.^{110,111} The analysis of post-treatment samples from these clinical studies indicated that combining RT in 3 fractions of 8 Gy each with a PD-1 blocker stimulates immune responses in primary tumors and (at least in some patients) causes the regression of non-irradiated lymph nodes bearing

metastatic disease. Dr. Gupta also noted that tumor biology, especially "cancer ecotypes" as defined by transcriptional profiles of the TME, may play a significant role in treatment response. In this context, optimizing RT dosing and delivery schedule with respect to ICIs stands out as a promising avenue to enhance efficacy in the context of acceptable toxicity.¹¹² Advances in digital pathology and spatial immune phenotyping are expected to assist the identification of biomarkers of response, ultimately leading to improved disease outcome in a number of oncological settings amenable to receive RT-IT combinations.¹¹³

Dr. Julien Sage (Stanford University) presented existing challenges for the treatment of small cell lung carcinoma (SCLC), a highly aggressive cancer associated with poor survival rates owing to its intrinsically metastatic and treatmentresistant nature.^{114,115} Indeed, as most SCLCs are poorly infiltrated by CD8⁺ CTLs and express reduced MHC Class I levels, ICIs targeting PD-1 or PD-L1 have provided only modest survival benefits to a limited number of patients with SCLC.¹¹⁶ In this context, tarlatamab - a bispecific T-cell engager (BiTE) targeting delta-like canonical Notch ligand 3 (DLL3) - appears to holds some promise for improved treatment outcomes.¹¹⁷ Dr. Sage also presented results from his team demonstrating that combining RT, a cornerstone in SCLC treatment, with inhibitors of the antiphagocytic ligand CD47 results in improved local and distant disease control along with improved macrophage-dependent anticancer immunity in preclinical models of SCLC.¹¹⁸⁻¹²⁰ Ongoing research focuses on optimizing RT dosing, exploring combinations with DLL3-targeting BiTEs, and identifying predictive biomarkers to overcome IT resistance and improve disease outcomes in patients with SCLC.

In continuation, Dr. Anna Wilkins (Institute of Cancer Research) discussed strategies to target the bladder TME to enhance RT sensitivity. More specifically, an association between an intratumoral enrichment of specific CAF populations and poor disease outcomes after RT, outlining that the most common CAF subpopulations found in bladder cancer that express high levels of podoplanin (PDPN), presumably independent of TGF- β signaling and immune exclusion.¹²¹⁻¹²³ Dr. Wilkins reinstated the promise of combining TGF-β inhibitors with RT to improve disease outcome across multiple cancer types, especially neoplasms characterized by an intense fibrotic response, by highlighting an abundant preclinical, preclinical¹⁰³⁻¹⁰⁶ and clinical^{108,109,124} literature. Moreover, she presented unpublished results demonstrating that RTinduced fibrosis can arise rapidly and independently of TGF- β via a lymphocyte-dependent mechanism that ultimately affects tumor infiltration by immune cells. In this setting, targeting PDPN may constitute an effective strategy to enhance therapeutic responses to RT in bladder cancer and other tumor types.

Role of radiotherapy and immunotherapy combinations in metastatic disease

Dr. Arta M. Monjazeb (UC Davis School of Medicine) reported on strategies combining RT with immunotherapeutic agents that extend beyond ICIs. First, he discussed

novel approaches focused on enhancing the immunostimulatory effects of RT as an *in situ* vaccine, including the intralesional delivery of recombinant IL2 to irradiated tumors, as currently investigated in preclinical settings as well as in the context of a clinical trial enrolling patients with metastatic NSCLC (NCT03224871). Strategies employing dendritic cell (DC)-activating immunotherapeutics were also discussed.¹²⁵ While RT per se may act indeed as in situ vaccine, exposing tumor-associated antigens and delivering potent immunostimulatory signals including type I interferon (IFN) via ICD induction,¹²⁶⁻¹²⁸ the RT-driven activation of adaptive anticancer immunity strictly depends on DCs,^{129,130} implying that boosting DC functions may enhance the immunostimulatory effects of RT. Such DC stimulators include CpG, a Toll-like receptor (TLR9) agonist that can be directly delivered to the tumor,¹³¹⁻¹³³ as well as epacadostat, an oral inhibitor of the immunosupenzyme indoleamine pressive 2,3-dioxygenase 1 (IDO1).^{134,135} Dr. Monjazeb highlighted the ability of both CpG and epacadostat to increase the local and distant tumor control in preclinical tumor models, correlating with reduced levels of intratumoral T_{REG} cells and immunosuppressive MDSCs.^{131,134,135} Results from a canine clinical trial lend further support to the validity of this approach to safely exacerbate the therapeutic activity of RT.¹³⁴ While these findings have already been translated into early-stage clinical trials with promising results, additional work is needed to fully elucidate the efficacy of DC-activating agents as combinatorial partners for RT.

Dr. Julie Constanzo (University of Montpellier) reported unpublished data on the ability of extracellular vesicles (EVs) released by cancer cells exposed to targeted radionuclide therapy (TRT) to promote beneficial immunological alterations of the TME that may be harnessed with ICIs.^{136–138} Recent data suggest indeed that TRT may represent a powerful inducer of ICD, hence representing an optimal therapeutic partner for ICIs.^{136,139,140} Her research demonstrates that mouse melanoma cells exposed to TRT in vitro release large EVs that can be used to elicit therapeutically relevant anticancer immune responses in vivo downstream of DC activation upon intratumoral administration. Dr. Constanzo went on to show that blocking EV release limits the anticancer activity of TRT, suggesting that EVs are crucial mediators of its immunostimulatory activity, hence representing potential targeting for the development of novel combinatorial partners thereof.

Dr. Sean Pitroda (University of Chicago) discussed the benefits of combining SBRT with dual PD-1 and cytotoxic T lymphocyte-associated protein 4 (CTLA4) blockade in patients with metastatic non-small cell lung carcinoma (NSCLC).¹⁴¹ Such an approach resulted in improved T cell activation, increased TCR diversity and superior local and distant tumor control, *de facto* outperforming SBRT alone.¹⁴² Of note, SBRT combined with dual PD-1 and CTLA4 blockade appeared particularly beneficial for patients bearing NSCLC with elevated degrees of aneuploidy, which typically correlates with immunosuppression and weaker responses to ICIs employed as standalone immunotherapeutics.^{141,143} Ongoing research is focusing on understanding the mechanisms

through which aneuploid cells evade anticancer immunosurveillance and refining strategies for best combining RT and ICIs in patients with NSCLC and other tumor types.

Cancer immunogenomics and RT

Dr. Timothy A. Chan (Cleveland Clinic) discussed the impact of genetic alterations and DNA repair defects on the efficacy of RT and IT in cancer. He explained that ICI sensitivity varies considerably across cancer types – for instance being high in melanoma and NSCLC, but limited in sarcoma and CRC correlating with tumor mutation burden (TMB), neoantigen expression, and immunological features of the TME.^{144–148} In this context, RT may enhance the sensitivity of some tumors to ICIs by reprogramming the TME toward an ICI-responsive status, although optimal biomarkers for combining RT and IT remain to be identified.¹⁴⁹ Importantly, clinical trials like NICHE-2 demonstrate the efficacy of neoadjuvant ICIs in patients with locally advanced CRC bearing defective mismatch repair (dMMR),¹⁵⁰ not only emphasizing the need for personalized treatment approaches to improve disease outcomes in response to IT, but also adding to an expanding literature revealing superior effects from neoadjuvant ICI administration.¹⁵¹ Such a change in treatment paradigm may have profound implications for the development of effective **RT-IT** combinations.

Dr. Anguraj Sadanandam (Institute of Cancer Research) highlighted the critical importance of dissecting tumor heterogeneity for the development of effective cancer regimens, not only intratumorally, but also across distinct neoplastic lesions from the same patient, in both the spatial and temporal dimension.^{64,152,153} Progress in this respect can only be achieved by state-of-the-art technologies that provide (spatial) single-cell resolution coupled with modern artificial intelligence/machine learning tools, ultimately enabling the integration of complex genomic, epigenomic, transcriptomic and proteomic spatial datasets from (whenever possible longitudinal) tissue samples.^{154–156} Such a highly integrated approach may indeed offer critical insights toward the development of personalized treatment strategies. Dr. Sadanandam presented examples of molecular stratifications in CRCs and pancreatic neuroendocrine tumors, illustrating their profound therapeutic implications.^{153,157,158} Ongoing research in this field aims at improving precision medicine through global collaborations and the development of public platforms with the goal of enhancing survival outcomes using biomarker-driven combination therapies.

Dr. Brian D. Brown (Icahn School of Medicine at Mount Sinai) concluded Day 2 by discussing the importance of the genetic factors that affect the TME in shaping tumor behavior and responses to therapy.¹⁵⁹ More specifically, he introduced Perturb-Map, a new technology that combines CRISPR screening with spatial proteomics and transcriptomics to study gene function in tissues, including the TME.^{160,161} This method enabled Dr. Brown's team to identify genes including suppressor of cytokine signaling 1 (*Socs1*) and transforming growth factor beta receptor 2 (*Tgfbr2*) as critical determinants of tumor infiltration by CD8⁺ CTLs in preclinical models of lung cancer,¹⁶⁰ as well as tumor-derived IL4 as a major driver of resistance to PD-1 blockers in preclinical models of ovarian carcinoma.¹⁶² This novel technology stands out as a powerful tool to identify novel determinants of resistance to RT-IT combinations.

Oral presentations (day 3)

Day 3 started with two oral presentations that were selected from the pool of abstracts submitted to this edition of ImmunoRad.

Dr. Sierra McDonald (University of North Carolina) presented the P-RAD clinical trial, a randomized phase II study testing neoadjuvant pembrolizumab with RT in patients with early-stage node-involved TNBC.¹¹¹ Preliminary results suggest that combining pembrolizumab with 3 RT fractions of 3 Gy or 9 Gy each not only causes a dose-dependent enrichment in transcriptional signatures associated with CD8⁺ CTL functions, macrophage activation and inflammation, but also improves pCR rates compared to pembrolizumab alone. Unpublished data from a genetically engineered mouse model of TNBC mimicking the P-RAD study appear to recapitulate these effects and hence may assist the identification of determinants of response and resistance. Dr. McDonald concluded that combining RT with pembrolizumab as a neoadjuvant intervention holds potential for improving disease outcome in patients with early-stage TNBC eligible to surgery.

