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## A Revised Prognostic Model for Patients with Acute Myeloid Leukemia and First Relapse

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#### Abstract:

Most patients with acute myeloid leukemia (AML) may obtain remission upon induction chemotherapy, but relapse is frequent and associated with poor survival. Previous prognostic models for outcomes after relapse lacked analysis of comprehensive molecular data. A validated prognostic model integrating clinical, cytogenetic, and molecular variables may support treatment decisions. We studied 943 AML patients who relapsed after first-line intensive induction treatment in a development cohort (HOVON-SAKK). A random survival forest algorithm was used to evaluate the association of clinical parameters, cytogenetic abnormalities, and molecular variables at diagnosis with overall survival (OS). Relapsing patients (n=377) who were enrolled in the NCRI-AML18 trial were used for model validation. In the development cohort, the median age at relapse was 58 years, and patients were classified as 2022 ELN favorable (22%), intermediate (31%), and adverse (48%) risk. One-third underwent allogeneic transplantation in first complete remission. Variable selection yielded nine variables significantly associated with 1-year OS, including relapse-free interval, age, white blood cell count, mutated TP53, FLT3-ITD, core-binding factor abnormalities, t(v;11q23)/KMT2A-rearranged and complex/monosomal karyotype, which were assigned points according to their estimated hazard ratios. Three prognostic groups were defined with distinct 1-year OS in both development (favorable:  $51\pm3$ %, intermediate:  $29\pm3$ % and poor:  $14\pm2$ %, respectively) and validation cohorts (51±4%, 26±5% and 14±3%, respectively). Independent validation confirmed the improved accuracy in predicting outcomes for AML patients in first relapse. The revised AML relapse model improved on previous classification systems for prognostication of outcomes after first AML relapse. It provides stratification which might support tailoring second line treatment.

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2 Relapse

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## 39 KEY POINTS

- 40 The revised AML relapse model includes nine predictors, grouped into three risk
- 41 categories which are each associated with distinct OS
- The model outperformed prior scoring systems and was validated in an independent
- 43 cohort of AML relapse patients

#### 44 ABSTRACT

45 Most patients with acute myeloid leukemia (AML) may obtain remission upon induction 46 chemotherapy, but relapse is frequent and associated with poor survival. Previous prognostic 47 models for outcomes after relapse lacked analysis of comprehensive molecular data. A 48 validated prognostic model integrating clinical, cytogenetic, and molecular variables may 49 support treatment decisions. We studied 943 AML patients who relapsed after first-line intensive 50 induction treatment in a development cohort (HOVON-SAKK). A random survival forest 51 algorithm was used to evaluate the association of clinical parameters, cytogenetic abnormalities, and molecular variables at diagnosis with overall survival (OS). Relapsing 52 53 patients (n=377) who were enrolled in the NCRI-AML18 trial were used for model validation. In 54 the development cohort, the median age at relapse was 58 years, and patients were classified as 2022 ELN favorable (22%), intermediate (31%), and adverse (48%) risk. One-third 55 56 underwent allogeneic transplantation in first complete remission. Variable selection yielded nine variables significantly associated with 1-year OS, including relapse-free interval, age, white 57 58 blood cell count, mutated TP53, FLT3-ITD, core-binding abnormalities, factor 59 t(v:11q23)/KMT2A-rearranged and complex/monosomal karyotype, which were assigned points according to their estimated hazard ratios. Three prognostic groups were defined with distinct 1-60 61 year OS in both development (favorable: 51±3%, intermediate: 29±3% and poor: 14±2%, 62 respectively) and validation cohorts (51±4%, 26±5% and 14±3%, respectively). Independent 63 validation confirmed the improved accuracy in predicting outcomes for AML patients in first relapse. The revised AML relapse model improved on previous classification systems for 64 65 prognostication of outcomes after first AML relapse. It provides stratification which might support tailoring second line treatment. 66

#### 67 INTRODUCTION

68 Disease recurrence is the most common cause of treatment failure in patients with acute 69 myeloid leukemia (AML). Treatment of relapsed AML remains challenging and outcome has been traditionally poor with median overall survival (OS) of less than 6 months.<sup>1-5</sup> Although 70 outcome has been improved with novel targeted treatments,<sup>6-9</sup> allogeneic hematopoietic cell 71 transplantation (allo-HCT) remains the only curative treatment for relapsing patients.<sup>10,11</sup> Risk 72 73 models for these patients might support decision making for reinduction treatment and 74 consolidation treatment with allo-HCT, whereas alternative, experimental strategies might be 75 considered for high-risk patients.

76 Prognostic factors associated with survival after relapse in AML have previously been identified.<sup>1,2,5,12-14</sup> These include the time between the first complete remission (CR1) and 77 78 relapse, age at relapse, the presence of favorable and unfavorable karvotypes, the presence of FLT3 internal tandem duplications (ITD), and prior allo-HCT.<sup>1,2,5,12-14</sup> Earlier developed 79 prognostic models<sup>1,2</sup> groups have highlighted these factors as significant predictors associated 80 with survival after first AML relapse. Since the development of these risk models, an increasing 81 82 number of molecular and cytogenetic alterations has been recognized impacting outcome in newly diagnosed AML patients,<sup>11,15</sup> which might also affect survival after first relapse. Models 83 84 that allow for risk stratification after first relapse using cytogenetic and molecular variables 85 established at initial diagnosis in a recently treated AML patient cohort are currently lacking.

We set out to study a large cohort of AML patients who relapsed after intensive induction treatment and for whom baseline comprehensive clinical, cytogenetic and molecular variables were available. We developed a revised stratification system which was validated in an independent cohort of older AML patients.

