TFOS DEWS III Diagnostic Methodology

James S. Wolffsohn, José Benítez-Del-Castillo, Denise Loya-Garcia, Takenori Inomata, Geetha Iyar, Lingyi Liang, Heiko Pult, Alfonso L. Sabater, Christopher E. Starr, Jelle Vehof, Michael TM Wang, Wei Chen, Jennifer P Craig, Murat Dogru, Victor L Perez Quinones, Fiona Stapleton, David A Sullivan, Lyndon Jones, TFOS collaborator group

 PII:
 S0002-9394(25)00275-2

 DOI:
 https://doi.org/10.1016/j.ajo.2025.05.033

 Reference:
 AJOPHT 13391

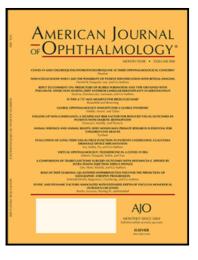
To appear in: American Journal of Ophthalmology

Received date: May 15, 2025 Accepted date: May 23, 2025

Please cite this article as: James S. Wolffsohn, José Benítez-Del-Castillo, Denise Loya-Garcia, Takenori Inomata, Geetha Iyar, Lingyi Liang, Heiko Pult, Alfonso L. Sabater, Christopher E. Starr, Jelle Vehof, Michael TM Wang, Wei Chen, Jennifer P Craig, Murat Dogru, Victor L Perez Quinones, Fiona Stapleton, David A Sullivan, Lyndon Jones, TFOS collaborator group, TFOS DEWS III Diagnostic Methodology, *American Journal of Ophthalmology* (2025), doi: https://doi.org/10.1016/j.ajo.2025.05.033

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2025 The Author(s). Published by Elsevier Inc. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/)



## TFOS DEWS III Diagnostic Methodology

James S. Wolffsohn,<sup>1</sup> José Benítez-Del-Castillo,<sup>2</sup> Denise Loya-Garcia,<sup>3,4</sup> Takenori Inomata,<sup>5</sup> Geetha Iyar,<sup>6</sup> Lingyi Liang,<sup>7</sup> Heiko Pult,<sup>8,9</sup> Alfonso L. Sabater,<sup>10</sup> Christopher E. Starr,<sup>11</sup> Jelle Vehof, <sup>12,13</sup> Michael T. M. Wang,<sup>14</sup> Wei Chen,<sup>15</sup> Jennifer P. Craig,<sup>14</sup> Murat Dogru,<sup>16</sup> Victor L. Perez Quinones,<sup>10</sup> Fiona Stapleton,<sup>17</sup> David A. Sullivan,<sup>18</sup> Lyndon Jones.<sup>19</sup> + TFOS collaborator group

- 1) Optometry and Vision Science, College of Health and Life Sciences, Aston University, Birmingham, UK
- 2) University Complutense, Hospital Clinico San Carlos, Clinica Rementeria, Madrid, Spain
- 3) Instituto de Oftalmología Fundación de Asistencia Privada Conde de Valenciana, I.A.P, Mexico City, Mexico.
- 4) Tecnologico de Monterrey, School of Medicine and Health Sciences, Monterrey, Mexico
- 5) Department of Ophthalmology, Juntendo University Graduate School of Medicine, Bunkyo-ku, Tokyo, Japan
- 6) CJ Shah Cornea Services/Dr G Sitalakshmi Memorial Clinic for Ocular Surface Disorders, Medical Research Foundation, Sankara Nethralaya, Chennai, Tamil Nadu, India
- 7) State key laboratory of Ophthalmology, Zhongshan Ophthalmic Center, Sun Yat-sen University, Guangzhou, China
- 8) College of Biomedical & Life Sciences, Optometry & Vision Sciences, Cardiff University, UK
- 9) Dr. Heiko Pult Optometry and Vision Research, Weinheim, Germany
- 10) Bascom Palmer Eye Institute, University of Miami Miller School of Medicine, Miami, FL, USA
- 11) Weill Cornell Medicine, New York-Presbyterian Hospital, New York, NY
- 12) Department of Ophthalmology and Epidemiology, University of Groningen, Groningen, the Netherlands
- 13) Department of Ophthalmology, Vestfold Hospital Trust, Tønsberg, Norway
- 14) Department of Ophthalmology, New Zealand National Eye Centre, The University of Auckland, Auckland, New Zealand
- 15) Eye Hospital of Wenzhou Medical University, National Eye Clinic Research Center, Peoples Republic of China.
- 16) Ichikawa General Hospital Tokyo Dental College Dept of Ophthalmology Ichikawa, Chiba, Japan
- 17) School of Optometry and Vision Science, UNSW Sydney, NSW, Australia
- 18) Tear Film & Ocular Surface Society, Boston, USA
- 19) Centre for Ocular Research & Education (CORE), School of Optometry and Vision Science, University of Waterloo, Waterloo, ON, Canada

## **TFOS Collaborator Group**

Reiko Arita, Itoh Clinic, University of Tokyo, Japan <u>ritoh@za2.so-net.ne.jp</u> Carlos Belmonte, Instituto de Neurociencias, Universidad Miguel Hernández-CSIC, Avda Santiago Ramón y Cajal s/n, Alicante, Spain <u>carlos.belmonte@umh.es</u> Robin L Chalmers, Clinical Trial Consultant, Atlanta, Georgia, USA chalmers2097@gmail.com

Anat Galor, Miami Veterans Affairs Medical Center Ophthalmology, Bascom Palmer Eye Institute, University of Miami Miller School of Medicine, Florida, USA

AGalor@med.miami.edu

Arkasubhra Ghosh, Molecular Signalling and Gene Therapy, GROW laboratory, Narayana Nethralaya Foundation (nnf.org.in), Bangalore, India

Arkasubhra@narayananethralaya.com

Marc Labetoulle, Ophthalmology Department, Bicêtre Hôpital, APHP, Paris-Saclay niversity, Kremlin-Bicêtre, France; Quinze-Vingts Hospital, Paris, France <u>marc.labetoulle@aphp.fr</u>

Kelly K. Nichols, School of Optometry, The University of Alabama at Birmingham, Alabama, USA <u>nicholsk@uab.edu</u>

Andrew D. Pucker, Eminent Ophthalmic Services, Milledgeville, Georgia, USA apucker@uab.edu

Eduardo M. Rocha, Dept of Ophthalmology, Otorhinolaryngology and Head & Neck Surgery Ribeirao Preto Medical School, University of Sao Paulo, Brazil <u>emrocha@fmrp.usp.br</u> Benjamin Sullivan, Bausch & Lomb, Bridgewater, New Jersey, USA; Lµbris BioPharma, Florida, USA <u>bdsulliv@me.com</u>

Piera Versura, Alma Mater Studiorum University of Bologna Cornea-Ocular surface Lab and translation research in Ophthalmology IRCCS Azienda Ospedaliero-Universitaria di Bologna, Bologna Italy <u>piera.versura@aosp.bo.it</u>

Mark D P Willcox, School of Optometry and Vision Science, UNSW, Sydney, Australia mdpwillcox@gmail.com

Corresponding author: James S. Wolffsohn, Vision Sciences, Aston University, Aston Triangle, Birmingham, B4 7ET, United Kingdom. J.s.w.wolffsohn@aston.ac.uk

## ABSTRACT

A standard approach to the diagnosis of dry eye disease across eye care practitioners is critical to reassuring the patient, providing consistency between practitioners and informing governments as to the true prevalence and resulting healthcare needs. The Tear Film & Ocular Surface Society (TFOS) Dry Eye Workshop (DEWS) III has reviewed the evidencebase since their previous reports published in 2017 and revised the definition to "Dry eye is a multifactorial, symptomatic disease characterized by a loss of homeostasis of the tear film and/or ocular surface, in which tear film instability and hyperosmolarity, ocular surface inflammation and damage, and neurosensory abnormalities are etiological factors." Key features from the definition include that dry eye disease is multifactorial, is a disease and not a syndrome and is always symptomatic. Differential diagnosis and ocular examination guidance is given along with the risk factors that should be discussed with the patient. The recommended screening questionnaire is the OSDI-6 with a cut-off score ≥4. A positive result together with a non-invasive breakup time <10s or alternatively tear film hyperosmolarity (≥308mOsm/L in higher eye or an interocular difference >8mOsm/L) gives a diagnosis of dry eye. In addition, the ocular surface should be stained and positive symptomology together with >5 corneal fluorescein and/or >9 conjunctival lissamine green punctate spots and/or lid margin lissamine green staining of ≥2mm length & ≥25 %width also gives a diagnosis of dry eye. Subclassification was separated into tear film (lipid, agueous and mucin/glycocalyx) and ocular surface and adnexa (anatomical misalignment, blink/lid closure, lid margin, neural dysfunction, ocular surface cell damage/disruption and primary inflammation/oxidative stress) components, with appropriate clinical tests and cut-offs provided to identify these etiological drivers in an individual, to inform appropriate management and therapy.

**Keyword:** dry eye disease (DED); definition; diagnosis; differential diagnosis; subtypes; subclassification; blepharitis, meibomian gland dysfunction (MGD)

Table of Contents	Tab	le of	Contents
-------------------	-----	-------	----------

1	Report	Aims7	
2	Definit	ions	
2	2.1 Dry	eye disease (DED)	8
	2.1.1	Multifactorial	8
	2.1.2	Disease, not a syndrome	8
	2.1.3 surface o	Dry eye is a subset of ocular surface disease and can co-exist with other ocu	
	2.1.3.1	Signs vs symptoms	9
	2.1.4	Ocular symptoms include vision	
	2.1.5	Pathophysiology elements	.10
3	History	y and Symptoms 10	
:		erential Diagnosis	.10
	3.1.1	Eyelid-related disorders	.12
	3.1.2	Conjunctival and corneal abnormalities	.12
	3.1.2.1	Conjunctival-related disorders	.12
	3.1.	2.1.1 Allergic conjunctivitis	.12
	3.1.	2.1.2 Viral conjunctivitis	.13
	3.1.	2.1.3 Bacterial conjunctivitis	.13
	3.1.	2.1.4 Cicatrizing conjunctivitis	.14
	3.1.	2.1.5 Conjunctivochalasis	.14
	3.1.	2.1.6 Pinguecula and pterygium	.14
	3.1.	2.1.7 Conjunctival concretions	.14
	3.1.2.2	2 Corneal-related disorders	.14
	3.1.	2.2.1 Neurotrophic keratitis	.14
	3.1.	2.2.2 Corneal dystrophies and degenerations	. 15
	3.1.	2.2.3 Bullous keratopathy	. 15
	3.1.	2.2.4 Infectious keratitis	. 15
	3.1.	2.2.5 Thygeson's superficial punctate keratitis	. 15
	3.1.3	Other mixed and miscellaneous ocular surface disorders	. 15
	3.1.3.1	Limbal stem cell deficiency	. 15
	3.1.3.2	2 Episcleritis	.16
	3.1.3.3	3 Mucus fishing syndrome	.16

3	3.1.3.4	4 Ocular neuropathic pain	16
3.1	.4 No	n-ocular surface disease disorders	16
3.1	.5 Su	mmary	16
3.2	Risl	k factors	17
3.3	Syn	nptomology	19
3.3	8.1	Routine questions based on variability of symptoms	19
3.3	8.2	Standardised questionnaires	19
3.3	8.3	Discordance between signs and symptoms	20
3.3	8.4	Paediatric considerations	23
3.4	Ocu	Ilar Examination	23
3.4		Diagnostic Homeostasis Test Battery	
-	3.4.1.1		
	3.4.1.2		
-	3.4.1.3		
3	3.4.1.4		
3	3.4.1.5	5 Tests to establish a loss of homeostasis of the tear film	24
3	3.4.1.6	5 Tests to establish a loss of ocular surface homeostasis	25
3	3.4.1.7	Practical diagnostic criteria considerations	26
3.4	.2	Advanced screening	28
3.4	.3	Severity rating	29
3.5	Sub	oclassification to identify DED etiological drivers	
3.5	5.1	Purpose of a DED subclassification	
3.5	5.2	Tear Film Deficiencies	
3	3.5.2.1	Lipid component	
		2.1.1 Interferometry	
	3.5.	2.1.2 Lipid turnover	31
	3.5.	2.1.3 Evaporimetry	31
	3.5.	2.1.4 Thermography	32
	3.5.	2.1.5 Meibum expressibility and quality (meibometry)	32
3	3.5.2.2	2 Aqueous	32
	3.5.	2.2.1. Meniscometry or tear meniscus assessment	32
	3.5.	2.2.2 Phenol red thread test	33
	3.5.	2.2.3 Schirmer test	33
	3.5.	2.2.4 Strip meniscometry	33

3.5.2.	2.5 Tea	ar clearance	
3.5.2.	2.6 Lac	rimal gland patency	
3.5.2.	2.7 Tea	ar proteins and other components	
3.5.2.3	Mucin /	glycocalyx	
3.5.2.	3.1 Mu	icins	
3.5.2.	3.2 Ros	se bengal and lissamine green (see section 3.4.1.7)	
3.5.2.	3.3 Cor	njunctival impression cytology	
3.5.2.	3.4 Fer	ning test	
3.5.3 E	Eyelid ano	malies	
3.5.3.1	Blink and	d lid closure anomalies	
3.5.3.2	Lid març	gin health	
3.5.3.	2.1 Ant	terior blepharitis	
3.5		emodex associated blepharitis	
3.5.3.		ibomian gland dysfunction (MGD)	
3.5.3.	2.3 Ocu	ular rosacea	
3.5.4 (		rface Abnormalities	
3.5.4.1	Anatomi	ical misalignment	40
3.5.4.2	Neural d	dysfunction	
3.5.4.3		surface cellular damage / disruption	
3.5.4.4	Primary	inflammation / oxidative stress	
3.5.4.	4.1 Ima	aging-based diagnostic tests	
		Ocular conjunctival redness	
3.5	.4.4.1.2	In vivo confocal imaging	
3.5.4.	4.2 Tea	Biomarker diagnostic tests	
3.5	.4.4.2.1	Matrix metalloproteinases	
3.5	.4.4.2.2	Cytokines and chemokines	
3.5	.4.4.2.3	Neurotrophic Factors and Neuropeptides	
3.5	.4.4.2.4	Ocular surface immune markers	
3.5	.4.4.2.5	Inflammasome markers	
3.5	.4.4.2.6	MicroRNAs	
3.5	.4.4.2.7	Oxidative stress markers	51
3.5	.4.4.2.8	Serum markers	51
3.5.5	Systemic d	diseases leading to dry eye	53
3.5.5.1	Autoimm	nune conditions	53
3.5.5.2	Hormon	al imbalance	53

	3	.5.5.3	Metabolic disease	53
	3	.5.5.4	Exposure	54
	3.6	Test	s for monitoring treatment	54
4	Pa	tient	s with only symptoms or ocular surface signs	
	4.1	Ocul	ar surface disease in the absence of symptoms	55
	4.1.	.1	Diagnosing Ocular Neurosensory Abnormalities	55
	4.1.	2	Diagnosing Neurotrophic Keratopathy	55
	4.2	Sym	ptoms in the absence of ocular surface disease	57
	4.2.	.1	Diagnosing corneal neuropathic pain	57
5	Fu		advances	
	5.1	Artif	cial intelligence	57
	5.2		biomarker testing of tears	
	5.3	Sust	ainability	58
	5.4	Need	I for experience-informed approach to un/under-researched areas	58
6	Su	mma	ry 59	
	6.1	Worl	flow and enhanced link to individualised management	59
	6.2	Patie	ent communication	61
	6.3	Key	diagnostic methodological changes from TFOS DEWS II	61
6	Re	ferer	nces	

## Abbreviations:

DED	Dry eye disease
DEWS	Dry eye workshop
IL	Interleukin
LIPCOF	Lid-parallel conjunctival folds
MGD	Meibomian gland dysfunction
MMP	Matrix metalloproteinase(s)
OCT	Optical coherence tomography imaging
OR	Odds ratio
OSDI	Ocular Surface Disease Index
TFOS	Tear Film & Ocular Surface Society

## 1 Report Aims

The Tear Film & Ocular Surface Society (TFOS) second dry eye workshop (DEWS II) Diagnostic Methodology report <sup>1</sup> provided practical, clinical diagnostic recommendations for dry eye disease (DED), based on the evidence available for tests with diagnostic potential to align with the revised definition of DED.<sup>2</sup> It also provided a clear rationale for the framework which informed the number and characteristics of the selected tests. Questions to inform a differential diagnosis were proposed. The need for subclassification post-diagnosis was emphasised, to inform management approaches. The purpose of the TFOS DEWS III Diagnostic Methodology report was to:

- Revisit the current definition <sup>2</sup> to ensure it aligns with current understanding of DED
  - Provide a rationale for the components of the definition
  - Highlight considerations when a patient only has symptoms or signs associated with DED
  - Define associated conditions
- Draw on risk / associated factors for DED (from the TFOS DEWS III Digest report) and masquerading diseases to guide appropriate history and symptomtaking
- Identify any updates required to the 2017 diagnosis of DED, reiterating the rationale for change and perceived challenges with regard to the available evidence
- Propose a new etiological driver-based approach to subclassification, identifying the tests that indicate the driver involved in an individual's dry eye, which can then be linked to management approaches by the TFOS DEWS III Management and Therapy report
- Discuss possible future directions that could help to inform further advances in DED diagnosis and subclassification

## 2 Definitions

## 2.1 Dry eye disease (DED)

In 2017, after careful consideration of the terminology including diction, word order, emphasis, and accepted meaning, DED was defined by a multidisciplinary and transnational committee as "a multifactorial disease of the ocular surface characterized by a loss of homeostasis of the tear film, and accompanied by ocular symptoms, in which tear film instability and hyperosmolarity, ocular surface inflammation and damage, and neurosensory abnormalities play etiological roles.". This was ratified by the full TFOS DEWS II authorship consisting of 150 clinical and basic science research experts from around the world, who utilized an evidence-based approach and a process of open communication, dialogue and transparency to consolidate the understanding of DED <sup>3</sup>.

It was the consensus of TFOS DEWS III that the definition did not require radical change based on our updated understanding of the disease pathology and the tear film <sup>4</sup>, but noted the intrinsic role of the ocular surface tissues as well as the tear film in homeostasis leading to repositioning of this aspect in the revised definition to read "Dry eye is a multifactorial, symptomatic disease characterized by a loss of homeostasis of the tear film and/or ocular surface, in which tear film instability and hyperosmolarity, ocular surface inflammation and damage, and neurosensory abnormalities are etiological factors." Key aspects of the definition include:

## 2.1.1 Multifactorial

There are many aspects of our lifestyle that can impact the ocular surface including digital environments <sup>5</sup>, environmental conditions <sup>6</sup>, nutrition <sup>7</sup>, use of cosmetics <sup>8</sup>, elective medication and procedures <sup>9</sup>, contact lenses <sup>10</sup>, societal factors <sup>11</sup> and general lifestyle <sup>12</sup>. All of these, therefore, need to be considered in the management of the disease and appropriate treatment or therapy is likely to include more than one approach or treatment. In addition, genetic factors have been found to play a role in DED. <sup>13-15</sup>

## 2.1.2 Disease, not a syndrome

When a combination of symptoms and physical findings are common to a group of patients, but the direct cause is not yet understood, it is referred to as a syndrome (from the Greek roots meaning 'running together')<sup>16</sup>. However, once causative agents or processes have been identified that have a moderately high degree of certainty, then it is appropriate to use the term 'disease' rather than a 'syndrome'.<sup>16</sup> Dry eye was once considered a syndrome due to insufficient understanding of its etiology. However, advances in medical knowledge have revealed its clearly identifiable diagnostic features <sup>1</sup> and disease progression <sup>4</sup> and response to specific treatments <sup>17</sup>. These insights support the current recognition of dry eye as a disease entity rather than a syndrome<sup>2</sup>. Defining dry eye as a disease is an important issue for patients (as this can affect reimbursement for clinical care and treatment as well as their understanding of their symptoms that are impacting their quality of life), and for eye care practitioners, since in some countries, certain professions are not permitted to treat 'disease'. The TFOS DEWS III consensus is that all eye care practitioners play an important role in managing patients with DED within the limits of their clinical competency, by providing evidence-based advice on lifestyle factors <sup>18</sup> and over-the counter treatments, as a minimum.

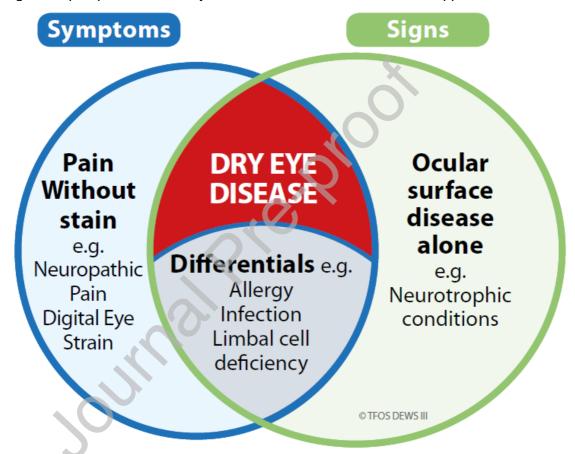
2.1.3 Dry eye is a subset of ocular surface disease and can co-exist with other ocular surface disease

DED is defined as a symptomatic disease and thus, by definition, must always be accompanied by ocular symptoms. Recognition of DED as a symptomatic disease is not new; the National Eye Institute/Industry Workshop in 1995 defined dry eye as "a disorder of the tear film … associated with symptoms of ocular discomfort" <sup>19</sup>. The first TFOS DEWS report identified dry eye as a "disease of the tears and ocular surface that results in symptoms of discomfort …" <sup>20</sup> and TFOS DEWS II as a "disease of the ocular surface

characterised by a loss of homeostasis of the tear film and accompanied by ocular symptoms ...."<sup>2</sup>. Hence, ocular surface signs in the absence of symptoms reflects the presence of ocular surface disease (Figure 1), but does not signify the existence of DED, specifically.

## 2.1.3.1 Signs vs symptoms

It is acknowledged that there are pathological conditions that result in symptoms without clinically significant signs (see Section 4.2), that present with signs but without symptoms (see Section 4.1), or that exhibit both dryness symptoms and signs but are not DED as they have a different pathophysiology (see Section 3.1). Such conditions can co-exist with dry eye disease; for example a patient with severe symptoms, but only mild signs, may have neuropathic pain in conjunction with mild DED. This is important to recognise from a management perspective as it may indicate the need for a multimodal approach <sup>17</sup>.



**Figure 1:** Venn diagram illustrating that dry eye disease requires the presence of both signs and symptoms, and exclusion of differential diagnoses. Dry eye disease can co-exist with other forms of ocular surface disease and symptomatic conditions.

## 2.1.4 Ocular symptoms include vision

Vision and vision-specific tasks can be significantly affected by DED due to destabilisation of the tear film disrupting the smooth air-tear interface between blinks or damage to the ocular surface affecting corneal transparency <sup>21-25</sup>. An unstable tear film results in light scatter and visual fluctuations between blinks <sup>26-28</sup>. Sensitivity to light and "spots in vision" are more commonly reported accompanying DED than glaucoma and/or cataract <sup>29</sup>. Hence, while vision is not specifically mentioned in the definition of DED (as there are many associated symptoms), visual disturbance is acknowledged to be encompassed within the terminology of "ocular symptoms". As also previously noted by TFOS DEWS II, the term "symptoms" is

considered to cover a wide range of possible patient-reported sensations associated with DED, which can include discomfort and / or visual disturbance <sup>3</sup>.

## 2.1.5 Pathophysiology elements

The component of the definition describing the disease pathology (see TFOS DEWS III Digest Pathophysiology section <sup>4</sup>) needs to exclude other conditions with both signs and symptoms, but with different pathophysiology and therefore appropriate management therapies. Clinically, these can be excluded through a process of differential diagnosis (See Section 3.1).

## 3 History and Symptoms

## 3.1 Differential Diagnosis

Both symptoms and signs of DED are heterogeneous. Therefore, the differential diagnosis of DED is extensive and encompasses most ocular surface disease (Figure 2). Several of the ocular surface changes caused by conditions identified as differential diagnoses of DED may increase the risk of DED, and DED may sometimes exacerbate other ocular surface diseases with similar symptoms (see section 3.2). Thus, many of the differential diagnoses are often comorbid with DED. There are also non-ocular surface disease diagnoses that may mimic dry eye symptoms, that are particularly important to consider when no ocular surface disease signs are present. Table X gives an overview of differential diagnoses of DED and their key differential features. The following sections focus on the most common differential diagnoses of DED.