Dr. Erik Wennerberg (Institute of Cancer Research) discussed the role of ADP-ribosyltransferase 1 (ART1) in cancer immune evasion. Signaling via purinergic receptor P2× 7(P2RX7) as driven by extracellular ATP and NAD is indeed critical for optimal T cell fitness in the tumor microenvironment,^{163,164} and ART1 expression by malignant cells actively interfere with such an immunostimulatory mechanism by promoting P2RX7 mono-ADPribosylation, a detrimental effect that is exacerbated when ADP-ribosyl cyclase CD38 is inhibited.¹⁶⁵ This is particularly relevant for RT-IT combinations - as demonstrated in NSCLC patients receiving SBRT plus a PD-L1 inhibitor¹⁶⁶ – because RT promotes ART1 upregulation in malignant cells. These findings point to ART1, CD38 and P2RX7 as potential targets to promote the efficacy of RT-IT combinations.

Novel approaches

Dr. Chandan Guha (Albert Einstein College of Medicine) presented the immunological consequences of carbon ion RT (CIRT).^{167,168} Specifically, he provided unpublished evidence suggesting that CIRT productively jumpstarts the cancerimmunity cycle,¹⁶⁹ resulting in effective release of tumorassociated antigens, delivery of immunostimulatory cues to DCs and ultimately cross-priming of a therapeutically relevant tumor-targeting immune response. By scRNAseq, Dr. Guha's team demonstrated that while conventional RT promotes the immunosuppressive expansion of component 1. q subcomponent, and alpha polypeptide (C1QA)-expressing TAMs infiltrating mouse KPC pancreatic tumors, CIRT limits the abundance of C1QA⁺ TAMs while increasing the expression of pro-inflammatory cytokines like C-X-C motif chemokine ligand 10 (CXCL10). Thus, CXCL10 secretion and the consequent reduction of C1QA⁺ TAMs appear to underlie the immunostimulatory effects of CIRT, at least in preclinical models of pancreatic cancer.

Drs. Aisha Hasan (Johnson & Johnson, USA) discussed the evolving role of RT in the era of IT, particularly for the clinical management of NSCLC. She reinstated the ability of RT to act as an in situ vaccine, especially when combined with IT,¹⁷⁰ highlighting promising clinical results as obtained by combining SBRT with ICIs targeting CTLA4, PD-1 or PD-L1 in patients with NSCLC.^{5,166,171,172} Finally, she commented on the potential for interventional oncology to promote the efficacy of RT in the context of reduced systemic toxicityfor instance, upon the intratumoral delivery of immunotherapeutics.89,90

Dr. Jalal Ahmed (Icahn School of Medicine at Mount Sinai) focused on the impact of DCs on CAR T cell therapy.^{173,174} Successfully employing CAR T cells and other adoptively transferred lymphocytes for the management of solid neoplasms faces indeed considerable challenges, largely reflecting the harsh metabolic conditions that most characterize their TME.^{80,173,175} At least in mouse tumor models, combining CAR T cell infusion with low-dose TBI has been shown promote disease control alongside improved CAR T cell expansion and (at least some degree of) cytotoxicity against antigen-loss cancer cell variants.^{40,41} Moreover, recent preclinical data suggest that focal LDRT can support the therapeutic activity of CAR T cells by engaging endogenous CD8⁺ CTLs upon DCdependent crosspriming.¹⁷⁶ Ongoing research is focusing at the development of safe and effective strategies to combine RT with CAR T cells in eligible patients toward the rapid implementation of innovative clinical studies.

Panel discussion

Day 3 concluded with a panel discussion with four panelists from industry that was moderated by the conference Chairs Dr. Silvia Formenti (Weill Cornell Medical College) and Dr. Eric Deutsch (Gustave Roussy).

Dr. Philippe Szapary (Johnson & Johnson) highlighted the value of interventional oncology as a novel approach to achieve superior therapeutic efficacy in the context of limited systemic toxicity via the rational combination of intratumorally administered therapeutics such as oncolytic viruses or chemotherapy depots combined with systemic IT with ICIs. Specifically, he emphasized the promising potential of combining RT with investigational radioenhancing agents like JNJ-1900 (a functionalized hafnium oxide nanoparticles also known as NBTXR3) for the management of HNSCC and NSCLC.^{30,31,177}

Dr. Howard Kaufman (Ankyra Therapeutics) introduced ANK-101, an aluminum hydroxide-anchored IL12 variant that exhibit superior retention within the TME upon intratumoral delivery, resulting not only in single agent activity across a variety of preclinical tumor models, but also in the activation of systemic immune responses effectively targeting distant tumors, especially in combination with otherwise inactive ICIs delivered systemically, pointing to ANK-101 as a promising agent for clinical development.¹⁷⁸

Dr. Sébastien Paris (Nanobiotix) further commented on JNJ-1900, which – while initially conceived as a radioenhancer¹⁷⁹ – has turned out to considerably enhance the ability of focal RT to elicit anticancer immune responses with systemic outreach, at least in preclinical tumor models, prompting clinical development in patients with a variety of cancer eligible to irradiation.^{180,181}

Dr. Dhan Chand (Agenus) discussed the promise of novel agents like botensilimab, a second generation CTLA4 inhibitor,^{182,183} AGEN1423, a dual inhibitor of CD73 and TGF- β ,¹⁸⁴ AGEN2373, an agonist of the immunostimulatory receptor TNF receptor superfamily member 9 (TNFRSF9, best known as CD137),^{185–187} and AgenT-797, an allogenic cell therapy showing promising results in combination with RT.^{188,189}

Concluding remarks

Dr. Formenti and Dr. Deutsch closed the meeting by emphasizing the importance of additional preclinical and clinical investigation for RT-IT combinations to be safely and successfully implemented into the clinical management of an increasing number of malignancies, as they recognized the progress that has been achieved over the past decade by the crosspollination between radiation oncology and clinical immunotherapy. The ImmunoRad Conference has considerably fostered such an advantageous interaction and will continue to do so at the Cordeliers Research Center in Paris, on September 17th to 19th 2025. You are all welcome to participate!

Acknowledgement

Conceptualization, L.G.,E. D., S. C. F;writing – original draft preparation;F. T, L. G; writing – review and editing; F. T., F. G., J. A., N. B. C., B. D. B., T. A. C., D. C., J. C., S. D., D. I. G., E. G., A. G., C. G., G. P. G., A. H., F. G. H., H. K., D. L., A. A. M., S. M., T. M., A. M. M., S. P., S. P., A. S., D. S., L. S., P. S., J. S., J. W. W., A. W., K. H. Y., E. W., L. Z., L. G., E. D. and S.C.F; All authors have read and agreed to the published version of the manuscript.

Disclosure statement

BDB sits in the scientific advisory board (SAB) of Noetik. DC is a full-time employee of Agenus Inc. and holds equity in the company. DG is a fulltime employee of AstraZeneca. EG declares research funding from Arcus Biosciences. CG sits in the data safety monitoring board (DSMB) for the Focused Ultrasound Foundation, is a co-founder of Bioconvergent Health and holds equity in the company. GPG declares funding from Merck, Breakpoint Therapeutics, the V Foundation, as well as ownership, intellectual property rights and royalties from Naveris Inc. AH is a full-time employee of Johnson & Johnson, hold stocks in the company, and declares royalties from Atara Biotherapeutics. FGH declares research funding from Accuray, Bioprotect, Bristol-Myers Squibb, Roche-ImFlame/ImCore, Nanobiotix, AstraZeneca, Eisai, MSD, Seagen, RaySearch Laboratories, as well as consultation fees, and travel expenses from Johnson & Johnson, Boehringer Ingelheim, Bristol-Myers Squibb, AstraZeneca, Eisai and Telix. HK is a full-time employee of Ankyra Therapeutics, owns stock in Replimune Inc. and serves on the SAB of Castle Biosciences, Immvira, Marengo Therapeutics, Tatum Biosciences, and Virogin. TM is a consultant for Immunos Therapeutics, Daiichi Sankyo, TigaTx, Normunity, and Pfizer, a co-founder and equity holder in IMVAQ Therapeutics, and declares research funding from Surface Oncology, Kyn Therapeutics, Infinity Pharmaceuticals, Peregrine

Pharmaceuticals, Adaptive Biotechnologies, Leap Therapeutics, Aprea Therapeutics, Bristol-Myers Squibb, Enterome SA, and Realta Life Sciences. TM is an inventor on patent applications related to oncolytic virotherapy, alpha virus - based vaccines, neoantigen modeling, CD40, GITR, OX40, PD-1, and CTLA-4. AMM declares research funding from Genentech, Bristol-Myers Squibb, Merck, Transgene, Incyte, Trisalus. SP is a full-time employee of Nanobiotix and co-founder of PersonaDx, and declares has research funding from LUNGevity Foundation, American Lung Association, Falk Medical Research Trust as well as patents related to the diagnosis and triage of patients with colorectal liver metastases, RNAs with tumor radio/chemosensitizing and immunomodulatory properties, methods and compositions relating to cancer therapy with DNAdamaging agents, and molecular subtyping of colorectal liver metastases to personalize treatment approaches. AS is a co-founder of Oncoassign and Diagnostring Laboratories. PS is a full-time employee of Johnson & Johnson. JS is an equity holder and advisor for DISCO Pharmaceuticals. JWW is affiliated with Accuray (SAB, consulting), Alpine Immune Science (SAB, consulting, equity), Boehringer Ingelheim (SAB, consulting), Checkmate Pharmaceuticals (SAB, consulting, equity), China Medical Tribune (SAB, consulting), Genentech (SAB, consulting, research), GI Innovation (SAB, consulting), Kezar Life Sciences (SAB, consulting), Legion Healthcare Partners (SAB, consulting), Life Science Dynamic Limited (SAB, consulting), McKesson Corporation (SAB, consulting), Molecular Match (equity), Nanorobotix (SAB, consulting), OligoImmune (founder), Roche (SAB, consulting), Roche Molecular Systems (SAB, consulting), Nanobiotix (research funding, travel expenses, SAB), Bristol-Myers Squibb (research funding), Merck (research funding), Varian (research funding, travel expenses, sponsored clinical research), Reflexion (research funding, travel expenses, stock options, SAB), Hotspot Therapeutics (research funding), Gilead (research funding), Novocure (SAB), Oncoresponse (SAB, stock options), AstraZeneca (consultant, research funding), Bayer Healthcare (research funding), Kiromic (research funding), Alkermes (SAB, research funding), Artidis (research funding), Sciclone (research funding), Takeda (research funding), Nurix (research funding). AW declares PhD funding from AstraZeneca, and clinical fellowship funding from Artera AI, as well research funding from Veracyte, Roche, and co-leadership of the TGFβ paradox ImCore Working Group. KHY declares research funding from Bristol-Myers Squibb, Bicara, Corbus, Eli Lilly, sits on the SAB of Synthis Therapeutics, and receives consulting fees from Corbus Pharmaceuticals. LZ is a founder and sits on the SAB of everImmune, sits on the SAB of Hookipa and the IHU Méditerranée Infections, declares research funding from 9 meters, Pileje, Biomérieux and Daiichi Sankyo, teaching contracts with Pierre Fabre and honoraria from everImmune, Kookipa, Pierre Fabre. LG is/has been holding research contracts with Lytix Biopharma, Promontory, and Onxeo, has received consulting/advisory honoraria from Boehringer Ingelheim, AstraZeneca, AbbVie, OmniSEQ, Onxeo, The Longevity Labs, Inzen, Imvax, Sotio, Promontory, Noxopharm, EduCom, and the Luke Heller TECPR2 Foundation, and holds Promontory stock options. ED receives research funding and/or personal fees from Roche Genentech, Boehringer Ingelheim, AstraZeneca, Merck, Bristol-Myers Squibb, and MSD. SCF has received research funding from Bristol Myers Squibb, Varian, Regeneron, Merck, Celldex, and Arcus, and has served as a consultant for Bayer, Bristol Myers Squibb, Varian, ViewRay, Elekta, Janssen, Regeneron, GlaxoSmithKline, Eisai, Astra Zeneca, MedImmune, Merck, Boehringer Ingelheim, EMD Serono, Roche Genentech, Nanobiotix, Telix, and EmBioSys.