#### 90 METHODS

#### 91 Clinical cohorts

92 The complete cohort consisted of 5086 patients aged 18 years and older enrolled in eight 93 consecutive HOVON-SAKK (HO42, HO42A, HO43, HO81, HO92, HO102, HO103, HO132, 94 Supplementary Materials) clinical trials between 2000 and 2018 with an intensive induction chemotherapy backbone for newly diagnosed AML and high-risk myelodysplastic syndrome 95 96 (MDS) with IPSS ≥1.5, IPSS-R risk score >4.5 or excess blasts of ≥10% (see Figure S1A-H for trial details).<sup>16-23</sup> Post-remission treatment with either additional chemotherapy, high-dose 97 98 chemotherapy followed by autologous HCT, or allo-HCT was based on AML risk in evolving classifications.<sup>11,24</sup> Patients were excluded because of no next-generation sequencing data 99 100 available at diagnosis (n=2543), no CR or CR with incomplete count recovery (CRi) following 101 two cycles of induction chemotherapy (n=416), and no relapse (n=1184). The final development 102 cohort consisted of 943 patients who developed a first relapse (Figure 1). The validation cohort 103 included newly diagnosed AML and high-risk MDS patients with excess blasts of ≥10% aged 60 104 years and older enrolled in the NCRI-AML18 trial who were treated with intensive induction chemotherapy between 2014 and 2018 (Figure S1I).<sup>25</sup> Patients were excluded because of no 105 106 CR or CR with incomplete count recovery (CRi) following two cycles of induction chemotherapy 107 (n=245), and no relapse (n=354). The final validation cohort consisted of 377 patients who 108 developed a first relapse (Figure 1). All trial participants provided written informed consent in 109 accordance with the Declaration of Helsinki.

#### 110 Genetic analysis

High molecular weight genomic DNA was extracted from bone marrow samples or peripheral
blood. Diagnostic samples at first AML diagnosis underwent gene re-sequencing using either a

113 97-gene panel (Oxford for UK-AML18) or a 54-gene panel (Erasmus MC, Rotterdam for 114 HOVON-SAKK), containing the most frequently mutated genes in myeloid malignancies. 115 Targeted next-generation sequencing (NGS) used liquid-phase capture of molecule-barcoded 116 libraries, with custom-designed probes targeting AML-associated genes for UK-AML18 (Roche, 117 Basel, Switzerland) or the Illumina TruSight Myeloid panel for HOVON-SAKK (Illumina, San 118 Diego, CA). Genomic libraries were sequenced on the Illumina platform, ensuring a minimum 119 mean target coverage of 500x. Details of cytogenetic analysis, NGS, and additional molecular analyses have been previously published.<sup>15</sup> 120

## 121 Statistical methods

122 The analysis adhered to the TRIPOD guideline for prediction model development and 123 validation.<sup>26</sup> A total of 45 variables (Table S1) were considered for the association with 1-year 124 OS in model development: age at relapse (≥60 vs. <60 years), relapse free interval (≤12 months 125 vs. >12 months) defined as the time between CR1 and relapse, number of treatment cycles needed to obtain CR1 (1 vs. >1), sex (male vs. female), white blood cell count (WBC) at 126 127 diagnosis (≥10 vs. <10 [x10<sup>9</sup>/L]), previous allo-HCT (yes vs. no), previous autologous HCT (yes 128 vs. no), 29 different gene mutations (presence vs. absence), and nine different cytogenetic 129 abnormalities (presence vs. absence) (see Table S1 for details of mutations and cytogenetic 130 variables). Complex and monosomal karyotype were grouped due to their frequent co-131 occurrence in this cohort (94% concordant). During variable selection, we evaluated different 132 thresholds for age, relapse free interval and WBC, and found that the most optimal cut-offs were 60 years, 12 months and 10 x10<sup>9</sup>/L, respectively. Gene mutations and cytogenetic abnormalities 133 134 were only evaluated if present in 10 or more patients (Table S1). OS was defined as the time 135 from the date of first relapse to either the date of death from any cause or the date of last 136 contact while still alive. OS curves were estimated using the Kaplan-Meier method and

137 compared using the log-rank test. Model development consisted of three sequential stages 138 which included: (1) variable selection using a random survival forest; (2) Cox regression 139 analysis with backward selection (P<0.10) to define the optimal model; and (3) assignment of 140 weights and defining groups based on significantly different OS compared to a lower score to 141 construct the final prognostic model. Further details are available in the Supplementary 142 Materials. The impact of allo-HCT after relapse was assessed using a time-dependent Cox 143 regression allowing the allo-HCT covariate to change the state at the time of allo-HCT. Fine-144 Gray models were used to estimate the probability of an event accounting for competing risks, 145 non-relapse mortality (NRM) and relapse. Prognostic accuracy of existing models and the 146 revised model was evaluated using Harrels' C-index. Cytogenetic data were not available in 38 147 patients (4.0%) due to failed karyotyping and 3 patients (0.3%) had missing WBC at diagnosis in 148 the development cohort. Missing cytogenetics data were uniformly imputed as absent, 149 consistent with real-world clinical practice, and with the median WBC count for missing WBC. All 150 analyses were performed in R version 4.4.1. The R script of the analyses can be found online 151 (https://github.com/niekvandermaas/AML-relapse-model).

## 152 **RESULTS**

## 153 Patient characteristics

154 The development cohort consisted of 943 patients who relapsed after having obtained CR1 155 upon initial intensive treatment (Table 1). In this cohort, 1-year OS was similar across treatment 156 periods (<2010 vs. ≥2010, Figure S2) and disease type (AML vs high-risk MDS, Figure S3). In 157 addition, no difference was observed for 1-year OS between missing and non-missing NGS 158 data (Figure S4). The median age of patients at relapse in this cohort was 58 (range 18-81 159 years, Table 1). At diagnosis, 22% of the patients were classified as having a favorable risk 160 according to the European LeukemiaNET (ELN) 2022 AML classification, 31% as intermediate risk, and 48% as adverse risk (Table 1).<sup>11</sup> In this cohort, 33% of patients received post-161 162 remission treatment with allo-HCT, while 14% underwent autologous HCT in first CR (Table 1). 163 A total of 95% relapsing patients had at least one cytogenetic abnormality or genetic mutation at 164 diagnosis (median 3), with 62% of patients having molecular mutations only, 3% having 165 karyotype alterations only, and 32% having both. Most mutations at diagnosis were found in the 166 following genes: DNMT3A (30%), NPM1 (27%), and FLT3-ITD (22%). Mutated TP53 was found 167 in 12% of patients. Complex and/or monosomal karyotype according to ELN 2022 was the most 168 frequent cytogenetic abnormality in these patients (19%), whereas core binding factor (CBF) abnormalities (t(16;16)/inv(16) [3%] and t(8;21) [2%]) were relatively infrequent (Figure 2).<sup>27</sup> The 169 170 development cohort had a median follow up from relapse of 49 months for patients alive (range: 171 0-128 months, Table 1). OS at 1 and 4 years after first relapse in the development cohort were 172 33±2% and 16±1%, respectively.