10

## The differential diagnoses of dry eye disease

Eyelid-related disorders	Key differential features	Non-ocular surface disorders	Key differential features	Other mixed and miscellaneous ocular surface disorders	Key differential features	
Anterior blepharitis (bacterial, fungal, parasitic, viral,	Crusts, collarettes, or debris on lashes, hyperaemic swollen lid margins, lid notching, marginal keratitis	Uncorrected refractive error	Blurry vision, headache, squinting, eye strain	surface disorders		
seborrhoic)		Digital eye strain	Eve strain, ache around the eves, blurred vision, headache,	Limbal stem cell deficiency	Stippled staining pattern, loss of clarity epithelium,	
Meibomian gland dysfunction	Altered meibornian gland secretion, meibornian gland plugging and pouting, lid margin telangiectasia, lid	shoulder and neck pain, associated with prolonged screen use			conjunctivalization of the cornea, superficial corneal vascularization	
Chalazion/hordeolum	notching Raised lesions on the eyelids	Intermittent angle closure	Shallow anterior chamber, high intraocular pressure, severe conjunctival hyperaemia, corneal edema, fixed mid-dilated pupil	Episcleritis and anterior scleritis	Diffuse or sectoral hyperaemia ocular surface, nodules, subacute presentation, boring pain (scleritis)	
Eyelid malposition (ectropion,	Part or whole eyelid everted from the globe (ectropion)	Posterior scleritis Severe pain, proptosis, vision loss, choroidal folds, exudative retinal detachment, papilledema		Mucus fishing syndrome	Repetitive eye-rubbing and self-extraction of mucus,	
entropion) Trichiasis and distichiasis	or inverted (entropion), tearing, mucus discharge. Eyelashes directed towards the globe (trichiasis),	Anterior uveitis	Anterior chamber cells and flare, keratic precipitates, pain and severe photophobia		epithelial damage of the nasal and temporal conjunctiva (lissamine green staining)	
	extra row of eyelashes more posterior (distichiasis), epitheliopathy where lashes touch eye	Tolosa hunt syndrome	Severe unilateral retrobulbar or periorbital pain, restricted eye movements	Ocular neuropathic pain	Increased perception of pain to non-painful stimuli (e.g. aircon or wind), history of refractive laser or ocular surge diagnosis of exclusion when symptoms outweigh signs	
Floppy eyelid syndrome	Severe eyelid laxity, spontaneous eversion of upper lid, papillary conjunctivitis, lash ptosis, history of sleep apnea	Thyroid eye disease	Lid retraction, proptosis, lid swelling, double vision, restricted eye movements, history of thyroid disease	Foreign body	Foreign body on cornea, conjunctiva, or under upper	
Ocular rosacea	Telangiectasia, erythema lid margin, MGD, history of rosacea (can present without skin involvement)	Carotid cavernous fistula	Pulsatile exophthalmos, orbital bruit, conjunctival chemosis and engorged vessels, restricted eye movements	Contact lens intolerance	eyelid, acute onset 3 and 9 o'clock staining, conjunctival hyperaemia,	
Blepharospasm	Episodic uncontrolled blinking or closure of the eyelids	Dacryocystitis	Purulent punctal discharge, painful erythematous swelling around lacrimal sac	and contact lens -induced conjunctivitis	improvement after discontinuing lens, (giant) papillae	
Bell's palsy and/or lagophthalmos	Inability to close eye due to VII nerve palsy, punctate epithelial defects in lower cornea.	Nasolacrimal duct	Continuous tearing and watery eyes, reflux with lacrimal	Toxic conjunctivitis and	Inferior papillary or follicular reaction, inferior conjunctiv	
		obstruction syringing keratopath			and corneal staining, use of (preserved) eye drops or othe	
Canaliculitis	Swollen and inflamed nasal lid, chronic punctal discharge	Sinusitis	Nasal drainage, hyposmia or anosmia, facial pain or pressure, nasal obstruction	Keratopatny Thermal burns, UV-radiation, traumatic and chemical injuries Superior limbic	triggering systemic medication Severe pain, history of trauma, welding or chemical injur epithelial defect on interpalpebral cornea	
Canaliculitis	Swollen and inflamed nasal lid, chronic punctal discharge		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-radiation, traumatic and chemical injuries	triggering systemic medication Severe pain, history of trauma, welding or chemical injur epithelial defect on interpalpebral comea Papillary reaction upper palpebral conjunctiva, sectorial conjunctival hyperaemia, keratinization and ciliary	
Conjunctiva-related	Swollen and inflamed nasal lid, chronic punctal discharge		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-radiation, traumatic and chemical injuries Superior limbic	triggering systemic medication Severe pain, history of trauma, welding or chemical injur epithelial defect on interpalpebral comea Papillary reaction upper palpebral conjunctiva, sectorial conjunctival hyperaemia, keratinization and ciliary	
Conjunctiva-related disorders	Key differential features Itch as main symptom, papillary conjunctivitis, atopic		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-radiation, traumatic and chemical injuries Superior limbic keratoconjunctivitis	triggering systemic medication Severe pain, history of trauma, welding or chemical injue epithelial defects on interpulpebral comea Papillary reaction upper palpebral conjunctiva, sectorial conjunctival hyperaemia, heratinization and cliary lineitoin, punctate staining or erosion of superior come Key differential features Decressed comeal entithicity comeal entithelial defects	
Conjunctiva-related disorders Allergic eye disease Infectious conjunctivitis (viral,	Key differential features		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns UV-radiation, traumatic and chemical injunes Superior limbic keratoconjunctivitis Cornea-related disorders	triggering systemic medication Severe pain, history of trauma, welding or chemical injue epithelial defects on interpulpebral comea Papillary reaction upper palpebral conjunctiva, sectorial conjunctival hyperaemia, heratinization and cliary lineitoin, punctate staining or erosion of superior come Key differential features Decressed comeal entithicity comeal entithelial defects	
Conjunctiva-related disorders Allergic eye disease Infectious conjunctivitis (viral, bacterial) Cicatricala conjunctivitis Gicatricala conjunctivitis	Key differential features Itch as main symptom, papilary conjunctivitis, atopic history, idi sveillar and chemosis Acute onest, conjunctival hypotenenia, discharge, papilae or folidez; neuro type regrospicory infection,		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-cadation, traumatic and chemical injunies Superior limbic keratoconjunctivitis Cornea-rolated disorders Neurotrophic keratopathy Corneal dystrophies and degenerations (e.g. Satrman and epithelia basement	triggering systemic medication Severe pain, history of trauma, welding or chemical injure epithelial defect on interpalpebral comes Papillary reaction upper palpebral conjunctiva, sectorial injection, practical salaring or erosion of superior come <b>Key differential features</b> Decreased corneal sensitivity, corneal epithelial defects and stronal ulcaration, history of herpes keratistis or LAS Corneal deposition, spacifica and/or nodelse, savija	
Conjunctiva-velated disorders Altergic eye disease Infectious conjunctivitis (viral, Gasticiala) Clasticial conjunctivitis Stevens-Johnson syndrome, colar actaricial pemphigold, rachema)	Key differential features Reha main symptom, pspillary conjunctivitis, atopic history, kid swelling and chemosia Acute onse, conjunctival hyperaremia, discharge, papilae or billicle, recent upper respiratory infection, hymbalenopathy. Inflammation and scarring of conjunctiva, subconjunctival fibrosis, fornis shortening, symblep hypon		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-adiation, traunatic and Chemical Injunes Superior Imbic keratoconjunctivitis Cormas-related disconders Neurotrophic keratopathy Cormad dystrophies and degenerations (e.g. Sufama and epithelial basement membiane dystrophyl)	triggering systemic medication Severe pain, history of trauma, welding or chemical injue epithelial defects on interpaiphetal comes Papillary machine upper pajohetal conjunctiva hypotenina, hardinationa and diag- injection, punctate staining or erosion of superior come Key differential features Decreased corneal sensitivity, corneal epithelial defects and stromal ulceation, history of herpes keratilis or LAS Corneal depositions, opacities and/or nodules, varying corneal ension or edema	
Conjunctiva-related disorders Mergic eye disease refectious conjunctivitis (viral, acaterial) Cicatricial conjunctivitis Suevers-Johns prodrome, cuclar cicatricial pemphigoid, Ligneous conjunctivitis	Key differential features Itch as main symptom, papellary conjunctivitis, atopic https://disward.chemosis Acute onest, conjunctival hypersemis, discharge, papellae of oblicity, sent upper responsion y intection, hymbadampathy interfammation and scarring of conjunctive, subconjunctival fibrosis, formis shortening, symblerphuron Fibrinous pseudomembranes on palpebral conjunctive		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-adation, traumatic and chemical injunes Superior limbic keratoconjunctivitis Cormae-velated disconders Neurotrophic keratopathy Cormael dystrophics and degenerations (ge Safeman and epithelial basement membrane dystrophy) Bullous keratopathy Infectious keratitis Thyseson superficial	triggering systemic medication Severe pain, history of trauma, welding or chemical injue spithelial defect on interpaipebratic cornes Papillary maction upper palpebrat conjunctive systemic injection, punctate staining or erosion of superior corne Koy differential features Decreased corneal sensitivity, corneal epithelial defects and stronal ulcoration, history of meps keratistis of XAC Corneal depositions, opacities and/or nodules, varying corneal erosion or edema Corneal edema, epithelial aduita, history of intracecular surgery or Fuchs endothelial corneal dystrephy. Context edema, epithelia lactices and corneal dystrephy. Context endothelial corneal dystrephy.	
Conjunctiva-related disorders Mergic eye disease refectious conjunctivitis (viral, acaterial) Cicatricial conjunctivitis Suevers-Johns prodrome, cuclar cicatricial pemphigoid, Ligneous conjunctivitis	Key differential features Reha main symptom, pspillary conjunctivitis, atopic history, kid swelling and chemosia Acute onse, conjunctival hyperaremia, discharge, papilae or billicle, recent upper respiratory infection, hymbalenopathy. Inflammation and scarring of conjunctiva, subconjunctival fibrosis, fornis shortening, symblep hypon		Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-adulon, traumatic and Chemical Ispaces Superior Imbic Kentocojunctivitis Correas voluted discortions Neurotrophic kentopathy Corread dystrophies and designifications (see a discortion) Bullous kentopathy Bullous kentopathy	triggering systemic medication Severe pain, history of trauma, welding or chemical injur- epithelial defect on interpalpebratic comes Papillary maction upper palpebrat conjunctival sys- injection, punctate staining or erosion of superior come Key differential foatures Decreased corneal sensitivity, corneal epithelial defects and attribuil alcostation, history of mespe karatistis or LSC Corneal depositions, opacities and/or nodules, varying corneal ensois on edema Corneal edema, epithelial bullae, history of intraocular supery or Fuch endothelial corneal dystrophy. Acute onset, infittate or ulcer in cornes, severe pain, conjunctical hypersensia, photophobia Discrete grey white focal corneal tesions, often central a elevated, luck of corgunctival hypersensia	
Canalicultis Canalicultis Canjunctive-related disorders Altergic-eye disase Infectious conjunctivitis (viral, bacterial) Cicatricial conjunctivitis (viral, bacterial) Cicatricial conjunctivitis Cicatricial conjunctivitis Cicatricial conjunctivitis Cicatricial conjunctivitis Cicatricial conjunctivitis Cicatricial conjunctivitis Cinductive-thalasis Pinguecula, pterygium	Key differential features Itch as main symptom, papilary conjunctivitis, atopic https://diseduction. Acute onset; conjunctival hypersenia; discharge papilae or ofilides; nexret upper registratory infection, hymphadenopathy inflammation and scarring of conjunctiva, subconjunctival fibrosis, fornis shortening, symblep hypon Fibrinous pseudomembranes on palpebral conjunctiva Loose, nedundent, non-oxdematous fidds in the conjunctival offen samptomatic. ULL parallel	Snutis	Nasal drainage, hyposmia or anosmia, facial pain or pressure,	Thermal burns, UV-adiation, traumatic and chemical injunits Supportor limbic: keratoconjunctivitis Cormas-related disorders Neurotrophic keratopathy Cormad dysrophics and dysrophical busement merichane dystrophy) Bullicos keratopathy Infectious kerathis Trypeson superficial punctate kerathis	triggering systemic medication Severe pain, history of trauma, welding or chemical injure sphihelial defect on interpalpebrial comea Papilary reaction upper palpebrial conjunctive sectorial conjunctive hyperamik, kerdinization and ciliary injection, punctate staining or erosion of superior correct Koy differential features Decreased corneal sensitivity, corneal epithelial defects and stronal ulcostaton, history of intraocular surgery or Fuchs endothelial corneal dystrophy. Corneal edema, epithelial bulka, history of intraocular surgery or Fuchs endothelial corneal dystrophy. Control edema, epithelia bulka, history of intraocular surgery or Fuchs endothelial corneal dystrophy. Control edema box of history bulka, history of intraocular surgery or Fuchs endothelial corneal dystrophy.	

Figure 2: The key differential diagnoses of dry eye disease (DED) grouped by anatomy, and main differential features compared to DED.

11

## 3.1.1 Eyelid-related disorders

While patients with anterior blepharitis or MGD do not always have DED, as these conditions can be asymptomatic, they are commonly a driver of DED (see section 3.5.3.2) Diseases affecting the eyelids, such as chalazion, hordeolum, entropion, ectropion, trichiasis/distichiasis, floppy eyelid syndrome, blepharospasm, ocular rosacea, Bell's palsy and canaliculitis may result in symptoms similar to DED including watery eyes as a neuroregulated response to dry or irritated ocular surface <sup>30</sup>, and so a thorough eyelid examination should be performed in every patient suspected of having DED <sup>31</sup>.

## 3.1.2 Conjunctival and corneal abnormalities

To distinguish between DED and other etiologies, a thorough patient history is vital; this is especially important in patients with a history of contact lens wear <sup>32</sup>, use of multiple eye drops, or exposure to toxic chemicals <sup>33, 34</sup>. Observation of the ocular surface provides further useful information. For example, a "curl" pattern of fluorescein staining (which may indicate epithelial stress for example due to medication toxicity)<sup>33</sup> or corneal conjunctivalization (which may be indicative of limbal stem cell deficiency) <sup>32, 35</sup>. Fluorescein staining in the superior cornea may point to superior limbic keratoconjunctivitis <sup>36</sup>, floppy eyelid syndrome <sup>37</sup> or can result from contact lens wear issues <sup>38,40</sup>. Non-responsiveness to standard DED therapies warrants consideration of concurrent systemic conditions such as mucus membrane pemphigoid (ocular cicatricial pemphigoid) or Stevens-Johnson syndrome <sup>41, 42</sup>. Even though DED can co-exist in these conditions, early diagnosis is vital as advanced therapies including systemic immunomodulatory therapy are often required. Superficial punctate keratitis can commonly be observed in a variety of corneal disorders. If it is primarily the superior cornea that is affected, possible causes include vernal keratoconjunctivitis, superior limbic keratoconjunctivitis, trachoma, poorly fitting contact lenses, floppy eyelid syndrome, trichiasis or distichiasis <sup>38</sup>. If it is primarily inferior in location, it might, more likely be attributable to ocular rosacea, atopic keratoconjunctivitis, allergic keratitis, blepharitis, exposure keratopathy, lower evelid margin lesions, topical medication toxicity, entropion or ectropion, trichlasis or distichlasis <sup>38</sup>. If the superficial punctate keratitis is primarily interpalpebral, it may be contact lens-related (chemical toxicity, tight lens syndrome, overwear syndrome), or due to exposure to ultraviolet light, neurotrophic keratopathy or DED <sup>38</sup>.

Filamentary keratitis is a persistent corneal condition, recognizable by the presence of fine strands (from mucin bound degenerated epithelial cells) and mucus which adhere to the corneal surface <sup>43-45</sup>. It may arise secondary to DED, causing discomfort symptoms and photophobia; blepharospasm and increased blinking are also common <sup>46</sup>.

## 3.1.2.1 Conjunctival-related disorders

## 3.1.2.1.1 Allergic conjunctivitis

The signs and symptoms of dry eye overlap with those of allergic conjunctivitis, and the conditions can coexist <sup>47, 48</sup>. In a study in of 689 randomly sampled patients from an ambulatory optometric practice in California, United States, 57% of those reporting itching had clinically significant dryness and 45% of those with dry eye reported itching <sup>49</sup>. Immunoglobulin E (IgE) antibodies against seasonal or perennial allergens are commonly evaluated by a blood test <sup>50</sup> and diagnostic tests are available to detect the presence of IgE in the tear film. Additionally, typical ocular signs of allergy, such as eyelid edema, and conjunctival papillae as well as conjunctival chemosis, help differentiate allergic conjunctivitis from DED <sup>51, 52</sup>. A strong family history of allergy, atopic dermatitis, and/or the presence of asthma is common in those patients <sup>53</sup>. Atopic and vernal keratoconjunctivitis, chronic, bilateral, inflammatory and visually threatening forms of the disease also have signs and symptoms similar to DED and can serve as a trigger for DED. Signs of inflammation can be found in the cornea, conjunctiva, and eyelids. Typically, symptoms are photophobia, burning, tearing, itching, mucous discharge, and hyperemia and papillary hypertrophy of the eyelids.

Some of the most common signs found in both atopic keratoconjunctivitis and DED (although generally less severely in DED) include superficial punctate keratitis, conjunctival injection or hyperemia, anterior blepharitis, meibomian gland dysfunction (MGD) and tear instability <sup>39, 54-57</sup>. Atopic keratoconjunctivitis should be considered if there are signs such as conjunctivitis (possibly with scarring) and periorbital eczema <sup>58</sup>, corneal neovascularization, symblepharon, keratoconus, and sometimes anterior polar cataracts <sup>59, 60</sup>. Again, a family history of allergy, atopic dermatitis (occurring in 95%), asthma (occurring in 87%) and periorbital eczema are common <sup>39, 61, 62</sup>.

#### 3.1.2.1.2 Viral conjunctivitis

While viruses cause approximately 80% of cases of acute infectious conjunctivitis in adults, they may be responsible for only around 20% of pediatric cases <sup>63, 64</sup>. Even though watery discharge is typical of viral conjunctivitis this may also be seen in about 25% of bacterial cases <sup>63, 64</sup>. Most viral conjunctivitis involves the highly contagious adenovirus (65–90%), <sup>65</sup> which has an incubation period of 4 to 10 days before being clinically observable <sup>66</sup> and may have associated pharyngoconjunctival fever and epidemic keratoconjunctivitis. Other causes of viral conjunctivitis include herpes viruses, picornaviruses, and several systemic viral infections <sup>39, 64</sup>. Unilateral herpetic keratitis can affect the tear film of both eyes <sup>67, 68</sup>. Even though viral conjunctivitis shares a number of findings with DED such as tearing, burning, redness, irritation, photophobia and blurred vision, the following signs and symptoms may help to differentiate a viral etiology <sup>39, 69</sup>:

- Acute onset of signs and symptoms
- Redness and irritation initially in one eye, which often spreads to the other eye within a few days.
- Recent upper respiratory tract infection or close contact with someone with a red eye.
- Crusting around the eyes in the morning.
- Examination findings are watery, mucoid discharge and red, edematous eyelids.
- Preauricular lymphadenopathy (swelling of the lymph nodes in front of the ears which drain lymph fluid from the area around the eyes, cheeks and surrounding scalp).

Epstein-Barr virus infects more than 90% of the adult population <sup>70</sup>. Even though the infection of ocular structures with the Epstein-Barr virus results most commonly in transient follicular conjunctivitis <sup>71</sup>, it can also present with signs and symptoms similar to DED, as well as with keratitis, uveitis, choroiditis, retinitis, ocular glandular syndrome, papillitis and ophthalmoplegia <sup>72</sup>. Several systemic viruses, including measles, rubella (German measles), mumps, and influenza, are also frequently associated with conjunctivitis <sup>69</sup>.

## 3.1.2.1.3 Bacterial conjunctivitis

Bacteria as a causative agent of conjunctivitis occurs more often in children than in adults (70% vs 20% of cases) <sup>63</sup> <sup>73</sup>. Purulent conjunctival discharge and morning eyelash crusting may suggest a bacterial involvement, but this does not rule out a viral cause <sup>63</sup>. Patients with bacterial conjunctivitis may complain of similar symptoms as DED, such as burning, stinging, irritation, foreign body sensation and photophobia. In contrast to dry eye, there is typically significant conjunctival hyperemia (often more than with viral conjunctivitis or DED) and discharge (typically moist and mucopurulent). Affected patients often complain about matting or clumping of the eyelashes, mostly in the morning. Bacterial conjunctivitis can occur in one or both eyes and systemic findings may be present, especially in children, such as purulent rhinorrhoea and respiratory infection, fever and malaise <sup>74</sup>. Chlamydial conjunctivitis, that is more prominent in the lower palpebral conjunctiva, and with mucopurulent discharge <sup>75</sup>.

## 3.1.2.1.4 Cicatrizing conjunctivitis

Cicatrizing conjunctivitis is characterized by inflammation and scarring of the conjunctiva. Cicatrization can range from mild and subtle, with only subconjunctival fibrosis, often seen as fine white lines at the palpebral conjunctiva, to extensive, with fornix shortening, multiple symblepharon and ankyloblepharon. It can induce ocular dryness and can lead to gross distortion of the anatomy of the eyelid and ocular surface, including entropion, trichiasis, limbal stem cell deficiency and keratinization of the ocular surface. Causes of cicatrizing conjunctivitis are many and include autoimmune disease such as ocular pemphigoid, epidermolysis bullosa, sarcoidosis, systemic sclerosis, Sjögren disease and lichen planus, bacterial and viral conjunctivitis such as trachoma and adenoviral conjunctivitis, thermal and chemical burns, graft versus host disease, Stevens-Johnson syndrome, ocular rosacea and atopic keratoconjunctivitis. Conjunctival biopsy is important in confirming a diagnosis and to identify the cause.

## 3.1.2.1.5 Conjunctivochalasis

Conjunctivochalasis is characterized by loose, redundant, non-oedematous folds in the conjunctiva, the more mild expression of which is lid-parallel conjunctival folds (LIPCOF). It is most often seen in the inferior bulbar conjunctiva, overlying the inferior lid margin. It is often asymptomatic, but can lead to tear film instability and symptoms consistent with DED, including irritation, dryness, foreign body sensation, mucus discharge, and tearing. The etiology of conjunctivochalasis is still unclear, but it is more common with increasing age, in patients with DED, and contact lens wearers. Symptoms may be increased during downward gaze or with vigorous blinking <sup>79, 80</sup>.

## 3.1.2.1.6 Pinguecula and pterygium

A pinguecula is a benign degeneration of the conjunctiva, that presents as a gray-white or yellowish lesion on the bulbar interpalpebral conjunctiva, adjacent to the limbus, which may cause localized disruption of the tear film due to a change in lid-globe juxtaposition. Wind, dust, and UV exposure are factors associated with pinguecula and pterygium development. Pinguecula are usually asymptomatic, but may present with mild foreign body sensation or itch <sup>81</sup>. A pterygium is a growth of epithelial and fibrovascular tissue from the conjunctiva migrating over the corneal limbus in a wing-shaped way. Pterygia have similar associated factors as pingueculae, and a pterygium is often preceded by, or is comorbid with, a pinguecula. Irregular astigmatism with a pterygium can lead to visual symptoms, and irritation may be present. Inflammation of pterygia and pingueculae is also possible, leading to conjunctival hyperemia and edema, and increased chance of irritation <sup>82, 83</sup>.

## 3.1.2.1.7 Conjunctival concretions

Conjunctival concretions are benign discrete yellow-white deposits, most commonly found in the palpebral conjunctiva or fornices. Most are idiopathic and the result of degenerative changes with ageing, but they may be secondary to conjunctival inflammation, such as that associated with allergic keratoconjunctivitis, trachoma and DED. They are normally asymptomatic, but when they erode through the conjunctiva they may irritate the cornea and bulbar conjunctiva<sup>84</sup>.

## 3.1.2.2 Corneal-related disorders

## 3.1.2.2.1 Neurotrophic keratitis

Neurotrophic keratitis is characterized by a decrease in corneal sensitivity and stems from dysfunction in the ophthalmic division of the trigeminal nerve, which can be triggered by

conditions such as diabetes mellitus, ocular herpes simplex/zoster, non-ocular surface neoplasia or ophthalmic surgery. This dysfunction ultimately leads to a decrease in aqueous tear production <sup>39</sup>.

#### 3.1.2.2.2 Corneal dystrophies and degenerations

Corneal dystrophies and degenerations present with similar symptoms as DED. They can also lead to recurrent corneal erosions. The more common cases seen in clinical practice are Salzmann's nodular degeneration, epithelial basement membrane dystrophy, and Fuchs endothelial corneal dystrophy. Salzmann's nodular degeneration is a non-inflammatory degeneration of the anterior cornea with blue-white-grayish sub-epithelial nodules of the cornea, usually mid-peripheral. It can lead to symptoms of decreased visual acuity and irritation, pain, foreign body sensation, and blepharospasm.<sup>85, 86</sup> epithelial basement membrane dystrophy, previously known as map-dot-fingerprint dystrophy affects around 42% of the general population and is characterized by abnormal epithelial turnover, maturation and production of basement membrane. Typical signs are gray patches, microcysts and fine lines in the corneal epithelium, and typical symptoms are blurred vision and pain. <sup>87, 88</sup> Fuchs' endothelial cornea dystrophy is a hereditary, progressive disease of the posterior cornea, with progressive decline of corneal endothelial cells, and the formation of extracellular matrix excrescences in Descemet's membrane. This can eventually lead to corneal edema, bullous keratopathy (see Section 3.1.2.2.3), loss of vision, photophobia, epiphora, and pain. 89

#### 3.1.2.2.3 Bullous keratopathy

Bullous keratopathy forms small vesicles or bullae in the cornea due to endothelial dysfunction. These blister-like formations can rupture painfully and disrupt vision. Treatment options may include 5% sodium chloride or other hyperosmotic eye drops to reduce swelling, amniotic membranes, bandage contact lenses for comfort, antiglaucoma medications (when associated with glaucoma) to decrease fluid flow into the cornea and corneal transplants to replace damaged tissue <sup>90</sup>.

## 3.1.2.2.4 Infectious keratitis

Infection of the cornea can be microbial (bacteria, fungal or parasitics), or viral (herpes simplex or zoster), and is a sight-threatening condition, that always needs to be ruled out in patients with ocular surface symptoms. Contact lens wear, topical steroid use, ocular trauma, previous keratitis and previous ocular surgery are important risk factors. Blurred vision, bulbar hyperaemia, pain, photophobia, tearing and discharge, and an acute, unilateral clinical picture should particularly raise suspicion.<sup>91</sup>.

## 3.1.2.2.5 Thygeson's superficial punctate keratitis

Thygeson's superficial punctate keratitis is a chronic and recurrent corneal epitheliopathy with episodes of foreign body sensation, tearing, photophobia and visual symptoms. Clinical signs are multiple, slightly elevated, round or oval, white-gray intraepithelial corneal lesions, usually with little or no conjunctival involvement. Etiology and optimal treatment is unknown as yet, but mild topical corticosteroids and immunomodulatory agents are often effective in causing regression of the keratitis.<sup>92</sup>

## 3.1.3 Other mixed and miscellaneous ocular surface disorders

## 3.1.3.1 Limbal stem cell deficiency

Limbal stem cell deficiency is characterized by loss or deficiency of the limbal epithelial stem cells, that are essential for the maintenance of the corneal surface and its physiological functioning. Diagnosis is primarily clinical, with varying signs depending on the stage, such as a stippled staining pattern of the epithelium, late fluorescein staining, loss of corneal

transparency, conjunctivalization of the cornea and superficial corneal vascularization. Symptoms include ocular redness, discomfort, pain, tearing, photophobia and decreased vision, the latter occurring particularly when the visual axis is involved.<sup>93</sup>

#### 3.1.3.2 Episcleritis

Episcleritis is a usually benign and self-limiting disease of the episcleral tissue that presents as superficial bulbar redness, most commonly in the interpalpebral area. In the majority of patients, the inflammation is limited to a single sector, but may involve the entire episclera, and it may present with a semi-mobile nodule. Symptoms are often not present, but may include irritation and tenderness.<sup>94</sup>

#### 3.1.3.3 Mucus fishing syndrome

Mucus fishing syndrome is caused by repetitive eye-rubbing and self-extraction of mucus discharge (for example that arises as a result of DED or allergic conjunctivitis) out of the eye, leading to chronic inflammation of the ocular surface, and even more mucus production, sparking a vicious cycle. An important clinical sign is epithelial damage (as seen by lissamine green staining) of the nasal and temporal conjunctiva.<sup>95, 96</sup>

## 3.1.3.4 Ocular neuropathic pain

Ocular neuropathic pain, or corneal neuropathic pain, is characterized by increased perception of pain in response to stimuli that are normally not painful. It is usually a diagnosis of exclusion, when symptoms outweigh clinical signs. Questionnaires that potentially could be used for the diagnosis of ocular neuropathic pain have been identified, although no gold standard questionnaire has been established. <sup>97</sup> Neuropathic pain can result from injury or disease of peripheral corneal nerves, but may also have a central nervous system origin. The use of an esthesiometer may help in determining hypersensitivity of the cornea <sup>98</sup>. A proparacaine challenge test may help distinguish between neuropathic pain of peripheral or central origin <sup>99</sup>. Reported symptoms may be very similar to those of DED, albeit often very severe, and treatment is often difficult and encompasses both regular ocular surface treatments including anti-inflammatory options, and systemic analgesics, tricyclic antidepressants, anticonvulsants (for example gabapentin and pregabalin), low dose naltrexone, and electrical neurostimulation to reduce pain. <sup>100-102</sup>.

## 3.1.4 Non-ocular surface disease disorders

Finally, the early stage of several non-ocular surface disease disorders of the eye, orbit and surrounding tissues may mimic symptoms of DED. These should be considered particularly when no ocular surface signs are visible. Ocular disorders include refractive error (such as latent hypermetropia), digital eye strain, intermittent angle closure, anterior scleritis and uveitis. Orbital disorders include Tolosa-Hunt-syndrome, thyroid eye disease (which may ultimately lead to severe ocular surface disease secondary to chronic exposure), and carotid cavernous fistula and disorders of the nasolacrimal duct that include dacryocystitis and nasolacrimal duct obstruction. In addition, sinusitis and headache disorders may also present with referred pain around the eye. <sup>103</sup>.

## 3.1.5 Summary

Many conditions can mimic symptoms and/or signs of DED. Failing to investigate possible comorbidities may lead to delayed diagnoses of source conditions and may impact the optimal treatment. To aid in the differential diagnoses, specific questions are listed below. For patients in whom the differential diagnostic history and symptoms suggest it may not be DED, a detailed anterior eye examination using a slit-lamp biomicroscope is warranted <sup>39</sup>, including assessment of:

- Eyelashes for blepharitis, trichiasis, distichiasis, milphosis, and/or poliosis
- Eyelids, including palpebral conjunctiva for irregularities or the presence of follicles, papillae or swelling, and eyelashes for crusting or cylindrical dandruff, and meibomian gland orifices for blockage, pouting or presence of Demodex tails
- Ocular surface for conjunctivochalasis, pinguecula, pterygium and any post treatment/surgery signs of corneal disruption (see section 3.5.4.1)
- Bulbar conjunctiva for redness and/or swelling
- Cornea, including staining, for signs of ulceration, marginal keratitis, erosion, lesions and possible trauma
- Anterior chamber for the presence of cells or keratic precipitates indicating intraocular inflammation.

Medications which can cause DED and the use of cosmetics should be considered (as reviewed by the TFOS Lifestyle reports <sup>8, 9</sup> as well as risk / associated factors (as reviewed in the TFOS DEWS III Digest <sup>4</sup>) including general health conditions.