Author contributions

CRediT: Fereshteh Talebi: Writing – original draft, Writing – review & editing; Fabiana Gregucci: Project administration, Writing – review & editing; Jalal Ahmed: Writing – review & editing; Nir Ben Chetrit: Writing – review & editing; Brian D. Brown: Writing – review & editing; Timothy A. Chan: Writing – review & editing; Dhan Chand: Writing – review & editing; Julie Constanzo: Writing – review & editing; Sandra Demaria: Writing – review & editing; Dmitry I. Gabrilovich: Writing – review & editing; Encouse Golden: Writing – review & editing; Andrew

Godkin: Writing - review & editing; Chandan Guha: Writing - review & editing; Gaorav P. Gupta: Writing - review & editing; Aisha Hasan: Writing - review & editing; Fernanda G. Herrera: Writing - review & editing; Howard Kaufman: Writing - review & editing; Donna Li: Writing - review & editing; Alan A. Melcher: Writing - review & editing; Sierra McDonald: Writing - review & editing; Taha Merghoub: Writing - review & editing; Arta M. Monjazeb: Writing - review & editing; Sébastien Paris: Writing - review & editing; Sean Pitroda: Writing review & editing; Anguraj Sadanandam: Writing - review & editing; Dörthe Schaue: Writing - review & editing; Laura Santambrogio: Writing - review & editing; Phillippe Szapary: Writing - review & editing; Julien Sage: Writing - review & editing; James W. Welsh: Writing - review & editing; Anna Wilkins: Writing - review & editing; Kristina H. Young: Writing - review & editing; Eric Wennerberg: Writing - review & editing; Laurence Zitvogel: Writing - review & editing; Lorenzo Galluzzi: Conceptualization, Writing - original draft, Writing - review & editing; Silvia C. Formenti: Conceptualization, Writing - review & editing.

Data availability statement

Data sharing is not applicable to this article as no data were created or analyzed in this study.

Funding

This work was supported by Janssen (Johnson & Johnson), Arcus, Ankyra Therapeutics, and the Department of Radiation Oncology at Weill Cornell Medicine. The sponsors had no role in the preparation, review, or approval of the manuscript. Additional support was provided through educational grants from Pfizer and Varian. No specific grant number is associated with this study.

ORCID

Fabiana Gregucci b http://orcid.org/0000-0002-9947-9650 Timothy A. Chan b http://orcid.org/0000-0002-9265-0283

References

- Galluzzi L, Aryankalayil MJ, Coleman CN, Formenti SC. Emerging evidence for adapting radiotherapy to immunotherapy. Nat Rev Clin Oncol. 2023;20(8):543–557. doi: 10.1038/s41571-023-00782-x.
- McLaughlin M, Patin EC, Pedersen M, Wilkins A, Dillon MT, Melcher AA, Harrington KJ. Inflammatory microenvironment remodelling by tumour cells after radiotherapy. Nat Rev Cancer. 2020;20(4):203–217. doi: 10.1038/s41568-020-0246-1.
- Cytlak UM, Dyer DP, Honeychurch J, Williams KJ, Travis MA, Illidge TM. Immunomodulation by radiotherapy in tumour control and normal tissue toxicity. Nat Rev Immunol. 2022;22 (2):124–138. doi: 10.1038/s41577-021-00568-1.
- Rodriguez-Ruiz ME, Vitale I, Harrington KJ, Melero I, Galluzzi L. Immunological impact of cell death signaling driven by radiation on the tumor microenvironment. Nat Immunol. 2020;21 (2):120–134. doi: 10.1038/s41590-019-0561-4.
- Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, Kurata T, Chiappori A, Lee KH, de Wit M, et al. Overall survival with Durvalumab after Chemoradiotherapy in stage III NSCLC. N Engl J Med. 2018;379(24):2342–2350. doi: 10.1056/ NEJMoa1809697.
- 6. Lee NY, Ferris RL, Psyrri A, Haddad RI, Tahara M, Bourhis J, Harrington K, Chang PMH, Lin J-C, Razaq MA, et al. Avelumab plus standard-of-care chemoradiotherapy versus chemoradiotherapy alone in patients with locally advanced squamous cell carcinoma of the head and neck: a randomised, double-blind,

placebo-controlled, multicentre, phase 3 trial. Lancet Oncol. 2021;22(4):450-462. doi: 10.1016/S1470-2045(20)30737-3.

- Lim M, Weller M, Idbaih A, Steinbach J, Finocchiaro G, Raval RR, Ansstas G, Baehring J, Taylor JW, Honnorat J, et al. Phase III trial of chemoradiotherapy with temozolomide plus nivolumab or placebo for newly diagnosed glioblastoma with methylated MGMT promoter. Neuro Oncol. 2022;24(11):1935–1949. doi: 10.1093/ neuonc/noac116.
- Omuro A, Brandes AA, Carpentier AF, Idbaih A, Reardon DA, Cloughesy T, Sumrall A, Baehring J, van den Bent M, Bähr O, et al. Radiotherapy combined with nivolumab or temozolomide for newly diagnosed glioblastoma with unmethylated MGMT promoter: an international randomized phase III trial. Neuro Oncol. 2023;25(1):123–134. doi: 10.1093/neuonc/noac099.
- Weichselbaum RR, Liang H, Deng L, Fu YX. Radiotherapy and immunotherapy: a beneficial liaison? Nat Rev Clin Oncol. 2017;14 (6):365–379. doi: 10.1038/nrclinonc.2016.211.
- Turchan WT, Pitroda SP, Weichselbaum RR. Radiotherapy and immunotherapy combinations in the treatment of patients with metastatic disease: current status and future focus. Clin Cancer Res. 2021;27(19):5188–5194. doi: 10.1158/1078-0432.CCR-21-0145.
- Viglione G. How scientific conferences will survive the coronavirus shock. Nature. 2020;582(7811):166–167. doi: 10.1038/d41586-020-01521-3.
- 12. Viglione G. A year without conferences? How the coronavirus pandemic could change research. Nature. 2020;579 (7799):327–328. doi: 10.1038/d41586-020-00786-y.
- Gregucci F, Spada S, Barcellos-Hoff MH, Bhardwaj N, Chan Wah Hak C, Fiorentino A, Guha C, Guzman ML, Harrington K, Herrera FG, et al. Updates on radiotherapy-immunotherapy combinations: proceedings of 6th annual ImmunoRad conference. Oncoimmunology. 2023;12(1):2222560. doi: 10.1080/2162402X. 2023.2222560.
- Laurent PA, André F, Bobard A, Deandreis D, Demaria S, Depil S, Eichmüller SB, Fernandez-Palomo C, Foijer F, Galluzzi L, et al. Pushing the boundaries of radiotherapy-immunotherapy combinations: highlights from the 7 th immunorad conference. Oncoimmunology. 2025;14(1):2432726. doi: 10.1080/2162402X. 2024.2432726.
- Joos R, Boucher K, Lavelle A, Arumugam M, Blaser MJ, Claesson MJ, Clarke G, Cotter PD, De Sordi L, Dominguez-Bello MG, et al. Examining the healthy human microbiome concept. Nat Rev Microbiol. 2025;23(3):192–205. doi: 10.1038/s41579-024-01107-0.
- Silva CAC, Fidelle M, Almonte AA, Derosa L, Zitvogel L. Gut microbiota-related biomarkers in immuno-oncology. Annu. Rev. Pharmacol Toxicol. 2025;65(1):333–354. doi: 10.1146/annurevpharmtox-061124-102218.
- Rodriguez J, Hassani Z, Alves Costa Silva C, Betsou F, Carraturo F, Fasano A, Israelsen M, Iyappan A, Krag A, Metwaly A, et al. State of the art and the future of microbiome-based biomarkers: a multidisciplinary delphi consensus. Lancet Microbe. 2024;6 (2):100948. doi: 10.1016/j.lanmic.2024.07.011.
- Bhutiani N, Wargo JA. Gut microbes as biomarkers of ICI response — sharpening the focus. Nat Rev Clin Oncol. 2022;19 (8):495-496. doi: 10.1038/s41571-022-00634-0.
- Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, Prieto PA, Vicente D, Hoffman K, Wei SC, et al. Gut microbiome modulates response to anti–PD-1 immunotherapy in melanoma patients. Science. 2018;359(6371):97–103. doi: 10.1126/ science.aan4236.
- 20. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, Molina DA, Salcedo R, Back T, Cramer S, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science. 2013;342 (6161):967–970. doi: 10.1126/science.1240527.
- Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillère R, Hannani D, Enot DP, Pfirschke C, Engblom C, Pittet MJ, et al. The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. Science. 2013;342(6161):971–976. doi: 10. 1126/science.1240537.

- Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, Rusakiewicz S, Routy B, Roberti MP, Duong CPM, et al. Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science. 2015;350(6264):1079–1084. doi: 10.1126/ science.aad1329.
- 23. Seo YD, Ajami N, Wargo JA. Using gut microorganisms to treat cancer. Nat Med. 2023;29(8):1910–1911. doi: 10.1038/s41591-023-02460-y.
- 24. Fernandes MR, Aggarwal P, Costa RGF, Cole AM, Trinchieri G. Targeting the gut microbiota for cancer therapy. Nat Rev Cancer. 2022;22(12):703–722. doi: 10.1038/s41568-022-00513-x.
- Finlay BB, Goldszmid R, Honda K, Trinchieri G, Wargo J, Zitvogel L. Can we harness the microbiota to enhance the efficacy of cancer immunotherapy? Nat Rev Immunol. 2020;20(9):522–528. doi: 10.1038/s41577-020-0374-6.
- Derosa L, Routy B, Desilets A, Daillère R, Terrisse S, Kroemer G, Zitvogel L. Microbiota-centered interventions: the next breakthrough in immuno-oncology? Cancer Discov. 2021;11 (10):2396–2412. doi: 10.1158/2159-8290.CD-21-0236.
- Chen J, Levy A, Tian A-L, Huang X, Cai G, Fidelle M, Rauber C, Ly P, Pizzato E, Sitterle L, et al. Low-dose irradiation of the gut improves the efficacy of PD-L1 blockade in metastatic cancer patients. Cancer Cell. 2025;43(3):361–379.e10. doi: 10.1016/j.ccell. 2025.02.010.
- Barsoumian HB, Ramapriyan R, Younes AI, Caetano MS, Menon H, Comeaux NI, Cushman TR, Schoenhals JE, Cadena AP, Reilly TP, et al. Low-dose radiation treatment enhances systemic antitumor immune responses by overcoming the inhibitory stroma. J Immunother Cancer. 2020;8(2):e000537. doi: 10.1136/jitc-2020-000537.
- 29. Chen D, Verma V, Patel RR, Barsoumian HB, Cortez MA, Welsh JW. Absolute lymphocyte Count predicts abscopal responses and outcomes in patients receiving combined immunotherapy and radiation therapy: analysis of 3 phase 1/2 trials. Int J Radiat Oncol Biol Phys. 2020;108(1):196–203. doi: 10.1016/j. ijrobp.2020.01.032.
- 30. Hu Y, Paris S, Sahoo N, Bertolet G, Wang Q, Wang Q, Barsoumian HB, Da Silva J, Huang A, Doss DJ, et al. Nanoparticleenhanced proton beam immunoradiotherapy drives immune activation and durable tumor rejection. JCI Insight. 2023;8(12): e167749. doi: 10.1172/jci.insight.167749.
- 31. Hu Y, Paris S, Sahoo N, Wang Q, Wang Q, Barsoumian HB, Huang A, Da Silva J, Bienassis C, Leyton CSK, et al. Superior antitumor immune response achieved with proton over photon immunoradiotherapy is amplified by the nanoradioenhancer NBTXR3. J Nanobiotechnol. 2024;22(1):597. doi: 10.1186/s12951-024-02855-0.
- Passelli K, Repáraz D, Kinj R, Herrera FG. Strategies for overcoming tumour resistance to immunotherapy: harnessing the power of radiation therapy. Br J Radiol. 2024;97 (1160):1378–1390. doi: 10.1093/bjr/tqae100.
- Ochoa-de-Olza M, Bourhis J, Coukos G, Herrera FG. Low-dose irradiation for reversing immunotherapy resistance: how to translate? J For Immunother Of Cancer. 2022;10(7):e004939. doi: 10.1136/jitc-2022-004939.
- 34. Herrera FG, Ronet C, Ochoa de Olza M, Barras D, Crespo I, Andreatta M, Corria-Osorio J, Spill A, Benedetti F, Genolet R, et al. Low-dose radiotherapy reverses tumor immune desertification and resistance to immunotherapy. Cancer Discov. 2022;12 (1):108–133. doi: 10.1158/2159-8290.CD-21-0003.
- Cassetta L, Pollard JW. A timeline of tumour-associated macrophage biology. Nat Rev Cancer. 2023;23(4):238–257. doi: 10.1038/ s41568-022-00547-1.
- 36. Levy A, Morel D, Texier M, Sun R, Durand-Labrunie J, Rodriguez-Ruiz ME, Racadot S, Supiot S, Magné N, Cyrille S, et al. An international phase II trial and immune profiling of SBRT and atezolizumab in advanced pretreated colorectal cancer. Mol Cancer. 2024;23(1):61. doi: 10.1186/s12943-024-01970-8.
- Bergeron P, Dos Santos M, Sitterle L, Tarlet G, Lavigne J, Liu W, Gerbé de Thoré M, Clémenson C, Meziani L, Schott C, et al. Non-

homogenous intratumor ionizing radiation doses synergize with PD1 and CXCR2 blockade. Nat Commun. 2024;15(1):8845. doi: 10. 1038/s41467-024-53015-9.

- Deutsch E, Chargari C, Galluzzi L, Kroemer G. Optimising efficacy and reducing toxicity of anticancer radioimmunotherapy. Lancet Oncol. 2019;20(8):e452-e463. doi: 10.1016/S1470-2045(19)30171-8.
- Arina A, Beckett M, Fernandez C, Zheng W, Pitroda S, Chmura SJ, Luke JJ, Forde M, Hou Y, Burnette B, et al. Tumor-reprogrammed resident T cells resist radiation to control tumors. Nat Commun. 2019;10(1):3959. doi: 10.1038/s41467-019-11906-2.
- 40. DeSelm C, Palomba ML, Yahalom J, Hamieh M, Eyquem J, Rajasekhar VK, Sadelain M. Low-dose radiation conditioning enables CAR T cells to mitigate antigen escape. Mol Ther: J Am Soc Gene Ther. 2018;26(11):2542–2552. doi: 10.1016/j.ymthe.2018. 09.008.
- Sugita M, Yamazaki T, Alhomoud M, Martinet J, Latouche J-B, Golden E, Boyer O, Van Besien K, Formenti SC, Galluzzi L, et al. Radiation therapy improves CAR T cell activity in acute lymphoblastic leukemia. Cell Death Dis. 2023;14(5):305. doi: 10.1038/ s41419-023-05829-6.
- 42. Buqué A, Bloy N, Perez-Lanzón M, Iribarren K, Humeau J, Pol JG, Levesque S, Mondragon L, Yamazaki T, Sato A, et al. Immunoprophylactic and immunotherapeutic control of hormone receptor-positive breast cancer. Nat Commun. 2020;11(1):3819. doi: 10.1038/s41467-020-17644-0.
- 43. Petroni G, Buqué A, Yamazaki T, Bloy N, Liberto MD, Chen-Kiang S, Formenti SC, Galluzzi L. Radiotherapy delivered before CDK4/6 inhibitors mediates Superior therapeutic effects in ER(+) breast cancer. Clin Cancer Res. 2021;27(7):1855–1863. doi: 10. 1158/1078-0432.CCR-20-3871.
- 44. Petroni G, Buqué A, Coussens LM, Galluzzi L. Targeting oncogene and non-oncogene addiction to inflame the tumour microenvironment. Nat Rev Drug Discov. 2022;21(6):440–462. doi: 10.1038/s41573-022-00415-5.
- Petroni G, Cantley LC, Santambrogio L, Formenti SC, Galluzzi L. Radiotherapy as a tool to elicit clinically actionable signalling pathways in cancer. Nat Rev Clin Oncol. 2022;19(2):114–131. doi: 10. 1038/s41571-021-00579-w.
- 46. Nanaware PP, Khan ZN, Clement CC, Shetty M, Mota I, Seltzer ES, Dzieciatkowska M, Gamboni F, D'Alessandro A, Ng C, et al. Role of the afferent lymph as an immunological conduit to analyze tissue antigenic and inflammatory load. Cell Rep. 2024;43 (6):114311. doi: 10.1016/j.celrep.2024.114311.
- 47. Zawieja DC, Thangaswamy S, Wang W, Furtado R, Clement CC, Papadopoulos Z, Vigano M, Bridenbaugh EA, Zolla L, Gashev AA, et al. Lymphatic cannulation for lymph sampling and molecular delivery. The J Immunol. 2019;203(8):2339–2350. doi: 10.4049/ jimmunol.1900375.
- Clement CC, Wang W, Dzieciatkowska M, Cortese M, Hansen KC, Becerra A, Thangaswamy S, Nizamutdinova I, Moon J-Y, Stern LJ, et al. Quantitative profiling of the lymph node clearance capacity. Sci Rep. 2018;8(1):11253. doi: 10.1038/s41598-018-29614-0.
- Yang K, Halima A, Chan TA. Antigen presentation in cancer mechanisms and clinical implications for immunotherapy. Nat Rev Clin Oncol. 2023;20(9):604–623. doi: 10.1038/s41571-023-00789-4.
- Jhunjhunwala S, Hammer C, Delamarre L. Antigen presentation in cancer: insights into tumour immunogenicity and immune evasion. Nat Rev Cancer. 2021;21(5):298–312. doi: 10.1038/ s41568-021-00339-z.
- Galassi C, Chan TA, Vitale I, Galluzzi L. The hallmarks of cancer immune evasion. Cancer Cell. 2024;42(11):1825–1863. doi: 10. 1016/j.ccell.2024.09.010.
- 52. Meissner M, Reichert TE, Kunkel M, Gooding W, Whiteside TL, Ferrone S, Seliger B. Defects in the human leukocyte antigen class I antigen processing machinery in head and neck squamous cell carcinoma: association with clinical outcome. Clin Cancer Res. 2005;11(7):2552–2560. doi: 10.1158/1078-0432.CCR-04-2146.
- 53. Yoshihama S, Roszik J, Downs I, Meissner TB, Vijayan S, Chapuy B, Sidiq T, Shipp MA, Lizee GA, Kobayashi KS. NLRC5/

MHC class I transactivator is a target for immune evasion in cancer. Proc Natl Acad Sci USA. 2016;113(21):5999–6004. doi: 10.1073/pnas.1602069113.