173 Patient outcomes by previous prognostic models

174 Previous prognostic models were assessed for their predictive performance in the development 175 cohort. The HOVON-SAKK model<sup>1</sup> was associated with a C-index of 0.65±0.016 for 1-year OS. 176 The majority of patients were classified as adverse risk in that model (684 out of 943, 73%). 1-177 year OS after relapse was clearly distinct estimating 74±7% for the favorable group, 54±3% for 178 the intermediate group, and 24±2% for the adverse group, respectively (Figure 3A). Similarly, the GOELAMS model<sup>2</sup> was associated with a C-index of 0.64±0.017, which outcomes were 179 180 distinct in the three risk groups associated with survival probabilities at 1-year 49±3% for the 181 favorable group, 29±2% for the intermediate group, and 11±3% for the adverse group (Figure 182 3B).

## 183 Development of a revised prognostic model for patients in first relapse

184 To determine the optimal patient-specific and genetic variables for risk stratification of OS, we used a random survival forest algorithm, which revealed the hierarchy of each clinical, genetic, 185 186 and cytogenetic variable for stratifying 1-year OS (details in Supplementary Materials). A total of 187 45 candidate variables were considered (Table S1), of which 18 were identified as significantly 188 impacting (P<0.01) 1-year OS (Figure S5). These were included in a multivariable Cox 189 regression analysis using a stepwise backward selection which resulted in a final model of 9 190 predictors. The three most important variables for 1-year OS after relapse were the absence of 191 CBF abnormalities (hazard ratio [HR] 2.07, 95% confidence interval [95%CI] 1.38-3.12, 192 P<0.001), mutated *TP53* (HR 1.99, 95%Cl 1.48-2.67, P<0.001) and relapse free interval ≤12 193 months (HR 1.76, 95%Cl 1.46-2.12, P<0.001) (Figure 4A). Notably after adjustment for 194 covariates, including mutated TP53, complex and/or monosomal karyotype was independently 195 correlated with adverse OS (HR 1.40 95%CI 1.08-1.82, P=0.01, Figure 4A). Each of the nine 196 variables was assigned points based on their rounded HRs (Figure 4A) resulting in 3 points for 197 either no CBF abnormalities or a TP53 mutation. Similarly, relapse free interval  $\leq 12$  months,

previous allo-HCT, t(v;11q23)/KMT2A-rearranged, and age at relapse  $\geq$ 60 years were each assigned 2 points, whereas WBC  $\geq$ 10 x10<sup>9</sup>/L, complex and/or monosomal karyotype and *FLT3*-ITD were assigned 1 point. These points were subsequently used to derive the total score for each patient. The median total score in the development cohort was 7 points (range 1-14). OS decreased with increasing scores (Figure S6). Based on significantly different OS, we collapsed the development cohort into three groups: favorable ( $\leq$ 6 points; n=389, 42%), intermediate (7 points; n=197, 21%), poor ( $\geq$ 8 points; n = 357, 38%, Figure S7).

205 Favorable risk patients in the revised model had a 1-year OS of 51±3% (Figure 4B). 206 Intermediate risk patients had a 1-year OS of 29±3%, which was 14±2% in the poor risk 207 subgroup (Figure 4B). The C-index of the prognostic model in the development cohort was 208 0.71±0.016 with excellent calibration (Figure S8). 4-year OS was also significantly different 209 amongst the three subgroups (29±2%, 11±2%, 5±1%, respectively, Figure S9A). The model 210 restratified 57% and 49% of patients from the previous HOVON-SAKK and GOELAMS risk 211 groups, respectively. The majority of patients in the large HOVON-SAKK poor risk group were 212 reclassified as favorable (26%) or intermediate (24%), whereas 30% and 42% of patients in the 213 large GOELAMS intermediate risk group were reclassified to the favorable and poor risk group 214 of the revised model, respectively (Figures 4C and 4D).

215 Model validation

The model was validated in an independent dataset derived from the NCRI-AML18 trial, consisting of 976 older newly diagnosed AML patients aged 60 years and older who also received intensive induction chemotherapy. A total of 377 relapsing patients were identified with a median age at relapse of 69 (range 60-81) years (Figure 5A). Sixty-two percent were classified as poor or very poor risk in the AML60+ classification at the time of diagnosis,<sup>15</sup> whereas 60% were adverse according to the 2022 ELN risk classification. Allo-HCT in CR1 was 222 applied in 14% as post-remission treatment (Figure 5A). A total of 97% of patients had at least 223 one cytogenetic or molecular abnormality (median 4), with 77% of patients having molecular 224 mutations only, 1% having karyotype alterations only, and 21% having both. The most 225 frequently mutated genes identified were DNMT3A (37%), NPM1 (27%) and ASXL1 (26%). 226 TP53 was mutated in 11% of patients. According to ELN 2022 criteria, the most common 227 cytogenetic abnormality was a complex and/or monosomal karyotype (14%), while CBF 228 abnormalities, specifically t(16;16)/inv(16) (1%) and t(8;21) (3%), were rare (Figure 5B). Median 229 follow up from relapse was 21 months for patients alive (range: 0-59 months, Figure 5A). OS at 230 1 and 4 years after first relapse year in the validation cohort were  $30\pm2\%$  and  $9\pm2\%$ , 231 respectively. The three groups defined in the development cohort were used to classify the 232 validation cohort: favorable (<6 points; n=137, 36%), intermediate (7 points; n=85, 23%), and 233 poor (≥8 points; n=155, 41%), which were associated with distinct 1-year and 4-year OS (1-year 234 OS: 51±4%, 26±5%, and 14±3%, respectively, Figure 5C and Figure S9B). The performance of 235 the revised prognostic model as measured by the C-index was 0.71±0.028 with excellent 236 calibration (Figure S10). The GOELAMS and HOVON-SAKK models had C-indices of 237 0.69±0.027 and 0.62±0.027 in the validation cohort, respectively. Similar to the development 238 cohort, the revised model restratified 59% from the original HOVON-SAKK risk groups and 35% 239 from the GOELAMS groups (Figures 5D-E). HOVON-SAKK poor risk patients were considered 240 favorable or intermediate in 24% and 26%, respectively, in the revised model.