Key clinical differential diagnosis questions, alongside asking about a patient's general health and medication, are:

- Do you feel eye pain rather than discomfort?
  - DED commonly presents with symptoms of discomfort, light sensitivity or blurred vision, rather than pain, especially in mild to moderate dry eye. If pain is present, investigate for signs of trauma / erosion / infection / ulceration / acute glaucoma and consider administering a pain questionnaire (see section 3.1.2.2).
- Do you have any facial flushing/redness, mouth dryness or enlarged salivary glands?
   Trigger for rosacea, sarcoidosis or Sjögren disease investigation.
- When did your symptoms start and can you recall any triggering event?
  - DED is a chronic condition. If there is increased dryness when waking up or dryness symptoms at night, consider incomplete lid closure, for example.
  - If the onset was sudden or linked with an event, examine the eyes for trauma / infection / ulceration.
  - If the symptoms are linked with contact lens use, refer to their contact lens practitioner to consider alternative approaches that might improve symptoms (such as changing lens material, fit or modality)<sup>104</sup>
- Is your vision affected and if so, does it improve on blinking?
  - A reduction in vision which does not improve with blinking, especially with sudden onset, requires an urgent complete ophthalmic examination to rule out conditions such as vascular occlusions.
- Are the symptoms or any redness much worse in one eye than the other?
  - DED is commonly bilateral. If signs and/or symptoms are significantly greater in one eye, investigate for signs of trauma / infection / ulceration.
- Do the eyes itch, are they swollen or crusty, or is there any discharge?
  - Ocular itching is more likely associated with allergies/history of atopy, while a mucopurulent discharge is associated with ocular infection. Reported itching, specifically along the lash line, warrants assessment for Demodex related anterior blepharitis.<sup>105</sup>

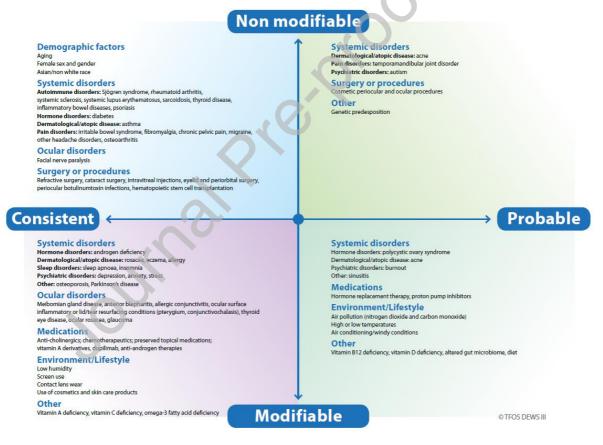
## 3.2 Risk factors

Identifying risk / associated factors may help in finding causes and pathophysiological mechanisms of DED, in determining subtypes, in setting a differential diagnosis, in identifying comorbidities, and in explaining possible discordance between symptoms and signs (see section 3.3.3). In addition, it may help in patient education, and many associated factors may be modifiable, offering the potential to improve symptoms and signs and help prevent/slow progression of disease. In clinical practice, administering a questionnaire to the patient (such as via paper or a digital application) for completion before the consultation is

recommended to save time and to ensure that all commonly associated factors have been addressed.

DED is a multifactorial and heterogeneous disease, and over 200 associated factors have been proposed in the literature. Not all these factors necessarily play a causative role, and some associated factors are linked more strongly with symptoms, while others are linked with signs. Figure 3 shows consistent and probable associated factors of DED as described in the TFOS DEWS III Digest Epidemiology section.<sup>4</sup> The following section briefly highlights the most important risk / associated factors by category. Irrespective of a possible causal mechanism, all associated factors described may be useful in elucidating the etiology of DED and seeking to identify the possible disease driver(s) in an individual. Table 1 presents frequently described risk factors that have inconclusive evidence (either directly conflicting information in peer-reviewed publications, or inconclusive information but with some basis for a biological rationale).

**Figure 3:** Consistent and probable associated factors of DED described in the TFOS DEWS III Digest Epidemiology section <sup>4</sup>.



**Table 1:** Frequently described risk factors that have inconclusive evidence (either directly conflicting information in peer-reviewed publications, or inconclusive information but with some biological rationale):

•	Smoking 106-108
•	Caffeine <sup>7, 109-112</sup>
•	Alcohol <sup>7,113,114</sup>
•	Water intake 7, 115, 116
•	Food restriction <sup>7</sup>
•	Mediterranean diet (possible positive effect) <sup>117-121</sup>
•	Menopause <sup>122, 123</sup>
•	Air pollution from particulate matter of $< 10 \mu m^{6}$
•	Oral contraceptives <sup>124-126</sup>

#### 3.3 Symptomology

3.3.1 Routine questions based on variability of symptoms

Patients with DED often report sensations of grittiness and burning alongside dryness, while contact lens wearers frequently complain of dryness, along with sensations of scratchiness and watery eyes, driving likelihood of failure for long term successful wear <sup>127-129</sup>. Individuals diagnosed with DED present with variable symptom severity, ranging from mild to severe, throughout the day.<sup>39</sup> Dryness symptoms are typically worse upon waking than later in the morning and tend to increase towards the end of the day in contact lens wearers <sup>130-133</sup>. Ocular allergies, exposure to air conditioning, and climatic factors that change between seasons may be associated with seasonal variations in symptoms and signs. However, a cross-sectional, retrospective cohort study in two Japanese clinics found that none of the symptoms examined (dryness, irritation, pain, fatigue, blurring and photophobia) showed significant seasonal variation <sup>134</sup>. This was confirmed by another study, which failed to show seasonal or weather-related variation in the severity of presenting signs or symptoms of DED over a period of 3 years in 652 people in Oslo, Norway <sup>135</sup>. Hence seasonality of DED symptoms, doesn't seem to warrant specific questioning beyond the differential diagnosis if the patient reports the eyes generally itch. However, itching along the lash line may indicate a Demodex blepharitis issue <sup>105</sup>

## 3.3.2 Standardised questionnaires

Around 25 questionnaires can be identified from a literature search using terms "dry eye" and "questionnaire" as well as by research articles cited in the identified publications. Each questionnaire warrants evaluation to assess its clinical relevance, known population validity, concurrent validity, internal consistency and reproducibility <sup>136, 137</sup>. Additionally, unidimensional evaluation validity should be confirmed using Rasch analysis and it should be confirmed whether "health-related quality of life" has been assessed.

Questionnaires such as the Ocular Surface Disease Index (OSDI), Impact of Dry Eye in Everyday Life (IDEEL), Dry Eye-Related Quality-Of-Life Score (DEQS), University of North Carolina Dry Eye Management Scale (UNC DEMS) and the 25-Item National Eye Institute Visual Function Questionnaire (NEI VFQ-25) are validated to a greater or lesser extent in regard to their utility in assessing the impact of DED on health-related quality of life <sup>137</sup>. These questionnaires are not interchangeable, underscoring the necessity of recommending a single, comprehensive, standardized tool in establishing robust diagnostic criteria <sup>137</sup>. A diagnostic questionnaire should have a low response burden on the patient, be simple to score for the clinician and contribute to the assessment of severity and monitoring treatment efficacy. TFOS DEWS II recommended the use of either the OSDI or the 5-item Dry Eye Questionnaire have shown poor comparability. <sup>139-141</sup>

The OSDI-6 was introduced in 2018 to reduce the response burden on patients. Researchers conducted a study that included Rasch analysis, to determine if the effectiveness of the 12 item OSDI questionnaire, could be maintained with a shortened version (Figure 4). The questionnaire was further adapted such that questions would reflect a recall period of one month versus only one week in the original OSDI. The resulting abbreviated version, OSDI-6, consisting of six questions, was found consistently to be more repeatable than either the full OSDI and DEQ-5<sup>142</sup>. Moreover, the scoring method that involves simply summing the item scores and using a diagnostic cutoff of ≥4 to indicate symptoms of DED, allows for a quicker evaluation by clinicians <sup>142</sup>. It is possible that this abbreviated format might also be more suitable for children, who are known to better tolerate questionnaires that require less time and assistance to complete. <sup>143</sup> Similarly to the original OSDI, the OSDI-6 results can be indexed against severity, as normal (0-3 points), mild-tomoderate DED (4-8 points), severe DED (>8)<sup>144</sup>. A Chinese translation of the OSDI-6 found it to be repeatable, valid and psychometrically responsive to DED in a Chinese population <sup>145</sup>. The OSDI-6 thus yields comparable outcomes to the full OSDI, requires less time to complete, and maintains similar variability and improved sensitivity to treatment effects <sup>146</sup>. Consequently, the OSDI-6 merits consideration as a suitable screening questionnaire for DED diagnostic purposes while other questionnaires may offer supplementary aid for risk factor identification and treatment selection, after confirming the initial diagnosis.

	Constantly	Mostly	Often	Sometimes	Never	
Have you experienced any of the following <i>during a typical day within the last month?</i>						
1. Eyes that are sensitive to light?	4	3	2	1	0	
2. *Vision blurring between blinks,	4	3	2	1	0	
with your refractive correction*?						
	Symptoms	and visual d	isturbance s	ubscale ⇔		
Have problems with your eyes limited	l you in perfo	rming any o	f the follow	ing <i>during a t</i>	ypical	
day within the last month?						
3. Driving or being driven at night?	4	3	2	1	0	
4. Watching TV, or a similar task?	4	3	2	1	0	
Visual function / tasks subscale ⇒						
Have your eyes felt uncomfortable in any of the following situations during a typical day within						
the last month?						
5. Windy conditions?	4	3	2	1	0	
6. Places or areas with low humidity?	4	3	2	1	0	
	E	Environmenta	l subscale ≓	>		

**Figure 4**: Questions of the OSDI-6 <sup>146, 147</sup>. The diagnostic cut-off is a summed score of  $\geq$ 4.

## 3.3.3 Discordance between signs and symptoms

It is well established that there is a poor correlation between symptoms and signs in DED. A systematic review including 33 population-based and cohort studies, with a total of 175 individual sign-symptoms associations, found the vast majority of correlations to range between +0.4 and -0.4 <sup>148</sup>. This means that dry eye signs generally explain only up to 15% of the variation in symptoms within a population.

There are numerous methodological, statistical, but also physiological reasons why the correlation between symptoms and signs in studies is low.

Firstly, measurement error is a common problem for many of the DED tests, and the order in which tests are performed or the instillation of fluorescein, for example, can impact test findings <sup>39</sup>. Outcomes are often subjectively scored, not only symptoms, but also purportedly objective signs such as fluorescein break-up time, staining of the ocular surface, and MGD.

- Secondly, DED is recognised to be a dynamic disease, in which symptoms can fluctuate over time, and where test outcomes may be affected by environmental factors, time of the day, and recent artificial tear use <sup>39</sup>. Furthermore, symptoms are often scored as an average over a defined period (such as 'during the last week' in the OSDI), leading to differences in time period over which signs are measured and symptoms are scored.
- Thirdly, DED is multifactorial, with potentially more than one etiology being identified in clinical practice <sup>47, 149</sup>. Studies that include different subgroups of DED in one analysis risk a dramatic fall in correlation values, such as when patients with neuropathic pain are included in studies focusing on more objective DED subtypes <sup>150</sup>. Indeed, when certain subgroups of patients with DED have been analysed independently, higher correlations between symptoms and signs have been found <sup>148</sup>.
- Fourth, neurosensory abnormalities and inflammation of the ocular surface, which are • considered two core mechanisms of dry eye, are not (directly) measured in common clinical dry eye tests. With time, changes in nerve status may effect changes in reported symptomology in DED. For example, a small study of patients with Sjögren disease found that those with advanced corneal staining counterintuitively reported fewer symptoms than patients with more mild corneal staining <sup>151</sup>. Increasing age can also lead to a reduction in corneal sensitivity <sup>152, 153</sup>. A study in the United States of America found correlations between a number of tear film measures and symptoms scores on the 5-item Dry Eye Questionnaire and OSDI to have coefficients all lower than r=0.18, while symptoms were much more strongly correlated with non-ocular pain, depression and post-traumatic stress disorder scores <sup>154</sup>. A large single-centre study in the Netherlands also found poor correlation between common dry eye signs and OSDI symptom scores (all r<0.30), and found correlations to be significantly lower in women than in men, indicating sex differences in symptom reporting in DED <sup>150</sup>. A wide variety of factors, including depression, stress, age, sex and gender can affect symptomology scores.
- Fifth, other comorbid ocular surface disease (that are often associated with DED) may also lead to symptoms (see section 3.1), risking obscuration of true correlations between dry eye signs and symptoms in analyses.
- Sixth, signs of DED may not necessarily lead to symptoms. For example, MGD is a common finding in asymptomatic persons <sup>155</sup>. Seventh, treatment may alter the correlation between symptoms and signs, with certain treatments influencing one variable more than another <sup>156</sup>. Frequently, clinical trials show a positive effect of a DED treatment on symptoms only, but not signs, or vice versa <sup>157-162</sup>. In addition, some longer duration trials have found signs to improve at a later stage than symptoms (for example with artificial tears), or vice versa (with cyclosporine)<sup>163, 164</sup>.
- Finally, despite the longitudinal nature of these clinical trials, there remains a lack of
  natural history studies that attempt to correlate dry eye signs and symptoms within the
  individual patient rather than between patients; there is therefore a need for within
  patient correlation studies across different time points, where it might be expected
  that relationships between signs and symptoms may be stronger.

In recent years, several studies have explored predictors of discordance between symptoms and signs in DED. In a study in the Netherlands, 648 patients were ranked based on a composite dry eye signs' severity score, and also ranked according to symptoms using the OSDI score. Next, a risk factor association analysis was performed with the difference between these two rank scores being the dependent variable. Factors associated with a finding of symptoms exceeding signs were chronic pain syndromes, a history of atopic diseases, allergies, use of antihistamines, depression, the use of antidepressants, and osteoarthritis. Predictors of fewer symptoms than signs were increased age, Sjögren disease and graft-versus-host disease <sup>165</sup>. A similar study in the United States of America with 326 patients replicated many of these factors, finding associations between increasing age and

fewer reported symptoms, while mental health and chronic pain disorders associated with more symptoms. In addition, the study found quantitative sensory testing scores that indicate hyperalgesia to be associated with more symptoms than signs <sup>166</sup>. A recent Taiwanese study with 1229 patients using a similar approach found female sex, and a history of cataract, pterygium and conjunctivochalasis surgery to be associated more with symptoms than signs, and people of older age and those using artificial tears to be relatively less symptomatic than their signs might suggest <sup>167</sup>.

Other studies looking at discordance between signs and symptoms in DED have found corneal microneuroma-like structures and increased corneal dendritic cell density <sup>168</sup>, but also decreased corneal nerve density <sup>169</sup>, and tear and conjunctival cytokines <sup>168, 170</sup> in patients where symptoms outweighed signs. Table 2 lists all the factors across studies that were associated with symptoms outweighing signs, and conversely, those with significantly fewer symptoms than signs. These factors may help in understanding outcomes in clinical practice, aid in patient education (such as in explaining about central sensitisation mechanisms or altered nerve status after surgery) and offer clues for differential diagnoses and comorbid disorders (for example atopy and allergic conjunctivitis) in patients with DED exhibiting discordance between signs and symptoms. The findings also emphasise the need for clinicians not to rely solely on symptom-reporting in older patients, or those with Sjögren disease or graft-versus-host disease, as symptoms may be understated in relation to the severity of effects on the ocular surface.

Predictors of more symptoms than signs	Predictors of fewer symptoms than signs				
Demographics					
450.467	405 407 474				
Female sex <sup>150, 167</sup>	Older age <sup>165, 167, 171</sup>				
Black race <sup>166</sup>					
Pathophysiolo	ogical Factors				
Tear and conjunctival cytokines (IL-10, IL-2, IL-6, IL-17a, tumor necrosis factor alpha) <sup>168, 170</sup>	Decreased corneal nerve density <sup>169</sup>				
Increased corneal dendritic cell density <sup>168</sup> Hyperalgesia as demonstrated with					
quantitative sensory testing <sup>166</sup> Non-ocular pain intensity <sup>166</sup>					
Comorbidi	ty Factors				
Post traumatic stress disorder <sup>166</sup> Allergy <sup>165</sup> Atopic disorders <sup>165</sup> Osteoarthritis <sup>165</sup> Depression <sup>165, 166</sup>	Sjögren disease <sup>165, 172</sup> Graft-versus-host disease <sup>165</sup> Benign prostatic hyperplasia <sup>166</sup> Hypertension <sup>166</sup>				
Anxiety <sup>166</sup> Chronic pain syndromes <sup>165, 166, 173-175</sup>					
Pharmaceuticals					
Use of analgesics <sup>166</sup> Use of anxiolytics <sup>166</sup> Use of antidepressants <sup>165, 166</sup> Use of antihistamines <sup>165</sup>	Use of artificial tears <sup>167</sup>				
Lifestyle Choices					
Current smoking <sup>166</sup>					

Surgery

Cataract surgery <sup>167</sup> Pterygium surgery <sup>167</sup> Conjunctivochalasis surgery <sup>167</sup>

**Table 2**: Factors associated with a discordance between symptoms and signs in dry eye disease. A higher number of studies showing an association is reflected by placement of the factor higher on the list.

#### 3.3.4 Paediatric considerations

While DED is common in children <sup>176</sup> (see TFOS DEWS III Digest epidemiology section),<sup>4</sup> diagnostic tests have largely been validated only in adults. One study examined the quantification of dry eye symptoms in children (6 to 15 years of age) using standardized questionnaires, identifying the completion time was ≤2 min for each individual questionnaire and while younger participants took longer to complete and required more assistance, especially with longer visual analog scales, repeatability was noted to remain high across the age range <sup>177</sup>.

3.4 Ocular Examination

## 3.4.1 Diagnostic Homeostasis Test Battery

#### 3.4.1.1 What is a diagnosis

As previously described, the ability to receive a diagnosis is critical for patients to acknowledge their symptoms and / or signs as real and that they have the attention of health care practitioners, as well as being required for healthcare insurance coverage, where available. For practitioners, diagnostic criteria inform which evidence-based clinical practice guidelines to follow and provide confidence in making patient diagnoses that are consistent with their peers. A standardised diagnosis is essential for industry and researchers to be able to target and validate the efficacy and safety of new products and to obtain regulatory approvals. Appropriate health resource allocation requires robust epidemiological and economic data that are based on consistent diagnoses. Hence diagnosis of a disease must be characterized by standardised, universally-adopted criteria, based on widely available and inexpensive tests with validated cut-off values. There are advantages in having a simple screening element that can rapidly identify those individuals who would benefit from further testing by a health care practitioner.

## 3.4.1.2 Need for standardisation

The definition of DED (see Section 2.1) dictates that symptoms must be present, and a loss of homeostasis of the tear film and / or ocular surface must be established. Identifying that the expected symptoms are present in a standardized way requires a validated questionnaire as discussed in Section 3.3.2. While TFOS DEWS II recommended two possible questionnaires that could be used, these questionnaires have since been shown to not be equivalent, risking variability in diagnosis depending on the instrument chosen, so it is appropriate to select a single questionnaire to be used in this setting. Based on evidence published since TFOS DEWS II, this has been identified as the OSDI-6 with a summed cut-off score of  $\geq$ 4 to be positive for dry eye symptoms (see section 3.3.2).

## 3.4.1.3 Other approaches since TFOS DEWS II

The Asian Dry Eye Society <sup>178, 179</sup> supported by the Japanese Dry Eye Society <sup>180</sup> suggested that DED should be diagnosed by confirming the presence of ocular symptoms (using any one of four possible questionnaires) along with identification of an unstable tear film (assessed with fluorescein tear breakup time of <5s). This approach shows similarities to

TFOS DEWS II, but lacks the standardisation that can be offered by a single questionnaire as proposed by TFOS DEWS III. Furthermore, changing the chemical composition and volume of the tear film by instilling fluorescein prior to assessment of its stability is not ideal. The Korean Dry Eye Society defined dry eye as "a disease of the ocular surface characterized by tear film abnormalities and ocular symptoms" without referring to the pathophysiology, or specifying the means by which to diagnose ocular symptoms (no questionnaire proposed). The diagnostic criteria included an unstable tear film in the form of fluorescein breakup time <7s (a test which is invasive in itself, and recommended without scientific justification for the cut-off), with a Schirmer test (<10 mm) and ocular surface staining considered "adjunctive criteria" <sup>181</sup>.

It has been suggested that diagnostic certainty (in the form of sensitivity and specificity) can be increased by requiring multiple discriminatory tests to be positive <sup>182</sup>; leading to the recommendation that both corneal AND conjunctival staining needed to be present to match a Bayesian-informed global prevalence of DED <sup>183</sup>. However, the reported global prevalence used had been generated from existing studies that relied on these various diagnostic criteria which introduces bias into the approach. Further, using the sensitivity and specificity of tests from multiple studies is also susceptible to bias <sup>184</sup> as previously identified in the TFOS DEWS II report <sup>1</sup>. There is increasing chance of misdiagnosing or under-diagnosing a disease if standardised diagnostic criteria are not used because it can require excessive time, consumables costs or equipment which is expensive or has limited availability. Corneal staining often occurs in later and in more severe stages of DED, so while requiring its presence will increase diagnostic specificity, it will exclude appropriate treatment for many individuals whom clinicians recognise to have a marked loss of quality of life as a result of their symptomatic ocular surface disease and would currently be identified as having DED <sup>185</sup>.

Attempts have been made to use dry eye tests to differentiate diseases that affect the ocular surface such as Sjögren disease, graft-versus-host-disease, Graves' orbitopathy, facial nerve palsy, non-proliferative diabetes mellitus and glaucoma treated with preserved topical drops, with mixed results <sup>186-188</sup>.

3.4.1.4 Use of sensitivity and specificity to select diagnostic tests

Without there being a recognised gold standard, and due to the well-established low correlation between signs and symptoms in DED <sup>189-191</sup>, the level of certainty in derived sensitivity and specificity values of new DED 'diagnostic' tests is low. Sensitivity and specificity depend on the inclusion and exclusion criteria for the 'healthy' and 'disease' groups, increasing with the severity of signs or symptoms required to be allocated to the DED group (spectrum bias). Individuals who do not meet the criteria of either group prevents generalisability of the results across the broad population (sampling bias). Selection bias occurs when the patient groups are selected with test(s) with a similar focus to the test being evaluated <sup>192</sup>. Parallel testing (requiring multiple tests to be positive) will increase the sensitivity and specificity of differentiating a selected 'healthy' and 'disease' group, but this approach is just as susceptible to the bias in group selection. Detailed discussion and examples of these issues are reported in the TFOS DEWS II Diagnostic Methodologies report <sup>1</sup>.

3.4.1.5 Tests to establish a loss of homeostasis of the tear film

While the diagnostic or screening potential of tests such as thermography <sup>193-195</sup>, interferometry <sup>196</sup>, lipid layer pattern/thickness <sup>197</sup> and tear evaporation rate <sup>198</sup> have been evaluated, they are not widely used in clinical or research settings. Artificial intelligence has been used to show that multiple factors including age, ocular surface staining and symptoms are the most important predictors of an unstable tear film, followed by meibomian gland drop-out and expressibility, blink frequency, osmolarity and meibum quality <sup>199</sup>, but it would not be practical to assess all of these in making a diagnosis of DED.

Tear film stability, as usually assessed by the tear film breakup time, is defined as the measured time interval between a blink and the appearance of the first discontinuity in the

tear film <sup>200, 201</sup>. While it is commonly assessed by instilling fluorescein, illuminating the ocular surface with blue light and observing the fluorescence through a yellow filter <sup>202</sup>, the application of fluorescein itself reduces the stability of the tear film and increases its volume so the tear film generally breaks up much more quickly <sup>203-205</sup> and the measurement may not be an accurate reflection of the natural tear film status <sup>206-208</sup>. Fluorescein breakup time is also limited as a measure of tear film stability due to the requirement for subjective assessment by the observer and, while an attempt has been reported to automate this measurement<sup>209</sup>, the required equipment is not available in clinical settings. Instead of the time taken for the first break in the tear film to be detected, a metric to describe overall disruption in tear film surface quality (assessed from a placido disc reflection) has been used to assess tear film instability <sup>210-212</sup>, but this has not been widely adopted. Non-invasive objective assessment, usually with a Placido disc reflected from the tear film surface, determines the time to the first detected break as well as mapping the locations of the tear film disruption and the velocity of the destabilised area. Clinically insignificant differences occur between the objective measurements by many automated devices to measure noninvasive breakup<sup>213, 214</sup>, but not all.<sup>215-217</sup> The detection algorithm results can be adjusted in the validation stage of new instruments (such as adjusting the threshold of discontinuity in the mires) to benchmark more closely to existing devices as there is no 'gold standard'. Controlling environmental factors such as temperature, humidity and air circulation during and immediately before the measurement is important, as well as instructing the patient to blink naturally several times and then to cease blinking until instructed to blink again. <sup>218</sup>. The Ocular Protection Index is the ratio of the tear film breakup time divided by the blink interval, but as with other signs of a loss of homeostasis of the tear film and/or ocular surface. levels deemed 'pathological' are poorly associated with patient reported DED symptoms <sup>219</sup>.

Increased osmolarity of the tear film occurs within the pathophysiology of dry eye <sup>4</sup> with estimated localized levels up of up to around 900 mOsm/kg predicted across the ocular surface at points of tear film destabilisation <sup>220</sup>. Point-of-care devices that measure osmolality are currently limited to sampling from the tear meniscus, as the volume of tears is too small to sample from the ocular surface. Osmolarity has also been reported to differ according to the location along the lid margin from which it is sampled <sup>221</sup>. While some studies have advocated for its diagnostic role <sup>222, 223</sup>, other studies suggest current instrumentation yields results that are not repeatable from a single *in vivo* measurement <sup>224</sup> and values vary with the device used <sup>225</sup>. Tear meniscus osmolarity may account for <5% of the variability in other tear film and ocular surface homeostasis signs <sup>226</sup> and is therefore unreliable in identifying individuals where DED is diagnosed <sup>227</sup>, or the DED severity is determined <sup>228</sup> with other dry eye metrics. The daily variation of osmolarity has been suggested to be a better marker of DED <sup>229</sup>, but this is impractical for diagnosic purposes in a clinical setting. Tear osmolarity was able to predict poor surgical outcomes and patient dissatisfaction in a cataract surgery population with mild ocular surface disease, again suggesting it provides additional information to tear stability and ocular surface damage <sup>230</sup> and hence is an important metric in diagnosing DED<sup>4</sup>. The inclusion of osmolarity amongst the possible signs of a loss of homeostasis of the tear film in the TFOS DEWS II diagnostic algorithm made less than 5% difference in the prevalence of DED <sup>231, 232</sup>. Similarly to osmolarity, tear composition alters where there is loss of homeostasis of the tear film, but not in a measurably consistent manner, and current point-of-care tests for other tear film biomarkers are of limited value (see Section 3.5.4.4.2)<sup>233</sup>.

## 3.4.1.6 Tests to establish a loss of ocular surface homeostasis

Loss of homeostasis of the ocular surface is most widely established by topically applying ophthalmic dyes such as fluorescein and lissamine green, and the resulting staining of the tissues is considered to be a diagnostic sign of DED <sup>234-236</sup>. Staining is one of the clinical signs most strongly associated with dryness symptoms in moderate-to-severe DED <sup>150, 237</sup>. Punctate staining, while not pathognomonic of DED, is commonly associated with desiccation stress, particularly when present in the inferior quadrant of the cornea <sup>238, 239</sup>.

Fluorescein staining occurs due to a loss of corneal epithelial cell integrity such as a disruption in superficial cell tight junctions or defective glycocalyx <sup>238, 240</sup>. Lissamine green has largely replaced the use of rose bengal as it is less toxic to the ocular surface <sup>241</sup>, staining epithelial cells if the cell membrane is damaged, irrespective of the presence of mucin <sup>238, 242-244</sup>. Staining of the eyelid margin conjunctiva that wipes the ocular surface during blinking (termed lid wiper epitheliopathy) possibly due to mechanical stress resulting from insufficient lubrication <sup>245</sup> is common in patients with DED and associated with poor lipid spread, poor tear film stability, abnormal lid anatomy and blink speed <sup>246, 247</sup>. It is also an earlier diagnostic sign than corneal and conjunctival staining in the natural history of DED pathophysiology <sup>248-250</sup>. A clinically detectable poor seal between the eyelids has also been identified as factor associated with symptoms of ocular discomfort <sup>251</sup>.

The diagnostic potential of point-of-care inflammation testing (MMP-9) has been reviewed, but while inflammation was common in severe DED, it was not sensitive to more mild to moderate DED which is more common in the general population.<sup>252</sup> Several studies have investigated the potential of epithelial thickness as a diagnostic test for DED; while the central epithelial thickness has largely to be found to be similar in DED compared to health controls, the superior epithelium is generally thinner, especially in more severe disease,<sup>253, 254</sup> but this finding is not consistent.<sup>255, 256</sup>

Squamous metaplasia and goblet cell density of the conjunctiva can be assessed using impression cytology and, as goblet cell density reduces in patients with dry eye, it has been suggested as a DED diagnostic technique <sup>257, 258</sup>. Impression cytology removes cells from the three most superficial layers of the epithelium, typically by applying cellulose acetate filters or biopore membranes; the cells can then be analyzed by techniques such as microscopy, flow cytometry, immunoblotting analysis, immunocytochemistry and polymerase chain reaction to meet the aims of the investigation <sup>259</sup>. It is a useful alternative to a biopsy, but the changes observed are not pathognomonic of DED <sup>260, 261</sup>.