- 54. Lee MH, Ratanachan D, Wang Z, Hack J, Adbulrahman L, Shamlin NP, Kalayjian M, Nesseler JP, Ganapathy E, Nguyen C, et al. Adaptation of the tumor antigen presentation machinery to ionizing radiation. J Immunol. 2023;211(4):693–705. doi: 10.4049/ jimmunol.2100793.
- 55. Tripathi SC, Peters HL, Taguchi A, Katayama H, Wang H, Momin A, Jolly MK, Celiktas M, Rodriguez-Canales J, Liu H, et al. Immunoproteasome deficiency is a feature of non-small cell lung cancer with a mesenchymal phenotype and is associated with a poor outcome. Proc Natl Acad Sci USA. 2016;113(11):E1555– 1564. doi: 10.1073/pnas.1521812113.
- 56. Ha YJ, Tak KH, Kim CW, Roh SA, Choi EK, Cho DH, Kim JH, Kim SK, Kim SY, Kim YS, et al. PSMB8 as a Candidate Marker of responsiveness to preoperative radiation therapy in rectal cancer patients. Int J Radiat Oncol Biol Phys. 2017;98(5):1164–1173. doi: 10.1016/j.ijrobp.2017.03.023.
- 57. Kalaora S, Lee JS, Barnea E, Levy R, Greenberg P, Alon M, Yagel G, Bar Eli G, Oren R, Peri A, et al. Immunoproteasome expression is associated with better prognosis and response to checkpoint therapies in melanoma. Nat Commun. 2020;11(1):896. doi: 10.1038/ s41467-020-14639-9.
- 58. Scurr M, Pembroke T, Bloom A, Roberts D, Thomson A, Smart K, Bridgeman H, Adams R, Brewster A, Jones R, et al. Low-dose cyclophosphamide induces antitumor T-Cell responses, which associate with survival in metastatic colorectal cancer. Clin Cancer Res. 2017;23(22):6771–6780. doi: 10.1158/1078-0432. CCR-17-0895.
- 59. Ghiringhelli F, Larmonier N, Schmitt E, Parcellier A, Cathelin D, Garrido C, Chauffert B, Solary E, Bonnotte B, Martin F. CD4 +CD25+ regulatory T cells suppress tumor immunity but are sensitive to cyclophosphamide which allows immunotherapy of established tumors to be curative. Eur J Immunol. 2004;34 (2):336-344. doi: 10.1002/eji.200324181.
- 60. Pilones KA, Hensler M, Daviaud C, Kraynak J, Fucikova J, Galluzzi L, Demaria S, Formenti SC. Converging focal radiation and immunotherapy in a preclinical model of triple negative breast cancer: contribution of VISTA blockade. Oncoimmunology. 2020;9(1):1830524. doi: 10.1080/2162402X.2020.1830524.
- Barry ST, Gabrilovich DI, Sansom OJ, Campbell AD, Morton JP. Therapeutic targeting of tumour myeloid cells. Nat Rev Cancer. 2023;23(4):216–237. doi: 10.1038/s41568-022-00546-2.
- 62. Ben-Chetrit N. Breast cancer macrophage heterogeneity and self-renewal are determined by spatial localization. bioRxiv. 2023;10.24.563749.
- 63. Hirschhorn D, Budhu S, Kraehenbuehl L, Gigoux M, Schröder D, Chow A, Ricca JM, Gasmi B, De Henau O, Mangarin LMB, et al. T cell immunotherapies engage neutrophils to eliminate tumor antigen escape variants. Cell. 2023;186(7):1432–1447.e17. doi: 10. 1016/j.cell.2023.03.007.
- 64. Vitale I, Shema E, Loi S, Galluzzi L. Intratumoral heterogeneity in cancer progression and response to immunotherapy. Nat Med. 2021;27(2):212–224. doi: 10.1038/s41591-021-01233-9.
- Roerden M, Spranger S. Cancer immune evasion, immunoediting and intratumour heterogeneity. Nat Rev Immunol. 2025;25 (5):353–369. doi: 10.1038/s41577-024-01111-8.
- Eruslanov E, Nefedova Y, Gabrilovich DI. The heterogeneity of neutrophils in cancer and its implication for therapeutic targeting. Nat Immunol. 2025;26(1):17–28. doi: 10.1038/s41590-024-02029-y.
- 67. Dai E, Chen X, Linkermann A, Jiang X, Kang R, Kagan VE, Bayir H, Yang WS, Garcia-Saez AJ, Ioannou MS, et al. A guideline on the molecular ecosystem regulating ferroptosis. Nat Cell Biol. 2024;26(9):1447–1457. doi: 10.1038/s41556-024-01360-8.
- Dixon SJ, Olzmann JA. The cell biology of ferroptosis. Nat Rev Mol Cell Biol. 2024;25(6):424–442. doi: 10.1038/s41580-024-00703-5.
- 69. Adjemian S, Oltean T, Martens S, Wiernicki B, Goossens V, Vanden Berghe T, Cappe B, Ladik M, Riquet FB, Heyndrickx L,

et al. Ionizing radiation results in a mixture of cellular outcomes including mitotic catastrophe, senescence, methuosis, and iron-dependent cell death. Cell Death Dis. 2020;11(11):1003. doi: 10.1038/s41419-020-03209-y.

- Wiernicki B, Maschalidi S, Pinney J, Adjemian S, Vanden Berghe T, Ravichandran KS, Vandenabeele P. Cancer cells dying from ferroptosis impede dendritic cell-mediated anti-tumor immunity. Nat Commun. 2022;13(1):3676. doi: 10.1038/s41467-022-31218-2.
- Catanzaro E, Demuynck R, Naessens F, Galluzzi L, Krysko DV. Immunogenicity of ferroptosis in cancer: a matter of context? Trends Cancer. 2024;10(5):407–416. doi: 10.1016/j.trecan.2024.01. 013.
- 72. Efimova I, Catanzaro E, Van der Meeren L, Turubanova VD, Hammad H, Mishchenko TA, Vedunova MV, Fimognari C, Bachert C, Coppieters F, et al. Vaccination with early ferroptotic cancer cells induces efficient antitumor immunity. J For Immunother Of Cancer. 2020;8(2):e001369. doi: 10.1136/jitc-2020-001369.
- Kim R, Hashimoto A, Markosyan N, Tyurin VA, Tyurina YY, Kar G, Fu S, Sehgal M, Garcia-Gerique L, Kossenkov A, et al. Ferroptosis of tumour neutrophils causes immune suppression in cancer. Nature. 2022;612(7939):338–346. doi: 10.1038/s41586-022-05443-0.
- Taniguchi K, Karin M. NF-κB, inflammation, immunity and cancer: coming of age. Nat Rev Immunol. 2018;18(5):309–324. doi: 10. 1038/nri.2017.142.
- Zhao Y, Simon M, Seluanov A, Gorbunova V. DNA damage and repair in age-related inflammation. Nat Rev Immunol. 2023;23 (2):75–89. doi: 10.1038/s41577-022-00751-y.
- Klapp V, Álvarez-Abril B, Leuzzi G, Kroemer G, Ciccia A, Galluzzi L. The DNA damage response and inflammation in cancer. Cancer Discov. 2023;13(7):1521–1545. doi: 10.1158/2159-8290.CD-22-1220.
- 77. Huang TT, Wuerzberger-Davis SM, Wu ZH, Miyamoto S. Sequential modification of NEMO/IKKγ by SUMO-1 and ubiquitin mediates NF-κB activation by genotoxic stress. Cell. 2003;115 (5):565–576. doi: 10.1016/S0092-8674(03)00895-X.
- Allard B, Allard D, Buisseret L, Stagg J. The adenosine pathway in immuno-oncology. Nat Rev Clin Oncol. 2020;17(10):611–629. doi: 10.1038/s41571-020-0382-2.
- Kraehenbuehl L, Weng CH, Eghbali S, Wolchok JD, Merghoub T. Enhancing immunotherapy in cancer by targeting emerging immunomodulatory pathways. Nat Rev Clin Oncol. 2022;19 (1):37–50. doi: 10.1038/s41571-021-00552-7.
- De Martino M, Rathmell JC, Galluzzi L, Vanpouille-Box C. Cancer cell metabolism and antitumour immunity. Nat Rev Immunol. 2024;24(9):654–669. doi: 10.1038/s41577-024-01026-4.
- Stagg J, Golden E, Wennerberg E, Demaria S. The interplay between the DNA damage response and ectonucleotidases modulates tumor response to therapy. Sci Immunol. 2023;8(85): eabq3015. doi: 10.1126/sciimmunol.abq3015.
- Wennerberg E, Spada S, Rudqvist N-P, Lhuillier C, Gruber S, Chen Q, Zhang F, Zhou XK, Gross SS, Formenti SC, et al. CD73 blockade promotes dendritic cell infiltration of irradiated tumors and tumor rejection. Cancer Immunol Res. 2020;8(4):465–478. doi: 10.1158/2326-6066.CIR-19-0449.
- Tsukui H, Horie H, Koinuma K, Ohzawa H, Sakuma Y, Hosoya Y, Yamaguchi H, Yoshimura K, Lefor AK, Sata N, et al. CD73 blockade enhances the local and abscopal effects of radiotherapy in a murine rectal cancer model. BMC Cancer. 2020;20(1):411. doi: 10.1186/s12885-020-06893-3.
- 84. Wainberg ZA, Han S-W, Lee S, Lee K-W, Kopetz S, Mizrahi J, Hong YS, Ghiringhelli F, Italiano A, Tougeron D, et al. ARC-9: a randomized study to evaluate etrumadenant based treatment combinations in previously treated metastatic colorectal cancer (mCRC). J Clin Oncol. 2024;42(16_suppl):3508–3508. doi: 10. 1200/JCO.2024.42.16_suppl.3508.
- 85. Kurago Z, Guo G, Shi H, Bollag RJ, Groves MW, Byrd JK, Cui Y. Inhibitors of the CD73-adenosinergic checkpoint as promising

combinatory agents for conventional and advanced cancer immunotherapy. Front Immunol. 2023;14:1212209. doi: 10.3389/fimmu.2023.1212209.

- Lindholm HT, Chen R, De Carvalho DD. Endogenous retroelements as alarms for disruptions to cellular homeostasis. Trends Cancer. 2023;9(1):55–68. doi: 10.1016/j.trecan.2022.09.001.
- Charpentier M, Spada S, Van Nest SJ, Demaria S. Radiation therapy-induced remodeling of the tumor immune microenvironment. Semin Cancer Biol. 2022;86:737–747. doi: 10. 1016/j.semcancer.2022.04.003.
- 88. Wilkins A, Fontana E, Nyamundanda G, Ragulan C, Patil Y, Mansfield D, Kingston J, Errington-Mais F, Bottomley D, von Loga K, et al. Differential and longitudinal immune gene patterns associated with reprogrammed microenvironment and viral mimicry in response to neoadjuvant radiotherapy in rectal cancer. J For Immunother Of Cancer. 2021;9(3):e001717. doi: 10.1136/jitc-2020-001717.
- Shalhout SZ, Miller DM, Emerick KS, Kaufman HL. Therapy with oncolytic viruses: progress and challenges. Nat Rev Clin Oncol. 2023;20(3):160–177. doi: 10.1038/s41571-022-00719-w.
- Kaufman HL, Maciorowski D. Advancing oncolytic virus therapy by understanding the biology. Nat Rev Clin Oncol. 2021;18 (4):197–198. doi: 10.1038/s41571-021-00490-4.
- 91. Galluzzi L, Guilbaud E, Schmidt D, Kroemer G, Marincola FM. Targeting immunogenic cell stress and death for cancer therapy. Nat Rev Drug Discov. 2024;23(6):445–460. doi: 10.1038/s41573-024-00920-9.
- 92. Wongariyapak A, Roulstone V, Melcher AA, Pedersen M, Harrington KJ. Combination strategies incorporating oncolytic viruses and immune checkpoint inhibitors for advanced melanoma: what is the evidence? Ann Transl Med. 2023;11(10):369. doi: 10.21037/atm-2023-5.
- 93. Chiu M, Armstrong EJL, Jennings V, Foo S, Crespo-Rodriguez E, Bozhanova G, Patin EC, McLaughlin M, Mansfield D, Baker G, et al. Combination therapy with oncolytic viruses and immune checkpoint inhibitors. Expert Opin Biol Ther. 2020;20 (6):635–652. doi: 10.1080/14712598.2020.1729351.
- 94. Crespo-Rodriguez E, Bergerhoff K, Bozhanova G, Foo S, Patin EC, Whittock H, Buus R, Haider S, Muirhead G, Thway K, et al. Combining BRAF inhibition with oncolytic herpes simplex virus enhances the immune-mediated antitumor therapy of BRAF-mutant thyroid cancer. J For Immunother Of Cancer. 2020;8(2):e000698. doi: 10.1136/jitc-2020-000698.
- Galluzzi L, Smith KN, Liston A, Garg AD. The diversity of CD8+ T cell dysfunction in cancer and viral infection. Nat Rev Immunol. 2025; doi: 10.1038/s41577-025-01161-6.
- Wakefield LM, Hill CS. Beyond TGFβ: roles of other TGFβ superfamily members in cancer. Nat Rev Cancer. 2013;13(5):328–341. doi: 10.1038/nrc3500.
- 97. Sancho E, Batlle E, Clevers H. Signaling pathways in intestinal development and cancer. Annu Rev Cell Dev Biol. 2004;20 (1):695–723. doi: 10.1146/annurev.cellbio.20.010403.092805.
- 98. Richardson L, Wilcockson SG, Guglielmi L, Hill CS. Contextdependent TGF β family signalling in cell fate regulation. Nat Rev Mol Cell Biol. 2023;24(12):876–894. doi: 10.1038/s41580-023-00638-3.
- Li MO, Wan YY, Sanjabi S, Robertson AK, Flavell RA. transforming growth factor-β regulation of immune responses. Annu Rev Immunol. 2006;24(1):99–146. doi: 10.1146/annurev.immunol.24. 021605.090737.
- 100. Nixon BG, Gao S, Wang X, Li MO. TGFβ control of immune responses in cancer: a holistic immuno-oncology perspective. Nat Rev Immunol. 2023;23(6):346–362. doi: 10.1038/s41577-022-00796-z.
- 101. Tauriello DVF, Sancho E, Batlle E. Overcoming TGFβ-mediated immune evasion in cancer. Nat Rev Cancer. 2022;22(1):25–44. doi: 10.1038/s41568-021-00413-6.
- 102. Chen Y, McAndrews KM, Kalluri R. Clinical and therapeutic relevance of cancer-associated fibroblasts. Nat Rev Clin Oncol. 2021;18(12):792–804. doi: 10.1038/s41571-021-00546-5.