241 Treatment after relapse

In the development cohort, reinduction treatment was used in 690 (73%) patients, which resulted in a second CR (CR2) in 348 (50%) of patients (Figure S11, Table S2). Across subgroups, CR2 after re-induction treatment was obtained in 65% and 44% of the favorable and intermediate risk patients, respectively, which was higher compared with the poor risk group

246 (32%, C-index 0.66, Table S2 and S3). Additionally, the revised model showed a distinct OS 247 and event-free survival (EFS) for patient that received reinduction treatment, and relapse-free 248 survival (RFS) for patients in CR2 (Figure S12 and Table S3). The revised model compared 249 favorably in terms of accuracy to these outcomes compared with the previous risk stratification 250 systems (Table S3). Allo-HCT was used as consolidation treatment of a CR2 in 159 (46%) 251 patients, and 38 (11%) received a donor lymphocyte infusion (DLI) after CR2 (Figure S11, Table S2). A total of 342 (50%) patients did not achieve a CR2 despite re-induction treatment 252 253 consisting of intensive chemotherapy (n=279, 82%), upfront allo-HCT (n=43, 13%), or DLI 254 (n=20, 6%) (Figure S11, Table S2).

A time-dependent analysis with allo-HCT as a time-varying covariate was performed for patients who had attained a CR2. OS was improved by allo-HCT compared with no allo-HCT (HR 0.54; 95%CI 0.37-0.79, P=0.001, Figure S13) and cumulative incidence of relapse was lower after allo-HCT compared with no allo-HCT (HR 0.58; 95%CI 0.38-0.88, P=0.011, Figure S13). In contrast, the cumulative incidence of NRM was higher in the allo-HCT group, although not significantly different (HR 1.30; 95%CI 0.61-2.80, P=0.499, Figure S13). Low patient numbers precluded an analysis of the impact of allo-HCT per risk group.

#### 262 **DISCUSSION**

Relapse in AML is frequent with low response rates to reinduction treatment and dismal OS.<sup>1-5,14</sup> 263 264 Recent prognostic models for relapsed AML patients incorporating comprehensive genetic data 265 are currently lacking. Therefore, it is necessary to reassess the prognostic value of clinical, 266 cytogenetic and molecular AML characteristics in patients with relapsed AML after intensive 267 induction treatment. We analyzed the genomic landscape and clinical outcomes of 943 AML 268 patients with a first relapse aiming to develop a simple prognostic classification system. Using a 269 machine-learning method for variable selection, nine variables were identified which were used 270 to stratify patients into three risk groups. Although age, relapse-free interval, CBF abnormalities, *FLT3*-ITD, and prior allo-HCT were confirmed from previous risk models,<sup>1,2,14</sup> we additionally 271 272 found WBC count, mutated TP53, t(v;11q23)/KMT2A-rearranged and complex/monosomal 273 karyotype. The prognostic model was associated with highly distinct OS in three risk groups. 274 The revised prognostic model was validated in an independent cohort of older AML patients 275 treated within the NCRI-AML18 trial indicating similar prognostic accuracy. It classifies patients 276 into other risk groups allowing for better discrimination of OS outcomes for AML patients 277 experiencing their first relapse compared with existing prognostic models, like those from the HOVON-SAKK and GOELAMS groups.<sup>1,2</sup> 278

Salvage treatment for patients with relapse AML includes high-dose chemotherapy, hypomethylating agents, or targeted treatments for patients with specific mutations followed by allo-HCT as consolidation treatment in patients who obtain CR2.<sup>10,11</sup> High-dose chemotherapy regimens (e.g., intermediate-dose cytarabine with or without anthracycline, or FLAG-Ida) are associated with remission rates of 20-65%, but also considerable toxicity and mortality (6-22%).<sup>28-33</sup> Although the type and intensity of chemotherapy-based reinduction treatment strategies were not available in our dataset, we observed a relatively high CR2 rate of 65% and

286 44% for favorable and intermediate risk patients, which was 32% for poor risk patients. 287 Alternative salvage treatment approaches might be considered such as targeted treatments with 288 or without hypomethylating agents. Gilteritinib has been approved for patients with relapsed FLT3 AML with CR rates of 34%.8 Similarly, both ivosidenib (IDH1 inhibitor) and enasidenib 289 290 (IDH2 inhibitor) offer 23-33% CR rates in refractory or relapsed patients with IDH1/2-mutated AML.<sup>6,7</sup> while menin-inhibitors show a CR rate of 30% in *KMT2A*-rearranged or *NPM1* mutated 291 leukemia.<sup>9</sup> Alternatively, combining venetoclax, a BCL2 inhibitor, with hypomethylating agents 292 293 presents as a potential alternative to attain CR2, despite the absence of trial data in the 294 relapsed setting.<sup>34-37</sup> Combination treatments of targeted drugs, hypomethylating agents and 295 venetoclax, or high-dose chemotherapy with venetoclax are currently investigated and early results have been encouraging.<sup>38-41</sup> Nevertheless, it remains largely unknown whether patient 296 297 outcomes will significantly improve with these novel treatment modalities. These recent 298 advancements may transform the therapeutic landscape of relapsed AML significantly in the 299 next years, making it a necessity to further validate this prognostic index to reflect emerging 300 treatment modalities in the future.