## 3.4.1.7 Practical diagnostic criteria considerations

Sections 3.4.1.5.and 3.4.1.6 highlight that the key tests to assess the homeostatic status of the tear film remain unchanged from those derived in TFOS DEWS II. The key homeostatic markers are thus non-invasive tear film breakup time, osmolarity and ocular surface staining. The sequence of diagnostic assessment can affect the results as restricting blinking and bright lights can stimulate reflex tearing. It is therefore recommended that tear film assessment tests are carefully ordered, from least to most invasive <sup>262</sup>.

The OSDI-6 is a short questionnaire, ideal for screening and is recommended to be conducted as the first component within routine eye examinations to identify those patients who would benefit from a fuller diagnostic evaluation to determine the likely drivers of disease.

As identified in section 3.4.1, it is critical that diagnosis follows a standard protocol. While combining more tests can improve sensitivity, this may be at the expense of clinical utility.

TFOS DEWS II recommended that one of three signs of a loss of homeostasis needs to be present:

- non-invasive breakup time (first break) <10s: as highlighted in Section 3.4.1.5, this test establishes a loss of homeostasis of the tear film There are now a range of affordable instruments available to the practitioner to avoid the adverse impacts of fluorescein dye on the robustness of the test result as is well documented (see Section 3.4.1.5). Where there is no access to such a device to allow non-invasive measurement of breakup time, fluorescein can be applied, but the volume instilled should be minimised and a cut-off of <5s applied as a positive sign of instability. <sup>204, 205</sup>
- osmolarity ≥308 mOsm/L in either eye or interocular difference > 8 mOsm/L (cut-offs established with the TearLab device only). This test serves as a marker of loss of homeostasis for both the tear film and ocular surface (see Section 3.4.1.5). Consideration of the inter-eye difference alone was found to be valid in one study <sup>263</sup>

but not in another (at least in relating to dryness symptoms) <sup>264</sup> and interocular osmolarity was found to have modest, but superior discriminative ability than absolute osmolarity (higher value of the two eyes) <sup>265</sup>.

ocular surface staining of >5 corneal or >9 conjunctival punctate spots or lid margin staining of ≥2 mm length and ≥25 % width following the instillation of both fluorescein and lissamine green dyes <sup>238, 266</sup>. It is more common to observe corneal and conjunctival staining in severe DED (see section 3.5.4.3)
 <u>Corneal observation</u>: The corneal surface, following the installation of fluorescein drops or application of a moistened fluorescein strip, should be observed using a blue light of 495 nm, as this is the peak excitation wavelength for fluorescein (rather than the 450 nm 'cobalt blue' peak of light filters historically used in slit-lamp biomicroscopes) <sup>202</sup>. An observation filter with a band pass at 500 nm limits visibility to the wavelength of the excited fluorescence molecules (emittance around 515 nm) while excluding those from the applied blue light <sup>202</sup>. Consensus on the ideal time after instillation for assessment is 1-4 minutes <sup>238, 262, 267</sup>.

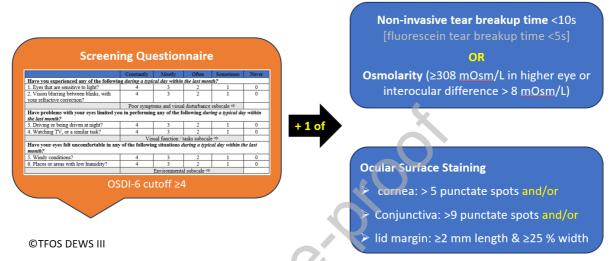
Conjunctival and lid margin observation: Lissamine green staining is critical for observation of conjunctival and lid margin staining assessment. A lissamine green strip (note, not all brands are equivalent)<sup>268, 269</sup> should be moistened with sterile saline with the whole drop applied to the eye after having been placed on the strip for at least 5s to allow the dye to be eluted for maximal concentration <sup>269, 270</sup>. The staining should be observed between 1 and 5 min post-instillation of lissamine green, <sup>269-271</sup> potentially through a red filter to aid visualization <sup>241, 272</sup>. Everting the eyelids multiple times should be avoided as this can stress the tissues and affect the degree of staining observed, whereas exposure time seems to be less impactful on the outcomes.<sup>273</sup> Assessment of lid wiper staining generally involves subjective estimation of the length and sagittal depth of staining, but may be optimised using semi-objective imaging techniques <sup>274</sup>.

<u>Grading</u>: It should be noted that if using a grading scheme such as the Oxford scale, applying half unit increments improves sensitivity and repeatability, while summing regional grading doesn't give a comparable score to global ocular surface staining and increases variability <sup>275</sup>. For diagnosis, counting punctate spots should result in higher consistency. Despite claims that objective assessment of corneal staining can be used as a highly successful automated dry eye diagnostic system <sup>276</sup>, a review of objective techniques suggest this still has limited reliability <sup>275</sup>. While there are challenges to grading staining of the lid margin <sup>277</sup>, a diagnostic criterion of 2mm or more length over at least 25% of the sagittal lid width still seems appropriate.

A study found that a substantial proportion of patients 'diagnosed' with DED by a consultant ophthalmologist without following any set criteria, reported symptoms and exhibited hyperosmolarity, but no other obvious signs of DED; however, tear film stability was assessed invasively and staining of only the corneal (not bulbar or lid margin conjunctiva) was included <sup>191</sup>.

In a UK population study, among the TFOS DEWS II diagnostic signs for DED, conjunctival staining (with lissamine green dye) occurred in most people diagnosed with DED, followed by reduced non-invasive tear breakup time, lower or upper lid wiper epitheliopathy staining, corneal staining and signs of tear hyperosmolarity.<sup>231</sup> The prevalence of DED was notably consistent if any one of the three markers indicating a loss of homeostasis was omitted from the TFOS DEWS II diagnostic algorithm, indicating its robustness.<sup>231</sup> In a larger study <sup>232</sup>, it was found that evaluating just one of the three TFOS DEWS II homeostatic signs resulted in between 12.3% and 36.2% of patients who would otherwise have met the DED diagnostic criteria, not being assigned this diagnosis; hence at least two signs need to be assessed, even although only one needs to be positive. While comprehensive ocular surface staining evaluation in combination with symptoms had the highest sensitivity (87.7%) of the three markers, the sensitivity dropped to 44.6% if corneal staining only was evaluated; hence

conjunctival and lid margin staining assessment (with lissamine green) is critical to diagnosing DED <sup>182, 232</sup>. Omitting either non-invasive tear breakup time or osmolarity each dropped the sensitivity by less than 5%. The prevalence of DED within the population was substantially reduced if diagnosis required symptoms plus two of the three signs to be positive (by between 43.7% to 61.2%) and by 65.9% if all three signs indicating a loss of tear film homeostasis were required <sup>232</sup>. The outcomes of this analysis did not change significantly across differing severities of DED symptoms, confirming the robustness of the DED diagnostic approach (Figure 5).





## 3.4.2 Advanced screening

Diagnosing and monitoring DED often relies on specialized equipment (such as a slit-lamp biomicroscope) and dyes, which are not readily available in non-eye care setting. A non-invasive and simplified test for DED could enable earlier diagnosis and intervention to help prevent disease onset or exacerbations <sup>278</sup>. Additionally, improved accessibility to DED screening could raise public awareness and encourage high-risk or undiagnosed individuals to seek attention by an eye care practitioner.

Blinking is a natural process that refreshes tear film, removes ocular surface debris, and maintains vision quality <sup>279</sup>. Altered blinking physiology is a common feature in DED and is implicated in its pathogenesis <sup>280-282</sup>. As such, DED-associated blinking patterns could potentially serve as a non-invasive biomarker. Blink rate, interblink interval and maximum blink interval (defined as the length of time that the participants can comfortably keep the eye open before blinking), have been reported to be useful in distinguishing between healthy participants and patients with DED <sup>283-288</sup>.

Maximum blink interval (cut-off 12.4 seconds) has demonstrated a sensitivity of 82.5% and specificity of 51.0% for a DED diagnosis based on OSDI symptoms and fluorescein tear breakup time) <sup>289</sup>. A smartphone application has been tested <sup>278, 287, 290, 291</sup>, showing comparable results with conventional paper-based OSDI and subjectively observed maximum blink interval <sup>292, 293</sup>. The app-based maximum blink interval had an area under the curve of 0.649, with a cut-off of 10.5 seconds on a digital device with an iOS, but a much shorter 7.0 seconds on an Android operating system, (compared to 12.4 seconds subjectively assessed) for a symptoms (perhaps due to differences in refresh rate and software algorithms) and fluorescein breakup time DED based diagnosis <sup>294</sup>. Similarly, interblink interval, with a cut-off of 3.1 seconds, has shown an approximately 80% sensitivity and 70% specificity for a DED diagnosis based on general symptoms and corneal staining <sup>283</sup>. Furthermore, patients with DED are reported to have higher blinking rates <sup>295</sup>, with a

greater proportion of incomplete blinks <sup>296</sup>. By analyzing such blinking characteristics, clinicians may be able to gain valuable insights into a patient's ocular surface status.

As maximum blink interval is dependent on an individual's tolerance to eye pain, an alternative approach is asking an individual to report when their eyes become uncomfortable after a blink while keeping their eyes open, which demonstrated a sensitivity of 66%, specificity of 88%, and an area under the curve of 0.77 (cut-off 10 seconds) compared to a full TFOS DEWS II diagnosis of DED, improving to 71%, 90%, and 0.81 respectively when considered in combination with the OSDI score <sup>288</sup>.

#### 3.4.3 Severity rating

The severity of a disease is rated for a diverse range of purposes, but primarily for evaluation and communication <sup>297</sup>:

- ➢ to predict prognosis
- to characterize the impact of the disease on the person's well-being at a given point in time
- > to establish the basis for treatment decisions
- > to evaluate disease activity and monitor response to treatment

Some classifications are based on pathological or physiological status, while others have used impairments or specific symptoms (such as pain) and still others have characterized severity based on exercise tolerance or functional status <sup>297</sup>. Symptoms and outcomes tend to be used in systems that are designed to reflect the patient experience. Pathological or physiological measures have been incorporated in systems used to predict prognosis and both tend to be used to guide treatment, or measure response to treatment <sup>297</sup>. Some severity classification systems also take into account the presence of other conditions, diseases, demographics or behaviors in their staging, if these are considered risk factors for a poorer prognosis.<sup>297</sup>

In other eye diseases, severity ratings vary from those based on the risk of advanced disease progression, for example in age-related macular degeneration <sup>298</sup>, to those variably combining signs and symptoms which aligns with clinician consensus, for example in keratoconus <sup>299-301</sup>. The former approach requires a large sample of natural history data <sup>302</sup> and the latter, a reasonably high association between subjective and objective clinical metrics <sup>303</sup>, neither of which are currently available for DED.

Severity grading in DED has been based on factors such as inflammatory cytokine concentrations <sup>304</sup>, corneal fluorescein staining scores <sup>305</sup> and doctor (clinical judgement) rating <sup>306</sup>. The ODISSEY group proposed a severity rating based on the expert opinion of 10 ophthalmologists <sup>307</sup> where the DED diagnosis relied on poor tear stability alone, and severe dry eye was considered to be those with an OSDI score  $\geq$ 33 and corneal fluorescein staining grade  $\geq$ 3 on the Oxford scale, or with additional criteria if there was less staining. An objective composite index of disease severity has been proposed using an independent component analysis approach <sup>308</sup>, however key non-invasive tests of the tear film and ocular surface were missing such as non-invasive tear breakup time, interferometry, tear meniscus height and lid wiper epitheliopathy and the cut-off for each test's severity grade was based on an "expert panel" of five; the amount of independent information provided by each test (eigenvalue) was used as its weighting for each component's contribution to a composite score, which was based on the sum of the squared measures, divided by the square-root of the sum of the weighting coefficients. A survey approach involving 37 corneal specialists in a hospital setting (non-representative of dry eye practitioners) of unknown location identified clinical tests and cut-offs they felt represented a diagnosis and reflected severity (mild, moderate, severe and very severe) of DED <sup>309</sup>; seven tests were identified and overall severity was based on an algorithm of combining the scores with equal weighting (identified as a limitation) which was based on clinician-ratings (not defined) of 50 patients. A recent study suggested observing corneal cell morphology and density with in vivo confocal microscopy in areas which stain with fluorescein and lissamine green dyes may be a reliable

basis for clinical grading of DED severity, but the cohort of 24 participants was classified into severity grades based on the Chinese Cornea Society criteria which also involved staining, as well as fluorescein tear breakup time, so the observation was not surprising.<sup>310</sup>

A recent review of how to "best diagnose severity levels of DED" erroneously described 'asymptomatic DED', only mentioned non-invasive tear breakup time as a form of interferometry and concluded, without proposing a clinical algorithm, that the evaluation of severity of the condition has often been difficult.<sup>311</sup> While the original TFOS DEWS report proposed a severity matrix <sup>312</sup> based on a prior Delphi panel of 17 DED specialists, <sup>313</sup> this was not adopted by TFOS DEWS II (2017) due to the limited association between the characteristics included and the lack of evidence for the tests included to inform their weighting in a composite algorithm. Likewise, the Asian Dry Eye Society did not propose a severity algorithm for DED <sup>179</sup>. However, the Korean Dry Eye Society proposed a severity matrix, but without scientific justification for the proposed tests, and the severity levels provided <sup>181</sup>. The revised American Academy of Ophthalmology Preferred Practice Pattern guidance <sup>314</sup> mentions the role of DED severity in informing management, but without guidance on how to rate severity. In a recent survey in the UK, patients with DED rated symptom frequency and severity along with tear film stability as the most desired aspects of their DED to improve with treatment, although other factors such as ocular surface, corneal nerve and tear gland damage followed by tear volume and constituents were rated only slightly less important.315

#### 3.5 Subclassification to identify DED etiological drivers

3.5.1 Purpose of a DED subclassification

Diagnostic subcategories (for diseases and most syndromes) are simply concepts. Their purpose is to segregate multifactorial aspects to allow a better characterization of patient outcomes and to guide decision-making regarding treatment <sup>316</sup>. TFOS DEWS and DEWS II confirmed the importance of subclassifying DED into aqueous deficient or evaporative forms, or a combination of the two<sup>2,312</sup>. However, several studies have confirmed that at least twothirds of those with DED exhibit the evaporative form <sup>317-321</sup> which is recognised to have a number of etiologies including lid-related and ocular surface-related,<sup>322</sup> which, without distinction, limits the ability to target treatment to the appropriate etiology. The selection of tests used to differentiate evaporative from aqueous deficient forms of DED varies between studies and a Delphi panel approach has attempted to establish agreement <sup>323</sup>. In addition, it has been noted that between 18 % and 29 % have no obvious signs of either a reduction in tear volume or disruption to meibomian gland structure and function, suggesting the need to acknowledge other subtypes of the disease <sup>191, 319-321, 324</sup>. DED is accepted as a multifactorial disease <sup>2, 20</sup> so addressing the different mechanisms leading to an individual's DED could impact treatment outcomes. Disease heterogeneity will reflect differences in the underlying pathophysiology, genetic risk and environmental contributors of affected individuals <sup>325</sup>. Clinical tests that identify the possible drivers of an individual's disease (which are not mutually exclusive), in turn, inform the appropriate treatment approach(es) <sup>326</sup>. The Asian Dry Eve Society <sup>179</sup> proposed four targets for therapy (lipid, aqueous and mucin layers along with ocular surface inflammation), but with a target of the epithelium consisting of membraneassociated mucins and epithelial (goblet) cells. Expanding on this approach, the following section outlines those clinical tests that inform the clinician about the contribution to DED from:

#### 3.5.2 Tear Film Deficiencies

The latest understanding on the tear film is reported in the TFOS DEWS III Digest.<sup>4</sup>. It has been proposed that differences observed in the fluorescein tear breakup patterns can inform the clinician about which tear film layer has been disrupted. Area break is thought to be associated with severe aqueous-deficient DED; spot, dimple, and line breaks with rapid expansion are associated with decreased wettability DED, with a line break thought to be associated with mild-to-moderate aqueous-deficient DED; and a random break appearance

is associated with increased evaporation DED <sup>327-329</sup>. However, to date, there has been only limited published evidence to support these hypotheses <sup>329</sup>.

#### 3.5.2.1 Lipid component

#### 3.5.2.1.1 Interferometry

Tear interferometry allows the tear film lipid layer thickness to be estimated, noninvasively <sup>330, 331</sup>. Due to nature of the thin lipid layer overlying a body of aqueous with different refractive index, reflections from the air-lipid and the lipid-aqueous interfaces create interference patterns, which can be analyzed quantitatively or semi-quantitatively<sup>332</sup>. Lipid layer interference pattern grades correlate with corneal staining and tear film breakup time <sup>3</sup>. Lipid layer thickness should be increased in hypersecretory MGD and decreased in obstructive meibomian MGD, but a direct association between the thickness of the lipid layer and the rate of tear evaporation has not been proven <sup>334, 335</sup>. There are various non-invasive diagnostic devices for assessing the tear film lipid layer <sup>336-342</sup>. Some devices attach to a slitlamp biomicroscope base while others are stand-alone instruments. Most require subjective grading of the lipid pattern <sup>343</sup>, which equates the pattern observed to an estimated thickness <sup>37</sup>. A dynamic lipid layer interference patterns test has been proposed, reporting the optimal number of blinks to observe a significant change in lipid pattern as being up to five forced followed by 10 natural blinks at 2s intervals; in patients with DED, the number of blinks required to change the lipid pattern (2.4 + 3.1 blinks) was statistically lower than in healthy subjects  $(18.1 \pm 5.9 \text{ blinks})^{344}$ . The LipiView interferometer has a sensitivity of around 68% and specificity of 64% if a cut-off value of 75 nm is used for MGD diagnosis <sup>332, 345</sup>; the coefficient of variability for inter-observer repeatability was 13 nm and the intra-observer repeatability 16 nm in healthy individuals, with values less than 60 nm considered pathological <sup>346</sup>. In a study of 221 participants, optimal diagnostic cut-offs for DED based on the TFOS DEWS II criteria were <72nm with the LipiView and a grade of  $\leq3$  subjectively. based on interferometric patterns (modified Guillon scale).<sup>197</sup> Lipid layer thickness values obtained with the LipiView instrument have been reported to correlate well with meibomian gland loss <sup>347</sup>. Another technique, using a spectrophotometer, claims an ability to directly image tear muco-aqueous and lipid layer thicknesses in vivo with nanometer axial resolution 348, 349

## 3.5.2.1.2 Lipid turnover

The turnover of the lipid terms assessed by fluorophotometry  $0.9 \pm 0.4$  % / min) is slower than the aqueous turnover ( $10.3 \pm 3.7$  % / min).<sup>350</sup> Contrast-enhanced optical coherence tomography (OCT) imaging has also been used to evaluate the clearance of lipids in human tears <sup>351</sup>. A system that combines simultaneous OCT and thicknessdependent fringes interferometry can be used for *in vivo* assessment, simultaneously imaging both the lipid layer thickness and overall tear film thickness <sup>352, 353</sup>. The analysis of the OCT's *en face* maps of the tear lipid layer provides complementary information to interferometry <sup>354</sup>. The tear film imager uses spectral interference to allow real-time evaluation of the rate of lipid thickness change and discontinuations over a large field of view at nanometer axial resolution <sup>330</sup>. The mucoaqueous thickness correlates with the Schirmer score and lipid or fluorescein tear breakup time <sup>349</sup>.

## 3.5.2.1.3 Evaporimetry

A meta-analysis of studies measuring evaporation found raised levels in patients with DED, particularly evaporative DED (normal,  $13.57 \pm 6.52 \times 10^{-7}$  g/cm<sup>2</sup>/s; aqueous deficient dry eye,  $17.91 \pm 10.49 \times 10^{-7}$  g/cm<sup>2</sup>/s; evaporative dry eye,  $25.34 \pm 13.08 \times 10^{-7}$  g/cm<sup>2</sup>/s)<sup>355</sup>. Different instruments have been used for assessing evaporation, although only one is currently commercially available <sup>356-359</sup>. Tear evaporation has been shown to be reduced by eyelid warming therapy <sup>360</sup>. Confounding factors are the sampling response rate to blinking, perspiration within the sampling area, palpebral aperture, variation in blink speeds and patterns, and in the level of chamber ventilation, and differences in resistance to evaporation caused by humid, and still air <sup>361, 362</sup>.

## 3.5.2.1.4 Thermography

Thermography uses a specialized camera to detect infrared radiation emitted from the ocular surface, mapping changes in the ocular surface temperature that are presumed to be caused by tear fluid evaporation <sup>363</sup>. The technique seems repeatable <sup>364</sup>. Thermal cooling of the ocular surface is a predictor of soft contact lens induced dryness symptoms <sup>365</sup>. Ocular surface temperature decreases more rapidly following a blink in individuals with adequate tear volume, but unstable tear films.<sup>366-368</sup> The temperature differential between the central cornea and limbus is higher in DED than in normal,<sup>369</sup> with evaporative DED associated with higher ocular surface temperature than aqueous deficient DED and patients with MGD having higher ocular surface temperature than those with healthy eyes.<sup>367, 370</sup>

## 3.5.2.1.5 Meibum expressibility and quality (meibometry)

Meibomian gland functionality is assessed by testing the expressibility of meibum and the quality of expressed meibum <sup>332, 371</sup>. Meibum quality is typically graded as 0 (clear fluid), 1 (cloudy fluid), 2 (cloudy particulate) or 3 (toothpaste-like). The expressibility of meibum from the meibomian glands of the upper and lower eyelids is graded after 10-15 seconds of applying pressure as 0 (all glands expressible), 1 (3 to 4 glands expressible), 2 (1 to 2 glands expressible) or 3 (no glands expressible). Meibomian gland expressor devices have been developed as a means of standardizing the pressure of 'diagnostic' expression, which aims to be equivalent to that of a natural blink <sup>371</sup>. Meibum quality and expressibility are correlated with gland loss and lipid layer thickness <sup>372-375</sup>.

Meibometry is a technique applied to assess lipid volume <sup>376</sup>, involving sampling lipid from the lower lid margin with a loop of translucent plastic tape. The tape is air-dried for 3 min to allow evaporation of any contaminating tear fluid and the optical density of the oil retained on the tape is assessed using a diode laser. A correlation has been reported between lower lid meibometry readings and expressibility of meibomian lipid from the central upper lid, as well as a reduction in volume in patients with MGD and an improvement after meibomian gland orifice probing <sup>377</sup>.

A test strip made of hemp oil-absorbing material, designed specifically to absorb tear lipid, has been developed. The folded end is placed in the conjunctival sac of lower eyelid and the length of infiltration (over 1 minute seemed optimal) measured and recorded, similar to the Schirmer test <sup>378</sup>.

Tear lipidomics are reviewed in Section 3.5.4.4.2.

## 3.5.2.2 Aqueous

As the aqueous component represents the majority of the tear film volume, techniques assessing tear volume are often used to quantify this tear component.

## 3.5.2.2.1. Meniscometry or tear meniscus assessment

Meniscometry involves non-invasive biometry of the lower eyelid tear meniscus, usually in the form of a central height in primary gaze. Subjective methods of tear meniscus height measurement, such as estimating the meniscus height relative to a height-adjusted slit-lamp beam scale, has shown poor inter-visit reproducibility <sup>379</sup>. Slit-image photography has also been used to quantify tear meniscus height, radius, width, as well as cross-sectional area and radius of meniscus curvature <sup>376</sup>. The TFOS DEWS II Diagnostic Subcommittee proposed tear meniscus height assessment as a differential factor for the subclassification of DED, describing a cut-off value of 0.20 mm or lower as an indicator of aqueous-deficient DED, which has subsequently been confirmed <sup>39, 320, 380</sup>. Tear meniscus height should be measured directly below the pupil midline (±1mm) as it is affected by varying lid morphologies more peripherally.<sup>381</sup> The timing of the assessment after a blink should be controlled, with 1.0 to 2.5s after two blinks found to be most robust; a single measure of tear meniscus height is sufficient, using either with infrared or visible white light (although these are not interchangeable).<sup>382</sup> Alternative meniscometry systems have been developed in

research settings, projecting a target to dynamically visualise the tear meniscus curvature, without the need for fluorescein instillation <sup>379, 383, 384</sup>.

OCT allows the cross-sectional area of the tear prism or even the volume along the lower lid to be quantified <sup>385, 386</sup>. Spectral-domain OCT has enabled higher resolution, greater imaging depth and faster acquisition (facilitating three-dimensional volume imaging) compared to time-domain OCT, improving image quality and measurement repeatability <sup>330, 387-389</sup>. The high repeatability allows changes in tear meniscus morphology after fluid instillation to be tracked, to determine tear clearance rate <sup>390</sup>. Meniscus measurements are instrument-dependent <sup>391</sup> and can be distorted by anatomical features such as lid parallel conjunctival folds, conjunctivochalasis, or other disruption to the shape of the lid margin or ocular surface <sup>392</sup>. Furthermore, analysis of the image may be complex, time-consuming and operator-dependent <sup>393</sup>.

## 3.5.2.2.2 Phenol red thread test

The slight alkalinity of the tear film (between pH 7 and 8)<sup>39</sup> allows the length of a thin cotton thread, with the folded end hooked over the temporal end of the eyelid, moistened by tear absorption over a 15 second period to be observed from a color change of yellow to red. Compared to the Schirmer test (see Section 3.5.2.2.3), the small profile and limited amount of pH indicator in the thread is expected to limit the chance of triggering substantial reflex tearing and as a result, intersession repeatability is good.<sup>394</sup> While the phenol red test is thought to indirectly measure the tear volume, it is only weakly correlated with other established methods such as fluorophotometry or tear meniscus height.<sup>394, 395</sup> In addition, it is weakly correlated with dry eye symptoms<sup>396</sup> Reported agreement with the Schirmer test is variable between studies.<sup>397, 398</sup> An arbitrary aqueous deficient DED cut-off value of 20 mm has been adopted in clinical practice <sup>399</sup> and values of <9mm in 15 seconds suggest more severe cases of aqueous deficient DED<sup>400</sup>. Due to issues with accessibility of a commercialized product, techniques to develop an equivalent test have been described <sup>401</sup>.

## 3.5.2.2.3 Schirmer test

The Schirmer test is an invasive test of tear volume, that involves assessing the length of a (Whatman 41) filter paper strip that becomes saturated by tears, 5 minutes after hooking the end of the strip, folded at the notch, over the lower lid margin, within the temporal one-third <sup>332</sup>. Technique variations, such as the use of anaesthetic which aims to differentiate basal from reflex tearing <sup>402</sup>, indicate poor repeatability, sensitivity and specificity <sup>1</sup>. Using the wetting of the Schirmer strip over the final 4 minutes out of the 5, seems to be more robust than other time intervals including assessment over the full 5 minutes, but not surprisingly, accounts for <3% of the variance in fluorescein breakup time, a key homeostatic marker for all DED subtypes, or meibomian gland secretion, a recognised marker of evaporative DED.<sup>403</sup>

## 3.5.2.2.4 Strip meniscometry

Strip meniscometry involves placing a strip with a 0.4mm diameter central duct into the lower lid tear meniscus for 5 seconds <sup>404-406</sup>. A cut-off of  $\leq$  2.5 mm has been adopted, with the results correlating with other tear film assessments and the values are moderately repeatable <sup>407, 408</sup>. The combination of strip meniscometry and fluorescein breakup time have been proposed as a more sensitive approach for assessing DED than either test alone.<sup>386</sup>

## 3.5.2.2.5 Tear clearance

Measuring the fluorescence of instilled quantities of fluorescein across the ocular surface (termed fluorophotometry) can be used to quantify tear turnover, reported as the decrease (in percent) per minute <sup>409</sup>. Alternative approaches to measuring tear flow include conducting a Schirmer test 5 minutes after instilling fluorescein with anesthetic, with both the length of wetting and the intensity of strip staining compared to a standard color plate after 5 minutes recorded <sup>410</sup>. Due to the various factors that can affect Schirmer test results, the tear function

index, which is the value obtained by dividing the value of the Schirmer test by the tear clearance rate, is a measure that has been used by a limited number of authors to assess patients with DED <sup>411-413</sup>. Other methods of tear clearance assessment include anterior segment OCT, which has been used to evaluate the early phase of tear clearance <sup>414</sup>. Lacrimal scintigraphy (tracking a small amount of radioactive material instilled into the conjunctival cul-de-sac) has also been used to measure tear clearance <sup>415, 416</sup>.