- 103. Young KH, Newell P, Cottam B, Friedman D, Savage T, Baird JR, Akporiaye E, Gough MJ, Crittenden M. TGFβ inhibition prior to hypofractionated radiation enhances efficacy in preclinical models. Cancer Immunol Res. 2014;2(10):1011–1022. doi: 10.1158/2326-6066.CIR-13-0207.
- 104. Vanpouille-Box C, Diamond JM, Pilones KA, Zavadil J, Babb JS, Formenti SC, Barcellos-Hoff MH, Demaria S. TGFβ is a Master regulator of radiation therapy-induced antitumor immunity. Cancer Res. 2015;75(11):2232–2242. doi: 10.1158/0008-5472. CAN-14-3511.
- 105. Rodríguez-Ruiz ME, Rodríguez I, Mayorga L, Labiano T, Barbes B, Etxeberria I, Ponz-Sarvise M, Azpilikueta A, Bolaños E, Sanmamed MF, et al. TGFβ blockade enhances radiotherapy abscopal efficacy effects in combination with anti-PD1 and anti-CD137 immunostimulatory monoclonal antibodies. Mol Cancer Ther. 2019;18(3):621–631. doi: 10.1158/1535-7163.MCT-18-0558.
- 106. Gunderson AJ, Yamazaki T, McCarty K, Fox N, Phillips M, Alice A, Blair T, Whiteford M, O'Brien D, Ahmad R, et al. TGFβ suppresses CD8+ T cell expression of CXCR3 and tumor trafficking. Nat Commun. 2020;11(1):1749. doi: 10.1038/s41467-020-15404-8.
- 107. Hamon P, Gerbé De Thoré M, Classe M, Signolle N, Liu W, Bawa O, Meziani L, Clémenson C, Milliat F, Deutsch E, et al. TGFβ receptor inhibition unleashes interferon-β production by tumor-associated macrophages and enhances radiotherapy efficacy. J For Immunother Of Cancer. 2022;10(3):e003519. doi: 10. 1136/jitc-2021-003519.
- 108. Yamazaki T, Gunderson AJ, Gilchrist M, Whiteford M, Kiely MX, Hayman A, O'Brien D, Ahmad R, Manchio JV, Fox N, et al. Galunisertib plus neoadjuvant chemoradiotherapy in patients with locally advanced rectal cancer: a single-arm, phase 2 trial. The Lancet Oncol. 2022;23(9):1189–1200. doi: 10.1016/S1470-2045(22)00446-6.
- 109. Young KH, Gunderson AJ, Gilchrist M, Whiteford M, Kiely MX, Hayman A, O'Brien DP, Ahmad R, Manchio JV, Brosnan E, et al. A phase II trial of TGF β type I receptor inhibitor, galunisertib, plus neoadjuvant chemoradiation in patients with locally advanced rectal cancer. J Clin Oncol. 2022;40(16_suppl):3617–3617. doi: 10.1200/JCO.2022.40.16_suppl.3617.
- 110. Ho AY, Shiao S, Kobald SA, Chen J, Duda DG, Ly A, Bossuyt V, Cho HL, Arnold B, Knott S, et al. PEARL: a phase Ib/II biomarker study of adding radiation therapy to pembrolizumab before neoadjuvant chemotherapy in human epidermal growth factor receptor 2-negative breast cancer. J Clin Oncol. 2024;42(36):4282-4293. doi: 10.1200/JCO.24.00003.
- 111. Connolly JJ, Spring LM, Taghian AG, Gadd M, Warren L, Garrido-Castro AC, King T, Mittendorf EA, Leone JP, Casey DL, et al. Abstract OT3-15-01: TBCRC-053: P-RAD: a randomized study of preoperative chemotherapy, pembrolizumab and No, low or high dose RADiation in Node-Positive, HER2-negative breast cancer. Cancer Res. 2023;83(5_Supplement):OT3-15-01-OT13-15-01. doi: 10.1158/1538-7445.SABCS22-OT3-15-01.
- 112. Petroni G, Galluzzi L. Impact of treatment schedule on the efficacy of cytostatic and immunostimulatory agents. Oncoimmunology. 2021;10(1):1889101. doi: 10.1080/2162402X. 2021.1889101.
- 113. Ho AY, Wright JL, Blitzblau RC, Mutter RW, Duda DG, Norton L, Bardia A, Spring L, Isakoff SJ, Chen JH, et al. Optimizing radiation therapy to boost systemic immune responses in breast cancer: a critical review for breast radiation oncologists. Int J Radiat Oncol Biol Phys. 2020;108(1):227–241. doi: 10.1016/j.ijrobp.2020.05.011.
- 114. Sen T, Takahashi N, Chakraborty S, Takebe N, Nassar AH, Karim NA, Puri S, Naqash AR. Emerging advances in defining the molecular and therapeutic landscape of small-cell lung cancer. Nat Rev Clin Oncol. 2024;21(8):610–627. doi: 10.1038/s41571-024-00914-x.
- Rudin CM, Brambilla E, Faivre-Finn C, Sage J. Small-cell lung cancer. Nat Rev Dis Primers. 2021;7(1). doi: 10.1038/s41572-020-00235-0.

- 116. Otano I, Ucero AC, Zugazagoitia J, Paz-Ares L. At the crossroads of immunotherapy for oncogene-addicted subsets of NSCLC. Nat Rev Clin Oncol. 2023;20(3):143–159. doi: 10.1038/s41571-022-00718-x.
- 117. Paz-Ares L, Champiat S, Lai WV, Izumi H, Govindan R, Boyer M, Hummel H-D, Borghaei H, Johnson ML, Steeghs N, et al. Tarlatamab, a first-in-class DLL3-targeted bispecific T-Cell engager, in recurrent Small-cell lung cancer: an open-label, phase I study. J Clin Oncol. 2023;41(16):2893–2903. doi: 10.1200/JCO.22.02823.
- 118. Nishiga Y, Drainas AP, Baron M, Bhattacharya D, Barkal AA, Ahrari Y, Mancusi R, Ross JB, Takahashi N, Thomas A, et al. Radiotherapy in combination with CD47 blockade elicits a macrophage-mediated abscopal effect. Nat Cancer. 2022;3 (11):1351–1366. doi: 10.1038/s43018-022-00456-0.
- Guilbaud E, Yamazaki T, Galluzzi L. T cell-independent abscopal responses to radiotherapy. Trends Cancer. 2023;9(2):93–95. doi: 10.1016/j.trecan.2022.12.005.
- 120. Hsieh RC, Krishnan S, Wu R-C, Boda AR, Liu A, Winkler M, Hsu W-H, Lin SH, Hung M-C, Chan L-C, et al. ATR-mediated CD47 and PD-L1 up-regulation restricts radiotherapy-induced immune priming and abscopal responses in colorectal cancer. Sci Immunol. 2022;7(72):eabl9330. doi: 10.1126/sciimmunol.abl9330.
- 121. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y, Kadel Iii EE, Koeppen H, Astarita JL, Cubas R, et al. TGF β attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells. Nature. 2018;554(7693):544–548. doi: 10.1038/nature25501.
- 122. Tauriello DVF, Palomo-Ponce S, Stork D, Berenguer-Llergo A, Badia-Ramentol J, Iglesias M, Sevillano M, Ibiza S, Cañellas A, Hernando-Momblona X, et al. TGFβ drives immune evasion in genetically reconstituted colon cancer metastasis. Nature. 2018;554 (7693):538–543. doi: 10.1038/nature25492.
- 123. Burley A, Rullan A, Wilkins A. A review of the biology and therapeutic implications of cancer-associated fibroblasts (CAFs) in muscle-invasive bladder cancer. Front Oncol. 2022;12:1000888. doi: 10.3389/fonc.2022.1000888.
- 124. Formenti SC, Lee P, Adams S, Goldberg JD, Li X, Xie MW, Ratikan JA, Felix C, Hwang L, Faull KF, et al. Focal irradiation and systemic TGFβ blockade in metastatic breast cancer. Clin Cancer Res. 2018;24(11):2493–2504. doi: 10.1158/1078-0432.CCR-17-3322.
- 125. Heras-Murillo I, Adán-Barrientos I, Galán M, Wculek SK, Sancho D. Dendritic cells as orchestrators of anticancer immunity and immunotherapy. Nat Rev Clin Oncol. 2024;21(4):257–277. doi: 10.1038/s41571-024-00859-1.
- 126. Lhuillier C, Rudqvist N-P, Yamazaki T, Zhang T, Charpentier M, Galluzzi L, Dephoure N, Clement CC, Santambrogio L, Zhou XK, et al. Radiotherapy-exposed CD8+ and CD4+ neoantigens enhance tumor control. J Clin Invest. 2021;131(5):e138740. doi: 10.1172/ JCI138740.
- 127. Yamazaki T, Kirchmair A, Sato A, Buqué A, Rybstein M, Petroni G, Bloy N, Finotello F, Stafford L, Navarro Manzano E, et al. Mitochondrial DNA drives abscopal responses to radiation that are inhibited by autophagy. Nat Immunol. 2020;21(10):1160–1171. doi: 10.1038/s41590-020-0751-0.
- 128. Rodriguez-Ruiz ME, Buqué A, Hensler M, Chen J, Bloy N, Petroni G, Sato A, Yamazaki T, Fucikova J, Galluzzi L. Apoptotic caspases inhibit abscopal responses to radiation and identify a new prognostic biomarker for breast cancer patients. Oncoimmunology. 2019;8(11):e1655964. doi: 10.1080/2162402X. 2019.1655964.
- 129. Vanpouille-Box C, Alard A, Aryankalayil MJ, Sarfraz Y, Diamond JM, Schneider RJ, Inghirami G, Coleman CN, Formenti SC, Demaria S. DNA exonuclease Trex1 regulates radiotherapy-induced tumour immunogenicity. Nat Commun. 2017;8(1):15618. doi: 10.1038/ncomms15618.
- 130. Rodriguez-Ruiz ME, Rodriguez I, Garasa S, Barbes B, Solorzano JL, Perez-Gracia JL, Labiano S, Sanmamed MF, Azpilikueta A, Bolaños E, et al. Abscopal effects of radiotherapy are enhanced by combined immunostimulatory mAbs and are dependent on CD8