301 Historically, allo-HCT in CR2 has been the preferred approach for long-term survival. Our 302 analysis confirms that allo-HCT in CR2 provides a survival benefit with reduced relapse 303 incidence compared with non-allo-HCT treatments. Nonetheless, the benefit of allo-HCT in 304 terms of relapse reduction can be compromised by NRM, particularly in older patients or those with underlying comorbidities.<sup>42-44</sup> Although the risk of a second failure without allo-HCT is high 305 306 (85% in our study), it needs to be balanced against the risk of NRM as assessed by risk 307 scores.<sup>42-44</sup> Ultimately, the decision to proceed with allo-HCT after AML relapse requires a 308 personalized approach, depending on the specific characteristics of each patient and their 309 disease. Of note, data from the recent ASAP trial suggested that immediate allo-HCT might be 310 an alternative for fit patients with non-proliferative refractory/relapsed AML who have a stem cell

donor available.<sup>45</sup> However, the small number of relapsing patients in this study preclude robust
conclusions.

313 Our study has several limitations. The optimal external validation cohort may be debated, but 314 TRIPOD guidelines have recommended using a cohort that differs in key characteristics (e.g., age), treatment approaches, or time period.<sup>26</sup> The revised model was validated in an 315 316 independent cohort, in which patients received different intensive chemotherapy regimens and 317 were older compared with the development cohort. The model was associated with strong 318 performance across both cohort populations, without any limitations imposed by narrow age 319 restrictions, highlighting broad applicability in relapsing AML patients after first-line intensive 320 treatment. Secondly, intensive chemotherapy and targeted therapies might exert a selective 321 pressure to the clonal landscape of AML, leading to both treatment-sensitive and treatment-322 resistant cells. For example, RAS mutations that are acquired at relapse may confer a particularly therapy-resistant disease.<sup>46,47</sup> Leukemic transformation by RAS mutations exhibit 323 324 resistance to the BCL2 inhibitor venetoclax, driving clinical resistance, relapse and worse OS after relapse following venetoclax-based therapy.<sup>37,46</sup> This clonal evolution of the AML is not 325 326 accounted for as molecular and cytogenetic data were only available from the time of initial 327 diagnosis. The predictive accuracy and discriminatory power of the model, as measured by the 328 C-index, indicate that the model performs relatively good in distinguishing between patients with 329 different risk levels. The C-index quantifies the risk classification performance based on the 330 predicted risk, with values ranging from 0.5 (no better than random chance) to 1.0 (perfect 331 discrimination). A higher C-index indicates better model performance in correctly identifying 332 which patients are more likely to experience an event. The revised relapse model improved 333 upon previous risk classification systems, and its predictive capability may be further enhanced 334 by incorporating molecular and cytogenetic data at the time of relapse. Additionally, the model 335 was developed in cohort of patients who did not receive FLT3 inhibitors as part of the first-line

336 induction regimen. Although 26% of patients (22 out of 85) with FLT3-ITD received guizartinib 337 added to the intensive induction backbone in the validation cohort, the model needs further 338 validation in the era of FLT3 targeted combination therapies. Lastly, there is an inherent bias 339 regarding treatment decision making in the patient population after first relapse. High risk 340 patients might not have received treatment because of their adverse risk factors (e.g., mutated 341 TP53, short relapse free interval, older age, other adverse cytogenetics). For example, in the 342 poor risk group, only 215 patients (60%) received reinduction treatment. Among those, 343 response rates were very low (32%), with only 20 patients proceeding to allo-HCT in CR2, 344 suggesting that this is a particularly difficult AML population to treat.

345 In conclusion, the revised prognostic classification system for adult patients with AML in first 346 relapse offers a useful and distinctive model for clinical practice. It identifies favorable or 347 intermediate risk patients who may benefit from reinduction strategies and consolidation with 348 allo-HCT taking into account the risk of NRM. Conversely, patients classified within the poor risk 349 group have dismal survival after first relapse and might be considered for novel treatment 350 strategies, experimental treatments or even best supportive care. To facilitate the integration of this prognostic tool into clinical practice, an online calculator has been developed.<sup>48</sup> Based on a 351 352 patient-specific hematologic, cytogenetic, and molecular profile, the tool provides a personalized 353 prediction score, assigns an associated risk group, and offers estimated OS projections. This 354 tool may aid clinicians in evaluating the relative benefits of salvage and experimental 355 treatments, balancing these against potential treatment-related risks.

## 356 AUTHOR CONTRIBUTION STATEMENT

- N.M. and J.V. designed the study, analyzed the data, and wrote the manuscript; D.B., T.P., A.T.,
- 358 B.J.B., J.K., C.E., O.V., M.C.V., C.G., J.M., S.K., M.D., S.F., I.T., G.H., C.C., P.V., N.R., G.O.,
- and B.L. provided patient data; N.M., P.G., H.B.B., P.J.V. and J.V. collected and assembled
- 360 clinical, laboratory, and genetic data; N.G. and J.V. conceived the statistical plan and performed
- the statistical analysis; N.G., J.C. and J.V. interpreted data and contributed to research
- 362 discussion; and all authors reviewed the manuscript and approved the submission. N.G. and
- 363 J.V. had full access to all data.
- 364 CONFLICT-OF-INTEREST DISCLOSURE
- 365 None of the authors has a relevant conflict of interest.
- 366 DATA SHARING SECTION
- 367 Requests for data sharing may be submitted to Jurjen Versluis (j.versluis.1@erasmusmc.nl).