## 3.5.2.2.6 Lacrimal gland patency

A clinical test for examining the patency of the palpebral lobe of the lacrimal gland has been described, which involves having the patient look inferonasally while the upper eyelid is retracted in the superotemporal direction. Dry 2% fluorescein ophthalmic strips are applied onto the exposed palpebral lobe multiple times over 20s. This allows the number and location of ductules per lobe as well as the tear flow rate to be assessed <sup>417</sup>. Alternatively the patency of the lacrimal gland can be assessed by stimulating the ocular surface with a pure CO<sub>2</sub> gas jet at 200 ml·min<sup>-1</sup> for 3s, delivered 5 mm from the cornea and measuring the increase in reflex tearing volume with a Schirmer strip <sup>418</sup>.

## 3.5.2.2.7 Tear proteins and other components

The presence of the antibacterial and anti-inflammatory lacrimal gland proteins, lipocalin, lactoferrin, and lysozyme, which possess anti-inflammatory and anti-bacterial properties <sup>419</sup> can be assessed as an indirect measure of lacrimal gland function. Tear protein concentration has generally been found to decrease with age <sup>420</sup>. Although lactoferrin has been proposed as a biomarker of DED <sup>421-423</sup>, low tear lactoferrin levels are also found in giant papillary conjunctivitis, vernal keratoconjunctivitis, and chronic meibomitis associated with acne rosacea <sup>424-426</sup>. Tear film urea levels are linearly related to Schirmer's test values; for diagnosing DED, a cutoff of  $\leq$  37.2 mg/dL has been reported to provide a sensitivity of 96% and a specificity of 76% <sup>427</sup>. Tear fluid proteomics are reviewed in section 3.5.4.4.2.7.

## 3.5.2.3 Mucin / glycocalyx

## 3.5.2.3.1 Mucins

Human conjunctival goblet cells synthesize and secrete the largest type of gel-forming, nonsurface-bound mucin in the eye, MUC5AC, which acts to protect and lubricate the ocular surface, mitigating friction during blinking <sup>4, 428</sup>. Patients with DED typically show reduced concentrations of soluble MUC5AC in the tear film <sup>429</sup>. Together with lipids, a concomitant increase in MUC5AC protein expression in tears in infants may contribute to their greater tear film stability <sup>430</sup>. Goblet cell count and tear MUC5AC protein are decreased in Graves´ ophthalmopathy, thought to be possibly due to ocular surface inflammation secondary to ocular surface exposure <sup>431</sup>.

Immunohistochemistry and immunoelectron microscopy have been used to examine binding of the HI85 antibody, which recognizes carbohydrate epitopes on the MUC16 mucin on the surfaces of apical conjunctival cells <sup>432, 433</sup>.

Other membrane-associated mucins, MUC1, MUC4, as well as MUC16 (glycocalyx) and gelforming mucin MUC5AC, have been studied using different techniques <sup>244, 434, 435</sup>. In other research large decreases in sialic acid (almost 7-fold) <sup>436</sup> and increases in galectin-3 (proxies of the glycocalyx/mucin) <sup>437</sup> have been observed in the tear film and shown to strongly correlate with clinically assessed disease severity <sup>438</sup>.

## 3.5.2.3.2 Rose bengal and lissamine green (see section 3.4.1.7)

Rose bengal has been shown to stain ocular surface epithelial cells that are unprotected by the mucin rich glycocalyx <sup>439</sup>, but it has been shown to suppress human corneal epithelial cell viability *in vitro*, indicating toxicity <sup>240</sup>.. Lissamine green is less toxic to the ocular surface and has largely replaced the use of rose bengal in clinical care and research. Lissamine

green is a vital dye that stains epithelial cells only if the cell membrane or intracell junctions are damaged, irrespective of the presence of mucin <sup>39, 440</sup>.

## 3.5.2.3.3 Conjunctival impression cytology

After the instillation of topical anesthetic, a filter strip is pressed against the bulbar conjunctival surface, usually upper, for 5-10 seconds using forceps. The samples are then fixed using 95% ethanol, stained with periodic acid-Schiff reagent and fixed on a slide to be viewed with a light microscope <sup>261</sup>. The Nelson classification system is used most frequently to grade the density, morphology, cytoplasmic staining affinity and nucleus/cytoplasm ratio of conjunctival epithelial and goblet cells across the ocular surface <sup>39</sup>. There is variation in goblet cell distribution across the conjunctival surface, with the lower forniceal conjunctiva goblet cell density higher than in the bulbar conjunctiva <sup>441</sup>. Recently, moxifloxacin-based fluorescence microscopy has emerged as a novel technique that enables efficient, non-invasive and *in vivo* animal imaging of goblet cells <sup>442</sup>. Confocal imaging, on the other hand, can be used to assess corneal cell morphology <sup>443</sup>, goblet cell density <sup>444</sup> and conjunctival squamous metaplasia (from nucleocytoplasmic ratios) <sup>445</sup> *in vivo* in humans.

## 3.5.2.3.4 Ferning test

Whole tears collected by one of many possible techniques <sup>446</sup> are transferred immediately to a small plastic centrifuge tube (0.5 ml or less); a sample (1–2 µl) is then pipetted onto a clean glass microscope slide and allowed to dry for 7–10 minutes under normal room temperature (20–26°C) and room humidity (up to 50 %) <sup>447</sup>. The slide then can be observed under a light or digital microscope with various magnifications <sup>446</sup>. Depending on the tear film composition, a variety of ferning patterns can be observed. The classification is typically: type I: uniform large arborization; type II: ferning abundant but of smaller size; type III: partially present incomplete ferning; and type IV: no ferning <sup>448</sup>. Healthy tear samples typically produce full dense ferning patterns (types I and II), while the ferning pattern are often fragmented or absent in patients with DED <sup>446</sup>.

The exact nature of what determines the ferning pattern is still not fully understood <sup>1, 449</sup>, though a causal link has been proposed between tear ferning pattern and the ocular mucins <sup>449</sup>. Hyperosmolarity affects the appearance of ferning patterns <sup>446</sup> as does introducing electrolytes into the tear film <sup>450</sup>. Tear ferning patterns have been found to be repeatable <sup>451</sup>. A correlation has been found between the ferning test grade, non-invasive breakup time <sup>452</sup> and low Schirmer test values, but no correlation has been reported between ferning pattern and various tear protein levels. <sup>453</sup>

## 3.5.3 Eyelid anomalies

## 3.5.3.1 Blink and lid closure anomalies

Blinking is a complex neuromuscular process that plays a vital role in maintaining ocular surface homeostasis and proper functioning of the tear film. Specifically, it facilitates the even distribution of tears, mucin, and lipids, essential for lubrication, protection from eye irritation, and removal of debris and foreign bodies <sup>289, 454-456</sup>. Blinking is primarily controlled by the orbicularis oculi muscle, innervated by the facial nerve (cranial nerve VII) <sup>457</sup>. The levator palpebrae superioris muscle, innervated by the oculomotor nerve (cranial nerve III), and Müller's muscle, innervated by sympathetic fibres, are also contributory <sup>458</sup>. Sensory input from the cornea and conjunctiva, relayed via the trigeminal nerve (cranial nerve V), modulates the blink reflex <sup>459</sup>.

Blinking can be categorized into three types <sup>460</sup>. The first is spontaneous blinking, which is the unconscious and coordinated closure of both upper eyelids occurring briefly and symmetrically without any evident stimulus. The second is reflex blinking, which is triggered by trigeminal, visual, and acoustic stimuli. Lastly, voluntary blinking is defined as the closure of eyelids consciously initiated by the individual. The normal rate for spontaneous blinking ranges from 10-15 blinks per minute <sup>295, 461</sup>. This rate can be influenced by multiple factors,

such as age, cognitive load, social activity, neurological and psychiatric diseases, fatigue, eye injury, medication, contact lens wear and dryness <sup>295, 460</sup>.

Abnormalities in blinking patterns have been implicated in DED development <sup>279, 282, 289</sup>. Blink rates, interblink interval and maximum blink interval can differentiate patients who are considered healthy from those with DED <sup>285, 291</sup>, likely due to the changes in ocular surface exposure and failure to restore tear film structure between blinks. Patients with DED often exhibit increased blink rates <sup>295</sup> relative to normal. However, reduced blink rates have also been observed both in individuals with and without DED, particularly during activities requiring prolonged visual attention (e.g., screen use), leading to insufficient tear film distribution and increased tear evaporation rates <sup>462</sup>. Similarly, studies have shown that patients with DED had shorter mean and maximum interblink intervals compared to healthy controls <sup>283, 289, 463</sup>. Notably, people with DED also demonstrated higher rates of incomplete blinking <sup>296</sup>, which in itself is associated with inadequate expression of meibomian gland secretions into the lipid layer on the ocular surface, exacerbating tear film instability 464-466. Given the multifactorial nature of DED, a comprehensive evaluation of blinking physiology is essential for understanding its pathophysiology and employing effective treatment strategies. Clinical monitoring is usually subjective and surreptitious to avoid a change in blink pattern from the patient <sup>467</sup>, but objective image analysis is becoming more widely available. Improving blink quality and frequency through behavioral modifications and therapeutic interventions can significantly benefit ocular surface health and patient comfort. Poor lid seal is also linked to symptomology in DED and can be detected by placing a pen torch or transilluminator against the relaxed, closed, outer upper evelids of semi-reclined patients and observing visible light emanating from the lid area between the lashes <sup>251</sup>.

#### 3.5.3.2 Lid margin health

#### 3.5.3.2.1 Anterior blepharitis

Anterior blepharitis has been defined as "an inflammation of the lid margin anterior to the gray line and concentrated around the lashes" which may be accompanied by squamous debris or cylindrical dandruff around the lashes (Figure 6), and inflammation may spill onto the posterior lid margin<sup>47, 468, 469</sup>. The term 'blepharitis' is often used by clinicians to describe anterior blepharitis, with posterior cases referred to by the more specific etiology, such as meibomian gland dysfunction. The pathophysiology of anterior blepharitis is multi-staged and relates to microbial changes that can culminate in inflammation: periocular bacteria build a defensive structure known as a biofilm, which predisposes to forming on the eyelid margin due to its moisture, nutrients and warmth. Biofilms are composed of a polysaccharide/protein matrix, that adhere strongly to surfaces due to proteins like adhesin produced by bacteria such as *Staphylococcus epidermidis* and *Staphylococcus aureus*. Biofilms allow bacteria to evade the impacts of desiccation and host defense mechanisms, facilitating gene activation and inflammatory virulence factors <sup>470</sup>. Dry eye symptoms <sup>471</sup> and signs, <sup>472</sup> as well as contact lens discomfort, <sup>473, 474</sup> are reduced on treatment anterior blepharitis, <sup>475</sup> implicating anterior blepharitis as one of the key triggers of the multifactorial disease.

Common clinical manifestations of anterior blepharitis include the presence of squamous debris or cylindrical dandruff around the base of the lashes, accompanied by vascular changes in the lid skin <sup>468, 476</sup> Anterior blepharitis is associated with ocular rosacea, seborrhea and hypersensitivity caused by staphylococcal toxins, infectious processes (bacterial or viral) or infestation by phthiriasis or *Demodex, or combinations* of these triggers <sup>477</sup>.

The most frequently identified species are *Staphylococcus epidermidis* (*in about one third of cases*), followed by *Pseudomonas* (approximately 20%), *Staphylococcus aureus*, *Propionibacterium*, *Corynebacteria*, and *Moraxella*<sup>478, 479</sup>. Immunologic mechanisms have been documented, with enhanced cell-mediated immunity to *Staphylococcus aureus* detected in 40% of patients with chronic blepharitis. Seborrheic blepharitis, in contrast, presents with greasy deposits and is commonly associated with seborrheic dermatitis of the eyebrows and scalp<sup>476, 480</sup>.

## 3.5.3.2.1.1 Demodex associated blepharitis

Anterior blepharitis can also result from the activity of *Demodex folliculorum* or *brevis*, parasites identified in 14% to 89% of the population (especially in older patients)<sup>105, 481-483</sup>. There are two types identified in the human eyelids: Demodex folliculorum and Demodex brevis<sup>484</sup>. Demodex folliculorum is typically found in the eyelash follicles of the eyelids and observed with high magnification slit-lamp microscopy<sup>485</sup>.

Characteristic features are apparent at the base of the lashes, and these can be present in asymptomatic individuals. *Demodex* mite presence is associated with changes in the anterior lid margin, such as increased scale intensity and cylindrical dandruff or sleeves. Cylindrical dandruff are considered pathognomonic for the presence of *Demodex* mites <sup>485</sup>,

<sup>486</sup>. Demodex can be detected by examining epilated eyelashes under white light microscopy at high magnification <sup>487, 488</sup> or on a slit-lamp at high magnification. Manipulating an eyelash with cylindrical dandruff around its axis in a circular motion <sup>489, 490</sup> or by applying lateral traction <sup>491</sup> using fine-tipped metal forceps, can reveal mite tails at the insertion point of the lashes. Detection of Demodex using *in vivo* confocal microscopy has also been described, but this technique is cumbersome, time-consuming and costly, limiting its diagnostic utility <sup>491, 492</sup>.

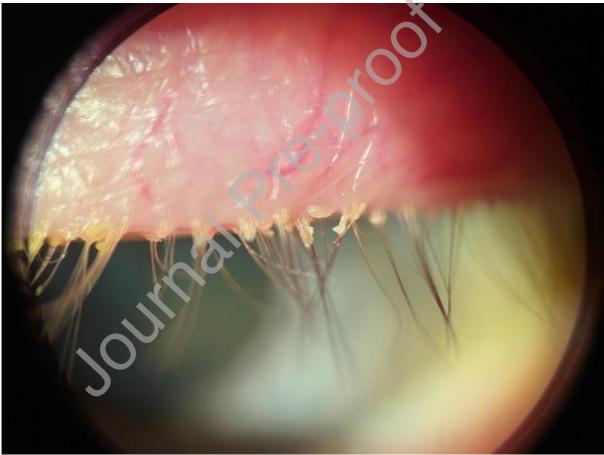


Figure 6: Anterior blepharitis.

### 3.5.3.2.2 Meibomian gland dysfunction (MGD)

MGD, a major contributing factor to DED, is described as "a chronic, diffuse abnormality of the meibomian glands, commonly characterized by terminal duct obstruction and/or qualitative/quantitative changes in the glandular secretion" in the TFOS International Report on Meibomian Gland Dysfunction Definition and Classification report published in 2011. <sup>468</sup>. The diagnosis of MGD is primarily clinically-based, focusing on the detection of signs indicative of altered meibomian gland secretions, lid margin changes, and meibomian gland dropout <sup>372</sup>. The severity of MGD is classified based on subjective symptoms, abnormal signs at the orifices (plugging, pouting and capping), vascularization or reddening of the

eyelid margin, anterior or posterior displacement of the mucocutaneous junction, eyelid margin irregularity and rounding, gland drop-out observed by meibography (where imaging both upper and lower lids is important <sup>493</sup>), corneal and tear film abnormalities (such as superficial punctate keratitis) and the quality of the expressed meibum <sup>494-496</sup>. Various clinical grading criteria for gland appearance have been proposed <sup>495, 497-499</sup>. Additionally, the term "plus disease" has been used to refer to co-existing or accompanying disorders of the ocular surface and/or eyelids <sup>331</sup>. Another scoring system combines subjective symptoms, slit-lamp microscopy findings, and tear test results to classify MGD severity into stages 1 to 4 and describes the relationship between meibomian gland dysfunction severity and serum lipids <sup>500</sup>. An alternative scoring system integrates subjective symptoms, *in vivo* confocal microscopy findings, slit-lamp microscopy, meibography, tear film breakup time and corneal staining scores, classifying MGD severity into three stages <sup>494, 500</sup>. Yet another scoring system elected to combine scores for secretion appearance and the digital pressure needed to express the meibum <sup>501</sup>.

The number of lower eyelid meibomian glands yielding liquid secretion has been assessed as a metric of MGD <sup>371</sup>. Assessing meibum expressibility, color and quantity is crucial <sup>502</sup>, either as the highest grade from eight expressed glands (total score range: 0 to 3) or summing all eight gland expressibility scores (total score range: 0 to 24) <sup>467, 468</sup>. Anatomically, there are more meibomian glands in the upper eyelid, with a median count of 31, compared to a median count of 26 in the lower eyelid <sup>469</sup>. The glands in the upper eyelid are notably longer and slimmer than those in the lower eyelid. Studies on lower and upper eyelids separately highlight secretion differences between the nasal and temporal sides. The lower eyelid <sup>467</sup>. Additionally, nasal glands appear more active, with activity diminishing toward the temporal margin <sup>503</sup>. Upper eyelid meibograpy has been proposed to be an integral component of comprehensive meibomian gland evaluation.<sup>493</sup>

Non-invasive infrared or transillumination photography of meibomian glands has advanced the assessment of two-dimensional gland silhouettes <sup>498, 504</sup>. Meibography enables direct visualization of meibomian gland morphology, highlighting partial or total non-visible meibomian tissue as dropout or loss <sup>372, 505</sup>. This can be assessed through subjective grading <sup>468, 506-508</sup>, or semi-quantitative or quantitative analyses <sup>509</sup>. Asymptomatic individuals have been found to have meibomian gland loss <16.9% for the upper and <28.7% for the lower eyelid <sup>498</sup>.Various gland morphological characteristics have been identified, including truncation, dilation and tortuosity <sup>510, 511</sup>, but none in isolation seems to be a good predictor of DED <sup>512</sup>.

Tortuosity of the meibomian glands in the upper eyelid has been observed to correlate negatively with tear film stability, while tortuosity in the lower eyelid correlated with dry eye symptoms <sup>508</sup>. However, its variability makes it less reliable as a standalone diagnostic parameter <sup>503, 508</sup> and it is only weakly correlated with meibomian gland expression <sup>372</sup>. Meibomian gland thickness increases with overall loss, potentially as a compensatory response, although this does not improve expressibility <sup>372, 503</sup>. On the other hand, distorted and thinned glands appear to be transitional phases before dropout <sup>468, 469, 502</sup>. However, meibomian gland length has been identified as the key morphological metric for function in terms of expressibility. <sup>372</sup>.

OCT has been used to image meibomian glands and may be more sensitive than traditional meibography techniques <sup>513</sup>. *In vivo* confocal microscopy can observe meibomian gland orifice tissues at a cellular level, enabling evaluation of acinar density, acinar diameter, enlarged meibomian gland orifice and conjunctival inflammatory cell density <sup>514</sup>; it can also allow assessment of glandular atrophy and peri-glandular fibrosis. However, the equipment is expensive, requires a learning curve to obtain good images, and the technique requires contact with the epithelium, posing disadvantages with respect to invasiveness and

prolonged examination burden. The technique is unable to permit visualization of the glands themselves in the human eyelid margin due to light attenuation at that tissue depth; the structures imaged are rete ridges located at the dermal–epidermal junction, with alterations believed to indicate a shift of the mucocutaneous junction <sup>515</sup>. Clinical use remains limited <sup>331, 494, 514, 516</sup>. Lipid quantification at the eyelid margins and biochemical analysis of gland secretions for lipid components and markers are still being researched (see Section 3.5.4.4.2).<sup>494</sup>

The inner border of the eyelid margin plays a crucial role in helping maintain ocular surface integrity by ensuring even spread of the thin tear film with each blink. Normally, the eyelid margin features a convex inner border with a keratinized epidermis, which ends abruptly behind the posterior margin of the meibomian orifices. This is followed by the mucocutaneous junction, creating a transition zone between the wet, non-keratinized conjunctival tissue of the ocular surface and the dry, keratinized tissue of the evelid margin <sup>517</sup>. This area, known as the lid wiper, comes into contact with the ocular surface to distribute the tear film and any morphological changes at this site may be associated with tear film instability and early signs of DED <sup>266, 518</sup>. The lid wiper extends from the mucocutaneous junction to the sub tarsal fold sagitally and from the medial punctum to the lateral canthus horizontally. Posterior migration of the mucocutaneous junction leads to lid-margin keratinization <sup>519</sup>. Mechanical factors between the eyelid and the ocular surface, contribute to diseases perceived to be friction-related (in the form of sheer forces), including superior limbic keratoconjunctivitis, lid wiper epitheliopathy and conjunctivochalasis <sup>520</sup>. Damage to the epithelium of the marginal conjunctiva at the lid wiper zone is a clinical sign indicative of DED <sup>271</sup> and its staining and grading is covered in Section 3.5.4.3.

Factors such as aging, inflammation, hormonal imbalance, bacterial growth, eye drops, and oral medications can induce hyperkeratinization of the meibomian gland ductal epithelium, altering meibum transparency and viscosity <sup>494</sup>. These changes are hypothesized to lead to gland obstruction and reduced secretion.

Keratinization of the lid margin can also result from long-term rigid contact lens use or severe systemic conditions such as Stevens-Johnson syndrome/toxic epidermal necrolysis. These conditions lead to loss of the mucocutaneous junction barrier, epidermalization, whitish keratin deposits over the lid wiper zone, dyskeratosis, T-cell and neutrophil infiltration and altered local microbiome.(Muntz et al., 2020, Singh et al., 2021)<sup>521, 522</sup>. Keratinization can also occur with androgen insensitivity <sup>523</sup> and androgen deficiency <sup>47, 123</sup>, Staphylococcus aureus over-colonization <sup>524</sup>, estradiol increases of cornulin <sup>525</sup>, stearoyl-CoA desaturase-1 deficiency and related upregulation of ceramides due to altered fatty acid metabolism <sup>526</sup>, hyperlipidemia <sup>527</sup> and isotretinoin <sup>528</sup>.



**Figure 7:** A) blocked meibomian glands; B) 'toothpaste' like meibum; C) upper lid meibography of an eye with meibomian gland truncation.

### 3.5.3.2.3 Ocular rosacea

Acne rosacea has long been recognized as an inflammatory disease resulting from a complex interaction of abnormalities of the innate and adaptive immune system,

accompanied by mast cell dysfunction and / or neurovascular compromise. However, the exact mechanisms and roles of these different components of the pathophysiology remain incompletely elucidated <sup>529</sup>. Acne rosacea is currently diagnosed <sup>530</sup> based on the presence of at least one 'diagnostic phenotype' (centro-facial erythema with periodic intensification or phymatous changes) or at least two 'major phenotypes' (papules and pustules, flushing, telangiectasia, or ocular rosacea); however, ocular rosacea is not well defined, with a list of features proposed from blepharitis and conjunctival injection as indicative of mild to moderate disease, to punctate keratitis, infiltrates, vascularization and scleritis indicating moderate to severe disease <sup>530</sup>. A recent review identified ten typical ocular signs and nine diagnostic steps for recognising ocular rosacea, but many of these overlap with other posterior blepharitic conditions, with the main differentiating features being concurrent signs of rosacea on the skin, recurrent hordeola/chalazia, corneal vascularization, corneal infiltrates/ulcers and anterior uveitis <sup>531</sup>.

#### 3.5.4 Ocular Surface Abnormalities

#### 3.5.4.1 Anatomical misalignment

Ultraviolet radiation and other chronic environmental exposures can cause changes in the corneal and conjunctival cells, leading to disruption of the smooth ocular surface, for example pterygia, <sup>532, 533</sup> pinguecula, <sup>534</sup>. LIPCOF (see section 3.1.4.6) <sup>392, 535</sup> and conjunctivochalasis <sup>535</sup>. Any such raised structures can affect the flow of the tear film, the position/function of the glands and the conformity between the eyelids and the ocular surface, reducing tear film stability and altering tear volume distribution <sup>536</sup>.

LIPCOF are undulations in the inferior bulbar conjunctiva, parallel to the margin of the lower lid. LIPCOF may be observed as the initial signs of conjunctivochalasis (possibly having the same aetiology),<sup>537</sup> but exhibit a smaller cross-sectional area <sup>392, 538</sup>, do not occur centrally <sup>539-541</sup> and have no apparent relationship with age.<sup>539</sup> LIPCOF have a moderately high predictive ability for differentiating symptomatic eyes with poor tear stability <sup>540, 542, 543</sup>, but not in all studies <sup>544</sup>. Independent groups have shown that OCT can clearly resolve LIPCOF morphology, such as LIPCOF area, that correlates well with subjective grading <sup>545, 546</sup>. While other signs are associated with DED, such as ocular/conjunctival redness, epithelial thickness <sup>547</sup>, corneal nerve damage, inflammatory cell migration into the cornea, a loss of corneal sensitivity, changes to the meibomian glands (morphology and expressibility) and blinking, there is not strong evidence for the role of each in the overall diagnosis of DED. These other tests are covered with respect to their role in determining the etiological drivers of DED (section 3.5).

#### 3.5.4.2 Neural dysfunction

Corneal neuropathic pain is complex with a pragmatic and systemised approach needed for management <sup>548</sup> Abnormal corneal sensitivity has been associated with signs and symptoms in individuals with ocular surface disease <sup>549, 550</sup>. The corneal nerves serve both sensing and nutritional functions. The sensing function is not only linked to the blink reflex, but also to tear secretion by the lacrimal gland. Corneal sensitivity is a measure of corneal nerve function and an indicator of the integrity of the protective mechanisms of the ocular surface <sup>551</sup>. Morphological and anatomical features can be directly observed by in vivo confocal imaging and objectively graded using software 552 553, whereas function is assessed with the aid of a contact or non-contact corneal esthesiometer to assess corneal sensitivity. Lid margin sensitivity determined by non-contact esthesiometry has been demonstrated to be strongly correlated to corneal sensitivity; <sup>550</sup> lid margin sensitivity thresholds exhibited marginally higher predictive performance than corneal sensitivity for clinical signs of DED, as defined by the TFOS DEWS II criteria, and were significantly correlated with non-invasive tear film breakup time, corneal, conjunctival and lid wiper staining.<sup>550</sup> Liquid jet esthesiometers have also been developed and corneal sensitivity to cold found to be related to digital eye strain.<sup>554</sup> The corneal sub-basal nerves may be evaluated in detail using non-invasive in vivo confocal microscopy 555. Corneal sub-basal

#### Journal Pre-proof

nerve plexus density and length tend to decrease and the tortuosity increase, whereas the loss of corneal endothelium is accelerated in patients with DED, indicating damaged nerve fibres (Figure 8)<sup>556-558</sup>. Moreover, the damage to the nerves in DED may prevent the nervous system from exerting its immunomodulatory role, leading to changes in corneal sensitivity <sup>559</sup>. Although many studies have shown that the number and density of the sub-basal nerves in patients with DED decreases significantly and strongly correlates with the decrease in corneal sensitivity <sup>560-563</sup>, some studies show no relationship, perhaps due to variations in DED subtypes and severity of disease being examined <sup>564</sup>. Some studies have also found that different clinical presentations of DED show corresponding corneal sensitivity changes.<sup>565</sup>

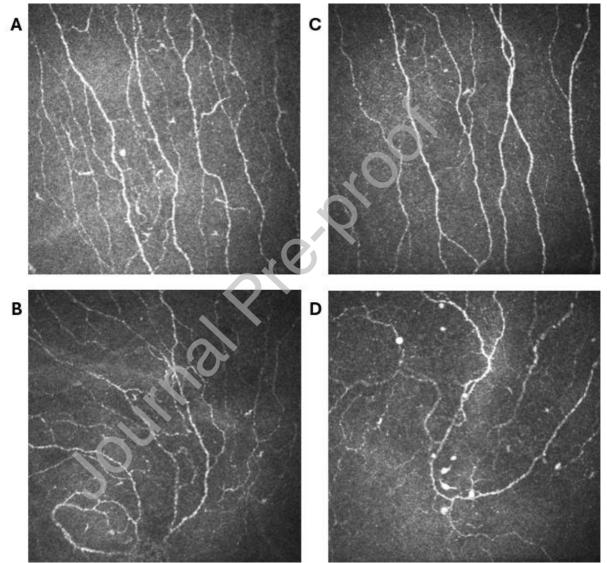


Figure 8: Corneal nerves observed by in vivo confocal microscopy (IVCM).(A) central corneal and (B) inferior whorl nerves in a health eye.(C) central corneal and (D) inferior whorl nerves in an eye with dry eye disease.