T cells and Crosspriming. Cancer Res. 2016;76(20):5994–6005. doi: 10.1158/0008-5472.CAN-16-0549.

- 131. Chavez M, Silvestrini MT, Ingham ES, Fite BZ, Mahakian LM, Tam SM, Ilovitsh A, Monjazeb AM, Murphy WJ, Hubbard NE, et al. Distinct immune signatures in directly treated and distant tumors result from TLR adjuvants and focal ablation. Theranostics. 2018;8(13):3611–3628. doi: 10.7150/thno.25613.
- 132. Vanpouille-Box C, Hoffmann JA, Galluzzi L. Pharmacological modulation of nucleic acid sensors — therapeutic potential and persisting obstacles. Nat Rev Drug Discov. 2019;18(11):845–867. doi: 10.1038/s41573-019-0043-2.
- 133. Iribarren K, Bloy N, Buqué A, Cremer I, Eggermont A, Fridman WH, Fucikova J, Galon J, Špíšek R, Zitvogel L, et al. Trial watch: immunostimulation with Toll-like receptor agonists in cancer therapy. Oncoimmunology. 2016;5(3):e1088631. doi: 10. 1080/2162402X.2015.1088631.
- 134. Monjazeb AM, Kent MS, Grossenbacher SK, Mall C, Zamora AE, Mirsoian A, Chen M, Kol A, Shiao SL, Reddy A, et al. Blocking indolamine-2,3-dioxygenase rebound immune suppression boosts antitumor effects of radio-immunotherapy in murine models and spontaneous canine malignancies. Clin Cancer Res. 2016;22 (17):4328–4340. doi: 10.1158/1078-0432.CCR-15-3026.
- 135. Li A, Barsoumian HB, Schoenhals JE, Caetano MS, Wang X, Menon H, Valdecanas DR, Niknam S, Younes AI, Cortez MA, et al. IDO1 inhibition overcomes radiation-induced "rebound immune Suppression" by reducing numbers of IDO1-expressing myeloid-derived suppressor cells in the tumor microenvironment. Int J Radiat Oncol Biol Phys. 2019;104(4):903–912. doi: 10.1016/j. ijrobp.2019.03.022.
- 136. Pouget JP, Chan TA, Galluzzi L, Constanzo J. Radiopharmaceuticals as combinatorial partners for immune checkpoint inhibitors. Trends Cancer. 2023;9(11):968–981. doi: 10.1016/j.trecan.2023.07.014.
- 137. Constanzo J. Immunomodulatory effects of targeted radionuclide therapy. Int Rev Cell Mol Biol. 2023;378:105–136.
- Buzas EI. The roles of extracellular vesicles in the immune system. Nat Rev Immunol. 2023;23(4):236–250. doi: 10.1038/s41577-022-00763-8.
- 139. Patel RB, Hernandez R, Carlson P, Grudzinski J, Bates AM, Jagodinsky JC, Erbe A, Marsh IR, Arthur I, Aluicio-Sarduy E, et al. Low-dose targeted radionuclide therapy renders immunologically cold tumors responsive to immune checkpoint blockade. Sci Transl Med. 2021;13(602):eabb3631. doi: 10.1126/scitranslmed. abb3631.
- 140. Lejeune P, Cruciani V, Berg-Larsen A, Schlicker A, Mobergslien A, Bartnitzky L, Berndt S, Zitzmann-Kolbe S, Kamfenkel C, Stargard S, et al. Immunostimulatory effects of targeted thorium-227 conjugates as single agent and in combination with anti-PD-L1 therapy. J For Immunother Of Cancer. 2021;9(10): e002387. doi: 10.1136/jitc-2021-002387.
- 141. Spurr LF, Martinez CA, Kang W, Chen M, Zha Y, Hseu R, Gutiontov SI, Turchan WT, Lynch CM, Pointer KB, et al. Highly aneuploid non-small cell lung cancer shows enhanced responsiveness to concurrent radiation and immune checkpoint blockade. Nat Cancer. 2022;3(12):1498–1512. doi: 10.1038/s43018-022-00467-x.
- 142. Bestvina CM, Pointer KB, Karrison T, Al-Hallaq H, Hoffman PC, Jelinek MJ, Juloori A, Melotek JM, Murgu S, Partouche J, et al. A phase 1 trial of concurrent or sequential ipilimumab, nivolumab, and stereotactic body radiotherapy in patients with stage IV NSCLC study. J Thorac Oncol. 2022;17(1):130–140. doi: 10.1016/ j.jtho.2021.08.019.
- 143. Spurr LF, Pitroda SP. Clinical and molecular correlates of tumor aneuploidy in metastatic non-small cell lung cancer. Sci Rep. 2024;14(1):19375. doi: 10.1038/s41598-024-66062-5.
- 144. Budczies J, Kazdal D, Menzel M, Beck S, Kluck K, Altbürger C, Schwab C, Allgäuer M, Ahadova A, Kloor M, et al. Tumour mutational burden: clinical utility, challenges and emerging improvements. Nat Rev Clin Oncol. 2024;21(10):725–742. doi: 10.1038/s41571-024-00932-9.

- 145. Teillaud JL, Houel A, Panouillot M, Riffard C, Dieu-Nosjean MC. Tertiary lymphoid structures in anticancer immunity. Nat Rev Cancer. 2024;24(9):629–646. doi: 10.1038/s41568-024-00728-0.
- 146. Holder AM, Dedeilia A, Sierra-Davidson K, Cohen S, Liu D, Parikh A, Boland GM. Defining clinically useful biomarkers of immune checkpoint inhibitors in solid tumours. Nat Rev Cancer. 2024;24(7):498–512. doi: 10.1038/s41568-024-00705-7.
- 147. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS, et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science. 2015;348(6230):124–128. doi: 10.1126/ science.aaa1348.
- 148. Alban TJ, Riaz N, Parthasarathy P, Makarov V, Kendall S, Yoo S-K, Shah R, Weinhold N, Srivastava R, Ma X, et al. Neoantigen immunogenicity landscapes and evolution of tumor ecosystems during immunotherapy with nivolumab. Nat Med. 2024;30 (11):3209–3222. doi: 10.1038/s41591-024-03240-y.
- 149. Lhuillier C, Vanpouille-Box C, Galluzzi L, Formenti SC, Demaria S. Emerging biomarkers for the combination of radiotherapy and immune checkpoint blockers. Semin Cancer Biol. 2018;52:125–134. doi: 10.1016/j.semcancer.2017.12.007.
- 150. Chalabi M, Verschoor YL, Tan PB, Balduzzi S, Van Lent AU, Grootscholten C, Dokter S, Büller NV, Grotenhuis BA, Kuhlmann K, et al. Neoadjuvant immunotherapy in locally advanced mismatch repair-deficient colon cancer. N Engl J Med. 2024;390(21):1949–1958. doi: 10.1056/NEJMoa2400634.
- 151. Kroemer G, Chan TA, Eggermont AMM, Galluzzi L. Immunosurveillance in clinical cancer management. CA Cancer J Clin. 2024;74(2):187–202. doi: 10.3322/caac.21818.
- 152. Herpels M, Ishihara J, Sadanandam A. The clinical terrain of immunotherapies in heterogeneous pancreatic cancer: unravelling challenges and opportunities. J Pathol. 2023;260(5):533–550. doi: 10.1002/path.6171.
- 153. Young K, Lawlor RT, Ragulan C, Patil Y, Mafficini A, Bersani S, Antonello D, Mansfield D, Cingarlini S, Landoni L, et al. Immune landscape, evolution, hypoxia-mediated viral mimicry pathways and therapeutic potential in molecular subtypes of pancreatic neuroendocrine tumours. Gut. 2021;70(10):1904–1913. doi: 10.1136/ gutjnl-2020-321016.
- 154. Method of the Year 2024: spatial proteomics. Nat Methods. 2024;21(12):2195-2196. doi: 10.1038/s41592-024-02565-3.
- 155. Method of the Year 2023: methods for modeling development. Nat Methods. 2023;20(12):1831–1832. doi: 10.1038/s41592-023-02134-0.
- 156. Method of the Year 2020: spatially resolved transcriptomics. Nat Methods. 2021;18(1):1–1. doi: 10.1038/s41592-020-01042-x.
- 157. Sadanandam A, Lyssiotis CA, Homicsko K, Collisson EA, Gibb WJ, Wullschleger S, Ostos LCG, Lannon WA, Grotzinger C, Del Rio M, et al. A colorectal cancer classification system that associates cellular phenotype and responses to therapy. Nat Med. 2013;19 (5):619–625. doi: 10.1038/nm.3175.
- 158. Guinney J, Dienstmann R, Wang X, de Reyniès A, Schlicker A, Soneson C, Marisa L, Roepman P, Nyamundanda G, Angelino P, et al. The consensus molecular subtypes of colorectal cancer. Nat Med. 2015;21(11):1350–1356. doi: 10.1038/nm.3967.
- Chen J, Larsson L, Swarbrick A, Lundeberg J. Spatial landscapes of cancers: insights and opportunities. Nat Rev Clin Oncol. 2024;21 (9):660–674. doi: 10.1038/s41571-024-00926-7.
- 160. Dhainaut M, Rose SA, Akturk G, Wroblewska A, Nielsen SR, Park ES, Buckup M, Roudko V, Pia L, Sweeney R, et al. Spatial CRISPR genomics identifies regulators of the tumor microenvironment. Cell. 2022;185(7):1223–1239.e20. doi: 10. 1016/j.cell.2022.02.015.
- 161. Wroblewska A, Dhainaut M, Ben-Zvi B, Rose SA, Park ES, Amir EAD, Bektesevic A, Baccarini A, Merad M, Rahman AH, et al. Protein barcodes enable high-dimensional single-cell CRISPR screens. Cell. 2018;175(4):1141–1155.e16. doi: 10.1016/j.cell.2018. 09.022.
- 162. Mollaoglu G, Tepper A, Falcomatà C, Potak HT, Pia L, Amabile A, Mateus-Tique J, Rabinovich N, Park MD,

LaMarche NM, et al. Ovarian cancer-derived IL-4 promotes immunotherapy resistance. Cell. 2024;187(26):7492–7510.e22. doi: 10.1016/j.cell.2024.10.006.