#### 368 **REFERENCES**

- Breems DA, Van Putten WLJ, Huijgens PC, et al: Prognostic index for adult patients with
   acute myeloid leukemia in first relapse. J Clin Oncol 23:1969-1978, 2005
- Chevallier P, Labopin M, Turlure P, et al: A new Leukemia Prognostic Scoring System
   for refractory/relapsed adult acute myelogeneous leukaemia patients: a GOELAMS
   study. Leukemia 25:939-944, 2011
- 374 3. Devillier R, Crocchiolo R, Etienne A, et al: Outcome of relapse after allogeneic stem cell 375 transplant in patients with acute myeloid leukemia. Leuk Lymphoma 54:1228-1234, 2013 376 Schmid C, de Wreede LC, van Biezen A, et al: Outcome after relapse of myelodysplastic 4. 377 syndrome and secondary acute myeloid leukemia following allogeneic stem cell 378 transplantation: a retrospective registry analysis on 698 patients by the Chronic 379 Malignancies Working Party of the European Society of Blood and Marrow 380 Transplantation. Haematologica 103:237-245, 2018
- 381 5. Ganzel C, Sun Z, Cripe LD, et al: Very poor long-term survival in past and more recent
  382 studies for relapsed AML patients: The ECOG-ACRIN experience. Am J Hematol
  383 93:1074-1081, 2018
- Stein EM, DiNardo CD, Pollyea DA, et al: Enasidenib in mutant IDH2 relapsed or
   refractory acute myeloid leukemia. Blood 130:722-731, 2017
- 386 7. DiNardo CD, Stein EM, de Botton S, et al: Durable Remissions with Ivosidenib in IDH1387 Mutated Relapsed or Refractory AML. N Engl J Med 378:2386-2398, 2018
- Perl AE, Martinelli G, Cortes JE, et al: Gilteritinib or Chemotherapy for Relapsed or
   Refractory -Mutated AML. N Engl J Med 381:1728-1740, 2019
- Issa GC, Aldoss I, DiPersio J, et al: The menin inhibitor revumenib in KMT2A-rearranged
   or NPM1-mutant leukaemia. Nature 615:920-924, 2023

- 392 10. DeWolf S, Tallman MS: How I treat relapsed or refractory AML. Blood 136:1023-1032,
  393 2020
- 394 11. Döhner H, Wei AH, Appelbaum FR, et al: Diagnosis and management of AML in adults:
  395 2022 recommendations from an international expert panel on behalf of the ELN. Blood
  396 140:1345-1377, 2022
- 397 12. Kantarjian HM, Keating MJ, Walters RS, et al: The characteristics and outcome of
   398 patients with late relapse acute myelogenous leukemia. J Clin Oncol 6:232-238, 1988
- 399 13. Keating MJ, Kantarjian H, Smith TL, et al: Response to salvage therapy and survival
  400 after relapse in acute myelogenous leukemia. J Clin Oncol 7:1071-1080, 1989
- 401 14. Craddock C, Versluis J, Labopin M, et al: Distinct factors determine the kinetics of
  402 disease relapse in adults transplanted for acute myeloid leukaemia. J Intern Med
  403 283:371-379, 2018
- 404 15. Versluis J, Metzner M, Wang A, et al: Risk Stratification in Older Intensively Treated
  405 Patients With AML. J Clin Oncol: JCO2302631, 2024
- 406 16. Löwenberg B, van Putten W, Theobald M, et al: Effect of priming with granulocyte
  407 colony-stimulating factor on the outcome of chemotherapy for acute myeloid leukemia. N
  408 Engl J Med 349:743-752, 2003
- 409 17. Löwenberg B, Beck J, Graux C, et al: Gemtuzumab ozogamicin as postremission
  410 treatment in AML at 60 years of age or more: results of a multicenter phase 3 study.
  411 Blood 115:2586-2591, 2010
- 412 18. Löwenberg B, Pabst T, Vellenga E, et al: Cytarabine dose for acute myeloid leukemia. N
  413 Engl J Med 364:1027-1036, 2011
- 414 19. Terwijn M, van Putten WLJ, Kelder A, et al: High Prognostic Impact of Flow Cytometric
  415 Minimal Residual Disease Detection in Acute Myeloid Leukemia: Data From the
  416 HOVON/SAKK AML 42A Study. J Clin Oncol, 2013

- 417 20. Löwenberg B, Pabst T, Maertens J, et al: Therapeutic value of clofarabine in younger
  418 and middle-aged (18-65 years) adults with newly diagnosed AML. Blood 129:1636-1645,
  419 2017
- 420 21. Ossenkoppele GJ, Breems DA, Stuessi G, et al: Lenalidomide added to standard
  421 intensive treatment for older patients with AML and high-risk MDS. Leukemia 34:1751422 1759, 2020
- 423 22. Lowenberg B, Pabst T, Maertens J, et al: Addition of lenalidomide to intensive treatment
  424 in younger and middle-aged adults with newly diagnosed AML: the HOVON-SAKK-132
  425 trial. Blood Adv 5:1110-1121, 2021
- 426 23. Janssen JJWM, Löwenberg B, Manz M, et al: Addition of the nuclear export inhibitor
  427 selinexor to standard intensive treatment for elderly patients with acute myeloid leukemia
  428 and high risk myelodysplastic syndrome. Leukemia 36:2189-2195, 2022
- 429 24. Döhner H, Estey E, Grimwade D, et al: Diagnosis and management of AML in adults:
  430 2017 ELN recommendations from an international expert panel. Blood 129:424-447,
  431 2017
- 432 25. Freeman SD, Thomas A, Thomas I, et al: Fractionated vs single-dose gemtuzumab
  433 ozogamicin with determinants of benefit in older patients with AML: the UK NCRI AML18
  434 trial. Blood 142:1697-1707, 2023
- 435 26. Moons KGM, Altman DG, Reitsma JB, et al: Transparent Reporting of a multivariable
  436 prediction model for Individual Prognosis or Diagnosis (TRIPOD): explanation and
  437 elaboration. Ann Intern Med 162:W1-73, 2015
- 438 27. Papaemmanuil E, Gerstung M, Bullinger L, et al: Genomic Classification and Prognosis
  439 in Acute Myeloid Leukemia. N Engl J Med 374:2209-2221, 2016