Severe neuropathic corneal pain, as an abnormality of corneal sensitivity with extreme effects, has attracted significant attention from researchers aiming to more fully understand its underlying nerve abnormalities (see TFOS DEWS III Digest pain and sensation section).<sup>4</sup> The decrease in corneal nerve density in neuropathic corneal pain is consistent with other types of DED. However, the relationship between microneuromas and nerve beading (Figure 9) with corneal nerve pain is still unclear, and further studies are required to confirm <sup>566, 567</sup>.

# Journal Pre-proof

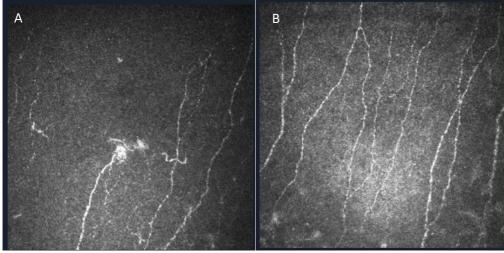


Figure 9 :(A) microneuromas in dry eye.(B) nerve beading in dry eye

decrease in all DEDKheirkhah et al. 2017 558N=20 DED N=13 controlretrospective longitudinalendothelial cell density and nerves lower in DEDLevy et al. 2017 559N=30 Sjögren [N=15 control]prospective, non- randomized treatment studye domothelial cyclosporin A 0.05% increased corneal sub- basal nerve density (only)Cardigos et al. 2019 556N=116 with non- Sjögren DED N=20 controlcross-sectional Sigoren DEDcross-sectional Scipren DEDCardigos et al. 2019 556N=116 with non- Sigoren DED N=20 controlcross-sectional Scipren DED N=20 controlcross-sectional Schirmer test score and tear breakup time				<u>s</u>
2017555obesityreproducibility for all corneal nerve parametersTepelus et al. 2017N=24 DED, N=44 Sjögren N=10 controlcross-sectional cross-sectional N=10 controldensity of nerve fibres decreased, primarily in Sjögren diseaseKheirkhah et al. 2017N=20 DED N=13 controlretrospective longitudinalendothelial cell density and nerve slower in DED loss is greater than literature reproducibility for all corneal sub- basal nerve density (only)Levy et al. 2017 559N=30 Sjögren [N=15 control]prospective, non- randomized treatment study6 months topical cyclosporin A 0.05% increased corneal sub- basal nerve density (only)Cardigos et al. 2019N=116 with non- Sjögren DED N=20 controlcross-sectional cross-sectional Sjögren DED N=20 controlcross-sectional cross-sectional Sjögren DED N=20 controlCardigos et al. 2019N=116 with non- Sjögren DED N=20 controlcross-sectional cross-sectional Sjögren DED N=20 controlcross-sectional cross-sectional Schirmer test score and tear breakup time	Study	Cohort	Design	Results
Tepelus et al. 2017N=24 DED, N=44 Sjögren N=10 controlcross-sectional•density of nerve fibres decreased, primarily in Sjögren diseaseN=10 controlN=10 control•density of nerve fibres decreased, primarily in Sjögren disease•nerve tortuosity and reflectivity decrease in all DED ••Kheirkhah et al. 2017N=20 DED N=13 controlretrospective longitudinal•endothelial cell density and nerve and cell changes correlated with symptomologyLevy et al. 2017 559N=30 Sjögren [N=15 control]prospective, non- randomized treatment study•6 months topical cyclosporin A 0.05% increased corneal sub- basal nerve density only)Cardigos et al. 2019N=116 with non- Sjögren DED N=20 controlcross-sectional Sjögren DED N=20 controlcross-sectional staning corneal sub-basal nerve plexus density and length lower, and tortuosity higher in DEDCardigos et al. 2019N=116 with non- Sjögren DED N=20 controlcross-sectional scoss-sectional Sjögren DED N=20 controlcross-sectional scoss-sectional Sjögren DED N=20 control			cross-sectional	reproducibility for all corneal nerve parameters
Kheirkhah et al. 2017N=20 DED N=13 controlretrospective longitudinalendothelial cell density and nerves lower in DEDLevy et al. 2017N=30 Sjögren [N=15 control]prospective, non- randomized treatment study6 months topical cyclosporin A 0.05% increased corneal sub- basal nerve density (only)Cardigos et al. 2019N=116 with non- Sjögren DED N=20 controlcross-sectional Sjögren DED N=20 controlcross-sectional Schirmer test score and tear breakup time		N=44 Sjögren	cross-sectional	<ul> <li>density of nerve fibres decreased, primarily in Sjögren disease</li> <li>nerve tortuosity and reflectivity decrease in all DED</li> <li>density of dendritic cells higher in DED groups</li> <li>nerve and cell changes</li> </ul>
<ul> <li><sup>559</sup> [N=15 control] randomized treatment study</li> <li><sup>659</sup> [N=15 control] randomized treatment study</li> <li><sup>6005%</sup> increased corneal subbasal nerve density (only)</li> <li><sup>6005%</sup> decreased dendritic cell numbers, more so in those with more severe baseline symptom and staining</li> <li><sup>6005%</sup> corneal sensitivity increased</li> <li><sup>6005%</sup> corneal subbasal nerve plexus density and length lower, and tortuosity higher in DED</li> <li><sup>6005%</sup> corneal subbasal nerve plexus density and length lower, and tortuosity higher in DED</li> <li><sup>6005%</sup> corneal subbasal nerve plexus strongly associated with Schirmer test score and tear breakup time</li> </ul>				<ul> <li>endothelial cell density and nerves lower in DED</li> <li>loss is greater than literature</li> </ul>
Cardigos et al. 2019 <sup>556</sup> N=116 with non- Sjögren DED N=20 control · cross-sectional N=20 control · corneal sub-basal nerve plexus strongly associated with Schirmer test score and tear breakup time			randomized	<ul> <li>0.05% increased corneal subbasal nerve density (only)</li> <li>decreased dendritic cell numbers, more so in those with more severe baseline symptoms and staining</li> </ul>
Ross et al. 2020 N=14 with severe cross-sectional	Cardigos et al. 2019 <sup>556</sup>	Sjögren DED	cross-sectional	<ul> <li>corneal sub-basal nerve plexus density and length lower, and tortuosity higher in DED</li> <li>corneal sub-basal nerve plexus strongly associated with Schirmer test score and tear</li> </ul>
	Ross et al. 2020	N=14 with severe	cross-sectional	sub-basal nerve density reduced

566	ocular pain >1year, 4 with neuropathic pain		<ul> <li>compared with controls</li> <li>more activated keratocytes and spindle, lateral and stump microneuromas in stroma in those with neuropathic pain</li> </ul>
Moein et al. 2020 <sup>567</sup>	N=30 DED N=25 neuropathic pain N=16 controls	retrospective, case-controlled	<ul> <li>similar lower nerve density and higher dendritic cell numbers in DED and neuropathic groups</li> <li>no difference in nerve beading</li> <li>microneuromas present only in neuropathic pain group</li> </ul>
Maity et al. 2024 <sup>568</sup>	N=28 Sjögren disease N=25 meibomian gland dysfunction	Cross-sectional	<ul> <li>similar dendritic cell density</li> <li>nerve fibre length, density and branching lower with Sjögren disease</li> <li>tear osmolarity weakly negatively correlated with corneal nerve parameters</li> </ul>

#### Table 3:

Confocal studies of corneal nerve abnormalities in DED.

Systemic diseases that are associated with DED can cause corneal hypoesthesia such as in patients with diabetes <sup>569-571</sup>. In addition to nerve reduction, a significant reduction in nerve beading frequency has also been reported, possibly due to reduced metabolomic activity in patients with diabetes <sup>572-574</sup>. Corneal nerve changes have also been observed in patients with Graves' ophthalmopathy (often associated with DED), perhaps due to nerve degeneration <sup>575</sup>. However, there appears to be no research on corneal sensitivity related to Graves' ophthalmopathy, and its correlation with structural alterations in nerves.

### 3.5.4.3 Ocular surface cellular damage / disruption

Punctate staining of the cornea and bulbar conjunctiva observed following the application of dyes, such as sodium fluorescein (Figure 10), rose bengal, and lissamine green, is a key diagnostic marker of numerous anterior segment conditions, including DED <sup>238</sup>. The distribution of punctate staining may provide an indication of potential aetiology <sup>238, 576</sup>, and DED is traditionally thought to be predominantly associated with interpalpebral or inferior staining <sup>238</sup>. In recent years, there has been growing interest in the utility of eyelid marginal conjunctival staining or lid wiper epitheliopathy in the diagnosis of DED <sup>1, 577</sup>.

To facilitate standardised recording of the severity of ocular surface staining and lid wiper epitheliopathy (Section 3.5.4.3), several grading systems have been devised, and the most commonly used are summarised in Table 4 <sup>578</sup>. Corneal and conjunctival staining grading systems include the van Bijsterveld system <sup>579</sup>, the National Eye Institute (NEI) Industry Workshop guidelines <sup>19</sup>, the Collaborative Longitudinal Evaluation of Keratoconus (CLEK) schema <sup>580</sup>, the Oxford scheme <sup>581</sup>, the Lexitas grading system <sup>582</sup>, the area–density combination index <sup>583</sup>, and the Sjögren's International Collaborative Clinical Alliance (SICCA) ocular staining score <sup>584</sup>. A recent study modified the Oxford scheme to incorporate half-unit increments for the assessment of corneal staining and reported improved sensitivity and repeatability <sup>275</sup>. The corneal and conjunctival staining component of the global consensus TFOS DEWS II diagnostic criteria was based on the SICCA grading system <sup>1</sup>. Lid wiper epitheliopathy is most commonly assessed relative to Korb's grading system, which combines the horizontal length of staining in mm, as well as the sagittal width relative to the eyelid margin <sup>266</sup>. The eyelid margin staining component of the global consensus TFOS DEWS II diagnostic criteria was based on this grading system.<sup>1</sup>

Key diagnostic accuracy studies evaluating the discriminative performance of ocular surface staining and lid wiper epitheliopathy in the detection of DED are summarised in Table 5. Overall, lid wiper epitheliopathy (C-statistic range, 0.69 to 0.80) demonstrated superior discriminative ability relative to corneal (C-statistic range, 0.52 to 0.57) and conjunctival staining (C-statistic range, 0.51 to 0.63) <sup>222, 248, 250, 540, 585</sup>, which would support the incorporation of all three staining parameters in the routine diagnostic workup of DED <sup>1</sup>. The reasons underlying the greater diagnostic performance of lid wiper epitheliopathy are not completely understood. Previous studies have reported that corneal and conjunctival staining more commonly present in patients with severe DED <sup>248, 250</sup> and demonstrate poorer correlation with other dry eye signs and symptoms in mild-to-moderate disease <sup>248, 250</sup>. Moreover, a number of epidemiological and diagnostic studies have suggested that lid wiper epitheliopathy might be an earlier clinical sign than corneal and conjunctival staining in the natural history of DED <sup>249, 586</sup> and it remains uncertain whether the greater exposure to shearing and viscosity-induced hydrodynamic forces during the blink cycle might predispose the lid wiper region to earlier damage <sup>537</sup>.

Grading system	Details
van Bijsterveld staining score 579	Corneal staining scoring: 1: sparsely scattered spots 2: densely scattered spots 3: confluent spots Conjunctival staining scoring: Divided into nasal and temporal zones. 1: few separated spots 2: many separated spots 3: confluent spots
NEI staining score <sup>19</sup>	Corneal staining scoring: Divided into five sectors (central, superior, inferior, nasal, and temporal), each scored from 0 to 3. Conjunctival staining scoring: Divided into superior paralimbal, inferior paralimbal, and peripheral area, both nasally and temporally, each scored 0 to 3.
CLEK staining score	Cornea staining scoring: Divided into five sectors (central, superior, inferior, nasal, and temporal), each scored 0 to 4 in 0.5 increments. Conjunctival scoring: Divided into four sectors (superior, inferior, nasal, and temporal), each scored 0 to 4 in 0.5 increments. Intra-class correlation coefficient <sup>587</sup> :
	Fluorescein: 0.76 Rose bengal: 0.40
Oxford staining score	Corneal staining scoring:

**Table 4:** Commonly used grading systems for ocular surface staining and lid wiper epitheliopathy.

581	Whole corneal area scored from 0 to 5 dependent on the intensity of punctate staining displayed pictorially, with the intensity of dots increasing on a logarithmic scale between grades.
	Conjunctival staining scoring: Whole conjunctival area scored from 0 to 5 dependent on the intensity of punctate staining displayed pictorially, with the intensity of dots increasing on a logarithmic scale between grades.
Korb grade <sup>266</sup>	Lid wiper epitheliopathy horizontal length grading: 0: <2 mm 1: 2 to 4 mm 2: 5 to 9 mm 3: ≥10mm
	Lid wiper epitheliopathy sagittal width grading: 0: <25% of the lid wiper 1: 25% to <50% of the lid wiper 2: 50% to <75% of the lid wiper 3: ≥75% of the lid wiper
Area–density combination index <sup>583</sup>	Corneal staining area scoring: A0: no punctate staining A1: $>\frac{1}{3}$ A2: $\frac{1}{3}$ to $\frac{2}{3}$ A3: $>\frac{2}{3}$
	Corneal staining density scoring: D0: no punctate staining D1: sparse D2: moderate D3: high with lesion overlap
SICCA staining score	Corneal fluorescein staining scoring: 0: 0 dots 1: 1 to 5 dots 2: 6 to 30 dots 3: >30 dots Extra points for confluent patches, staining within the pupil or filaments
	Conjunctival lissamine green staining scoring: Divided into nasal and temporal zones 0: 0 to 9 dots 1: 10 to 32 dots 2: 33 to 100 dots 3: >100 dots
	Intra-class correlation coefficient: 0.90 to 0.91 588

**Table 5:** Key diagnostic accuracy studies assessing the discriminatory performance of ocular surface staining and lid wiper epitheliopathy in detecting dry eye disease.

Study	Methods	Outcomes
Lemp et	Sample size: 314	Corneal staining:
al. 2011		C-statistic: 0.77
222	Index tests:	Sensitivity: 54%
	Corneal staining (NEI score)	Specificity: 89%
	Conjunctival staining (NEI score)	
		Conjunctival staining:
	Reference standard:	C-statistic: 0.88
	Composite disease severity index, derived	Sensitivity: 60%
	from the TFOS DEWS severity scale (clinical signs only).	Specificity: 91%
	Incorporation bias:	
	Ocular surface staining formed part of both	
	index testing and reference standard.	<u> </u>
Pult et	Sample size: 47	Corneal staining:
al. <sup>540</sup>	(	C-statistic: 0.52
	Index tests:	Sensitivity: not reported
	Corneal staining with fluorescein (CCLRU scale)	Specificity: not reported
	Conjunctival staining with lissamine green	Conjunctival staining:
	(CCLRU scale)	C-statistic: 0.51
	Lid wiper epitheliopathy with lissamine green	Sensitivity: not reported
	(Korb grading)	Specificity: not reported
	Reference standard:	Lid wiper epitheliopathy:
	OSDI score (clinical symptoms only).	C-statistic: 0.75
		Sensitivity: 48%
	Incorporation bias:	Specificity: 96%
	None.	opcomony. co/c
Wang	Sample size: 552	Corneal fluorescein
et al.		staining:
2019 <sup>248</sup>	Index tests:	C-statistic: 0.56
	Corneal fluorescein staining (Oxford score)	Sensitivity: 25%
	Conjunctival lissamine green staining (Oxford	Specificity: 86%
	score)	
	Superior lid wiper epitheliopathy (lissamine	Conjunctival lissamine
		•
	green) (Korb grading)	green staining:
	green) (Korb grading) Inferior lid wiper epitheliopathy (lissamine	green staining: C-statistic: 0.52
	Inferior lid wiper epitheliopathy (lissamine	C-statistic: 0.52
		•
	Inferior lid wiper epitheliopathy (lissamine	C-statistic: 0.52 Sensitivity: 11%
	Inferior lid wiper epitheliopathy (lissamine green) (Korb grading)	C-statistic: 0.52 Sensitivity: 11%
	Inferior lid wiper epitheliopathy (lissamine green) (Korb grading) Reference standard: TFOS DEWS II criteria (excluding staining	C-statistic: 0.52 Sensitivity: 11% Specificity: 94% Superior lid wiper
	Inferior lid wiper epitheliopathy (lissamine green) (Korb grading) Reference standard:	C-statistic: 0.52 Sensitivity: 11% Specificity: 94%
	Inferior lid wiper epitheliopathy (lissamine green) (Korb grading) Reference standard: TFOS DEWS II criteria (excluding staining	C-statistic: 0.52 Sensitivity: 11% Specificity: 94% Superior lid wiper epitheliopathy:

Wang Sample size: 2,066	
	Corneal fluorescein staining:
	C-statistic: 0.57
÷ · · · ·	Sensitivity: 38% Specificity: 76%
,	Conjunctival lissamine
	green staining:
5 1	C-statistic: 0.63
	Sensitivity: 58%
	Specificity: 64%
Reference standard: TFOS DEWS II criteria (excluding staining	Superior lid wiper
	epitheliopathy:
	C-statistic: 0.72
Incorporation bias:	Sensitivity: 72%
None.	Specificity: 66%
	Inferior lid wiper epitheliopathy:
	C-statistic: 0.71
	Sensitivity: 77%
	Specificity: 60%
	Fluorescein lid wiper epitheliopathy:
	C-statistic: 0.80
	Sensitivity: 44%
	Specificity: 93%
Rose bengal lid wiper epitheliopathy (Korb	
	Rose bengal lid wiper
	epitheliopathy:
(	C-statistic: 0.78
	Sensitivity: 43%
OSDI score (clinical symptoms only).	Specificity: 90%
	Lissamine green lid
	wiper epitheliopathy:
•	C-statistic: 0.76
	Sensitivity: 39%
	Specificity: 90%

# Journal Pre-proof



**Figure 10:** A) Fluorescein corneal staining illuminated with a blue light; B) Lissamine green bulbar conjunctival staining under white light; C) Lissamine green lower lid wiper staining under white light

### 3.5.4.4 Primary inflammation / oxidative stress

Inflammation plays an etiological role in the pathophysiology of DED <sup>149</sup>. Assessing the presence and intensity of inflammation is essential to determine the severity of the disease <sup>589</sup>, the risk of progression <sup>590</sup> and to inform its management. Inflammation at the ocular surface can be both a cause and a consequence of DED <sup>591</sup>. Ocular surface inflammation can occur due to ocular surface damage <sup>592</sup>, however autoimmune diseases are intrinsically significant contributors to DED <sup>593</sup>. In systemic immune-mediated conditions such as Sjögren disease, lymphocyte infiltration in the lacrimal gland can result in damage and fibrosis , resulting in reduced tear secretion and elevated inflammatory cytokine levels in tears <sup>596</sup>. Therefore, the assessment of inflammation in DED frequently includes both local and systemic investigations to help elucidate the source of inflammation. As an example, it has been reported that MMP-9-positive patients respond more favorably to topical cyclosporine than MMP-9–negative patients <sup>597</sup>, and the use of topical anti-inflammatory therapy has been associated with a reduction in HLA-DR <sup>598</sup> and MMP-9 tear levels <sup>599, 600</sup>. Treatments targeting novel inflammatory pathways in DED are continuously being explored and developed <sup>601, 602</sup>, although many to date have failed. Current diagnostic inflammatory tests have some limitations <sup>603, 604</sup>. While image-based comparative scales and noncomparative methods based purely on clinical observation remain valid assessment tools for use in the clinic, objective conjunctival redness quantification is substantially more sensitive and reliable than subjective grading <sup>605, 606</sup> and can be performed using a smartphone.<sup>607</sup> Moreover, enhancing the user-friendliness of confocal microscopy by developing a noncontact imaging technique with adequate resolution or providing a wider field of imaging would probably facilitate its adoption. In terms of molecular-based diagnostic tests, the lack of standardized methods for tear fluid collection and biomarker quantification <sup>608, 609</sup>, as well as the absence of normal reference values <sup>610</sup>, are key aspects that may limit the accuracy, reproducibility and overall implementation of these techniques. The cost and technical complexity of some of these diagnostic tests may also be an important limitation to their implementation. However, improving clinical outcomes, avoiding unnecessary therapies, and accelerating patients' recovery not only stands to benefit patients but also to save costs.

## 3.5.4.4.1 Imaging-based diagnostic tests

## 3.5.4.4.1.1 Ocular conjunctival redness

Conjunctival ocular hyperemia occurs with dilation of the microvasculature arising from a multitude of etiologies <sup>606, 611, 612</sup>. The vasodilatation of conjunctival microvessels plays a critical role in the efferent component of the immune system, providing both soluble mediators and cellular elements to the site of inflammation <sup>613</sup>. Accurate assessment of the underlying causes of conjunctival hyperemia is key in differentiating systemic causes from a localized inflammatory response <sup>614</sup>. Both descriptive <sup>615</sup> and reference image-based <sup>616-618</sup> subjective grading scales can facilitate the detection and monitoring of changes in the conjunctival microvasculature during follow-up, supporting decision-making in modifying the treatment plan. Multiple studies have shown that these scales have limited inter- and intra-observer repeatability <sup>605, 619</sup>, which prompted the development of computer-based

photograph-analysis techniques to allow objective grading of conjunctival redness allowing higher precision and repeatability <sup>605, 606, 620-627</sup>.

3.5.4.4.1.2 In vivo confocal imaging

*In vivo* confocal imaging can aid in identifying characteristic structural changes in the cornea, conjunctiva, meibomian glands and lacrimal gland in patients with DED <sup>330, 628</sup>. Conflicting findings in corneal and conjunctival epithelial cell changes in DED studies may result from DED disrupting cell renewal, but simultaneously promoting cell repair at the same time, affecting the apoptosis-proliferation balance <sup>629</sup>.

Previous studies have reported characteristic changes in the corneal stroma of patients with DED such as a significant increase in anterior corneal stromal keratocyte density <sup>630</sup> and abnormal stromal hyper-reflectivity indicating increased activity <sup>631</sup>. An increase in dendritic cell density has been reported in patients with DED <sup>630, 632, 633</sup>. Mature and immature dendritic cells have been found in the corneal stroma of these patients <sup>634</sup>. Interestingly, increased dendritic cell density has been correlated with severe symptoms <sup>557</sup> as well as with aqueous deficient DED due to immune disease <sup>635</sup>. Dendritic cell and activated keratocyte density, as well as reduced corneal sub-basal nerve fibre length have shown an indirect association with inflammation on the ocular surface, through a significant reduction following treatment with topical corticosteroids <sup>636, 637</sup>. Similarly, long-term therapy with topical cyclosporine for more than 6 months has shown a positive impact on corneal epithelial, stromal, dendritic, and nerve confocal imaging parameters <sup>559, 638</sup> (see TFOS DEWS III Digest Pathophysiology section).<sup>4</sup>

# 3.5.4.4.2 Tear Biomarker diagnostic tests

### 3.5.4.4.2.1 Matrix metalloproteinases

Matrix metalloproteinases (MMPs) are a family of enzymes that are core to several ocular and systemic inflammatory processes <sup>639, 640</sup>. MMPs are generated by connective tissues and pro-inflammatory cells <sup>641</sup>, and can be detected in tears of patients with DED <sup>642, 643</sup>. In DED, corneal epithelial damage can result in a local inflammatory reaction that leads to increased secretion of MMPs <sup>591</sup>. MMP-1, -3, -9, -10, and -13 are the MMPs most notably elevated at the corneal surface, splitting epithelial basement membrane components and tight junction proteins (such as ZO-1 and occludin) that maintain corneal epithelial barrier function <sup>643, 644</sup>. Studies have reported a significant correlation between MMP-9 degree of positivity and ocular surface fluorescein staining <sup>228, 597</sup>. However, tear volume has an impact on the assay indicator, and therefore, a MMP-9 test degree of positivity may not correlate as strongly with MMP-9 tear concentration in cases of either low tear volume or reflex tearing <sup>645</sup> 3.5.4.4.2.2 Cytokines and chemokines

Measurement of cytokines and chemokines may enable differentiation of ocular surface inflammation caused through the innate immune response, and adaptive immune response (see TFOS DEWS III Digest Pathophysiology section).<sup>4</sup> Further, within the adaptive immune response, these markers may separate into Th1, Th2, Th17, and Treg-mediated responses <sup>646</sup>. IFN-γ, the dominant cytokine associated with the Th1 response, has been associated with goblet cell loss and squamous epithelial hyperplasia. Consequently, some clinical studies demonstrated correlation between higher IFN-y and tear deficiency, though other studies found contradictory results <sup>647-649</sup>, possibly due to inherent differences in assay methods, or population characteristics. IFN-y may be specifically associated with an increase in osmolarity <sup>650</sup>. Th17-mediated ocular inflammation may be induced through IL-17 signaling. IL-17 activates MMP-9 which contributes to damage to the corneal epithelial barrier <sup>646, 651</sup>. A significant correlation between both corneal and conjunctival staining scores and presence of Th cells has been reported, although Th subtypes such as Th1 and Th17, were not detectable at high enough levels for establishing correlations with tear film stability and volume in patients with DED<sup>652</sup>; in addition, no correlation was found between DED and the detection of IL-1B, II-6, IL-8, IL-10, IL-17A, IFNy, and tumor necrosis factor alpha in tears <sup>653</sup>, although cytokine upregulation has been detected in patients with Sjögren disease <sup>654</sup>. However higher MMP levels are found in patients with DED <sup>655</sup>, likely from episodic flares <sup>656</sup> and MMPs have been found to correlate with osmolarity and tear volume, more strongly than with tear stability and symptoms <sup>657</sup>. The variable nature of cytokine levels and DED corresponds with the heterogeneous nature of ocular surface inflammation <sup>658-660</sup>, changes over time <sup>661</sup> and the location of sampling <sup>662</sup>. This confirms that inflammation is more often downstream (a consequence) rather than intrinsic (a driver) in DED (see TFOS DEWS III Digest Pathophysiology section).<sup>4</sup>

3.5.4.4.2.3 Neurotrophic Factors and Neuropeptides

Neuropeptides and neurotrophic factors have a role in mediating sensory information and in regulating aspects of neuronal function and cell survival. <sup>649</sup>.

Serotonin, which is a peripheral nerve sensitizer, is found at a higher concentration in tears of patients with DED than in those of normal eyes and correlates with symptoms <sup>663</sup>. Serotonin is activated by inflammation and sensitizes peripheral nerves, perhaps playing a role in the development of corneal hypersensitization in DED <sup>664</sup>. Increased nerve growth factor has a protective role in DED, improving the integrity of the epithelial cell layer and tear secretion <sup>665, 666</sup>. Lacrimal gland dysfunction has been associated with decreased calcitonin gene-related peptide <sup>665, 666</sup>. Substance P has been found to be raised in tears of DED patients after refractive surgery <sup>667</sup> and nerve growth factor has also been found to be raised in tear fluid of patients with neuropathic pain <sup>668</sup>. Hence these neurotrophic factors and neuropeptides appear to regulate tear aqueous production such that deficiencies in an individual with DED may indicate a strategy for improving tear secretion.