- 163. Kepp O, Bezu L, Yamazaki T, Di Virgilio F, Smyth MJ, Kroemer G, Galluzzi L. ATP and cancer immunosurveillance. The EMBO J. 2021;40(13):e108130. doi: 10.15252/embj.2021108130.
- 164. Vardam-Kaur T, Sun J, Borges da Silva H. Metabolic regulation of tissue-resident memory CD8(+) T cells. Curr Opin In Pharmacol. 2021;57:117–124. doi: 10.1016/j.coph.2021.02.004.
- 165. Wennerberg E, Mukherjee S, Spada S, Hung C, Agrusa CJ, Chen C, Valeta-Magara A, Rudqvist N-P, Van Nest SJ, Kamel MK, et al. Expression of the mono-ADP-ribosyltransferase ART1 by tumor cells mediates immune resistance in non-small cell lung cancer. Sci Transl Med. 2022;14(636):eabe8195. doi: 10.1126/scitranslmed. abe8195.
- 166. Altorki NK, McGraw TE, Borczuk AC, Saxena A, Port JL, Stiles BM, Lee BE, Sanfilippo NJ, Scheff RJ, Pua BB, et al. Neoadjuvant durvalumab with or without stereotactic body radiotherapy in patients with early-stage non-small-cell lung cancer: a single-centre, randomised phase 2 trial. The Lancet Oncol. 2021;22 (6):824–835. doi: 10.1016/S1470-2045(21)00149-2.
- 167. Zhou H, Tu C, Yang P, Li J, Kepp O, Li H, Zhang L, Zhang L, Zhao Y, Zhang T, et al. Carbon ion radiotherapy triggers immunogenic cell death and sensitizes melanoma to anti-PD-1 therapy in mice. Oncoimmunology. 2022;11(1):2057892. doi: 10.1080/ 2162402X.2022.2057892.
- 168. Zhou H, Yang P, Li H, Zhang L, Li J, Zhang T, Sheng C, Wang J. Carbon ion radiotherapy boosts anti-tumour immune responses by inhibiting myeloid-derived suppressor cells in melanoma-bearing mice. Cell Death Discov. 2021;7(1):332. doi: 10.1038/s41420-021-00731-6.
- 169. Mellman I, Chen DS, Powles T, Turley SJ. The cancer-immunity cycle: indication, genotype, and immunotype. Immunity. 2023;56 (10):2188–2205. doi: 10.1016/j.immuni.2023.09.011.
- 170. Golden EB, Marciscano AE, Formenti SC. Radiation therapy and the in situ vaccination approach. Int J Radiat Oncol Biol Phys. 2020;108(4):891–898. doi: 10.1016/j.ijrobp.2020.08.023.
- 171. Altorki NK, Walsh ZH, Melms JC, Port JL, Lee BE, Nasar A, Spinelli C, Caprio L, Rogava M, Ho P, et al. Neoadjuvant durvalumab plus radiation versus durvalumab alone in stages I–III nonsmall cell lung cancer: survival outcomes and molecular correlates of a randomized phase II trial. Nat Commun. 2023;14(1):8435. doi: 10.1038/s41467-023-44195-x.
- 172. Iyengar P, Hu C, Gomez DR, Timmerman RD, Simone CB, Robinson CG, Gerber DE, Waqar SN, Donington J, Swisher S, et al. NRG-LU002: randomized phase II/III trial of maintenance systemic therapy versus local consolidative therapy (LCT) plus maintenance systemic therapy for limited metastatic non-small cell lung cancer (NSCLC). J Clin Oncol. 2024;42(16_suppl):8506--8506. doi: 10.1200/JCO.2024.42.16_suppl.8506.
- 173. Diorio C, Teachey DT, Grupp SA. Allogeneic chimeric antigen receptor cell therapies for cancer: progress made and remaining roadblocks. Nat Rev Clin Oncol. 2025;22(1):10–27. doi: 10.1038/ s41571-024-00959-y.
- 174. Albelda SM. CAR T cell therapy for patients with solid tumours: key lessons to learn and unlearn. Nat Rev Clin Oncol. 2024;21 (1):47-66. doi: 10.1038/s41571-023-00832-4.
- 175. Tong L, Jiménez-Cortegana C, Tay AHM, Wickström S, Galluzzi L, Lundqvist A. NK cells and solid tumors: therapeutic potential and persisting obstacles. Mol Cancer. 2022;21(1):206. doi: 10.1186/ s12943-022-01672-z.
- 176. Kostopoulos N, Costabile F, Krimitza E, Beghi S, Goia D, Perales-Linares R, Thyfronitis G, LaRiviere MJ, Chong EA, Schuster SJ, et al. Local radiation enhances systemic CAR T-cell efficacy by augmenting antigen crosspresentation and T-cell infiltration. Blood Adv. 2024;8(24):6308–6320. doi: 10.1182/bloodadvances. 2024012599.
- 177. Jimenez-Labaig P, Rullan A, Braña I, Hernando-Calvo A, Moreno V, Doger B, Bitar G, Ap Dafydd D, Melcher A, Harrington KJ. Intratumoral therapies in head and neck squamous

cell carcinoma: a systematic review and future perspectives. Cancer Treat Rev. 2024;127:102746. doi: 10.1016/j.ctrv.2024.102746.

- 178. Battula S, Papastoitsis G, Kaufman HL, Wittrup KD, Schmidt MM. Intratumoral aluminum hydroxide-anchored IL-12 drives potent antitumor activity by remodeling the tumor microenvironment. JCI Insight. 2023;8(23):e168224. doi: 10.1172/jci.insight.168224.
- 179. Bonvalot S, Le Pechoux C, De Baere T, Kantor G, Buy X, Stoeckle E, Terrier P, Sargos P, Coindre JM, Lassau N, et al. Firstin-human study testing a New Radioenhancer using Nanoparticles (NBTXR3) activated by radiation therapy in patients with locally advanced soft tissue sarcomas. Clin Cancer Res. 2017;23 (4):908–917. doi: 10.1158/1078-0432.CCR-16-1297.
- 180. Zhang P, Darmon A, Marill J, Mohamed Anesary N, Paris S. radiotherapy-activated hafnium oxide nanoparticles produce abscopal effect in a mouse colorectal cancer Model. Int J Nanomed. 2020;15:3843–3850. doi: 10.2147/IJN.S250490.
- 181. Hu Y, Paris S, Barsoumian H, Abana CO, He K, Wasley M, Younes AI, Masrorpour F, Chen D, Yang L, et al. Radiation therapy enhanced by NBTXR3 nanoparticles overcomes anti-PD1 resistance and evokes abscopal effects. Int J Radiat Oncol Biol Phys. 2021;111(3):647–657. doi: 10.1016/j.ijrobp.2021.06.041.
- 182. Chand D, Savitsky DA, Krishnan S, Mednick G, Delepine C, Garcia-Broncano P, Soh KT, Wu W, Wilkens MK, Udartseva O, et al. Botensilimab, an Fc-enhanced Anti–CTLA-4 antibody, is effective against tumors poorly responsive to conventional immunotherapy. Cancer Discov. 2024;14(12):2407–2429. doi: 10.1158/ 2159-8290.CD-24-0190.
- 183. Bullock AJ, Schlechter BL, Fakih MG, Tsimberidou AM, Grossman JE, Gordon MS, Wilky BA, Pimentel A, Mahadevan D, Balmanoukian AS, et al. Botensilimab plus balstilimab in relapsed/ refractory microsatellite stable metastatic colorectal cancer: a phase

1 trial. Nat Med. 2024;30(9):2558–2567. doi: 10.1038/s41591-024-03083-7.

- 184. Tolcher AW, Gordon M, Mahoney KM, Seto A, Zavodovskaya M, Hsueh C-H, Zhai S, Tarnowski T, Jürgensmeier JM, Stinson S, et al. Phase 1 first-in-human study of dalutrafusp alfa, an anti–CD73-TGF-β-trap bifunctional antibody, in patients with advanced solid tumors. J For Immunother Of Cancer. 2023;11(2):e005267. doi: 10. 1136/jitc-2022-005267.
- 185. Aranda F, Vacchelli E, Eggermont A, Galon J, Fridman WH, Zitvogel L, Kroemer G, Galluzzi L. Trial watch: immunostimulatory monoclonal antibodies in cancer therapy. Oncoimmunology. 2014;3(2):e27297. doi: 10.4161/onci.27297.
- 186. Barve MA, Tolcher AW, Carvajal RD, Izar B, El-Khoueiry AB, Hanna DL, Tarhini AA, Whitman ED, Bhatia S, Davar D, et al. A phase 1 study of AGEN2373, a novel CD137 agonist antibody designed to avoid hepatoxicity, in patients with advanced solid tumors. J Clin Oncol. 2023;41(16_suppl):2524–2524. doi: 10. 1200/JCO.2023.41.16_suppl.2524.
- 187. Tolcher AW, Carvajal RD, El-Khoueiry AB, Ortuzar Feliu W, Zang H, Ancukiewicz M, Shapiro I, Strauss JF. Initial findings of the first-in-human phase I study of AGEN2373, a conditionally active CD137 agonist antibody, in patients (pts) with advanced solid tumors. J Clin Oncol. 2021;39(15_suppl):2634–2634. doi: 10.1200/JCO.2021.39.15_suppl.2634.
- 188. Hadfield MJ, Safran H, Purbhoo MA, Grossman JE, Buell JS, Carneiro BA. Overcoming resistance to programmed cell death protein 1 (PD-1) blockade with allogeneic invariant natural killer T-cells (iNKT). Oncogene. 2024;43(10):758–762. doi: 10.1038/ s41388-024-02948-y.
- Yigit B. 164 AgenT-797, a novel allogenic and 'off-the shelf' iNKT cell therapy promotes effective tumor killing. J For Immunother Of Cancer. 2020;8:A98–A98.