- 440 28. Herzig RH, Lazarus HM, Wolff SN, et al: High-dose cytosine arabinoside therapy with
  441 and without anthracycline antibiotics for remission reinduction of acute nonlymphoblastic
  442 leukemia. J Clin Oncol 3:992-997, 1985
- 443 29. Amadori S, Arcese W, Isacchi G, et al: Mitoxantrone, etoposide, and intermediate-dose
  444 cytarabine: an effective and tolerable regimen for the treatment of refractory acute
  445 myeloid leukemia. J Clin Oncol 9:1210-1214, 1991
- 44630.Vogler WR, McCarley DL, Stagg M, et al: A phase III trial of high-dose cytosine447arabinoside with or without etoposide in relapsed and refractory acute myelogenous
- 448 leukemia. A Southeastern Cancer Study Group trial. Leukemia 8:1847-1853, 1994
- 449 31. Parker JE, Pagliuca A, Mijovic A, et al: Fludarabine, cytarabine, G-CSF and idarubicin
  450 (FLAG-IDA) for the treatment of poor-risk myelodysplastic syndromes and acute myeloid
  451 leukaemia. Br J Haematol 99:939-944, 1997
- 452 32. Litzow MR, Wang XV, Carroll MP, et al: A randomized trial of three novel regimens for
  453 recurrent acute myeloid leukemia demonstrates the continuing challenge of treating this
  454 difficult disease. Am J Hematol 94:111-117, 2019
- 455 33. Mühleck R, Scholl S, Hilgendorf I, et al: Outcome of patients with relapsed or refractory
  456 acute myeloid leukemia treated with Mito-FLAG salvage chemotherapy. J Cancer Res
  457 Clin Oncol 148:2539-2548, 2021
- 458 34. Brancati S, Gozzo L, Romano GL, et al: Venetoclax in Relapsed/Refractory Acute
  459 Myeloid Leukemia: Are Supporting Evidences Enough? Cancers 14:22, 2021
- 460 35. Kwag D, Cho B-S, Bang S-Y, et al: Venetoclax with decitabine versus decitabine
  461 monotherapy in elderly acute myeloid leukemia: a propensity score-matched analysis.
  462 Blood Cancer J 12:169, 2022

- 463 36. Angotzi F, Lessi F, Leoncin M, et al: Efficacy and safety of venetoclax plus
  464 hypomethylating agents in relapsed/refractory acute myeloid leukemia: a multicenter
  465 real-life experience. Front Oncol 14:1370405, 2024
- 37. Stahl M, Menghrajani K, Derkach A, et al: Clinical and molecular predictors of response
  and survival following venetoclax therapy in relapsed/refractory AML. Blood Adv 5:15521564, 2021
- Atluri H: Phase Ib/2 Study of Oral Decitabine/Cedazuridine (ASTX727) and Venetoclax
  in Combination with the Targeted Mutant IDH1 Inhibitor Ivosidenib or the Targeted
  Mutant IDH2 Inhibitor Enasidenib: 2023 Update. Presented at the 65th ASH Annual
  Meeting & Exposition, 2023/12/11, 2023
- 473 39. Briski R: A Phase I/II Study of Combination of ASTX727, Gilteritinib and Venetoclax in
  474 Patients with Relapsed/Refractory FLT3- Mutated Acute Myeloid Leukemia (AML).
  475 Presented at the 65th ASH Annual Meeting & Exposition, 2023/12/10, 2023
- 476 40. Yilmaz M: Phase I/II Study of Quizartinib, Venetoclax, and Decitabine Triple
  477 Combination in FLT3-ITD Mutated AML. Presented at the 65th ASH Annual Meeting &
  478 Exposition, 2023/12/9, 2023
- 479 41. Al-Shaibani E: FLAG-IDA Plus Venetoclax in High-Risk Newly Diagnosed and Relapsed
  480 or Refractory Acute Myeloid Leukemia: The Princess Margaret Cancer Center
  481 Experience. Presented at the 66th ASH Annual Meeting & Exposition, 2024
- 482 42. Cornelissen JJ, Gratwohl A, Schlenk RF, et al: The European LeukemiaNet AML
  483 Working Party consensus statement on allogeneic HSCT for patients with AML in
  484 remission: an integrated-risk adapted approach. Nat Rev Clin Oncol 9:579-590, 2012
- 485 43. Versluis J, Labopin M, Niederwieser D, et al: Prediction of non-relapse mortality in
  486 recipients of reduced intensity conditioning allogeneic stem cell transplantation with AML
  487 in first complete remission. Leukemia 29:51-57, 2015

488 44. Hermans SJF, Versluis J, Labopin M, et al: Prediction of Nonrelapse Mortality in Patients
489 With Acute Myeloid Leukemia and Acute Lymphoblastic Leukemia Receiving Allogeneic
490 Stem Cell Transplantation With Posttransplantation Cyclophosphamide-based Graft
491 Versus Host Disease Prophylaxis. HemaSphere 7:e846, 2023

- 492 45. Stelljes M, Middeke JM, Bug G, et al: Remission induction versus immediate allogeneic
  493 haematopoietic stem cell transplantation for patients with relapsed or poor responsive
  494 acute myeloid leukaemia (ASAP): a randomised, open-label, phase 3, non-inferiority
  495 trial. The Lancet Haematology 11:e324-e335, 2024
- 496 46. Sango J, Carcamo S, Sirenko M, et al: RAS-mutant leukaemia stem cells drive clinical
  497 resistance to venetoclax. Nature 636:241-250, 2024
- 498 47. McMahon CM, Ferng T, Canaani J, et al: Clonal Selection with RAS Pathway Activation
  499 Mediates Secondary Clinical Resistance to Selective FLT3 Inhibition in Acute Myeloid
  500 Leukemia. Cancer Discov 9:1050-1063, 2019
- 50148.Survival Prediction for Patients with AML and First Relapse, <a href="https://hovon-</a>502aml.shinyapps.io/AML\_Relapse\_Score/, 2024