3.5.4.4.2.4 Ocular surface immune markers

Major histocompatibility complex based markers have long been identified as a risk factor for DED, particularly with Sjögren disease <sup>669, 670</sup>. Antigen presentation through the major histocompatibility process seems to play an intermediary role in T-cell activation and the cytokine-based inflammatory cascade <sup>646</sup>. While both major histocompatibility complex class I and class II have been connected to inflammation in DED, HLA-DR, a member of the class II family, has been more thoroughly investigated <sup>671, 672</sup>. A gene expression analysis of mRNA transcripts observed from conjunctival impression cytology sampling discovered a correlation between HLA-DR, CD40, and IFN. This connection further links inflammatory DED with T cell activation <sup>671</sup>. Conjunctival impression cytology samples collected on a heterogenous group of patients with DED revealed that the percentage total cells expressing HLA-DR was positively correlated with conjunctival and corneal staining scores <sup>672</sup> and weakly with tear volume <sup>673</sup>; although HLA-DR percentage analysis was not a sensitive diagnostic marker for DED in itself, it may represent a means of helping identifying specific dry eye subtypes based on the lymphocytic response responsible for the ocular surface inflammation in a particular patient, and guiding therapeutic decisions. Neutrophils, macrophages, mast cells, T-cells and dendritic cells have been found to increase in DED across several studies, particularly in more severe levels of dry eye found in autoimmune disease such as Sjögren disease and graft versus-host disease. <sup>674</sup>,

3.5.4.4.2.5 Inflammasome markers

The inflammasomes are innate immune system sensors that induce an inflammatory form of cell death, known as pyroptosis, in response to harmful stimuli such as pathogens or oxidative stress, among others <sup>675, 676</sup> (see TFOS DEWS III Digest Pathophysiology section).<sup>4</sup> Reactive oxygen species are involved in the pathogenesis of DED <sup>677</sup>, and have been suggested as a priming signal for inflammasome activation <sup>602</sup>. NOD-like receptor protein-3 (NLRP3) inflammasome, a key driver in the innate immune system, has a role in DED pathogenesis <sup>677, 678</sup>, is upregulated in the tear film of people with Sjögren disease <sup>678</sup> and is activated by hyperosmolarity <sup>677, 679</sup>. Tear levels of caspase-1, a molecule involved in the inflammasome cascade, and various clinical signs of ocular surface damage in patients with DED and patients using topical glaucoma medications have been found to be correlated <sup>680</sup>. Moreover, tear levels of Gasdermin-D, a pyroptosis executor, are also elevated in patients with DED <sup>681</sup>

### 3.5.4.4.2.6 MicroRNAs

MicroRNAs are non-coding RNAs that serve as significant regulators in a variety of molecular pathways <sup>682</sup> (see TFOS DEWS III Digest pain and sensation section).<sup>4</sup> Several

studies have identified tear microRNAs as potential biomarkers for ocular diseases, including Sjögren disease <sup>683</sup> and DED <sup>684-686</sup>, among others <sup>608, 687</sup>. Nine tear microRNAs (miR-127-5p, miR-1273h-3p, miR-1288-5p, miR-130b-5p, miR-139-3p, miR-1910-5p, miR-203b-5p, miR-22-5p, and miR-4632-3p) associated with inflammation have been found to be upregulated in the tears of patients with DED.

3.5.4.4.2.7 Oxidative stress markers

Oxidative stress, an imbalance of free radicals and antioxidants that leads to cell damage, may play a role in the pathogenesis of DED <sup>688</sup>. Proteomic analysis of tears from patients with DED shows an upregulation of proteins associated with oxidative stress injury <sup>689</sup>. It is well established that oxidative damage triggers an inflammatory response, resulting in ocular surface dysfunction <sup>690-692</sup>. Moreover, oxidative stress may cause the progression of DED by exacerbating inflammation by triggering the vicious circle of DED. Oxidative stress biomarkers, which indicate the degree of oxidative stress, have been found elevated in the tears and conjunctiva of patients with DED <sup>693</sup>. The detection of oxidative stress biomarkers through tear film or conjunctival impression cytology samples may be undertaken to evaluate DED status, monitor the efficacy of drugs or evaluate disease progression. Oxidative markers such as lactoferrin (tears), peroxiredoxin 2 (tears), SOD (tears), CAT (tears), and GSH-Px (tears) are downregulated in DED 689, 694. In contrast, markers such as S100A8 (tears), S100A9 (tears), reactive oxygen species (conjunctiva), LPO (conjunctiva), 4-HNE (conjunctiva), MDA (conjunctiva) and HEL (tears) are upregulated in these patients 692, 695-698. A correlation between ocular surface oxidative stress markers and topical treatments has been described by several authors 696, 699, 700, indicating the utility of these markers in monitoring response to anti-oxidant therapies.

3.5.4.4.2.8 Serum markers

DED is associated with chronic inflammatory systemic conditions including, collagen vascular diseases <sup>701</sup>, rheumatoid arthritis <sup>702</sup>, or Sjögren disease <sup>703</sup>, among others <sup>704</sup>. Acute phase reactants such as erythrocyte sedimentation rate and C-reactive protein indicate active systemic inflammation <sup>705</sup>. However, previous studies found that levels of these reactants do not correlate with ocular surface symptoms or tear parameters in DED <sup>701, 706</sup>, although serum inflammatory markers PM-Scl100 and Sm were associated with more severe DED symptoms, while inflammatory markers U2SnRNP A', Ro52, La, DNA, and Ro60 were associated with more severe ocular surface disease signs <sup>707, 708</sup>. Other serum inflammatory-related markers such as antinuclear antibody and IL-2 receptor(sIL-2R), or anti-double-strand DNA antibody, have been associated with DED in primary Sjögren disease <sup>709, 710</sup> or systemic lupus erythematosus <sup>701</sup>. Similarly, serum levels of IL-17, a proinflammatory cytokine <sup>711</sup>, are significantly increased in patients with DED and high fluorescein staining scores <sup>712</sup>. S100A8/A9 and granulysin have been found to be higher in more severe Stevens-Johnson Syndrome <sup>713</sup>. In addition, in a large population-based study, decreased serum androgens were found to be highly associated with DED diagnosis and symptoms <sup>714</sup>.

 Table 6:
 Subclassification of DED etiological drivers recommended tests and cut-offs [where available]

		Standard testing	Advanced testing	
	Lipid	Interferometry – grade ≤ 3 (non-amorphous or colored pattern) or <72nm on LipiView <sup>197, 320, 343</sup>		
		Meibum expressibility/quality – meibum not clear or limited expressibility <sup>332, 371, 715</sup> .		
Tear Film	Aqueous	Meniscometry - tear meniscus height ≤0.20mm <sup>39, 320, 380</sup> .	Strip meniscometry - $\leq$ 2.5mm wetting length <sup>404-406</sup> .	
Deficiencies	Aqueous		Tear proteins and other chemical components testing	
	Mucin / glycocalyx	Rose bengal or lissamine green staining - >9 punctate spots <sup>581</sup>	Immunohistochemistry and immunoelectron microscopy o tear film	
	Muchin' grycocalyx		Impression cytology – goblet cell density and epithelial cel morphology	
	Blink / lid closure	Partial blinking observation - >40% occurrence 463		
	Blink / lid closure	Lagophthalmos / inadequate lid seal - observed		
		Anterior blepharitis observation		
Eyelid Anomalies	Lid margin	MGD Meibography – gland length <75% <sup>320,</sup>		
		Gland plugging - observed <sup>715</sup> Telangiectasia - observed <sup>715</sup> Gland expressibility		
		Keratinization Slit-lamp biomicroscopy		
		Ocular rosacea Slit-lamp biomicroscopy	-	
	Anatomical misalignment	Slit-lamp biomicroscopy	Corneal topography	
Ocular Surface Abnormalities	Neural dysfunction	Puff or physical sensation - corneal and lid margin sensitivity thresholds ≥0.8 mbar <sup>550</sup> although instruments are not comparable <sup>716</sup>	<i>In vivo</i> confocal microscopy – normative values available for nerve length, branch and density metrics <sup>717</sup>	
	Ocular surface cellular damage / disruption	Corneal fluorescein staining - >5 punctate spots <sup>250, 581</sup> Conjunctival lissamine green staining - >9 punctate spots <sup>250, 581</sup> Lid wiper staining - >2mm length and 25% width <sup>250</sup>		
	Primary inflammation /	Bulbar conjunctival hyperaemia $->1.5$ Efron scale or	In vivo confocal microscopy	
	oxidative stress	>0.95 objective JENVIS <sup>604</sup>	Tear film and ocular surface molecular testing	
Systemic Drivers		Check for systemic conditions		

52

## 3.5.5 Systemic diseases leading to dry eye

Many systemic diseases significantly contribute to DED through inflammation, autoimmunity, metabolic dysregulation and ocular surface exposure. Interdisciplinary collaboration in the management of DED patients with underlying systemic conditions is important, such as coordinating care with rheumatologists, endocrinologists, or other relevant specialists to optimize both ocular and systemic outcomes.

#### 3.5.5.1 Autoimmune conditions

Systemic inflammatory diseases such as rheumatoid arthritis, systemic lupus erythematosus, Stevens-Johnson syndrome and Sjögren disease are closely associated with DED. Chronic inflammation leads to the infiltration of immune cells in the lacrimal gland, reducing tear production and altering tear film composition <sup>718</sup>. Key inflammatory cytokines include IL-1, tumor necrosis factor-alpha and MMPs <sup>719</sup>. Patients with ankylosing spondylitis treated with tumor necrosis factor inhibitors showed improvement in clinical and laboratory disease parameters, tear production, DED severity, and impression cytology scores, suggesting tumor necrosis factor inhibitors may restore lacrimal gland acinar cells affected by proinflammatory cytokines <sup>720</sup>. In rheumatoid arthritis, the activation of the NF-κB pathway and overexpression of tumor necrosis factor alpha and IL-6 lead to systemic and ocular inflammation <sup>721</sup>. The full potential of tumor necrosis factor alpha inhibitors to reduce ocular surface inflammation and improve tear production in rheumatoid arthritis remains uncertain <sup>722, 723</sup>.

Autoimmune diseases, particularly Sjögren disease, are closely associated with DED. Sjögren disease involves lymphocytic infiltration of exocrine glands, affecting the production of aqueous tears and saliva, respectively, and leading to dry eye and dry mouth. Detection of antibodies, such as anti-Ro/SSA and anti-La/SSB, support its autoimmune nature <sup>724</sup>. Gene analysis of conjunctiva imprints revealed 53 differentially expressed genes in Sjögren disease compared to healthy controls, that indicated immune activation in patients with Sjögren disease <sup>725</sup>. Higher percentages of antigen-presenting cells and mature dendritic cells in the conjunctiva are associated with more severe keratoconjunctivitis sicca in Sjögren disease, which may contribute to goblet cell loss.<sup>726</sup>

Multidisciplinary treatment for both the underlying systemic disease as well as managing the residual ocular symptoms of DED is generally appropriate. In rheumatoid arthritis, for example, disease-modifying antirheumatic drugs such as methotrexate and biologics like tumor necrosis factor inhibitors help control systemic inflammation and improve ocular symptoms.<sup>721</sup>

### 3.5.5.2 Hormonal imbalance

Hormonal changes can trigger DED by affecting tear production and quality, especially in women, who experience hormonal fluctuations throughout their lives <sup>727</sup>. Hormonal imbalance and its management in patients with DED is covered in the TFOS DEWS III Digest.<sup>4</sup>

### 3.5.5.3 Metabolic disease

Metabolic diseases such as diabetes mellitus significantly impact the ocular surface and can lead to DED <sup>4, 7</sup>. Hyperglycemia and advanced glycation end-products contribute to microvascular damage and neuropathy, affecting the lacrimal glands and corneal nerves <sup>728</sup>. Patients with diabetes often exhibit reduced tear secretion, increased tear osmolarity, and altered corneal sensitivity <sup>729</sup>. Antioxidant therapy, along with strict glycemic control, can mitigate the effects of diabetes on the body <sup>730</sup> and the ocular surface <sup>731, 732</sup>. The impact of medications and procedures related to managing metabolic disease on the ocular surface are covered in the TFOS Lifestyle reports.<sup>7, 9</sup>

#### 3.5.5.4 Exposure

A multitude of diseases as well as trauma can cause intermittent (for example nocturnal) or constant lagophthalmos (such as facial nerve palsy), leading to exposure of the ocular surface and DED <sup>733</sup>. A number of these conditions and the impact of medications and procedures related to ocular exposure on the ocular surface are covered in the TFOS Lifestyle reports <sup>9, 12</sup>.

#### 3.6 Tests for monitoring treatment

In monitoring treatment effects over time, it is important to consider indicators that may lag (such as corneal staining <sup>249</sup>), those that can change more rapidly (such as symptomology) and the treatment's mechanism of action <sup>734</sup>. It is therefore recommended that practices adopt a standardised, reproducible and repeatable DED protocol that involves validated questionnaires, diagnostic tests and clinical exam that remains consistent from visit to visit. While every real-life patient encounter will undoubtedly not be as clear cut as this hypothetical example, clinicians are encouraged to make diagnostic and treatment decisions based on scientific evidence and on tracking subjective and objective data over time, and with the aid of clinical judgment, experience and acumen as outlined in this report.

As the number of testing options increases, the volume of clinical data that is required to be processed at each patient visit also increases and can be, at times, overwhelming. This becomes increasingly challenging as physician time becomes progressively constrained and face-to-face consultation times are shortened. With handwritten paper records, processing diagnostic data trends over long periods of time encompassing multiple office visits was challenging, time consuming and rarely done thoroughly. However, in the current electronic medical record era, the ability to seamlessly and instantaneously process, summarize, plot and chart large volumes of disparate diagnostic data over long periods of time is much easier to facilitate. DED/ocular surface disease-specific electronic medical record platforms are now available and enhance the ability to track and monitor DED symptoms, objective tests, exam findings and treatment outcomes <sup>735</sup> as well as real-world registries which also provide the facility to monitor patient progress and benchmark against peers <sup>736</sup>. Additionally, with the ubiquity of smartphones and the burgeoning emergence of smart glasses and wearable health monitors/sensors, there is unprecedented potential for gathering and analyzing real-time continuous data in-between office visits data<sup>292, 737-739</sup>. Machine learning and artificial intelligence (see section 5.1), when integrated into electronic medical record platforms and smart devices, with access to process reliable data from disparate physicians, practices and geographic locations, may possibly provide novel insights into DED that are as yet unknown, facilitating more accurate diagnoses, better treatments and more strategic clinical trial designs <sup>740</sup>. With the assistance of machine learning and artificial intelligence, historically-challenging diagnoses such as neurotrophic keratopathy and neuropathic corneal pain will likely be made earlier, with the potential to improve patient outcomes and reduce late-stage sequelae.

# 4 Patients with only symptoms or ocular surface signs

There are numerous explanations highlighted in section 3.3.3 for the, often vexing, clinical scenario of discordant signs and symptoms. Significant corneal staining in a pain-free patient, colloquially referred to as 'stain without pain', may indicate neurotrophic keratopathy.<sup>741</sup> Conversely, when the clinical signs of ocular surface dysfunction are mild or subtle in the absence of patient-reported symptoms it might indicate an early, preclinical or situational DED <sup>155</sup>. In the extreme, a clinical scenario involving significant symptomology in the absence of, or out of proportion to, clinical signs, colloquially referred to as 'pain without stain' might be indicative of neuropathic corneal pain <sup>175</sup>.

## 4.1 Ocular surface disease in the absence of symptoms

It is recommended to treat any significant ocular surface disease prior to a patient undergoing any type of eye surgery (such as laser vision correction or cataract removal), initiating contact lens wear, starting any high risk topical or systemic medications and/or any other intervention known to cause or exacerbate DED (see TFOS DEWS III Digest latrogenic section).<sup>4</sup> The surgical informed consent process should include a discussion of the risk of worsening signs and/or symptoms of DED, including visual symptoms, which may require intensive long-term treatment to control. The importance of identifying, offering education about and treating early preclinical DED/ocular surface disease is particularly critical in the setting of refractive, cataract or laser vision correction surgery where patient expectations tend to be particularly high.<sup>742</sup>

There is a high prevalence of ocular surface disease in presurgical cataract patients, many of whom have few or no reported symptoms <sup>743, 744</sup>. It is speculated that the stark disconnect between signs and symptoms in this older population is due to a combination of generational stoicism, inherent bias to focus on the perceived 'bigger' problem (for example their poor vision due to cataract) and/or age-related reductions in corneal nerve density and sensation <sup>152, 153, 165, 167, 171, 745</sup>. In the younger patient population, contact lens intolerance from DED is a common reason for seeking laser vision correction surgery,<sup>746</sup> but can also place the patient at higher risk of postsurgical complications if not identified, discussed and managed preoperatively.

Many studies have shown that DED, especially when significant corneal staining is present, can adversely affect the accuracy of preoperative measurements (such as keratometry, topography, optical pachymetry and aberrometry) potentially leading to post-surgical refractive error misses, poor and/or fluctuating visual quality (especially when multifocal or extended-depth of focus intraocular lenses are implanted) and overall lower patient satisfaction <sup>28, 747-751</sup>. Due to the high prevalence of ocular surface disease in this patient population, an algorithm has been designed by the American Society of Cataract and Refractive Society to identify and treat visually significant ocular surface disease preoperatively <sup>752</sup>.

## 4.1.1 Diagnosing Ocular Neurosensory Abnormalities

A medical history questionnaire or intake form should include often missed non-ophthalmic risk factors for neurotrophic keratopathy (diabetes, herpes, Parkinson's, multiple sclerosis, prior brain surgery such as acoustic neuroma, cerebrovascular accidents and congenital dysautonomia) and for neuropathic corneal pain (such as small fibre peripheral neuropathies, fibromyalgia, migraine, irritable bowel, anxiety, depression and post-traumatic stress disorder)<sup>753</sup>. Clinical suspicion for neurosensory abnormalities should be raised in any patient with marked discordance of signs and symptoms, risk factors and/or positive answers to targeted triaging questions. Corneal surgical incisions, laser corneal ablation, suction prior to flap creation by microkeratome or femtosecond laser, phototoxicity from the operating microscope, benzalkonium chloride-preserved postoperative drops amongst other factors can exacerbate preexisting or induce new corneal neurosensory abnormalities, potentially leading to visually significant post-operative ocular surface disease<sup>9, 33, 107, 754-761</sup>.

### 4.1.2 Diagnosing Neurotrophic Keratopathy

Neurotrophic keratopathy is considered a rare disease, but is likely underdiagnosed <sup>762</sup>. It is caused by a unilateral or bilateral abnormality of the trigeminal nerve, resulting in decreased or absent corneal sensation, and leading to diffuse corneal punctate epithelial defects (Mackie stage 1), persistent epithelial defects with characteristic smooth rolled edges (stage 2), and stromal melting with the potential for corneal perforation (stage 3) <sup>763, 764</sup>. A more nuanced and detailed 6-stage grading system has recently been proposed by a Neurotrophic Keratopathy Study Group to better refine the stages of progression and to allow practitioners

to identify NK at earlier stages <sup>741</sup>. Because early stages of neurotrophic keratopathy can involve corneal epitheliopathy with staining, increased mucous viscosity and decreased tear film stability, it is often misdiagnosed and treated as moderate-to-severe DED. The pattern of corneal staining in neurotrophic keratopathy is often diffuse involving the entire cornea in contrast to the inferior or interpalpebral staining typically seen in DED <sup>551, 765</sup>. Patients with neurotrophic keratopathy will also typically have lower blink rates and poorer blink quality <sup>762</sup> whereas patients with DED and poor blink quality, tend to blink more frequently <sup>766</sup>. Patients with neurotrophic keratopathy typically don't self-report symptoms of pain or discomfort, inconsistent with the level of corneal staining, but they may complain about reduced visual acuity, quality, stability and performance. This clinical scenario of 'stain without pain' should alert clinicians to suspect the possibility of NK as early as possible, prior to the patient failing long-term DED treatment, and, ideally well before potentially blinding stromal breakdown occurs in later stages.

Reduced or absent corneal sensitivity is suggestive of neurotrophic keratopathy and therefore corneal sensation should be assessed as soon as there is a clinical suspicion and prior to instillation of anaesthetic drops. Various methods are available for assessing corneal sensitivity. The Cochet-Bonnet esthesiometer is quantitative and makes contact with the ocular surface while the gas esthesiometers are quantitative, non-contact, and able to assess chemical, thermal and/or mechanical corneal sensitivity 716, 767-770. Both are commonly used in research and in specialty referral centres but historically have been costly and impractical for most general eye care practitioners. Cheaper and simpler methods involving contact with the eye's surface that are commonly used in practice include a wisp from a cotton-tip applicator, a corner of a disposable facial tissue and/or unwaxed and unflavored dental floss and can provide a quick qualitative assessment of corneal sensation in a primary eye care setting <sup>741</sup>. A patient with normal sensation will blink and report discomfort when the central cornea is touched, a patient with a hypoesthetic cornea may blink but not report sensation and one with an anaesthetic cornea typically will not register a blink or any sensation to the stimulus <sup>741</sup>. A modified-Delphi expert panel on neurotrophic keratopathy strongly recommended corneal sensitivity testing for persistent epithelial defects after 14 days, new painless epithelial defects, a history of herpetic eye disease or procedures that may injure the trigeminal nerve, and pain in an eye with multiple concurrent risk factors for neurotrophic keratopathy such as poorly controlled diabetes and either reduced blink rate or a history of corneal surgery <sup>771</sup>. As more modalities for non-contact, reproducible, quantifiable and inexpensive esthesiometry become available.<sup>769, 772-774</sup> corneal sensation can be performed earlier in the diagnostic subtyping process and ideally be incorporated into office-based routine DED/ocular surface disease protocols (see section 6.1).

Once clinical suspicion for neurotrophic keratopathy and reduced corneal sensation are identified, further investigation with corneal *in vivo* confocal microscopy can be diagnostically confirmatory (see sections 3.5.3.6.1.2 and 3.5.3.4). Studies have consistently demonstrated significant alterations in the corneal nerves, epithelial cells and corneal stroma in patients with neurotrophic keratopathy <sup>775-778</sup>. Of note, corneal sensation may remain relatively normal despite significant reductions (of 50 to 80%) in sub-basal nerve density accompanied by morphological changes such as increased tortuosity and beading <sup>779</sup>, and may be clinically detectable only when the nerve density drops below 1000 µm nerve length /frame <sup>780, 781</sup>. Epithelial abnormalities include enlarged and irregular cell shapes, decreased cell density and squamous metaplasia correlating with disease severity <sup>780, 781</sup>. Severe disorganization, altered keratocyte morphology and presence of hyperreflective cells can be observed in stage 3 along with increased dendritiform cell density, particularly in the central cornea, suggesting a possible inflammatory component <sup>782</sup>. Substantial reductions in sub-basal nerve density are commonly seen in patients with neurotrophic keratopathy secondary to herpetic eye disease <sup>777, 778, 781</sup>. Interestingly, patients with unilateral herpes simplex and zoster ophthalmicus may exhibit contralateral reductions in sub-basal nerve density <sup>67, 780</sup>.

## 4.2 Symptoms in the absence of ocular surface disease

Unlike nociceptive pain that involves the triggering of nociceptors from local tissue damage. neuropathic pain, which can be peripheral or central in origin, is caused by an abnormality in the somatosensory nervous system <sup>783</sup>. Diagnosing neuropathic corneal pain is primarily clinical and exclusionary and based heavily on clinical history, risk factors and the discordance in terms of the level of symptoms in the absence of corresponding clinical signs, colloquially termed 'pain without stain' 784. While DED is commonly the initial diagnosis and treatment target for many of these patients, neuropathic pain does not respond to conventional treatments in the same way, and artificial tears do not provide the same temporary symptom relief <sup>785</sup>. DED and neuropathic corneal pain can coexist, requiring treatment for both. Indeed, DED may have been the trigger, leading to neuropathic corneal pain, often in patients with comorbid chronic pain (such as migraine and fibromyalgia), psychiatric and/or mental health disorders <sup>175, 786, 787</sup>. Patients with Sjögren disease also have a higher risk of chronic ocular pain with neuropathic features <sup>788</sup>. Other risk factors and associated comorbidities of neuropathic ocular pain include DED, diabetes, sarcoidosis, small fibre neuropathies, herpetic eye disease, prior eye surgery, infection, trauma, contact lens wear and radiation keratopathy, and many of these overlap with risk factors for neuropathic keratopathy <sup>789</sup>. An association with long-COVID has also been identified <sup>790, 791</sup>. To confound the inherent diagnostic challenges even further, a subset of patients with stage 1 neurotrophic keratopathy and concomitant neuropathic corneal pain has also been reported <sup>792</sup>. If neuropathic corneal pain is suspected on the basis of clinical history, symptoms and a lack of signs on examination, a pain-specific validated questionnaire such as the Ocular Pain Assessment Survey or Neuropathic Pain Symptom Inventory, which has been modified for the eye, can be utilized to score symptoms 793, 794.

### 4.2.1 Diagnosing corneal neuropathic pain

If the patient's eyes are painful despite lubricating drops and the pain is incited by light, wind or other triggers, neuropathic corneal pain should be suspected. In such a situation, a proparacaine challenge test can be performed to aid in both diagnosis and differentiation between peripheral and central etiologies <sup>789, 795, 796</sup>. If the pain is completely ameliorated by an anaesthetic drop the patient likely suffers from peripheral or nociceptive corneal pain whereas, if the pain persists unchanged afterwards, then a central neuropathic pain mechanism is likely; if only partial relief is achieved then a mixed mechanism of peripheral and central neuropathic corneal pain is likely <sup>795</sup>. Differentiating peripheral from central neuropathic corneal pain is important as the treatment strategies differ significantly between the two <sup>797, 798</sup>. While corneal aesthesiometry is a critical test for neurotrophic keratitis it is less useful in the workup of neuropathic corneal pain as studies have revealed both higher and lower sensitivities in these patients and overall poor diagnostic correlation 152, 796, 799, 800 although more objective, non-contact techniques offer promise in identifying neuropathic pain <sup>769</sup>. Reduced density of sub-basal nerves is a finding common to both neurotrophic keratitis and neuropathic corneal pain, but the increased presence of sub-basal and stromal microneuromas, detectable by confocal microscopy, is more common in neuropathic corneal pain<sup>168, 169, 566, 567, 801</sup>. Further studies are needed to establish whether confocal microscopy can be used reliably for differentiating the etiological drivers in a patient with DED.

## 5 Future advances

## 5.1 Artificial intelligence

Artificial Intelligence, a term coined by emeritus Stanford Professor John McCarthy in 1955, was defined by him as "the science and engineering of making intelligent machines." A wide range of definitions have now been proposed, making it difficult to assess claims on its use. It has been suggested that it is already widely used in DED clinical tests and research <sup>802</sup>, but it could be argued whether an algorithm to detect a change in pixel contrast or an 'edge'

of a placido ring is truly "intelligence". Machine learning thorough training has been used to try and predict video frames that a specialist identified as showing breakup, with a sensitivity of 78% and specificity of 86% <sup>803</sup>. There is the potential that machine learning could make a clinical test used in the diagnosis of dry eye more objective, but as such a diagnosis would require a gold standard disease group against which to compare the individual, and a consensus of which tests are most suitable to identify this group based on available evidence, along with practical considerations such as cost and equipment availability, as artificial intelligence cannot in itself drive development or change in the diagnostic algorithm for dry eye. It certainly can aid in image analysis however, and particularly structural segmentation such as in meibography <sup>804-809</sup>. Rating of corneal fluorescein staining, on the other hand, remains more challenging <sup>810</sup>. Potential barriers to widespread adoption of artificial intelligence include cost, accessibility, regulatory and ethical considerations, training requirements and integration with existing diagnostic protocols <sup>811</sup>.

#### 5.2 Tear biomarker testing of tears

MMPs are one of many classes of proteases secreted into the tears in DED. Since MMPs can destroy tight junctions in the ocular surface epithelium, increased levels of MMPs reflects loss of ocular surface barrier function, <sup>812-814</sup>. MMPs are produced as inactive proenzymes and can be cleaved to become active enzymes <sup>815</sup>. It is therefore important for future MMP tests to detect enzyme activity levels and not simply total tear protein levels. MMP-9 is detected more commonly in severe DED and has been proposed as a potential means of monitoring the success of DED management; patients with a positive MMP-9 test showed a greater benefit from topical cyclosporine than those who were MMP-9-negative <sup>597</sup>. A silicon nanowire-based field-effect transistor MMP-9 tear film biosensor was found to have a sensitivity of 87% and specificity of 90% for DED <sup>816</sup>. More advanced point-of-care tear proteomic test kits for identifying DED subtype drivers are needed along with independent validation / replication studies.

Lymphotoxin-alpha (LT- $\alpha$ ), a member of the tumor necrosis factor superfamily, is expressed by T cells, B cells, and natural killer cells, playing a crucial role in immune system development and function <sup>817-819</sup>; this includes the formation of lymphoid organs, maintenance of lymphatic microenvironments, host defense, and modulation of inflammation. Despite its established association with inflammation, emerging research has identified a negative correlation between LT- $\alpha$  levels in the blood and fatigue symptoms (often linked to proinflammatory processes) in patients with primary Sjögren disease <sup>820, 821</sup>. This suggests that LT- $\alpha$ 's role in inflammation might be more complex than previously thought. Levels of multiple tear protein markers (TNF- $\alpha$ , IL-10, IL-1 $\beta$ , IL-1Ra, IL-17A, and IL-12/23 p40) were elevated in patients with DED with high LT- $\alpha$  (>700 pg/mL) compared to those with lower LT- $\alpha$  (<700 pg/mL), indicating possible differences in pathogenesis <sup>822</sup>.