503

	Development cohort (n=943)
Age at relapse	
Median (range)	58.0 (18.0-81.0)
Sex	
Male	523 (55.5%)
Female	420 (44.5%)
WBC (at diagnosis;[x10 <sup>9</sup> /L])	
Median (range)	8.45 (0-510)
ELN risk at diagnosis	
Favorable	204 (21.6%)
Intermediate	289 (30.6%)
Adverse	450 (47.7%)
Best response within 2 induction cycles	
CR	889 (94.3%)
CRi	54 (5.7%)
HCT in first CR(i)	
No	506 (53.7%)
Allogeneic	310 (32.9%)
Autologous	127 (13.5%)
Relapse free interval (months)	
Median (range)	8 (0-124)
Follow-up of patients alive after first relapse	
(months)	
Median (range)	49 (0-128)

504 **Table 1.** Patient characteristics at first AML diagnosis

505

506 Abbreviations: ELN, European Leukemia Network; CR, complete remission; CRi, complete

- 507 remission with incomplete count recovery; HCT, hematopoietic cell transplantation; WBC, white
- 508 blood cell count.
- 509
- 510

#### 511 FIGURE LEGENDS

512 **Figure 1.** Consort diagram of patients included in this study

513 Abbreviations: AML, acute myeloid leukemia.

514

515 **Figure 2.** Mutational landscape of the development cohort at first AML diagnosis

516 Driver events found at diagnosis in 943 patients with relapsed AML. Each bar represents a

517 driver lesion, including gene mutations, and chromosomal abnormalities. Abbreviations: abn,

abnormality in; ck, complex karyotype (annotated according to the 2022 ELN risk classification);

519 del, deletion in; inv, inversion of; ITD, internal tandem duplication; mk, monosomal karyotype; m,

520 monosomy; t, translocation of; TKD, tyrosine kinase domain.

521

522 **Figure 3A-B.** Overall survival after 1 year by previous prognostic models

523 Overall survival in the development cohort by (A) HOVON-SAKK and (B) GOELAMS prognostic 524 models.

525

526 **Figure 4A-D.** Revised prognostic model for AML after first relapse

527 Hazard ratios with 95% confidence interval and P-value of final model in the- development 528 cohort with points for a clinical prediction score (A) and overall survival after 1 year for patients 529 in first relapse according to risk categories of the revised prognostic model in the development 530 cohort (B). Figure S9A shows the 4 year overall survival rates for patients from their first 531 relapse, categorized by the revised prognostic model. Restratification of patients from HOVON-532 SAKK (C) and GOELAMS (D) model to the revised prognostic model in the development cohort. 533 Abbreviations: allo-HCT, allogeneic hematopoietic cell transplantation; CBF, core-binding factor; 534 ITD: internal tandem duplication; mk, monosomal karyotype; t, translocation of.

535

537 Patient characteristics (A) with the molecular and cytogenetic landscape at diagnosis of the 538 validation cohort (B) and overall survival after 1 year for patients in first relapse according to risk 539 categories of the revised prognostic model (C). Figure S9B shows the 4 year overall survival 540 rates for patients at their first relapse, categorized by the revised prognostic model. 541 Restratification of patients from HOVON-SAKK (D) and GOELAMS (E) model to the revised prognostic model in the validation cohort. Abbreviations: abn, abnormality in; ck, complex 542 543 karyotype (annotated according to the 2022 ELN risk classification); CR, complete remission; 544 CRi, complete remission with incomplete count recovery; del, deletion in; ELN, European 545 Leukemia Network; HCT, hematopoietic cell transplantation; inv, inversion of; ITD, internal tandem duplication; mk, monosomal karyotype; m, monosomy; t, translocation of; TKD, tyrosine 546 547 kinase domain; WBC, white blood cell count.

## Development cohort









Figure 3A-B



В

# Figure 4A-D

А

Predictor variable	HR	95% CI	P-value	Points
No CBE abnormalities	2.07	1.38-3.12	<0.001	3
Mutated TP53	1.99	1.48-2.67	<0.001	3
Relapse free interval ≤12 months	1.76	1.46-2.12	<0.001	2
Previous allo-HCT	1.65	1.38-1.96	<0.001	2
t(v;11q23)/KMT2A-rearranged	1.54	1.10-2.14	0.01	2
Age at relapse ≥60 years	1.53	1.29-1.82	<0.001	2
Complex karyotype (inc. MK)	1.40	1.08-1.82	0.01	1
FLT3-ITD	1.37	1.11-1.69	0.003	1
White blood cell count ≥10 x10 <sup>9</sup> /L	1.27	1.07-1.52	0.01	1







D

Figure	5A-E
0	Validation cohort (n=377)
Age at relapse	
(median [range])	69.0 [60.0-81.0]
Sex	

Sex	
Male	226 (59.9%)
Female	151 (40.1%)
WBC (at diagnosis;[x10 <sup>9</sup> /L])	
(median [range])	6.4 [0-365]
ELN risk 2022 at diagnosis	
Favorable	58 (15.4%)
Intermediate	92 (24.4%)
Adverse	227 (60.2%)
AML60+ classification at diagnosis	
Favorable	25 (6.6%)
Intermediate	117 (31.0%)
Poor	178 (47.2%)
Very Poor	57 (15.1%)
Best response on protocol treatment	
CR	333 (88.3%)
CRi	44 (11.7%)
SCT in first CR(i)	
No	326 (86.5%)
Allogeneic	51 (13.5%)
Autologous	0 (0%)
Relapse free interval (months)	
(median [range])	12.1 [1-61]
Follow-up of patients alive after first	
relapse (months)	
(median [range])	21 [0-59]