### 5.3 Sustainability

To date, there has been a significant paucity of literature examining the sustainability implications of diagnostic testing and practices for DED <sup>823</sup>. Future research is required to characterise the potential sustainability implications, environmental effects, and carbon footprint of the production, use, and disposal of different types of diagnostic instruments, dyes, consumables, treatments and packaging.

5.4 Need for experience-informed approach to un/under-researched areas Where research on best practice is either limited, conflicting or logistically or ethically difficult to obtain, a group process using collective intelligence may help <sup>824</sup>. This can for example be applied to achieve consensus on the best clinical criteria for diagnosis or to initiate treatment <sup>825</sup>. The Delphi technique is a systematic process designed to establish consensus in a group of experts. A series of questionnaires is distributed, and controlled feedback with group statistical responses is given each time, until answers are converged and a predefined criterion is reached to bring the process to a close. Important aspects, in order to obtain valid outcomes, include systematic identification of the problem area, the selection of panel members based on objective and predefined criteria, anonymity of panelists and responses, controlled feedback, and stability of results including *a priori* defined closing criteria<sup>824</sup>.

A Delphi approach has been used to define ocular surface disease activity and damage indices <sup>826</sup>. Several Delphi or other group process approaches have been conducted in the past two decades to establish a best practice on a diagnostic aspect of DED and blepharitis. The ODISSEY European Consensus group defined a two-step scoring algorithm for diagnosing DED, but only at a severe level. Symptom-based assessment and corneal fluorescein staining were considered to be the two most important criteria. In case of discordance between these two tests, identification of additional criteria was recommended.<sup>827</sup> Separately, the DIDACTIC study used a Delphi approach to categorize signs and symptoms to identify DED pathophysiology. A total of 19 items were deemed indicative of evaporative dry eye, and 12 items of aqueous deficient DED.<sup>323</sup>. Using nominal group and Delphi techniques, a group of Italian ophthalmologists reached consensus on criteria for classification of DED. Three types were classified: a transient and reversible form, a recurrent form, and a chronic form, each with its own clinical characteristics <sup>828</sup>. Recent Delphi panels on Demodex-associated blepharitis have achieved consensus on it being chronic and recurrent <sup>486</sup>, with the presence of cylindrical dandruff at the base of the evelashes, visible Demodex mites, lid margin telangiectasia, and a previous history of anterior blepharitis not responding to treatment, being proposed as the most indicative independent signs <sup>485</sup>. Cylindrical dandruff is considered pathognomonic for Demodex blepharitis, with the suggestion that patients with >10 collarettes should be treated even in the absence of symptoms and that treatment efficacy can be tracked by the extent of cylindrical dandruff resolution <sup>829</sup>.

Examples of areas in the field of diagnostic methodology of DED that lack scientific evidence and could benefit from a future Delphi approach include:

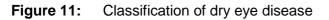
- how should DED severity be graded
- is site-specific itch useful in differentiating dry eye from allergy
- what cut-offs should be used for etiological drivers without current evidence-based diagnostic thresholds
- the best sequence for instilling fluorescein and lissamine green to assess ocular surface staining

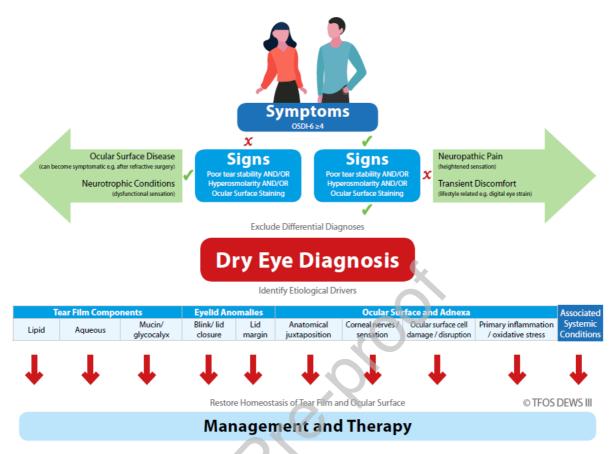
### 6 Summary

A standardised approach is critical to providing robust epidemiological information on DED in future.<sup>4</sup> It is critical that all practitioners and researchers adopt the same approach for the field to move forward for the benefit of patients.

### 6.1 Workflow and enhanced link to individualised management

In those patients that identify dryness type symptoms, the OSDI-6 screening questionnaire should be used to quantify these (see Section 3.3.2; Figure 4) and a score of  $\geq$ 4 used as a prompt for further investigation. Risk factors should be explored (see Section 3.2; Figure 3; Table 1) and a differential diagnosis conducted (see Section 3.1; Figure 2). If the practitioner lacks the expertise and access to instrumentation to facilitate a detailed examination of the eye, the triaging questions (see Section 3.1.5) will help to identify patients for whom referral is appropriate. Diagnosis of DED requires evaluation of ocular surface staining (including the cornea, conjunctiva and lid margin) along with tear film instability and/or hyperosmolarity (Figure 5), and the criteria met by demonstrating a positive score in at least one of these three indices of tear film and ocular surface homeostasis. In those diagnosed with DED, it is important to identify the etiological driver(s) of the individual's disease (see Section 3.5; Table 6) to inform the most appropriate management and therapy option(s) (as described in the TFOS DEWS III Management and Therapy report;<sup>17</sup> Figure 11)





An example of a test sequence to diagnose and identify the drivers (subtypes) of dry eye disease based on key tests / observations is presented in Table 7. Tests should be ordered from least to most invasive, to best maintain the integrity of the assessments and minimise impact on subsequent test results. Slight variations in test order may be expected where tests applied differ, but, in any case, the same test order should be utilised consistently on each patient at any one site, to enhance consistency in diagnosis and subclassification. Diagnosis and assessment for subtype drivers may be conducted in a single visit (if time allows) or may be separated into two visits to include a rapid diagnosis as part of routine testing, with follow up for subtype driver analysis at a separate visit. Test order will be adjusted accordingly, always from least to most invasive.

Table 7:An example of a test sequence (from least to most invasive) to diagnose(second column) and identify the drivers (subtypes – third column) of dry eye disease based on keytests / observations in the same appointment.

Sequence	Diagnosis	Subtype Drivers
1	Symptoms Screening - OSDI-6	
2		Blink / lid closure - rate / completeness
		/ lid seal
3		Aqueous – tear meniscus height (using
		infrared illumination)
4	Non-invasive tear breakup time	
5		Anatomical lid/globe misalignment –

		features such as pterygia
6		Inflammation – redness
7		Lipid – interferometry
8	Osmolarity	
9		Lid margin – eyelashes, lid margin,
		diagnostic expression
[10]	[Fluorescein tear breakup time]	
11	Ocular surface staining	Ocular surface damage- corneal,
		conjunctival and lid wiper staining
		Mucin – conjunctival staining
		Lid margin – keratinisation staining
12		Lid margin – meibography
13		Neural dysfunction - corneal nerves /
		sensation (if contact methods are
		used)

### 6.2 Patient communication

When communicating with a patient experiencing dry eye, active, two-way communication should be prioritized by openly discussing their symptoms, addressing concerns, educating them on lifestyle modifications to manage the condition, and empowering them to actively participate in their treatment plan. Key aspects include: explaining the chronic nature of DED, setting realistic expectations, discussing environmental triggers, promoting proper eye hygiene practices and recommending appropriate artificial tears or other treatments based on their individual needs and as indicated by their individual identified etiological drivers. Patients are usually highly engaged in their desired outcomes<sup>315</sup> and if DED is suspected, a follow up appointment is likely to be needed to perform a differential diagnosis, conduct the diagnostic algorithm, identify the etiological drivers (with the potential for a technician to conduct these measurements) and to discuss these with the patient and ideally to show them the images capture, to make shared decisions on appropriate management and therapy.<sup>17</sup>

### 6.3 Key diagnostic methodological changes from TFOS DEWS II

The revised definition: "Dry eye is a multifactorial, symptomatic disease characterized by a loss of homeostasis of the tear film and/or ocular surface, in which tear film instability and hyperosmolarity, ocular surface inflammation and damage, and neurosensory abnormalities are etiological factors." Key features of DED from the definition that have been reemphasised include that DED is multifactorial, a disease not a syndrome and always symptomatic.

The recommended screening questionnaire is the OSDI-6 with a cut-off of a score  $\geq$ 4 (Figure 4). Other questionnaires can be used as desired to gain further understanding of the environmental risk factors, but the standardised diagnostic questionnaire is necessary to achieve diagnostic consistency for all patients.

Key clinical differential diagnosis questions, alongside asking about a patient's general health and medication, are:

- Do you feel eye pain rather than discomfort?
- Do you have any facial flushing/redness, mouth dryness or enlarged salivary glands?
- When did your symptoms start and can you recall any triggering event?
- Is your vision affected and if so, does it improve on blinking?
- Are the symptoms or any redness much worse in one eye than the other?
- Do the eyes itch, are swollen, crusty or have given off any discharge?

A detailed examination of the ocular surface is recommended where the responses to these questions suggest possible presence of eye conditions that might masquerade as DED, with a guide to the ocular examination presented in Figure 2.

Known DED risk / associated factors (Figure 3) have been updated based on the scientific evidence<sup>4</sup> and factors commonly assumed to be associated with, or even cause, DED for which the evidence is equivocal, are reported (Table 1).

The diagnostic algorithm for DED has been refined (Figure 5) and in mitigating the established variability between questionnaires, only one (the OSDI-6) is recommended for the diagnostic algorithm. The impact on the diagnosis of each individual signs of a loss of homeostasis of the tear film and ocular surface recommended in TFOS DEWS II has been examined demonstrating that the lack of non-invasive breakup time or osmolarity only has a minor effect. Hence the revised, TFOS DEWS III approach has been shown to be robust<sup>232</sup> and further simplifies the procedure for application in clinical practice.

In aligning treatment strategies that possess different mechanisms of action with the multiple established drivers of dry eye disease, it has become clear that a greater number of distinct subtypes than simply aqueous and evaporative need to be acknowledged to ensure optimal patient care. This report has compiled the evidence on a more detailed subclassification of the disease based on the etiological drivers, so that these can be identified for an individual patient for the purpose of informing appropriate management and therapy. Cut-offs for the identified clinical tests have been provided where available (Table 6). Many of these clinical tests are already part of a standard clinical DED routine (Table 7), with the TFOS DEWS III approach offering structure and links to management and therapy (Figure 11).<sup>17</sup>

#### Acknowledgments

Sarp Orgul and Arthur Chan are acknowledged for their support in specific sections of the report.

### Funding

The TFOS DEWS III effort was supported by unrestricted donations from Alcon, Bausch + Lomb, Azura, AbbVie, CooperVision, Dompé, Espansione Group, Harrow, Laboratoire Théa, SIFI, SINQI, Tarsus, Topcon and Trukera.

## 6 References

1. Wolffsohn JS, Arita R, Chalmers R, et al. TFOS DEWS II Diagnostic methodology report. Ocular Surface 2017;15:539-574.

2. Craig JP, Nichols KK, Akpek E, et al. TFOS DEWS II Definition and Classification Report. Ocular Surface 2017;15:276-283.

3. Craig JP, Nelson JD, Azar DT, et al. TFOS DEWS II Report Executive Summary. The ocular surface 2017;15:802-812.

4. Stapleton F, Argueso P, Azar D, et al. TFOS DEWS III Digest Report. American Journal of Ophthalmology 2025 HOLDER.

5. Wolffsohn JS, Lingham G, Downie LE, et al. TFOS Lifestyle: Impact of the digital environment on the ocular surface. The ocular surface 2023;28:213-252.

6. Alves M, Asbell P, Dogru M, et al. TFOS Lifestyle Report: Impact of environmental conditions on the ocular surface. The ocular surface 2023;29:1-52.

7. Markoulli M, Ahmad S, Arcot J, et al. TFOS Lifestyle: Impact of nutrition on the ocular surface. The ocular surface 2023;29:226-271.

8. Sullivan DA, da Costa AX, Del Duca E, et al. TFOS Lifestyle: Impact of cosmetics on the ocular surface. The ocular surface 2023;29:77-130.

9. Gomes JAP, Azar DT, Baudouin C, et al. TFOS Lifestyle: Impact of elective medications and procedures on the ocular surface. The ocular surface 2023;29:331-385.

10. Jones L, Efron N, Bandamwar K, et al. TFOS Lifestyle: Impact of contact lenses on the ocular surface. The ocular surface 2023;29:175-219.

11. Stapleton F, Abad JC, Barabino S, et al. TFOS lifestyle: Impact of societal challenges on the ocular surface. The ocular surface 2023;28:165-199.

12. Galor A, Britten-Jones AC, Feng Y, et al. TFOS Lifestyle: Impact of lifestyle challenges on the ocular surface. The ocular surface 2023;28:262-303.

Hsu CC, Chuang HK, Hsiao YJ, et al. Predicting Risks of Dry Eye Disease Development Using a Genome-Wide Polygenic Risk Score Model. Translational vision science & technology 2024;13:13.
 Vehof J, Wang B, Kozareva D, Hysi PG, Snieder H, Hammond CJ. The heritability of dry eye

disease in a female twin cohort. Investigative ophthalmology & visual science 2014;55:7278-83.

15. Vehof J, Zavos HMS, Lachance G, Hammond CJ, Williams FMK. Shared genetic factors underlie chronic pain syndromes. Pain 2014;155:1562-1568.

16. Spranger J. [Disease, syndrome, sequence]. Monatsschrift Kinderheilkunde : Organ der Deutschen Gesellschaft fur Kinderheilkunde 1989;137:2-7.

17. Jones L, Craig JP, Markoulli M, et al. TFOS DEWS III Management and Therapy Report. American Journal of Ophthalmology 2025 HOLDER.

18. Craig JP, Alves M, Wolffsohn JS, et al. TFOS Lifestyle Report Introduction: A Lifestyle Epidemic - Ocular Surface Disease. The ocular surface 2023;28:304-309.

19. Lemp MA. Report of the National eye institute/ industry workshop on clinical trials in dry eyes. CLAO J 1995;21:221-232.

20. The definition and classification of dry eye disease: report of the Definition and Classification Subcommittee of the International Dry Eye WorkShop (2007). The ocular surface 2007;5:75-92.

21. Tong L, Waduthantri S, Wong TY, et al. Impact of symptomatic dry eye on vision-related daily activities: the Singapore Malay Eye Study. Eye (Lond) 2010;24:1486-91.

22. Eroy AA, Utheim TP, Sundling V. Cross-sectional Study Exploring Vision-related Quality of Life in Dry Eye Disease in a Norwegian Optometric Practice. Optom Vis Sci 2023;100:679-687.

23. Karakus S, Mathews PM, Agrawal D, Henrich C, Ramulu PY, Akpek EK. Impact of Dry Eye on Prolonged Reading. Optom Vis Sci 2018;95:1105-1113.

24. Morthen MK, Magno MS, Utheim TP, et al. The vision-related burden of dry eye. The ocular surface 2022;23:207-215.

25. Boboridis KG, Messmer EM, Benítez-del-Castillo J, et al. Patient-reported burden and overall impact of dry eye disease across eight European countries: a cross-sectional web-based survey. BMJ Open 2023;13:e067007.

26. D'Souza S, Annavajjhala S, Thakur P, Mullick R, Tejal SJ, Shetty N. Study of tear film optics and its impact on quality of vision. Indian J Ophthalmol 2020;68:2899-2902.

27. Habay T, Majzoub S, Perrault O, Rousseau C, Pisella PJ. [Objective assessment of the functional impact of dry eye severity on the quality of vision by double-pass aberrometry]. J Fr Ophtalmol 2014;37:188-94.

28. Sullivan BD, Palazón de la Torre M, Yago I, et al. Tear Film Hyperosmolarity is Associated with Increased Variation of Light Scatter Following Cataract Surgery. Clinical ophthalmology 2024;18:2419-2426.

29. Zhao DX, Quintero M, Mihailovic A, et al. Utilizing visual symptoms to distinguish dry eye from glaucoma, cataract, and suspect glaucoma patients: a cross-sectional study. BMC Ophthalmol 2024;24:16.

30. Hartong DT, van der Meeren SW, Coumou AD, Kloos R, Saeed P. [Watery eyes: diagnostic and therapeutic considerations]. Nederlands tijdschrift voor geneeskunde 2023;167.

31. Gordon AA, Danek DJ, Phelps PO. Common inflammatory and infectious conditions of the eyelid. Disease-a-month : DM 2020;66:101042.

32. Kim BY, Riaz KM, Bakhtiari P, et al. Medically Reversible Limbal Stem Cell Disease: Clinical Features and Management Strategies. Ophthalmology 2014;121:2053-2058.

33. Gomes JAP, Azar DT, Baudouin C, et al. TFOS DEWS II iatrogenic report. The ocular surface 2017;15:511-538.

34. Pisella PJ, Pouliquen P, Baudouin C. Prevalence of ocular symptoms and signs with preserved and preservative free glaucoma medication. British Journal of Ophthalmology 2002;86:418-423.

35. Dua HS, Watson NJ, Mathur RM, Forrester JV. Corneal epithelial cell migration in humans: 'Hurricane and blizzard keratopathy'. Eye 1993;7:53-58.

36. Nelson JD. Superior limbic keratoconjunctivitis (SLK). Eye 1989;3:180-189.

37. De Gregorio A, Cerini A, Scala A, Lambiase A, Pedrotti E, Morselli S. Floppy eyelid, an underdiagnosed syndrome: a review of demographics, pathogenesis, and treatment. Ther Adv Ophthalmol 2021;13:25158414211059247.

38. Rapuano CJ, editor. Color Atlas & Synopsis of Clinical Ophthalmology: Cornea, 3rd\_Edition ed: Wolters Kluwer PE, 2019:2-421.

39. Wolffsohn JS, Arita R, Chalmers R, et al. TFOS DEWS II Diagnostic Methodology report. The ocular surface 2017;15:539-574.

40. Wolffsohn JS, Dumbleton K, Huntjens B, et al. CLEAR - Evidence-based contact lens practice. Contact Lens and Anterior Eye 2021;44:368-397.

41. Saw VPJ, Dart JKG. Ocular Mucous Membrane Pemphigoid: Diagnosis and Management Strategies. The Ocular Surface 2008;6:128.

42. Kohanim S, Palioura S, Saeed HN, et al. Stevens-Johnson Syndrome/Toxic Epidermal Necrolysis – A Comprehensive Review and Guide to Therapy. I. Systemic Disease. The Ocular Surface 2016;14:2-19.

43. Zaidman GW, Geeraets R, Paylor RR, Ferry AP. The Histopathology of Filamentary Keratitis. Archives of Ophthalmology 1985;103:1178-1181.

44. Roy A, Priyadarshini SR, Das S. Filamentary keratitis: A review. The ocular surface 2024;34:22-29.

45. Weiss M, Molina R, Ofoegbuna C, Johnson DA, Kheirkhah A. A review of filamentary keratitis. Survey of Ophthalmology 2022;67:52-59.

46. Albietz J, Sanfilippo P, Troutbeck R, Lenton LM. Management of filamentary keratitis associated with aqueous-deficient dry eye. Optom Vis Sci 2003;80:420-30.

47. Bron AJ, de Paiva CS, Chauhan S, et al. TFOS DEWS II Pathophysiology Report. Ocular Surface 2017;15:438-510.

48. Zemba M, Ionescu MA, Pîrvulescu RA, et al. Biomarkers of ocular allergy and dry eye disease. Romanian journal of ophthalmology 2023;67:250-259.

49. Hom MM, Nguyen AL, Bielory L. Allergic conjunctivitis and dry eye syndrome. Annals of allergy, asthma & immunology : official publication of the American College of Allergy, Asthma, & Immunology 2012;108:163-6.

50. Bonini S. Atopic keratoconjunctivitis, 78 ed. Allergy: European Journal of Allergy and Clinical Immunology, Supplement, 2004:71-73.

51. Kubicka-Trzaska A, Romanowska-Dixon B. Dry eye syndrome and allergic conjunctivitis-epidemics of XXI century--diagnostic problems and management. Przegląd lekarski 2009;66:967-971.

52. Akil H, Celik F, Ulas F, Kara IS. Dry Eye Syndrome and Allergic Conjunctivitis in the Pediatric Population. Middle East African journal of ophthalmology 2015;22:467-71.

53. Bonini M, Gramiccioni C, Fioretti D, et al. Asthma, allergy and the olympics: A 12-year survey in elite athletes. Current opinion in allergy and clinical immunology 2015;15:184-192.

54. Chen JJ, Applebaum DS, Sun GS, Pflugfelder SC. Atopic keratoconjunctivitis: A review. Journal of the American Academy of Dermatology 2014;70:569-575.

55. Akpek EK, Dart JK, Watson S, et al. A randomized trial of topical cyclosporin 0.05% in topical steroid–resistant atopic keratoconjunctivitis. Ophthalmology 2004;111:476-482.

56. Akova YA, Rodriguez A, Foster CS. Atopic keratoconjunctivitis. Ocular Immunology and Inflammation 1994;2:125-144.

57. Cornish KS, Gregory ME, Ramaesh K. Systemic Cyclosporin a in Severe Atopic Keratoconjunctivitis. European journal of ophthalmology 2010;20:844-851.

58. Dogru M, Katakami C, Nakagawa N, Tetsumoto K, Yamamoto M. Impression cytology in atopic dermatitis11The authors have no proprietary interest in any of the products or equipment mentioned in this article. Ophthalmology 1998;105:1478-1484.

59. Power WJ, Tugal-Tutkun I, Foster CS. Long-term follow-up of patients with atopic keratoconjunctivitis. Ophthalmology 1998;105:637-642.

60. Guglielmetti S, Dart JK, Calder V. Atopic keratoconjunctivitis and atopic dermatitis. Current opinion in allergy and clinical immunology 2010;10:478-485.

61. Bielory B, Bielory L. Atopic Dermatitis and Keratoconjunctivitis. Immunology and Allergy Clinics of North America 2010;30:323-336.

62. Friedman T, Friedman Z, Neumann E. Giant papillary conjunctivitis following cataract extraction. Annals of Ophthalmology 1984;16:50-52.

63. Durand ML, Barshak MB, Sobrin L. Eye Infections. N Engl J Med 2023;389:2363-2375.

64. Muto T, Imaizumi S, Kamoi K. Viral Conjunctivitis. Viruses 2023;15.

65. Maranhão AG, Soares CC, Albuquerque MCM, Santos N. Molecular epidemiology of adenovirus conjunctivitis in Rio de Janeiro, Brazil, between 2004 and 2007. Revista do Instituto de Medicina Tropical de Sao Paulo 2009;51:227-229.

66. Lynch JP, Fishbein M, Echavarria M. Adenovirus. Semin Respir Crit Care Med 2011;32:494-511.

67. M'Garrech M, Rousseau A, Kaswin G, et al. Impairment of lacrimal secretion in the unaffected fellow eye of patients with recurrent unilateral herpetic keratitis. Ophthalmology 2013;120:1959-67.

68. Cavalcanti BM, Cruzat A, Sahin A, Pavan-Langston D, Samayoa E, Hamrah P. In vivo confocal microscopy detects bilateral changes of corneal immune cells and nerves in unilateral herpes zoster ophthalmicus. The ocular surface 2018;16:101-111.

69. Alfonso SA, Fawley JD, Alexa Lu X. Conjunctivitis. Primary Care: Clinics in Office Practice 2015;42:325-345.

70. Henle G, Henle W, Clifford P, et al. Antibodies to epstein-barr virus in burkitt's lymphoma and control groups. Journal of the National Cancer Institute 1969;43:1147-1157.

71. Wong KW, D'Amico DJ, Hedges TR, III, Soong HK, Schooley RT, Kenyon KR. Ocular Involvement Associated With Chronic Epstein-Barr Virus Disease. Archives of Ophthalmology 1987;105:788-792.

72. Matoba AY. Ocular disease associated with Epstein-Barr virus infection. Survey of Ophthalmology 1990;35:145-150.

73. Orden Martínez B, Martínez Ruiz R, Millán Pérez R. Conjuntivitis bacteriana: patógenos más prevalentes y sensibilidad antibiótica. Anales de Pediatría 2004;61:32-36.

74. Sugita G, Hotomi M, Sugita R, et al. Genetic characteristics of Haemophilus influenzae and Streptococcus pneumoniae isolated from children with conjunctivitis-otitis media syndrome. Journal of Infection and Chemotherapy 2014;20:493-497.

75. Azari AA, Barney NP. Conjunctivitis: a systematic review of diagnosis and treatment. JAMA 2013;310:1721-9.

76. Faraj HG, Hoang-Xuan T. Chronic cicatrizing conjunctivitis. Curr Opin Ophthalmol 2001;12:250-7.

77. Vazirani J, Donthineni PR, Goel S, et al. Chronic cicatrizing conjunctivitis: A review of the differential diagnosis and an algorithmic approach to management. Indian J Ophthalmol 2020;68:2349-2355.

78. Branisteanu DC, Stoleriu G, Branisteanu DE, et al. Ocular cicatricial pemphigoid (Review). Exp Ther Med 2020;20:3379-3382.

79. Marmalidou A, Kheirkhah A, Dana R. Conjunctivochalasis: a systematic review. Surv Ophthalmol 2018;63:554-564.

80. Yvon C, Patel B, Malhotra R. Conjunctivochalasis. Int Ophthalmol Clin 2023;63:209-223.

81. Somnath A, Tripathy K. Pinguecula. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Koushik Tripathy declares no relevant financial relationships with ineligible companies., 2024.

82. Ghiasian L, Samavat B, Hadi Y, Arbab M, Abolfathzadeh N. Recurrent Pterygium: A Review. J Curr Ophthalmol 2021;33:367-378.

83. Van Acker SI, Van den Bogerd B, Haagdorens M, et al. Pterygium-The Good, the Bad, and the Ugly. Cells 2021;10.

84. Sood G, Patel BC. Conjunctival Concretions. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Bhupendra Patel declares no relevant financial relationships with ineligible companies., 2024.

Wang K, See CW. Salzmann's nodular degeneration. Exp Eye Res 2021;202:108351.
Maharana PK, Sharma N, Das S, et al. Salzmann's Nodular Degeneration. Ocul Surf 2016;14:20-30.

87. Lin SR, Aldave AJ, Chodosh J. Recurrent corneal erosion syndrome. Br J Ophthalmol 2019;103:1204-1208.

88. Laibson PR. Recurrent corneal erosions and epithelial basement membrane dystrophy. Eye Contact Lens 2010;36:315-7.

89. Ong Tone S, Kocaba V, Bohm M, Wylegala A, White TL, Jurkunas UV. Fuchs endothelial corneal dystrophy: The vicious cycle of Fuchs pathogenesis. Prog Retin Eye Res 2021;80:100863.
90. Siu GDJY, Young AL, Jhanji V. Alternatives to corneal transplantation for the management of

bullous keratopathy. Current Opinion in Ophthalmology 2014;25:347-352.

91. Cabrera-Aguas M, Khoo P, Watson SL. Infectious keratitis: A review. Clin Exp Ophthalmol 2022;50:543-562.

92. Mandal N, Yeung SN, Tadrous C, Iovieno A. Thygeson's superficial punctate keratitis. Graefes Arch Clin Exp Ophthalmol 2022;260:1837-1841.

93. Kate A, Basu S. A Review of the Diagnosis and Treatment of Limbal Stem Cell Deficiency. Front Med (Lausanne) 2022;9:836009.

94. Tappeiner C, Walscheid K, Heiligenhaus A. [Diagnosis and treatment of episcleritis and scleritis]. Ophthalmologe 2016;113:797-810.

95. Chiew RLJ, Au Eong DTM, Au Eong KG. Mucus fishing syndrome. BMJ Case Rep 2022;15.

96. Slagle WS, Slagle AM, Brough GH. Mucus fishing syndrome: case report and new treatment option. Optometry (St Louis, Mo) 2001;72:634-40.

97. Le DT, Kandel H, Watson SL. Evaluation of ocular neuropathic pain. The ocular surface 2023;30:213-235.

98. Tagawa Y, Noda K, Ohguchi T, Tagawa Y, Ishida S, Kitaichi N. Corneal hyperalgesia in patients with short tear film break-up time dry eye. The ocular surface 2019;17:55-59.

99. Kim J, Yoon HJ, You IC, Ko BY, Yoon KC. Clinical characteristics of dry eye with ocular neuropathic pain features: comparison according to the types of sensitization based on the Ocular Pain Assessment Survey. BMC Ophthalmol 2020;20:455.

100. Galor A, Levitt RC, Felix ER, Martin ER, Sarantopoulos CD. Neuropathic ocular pain: an important yet underevaluated feature of dry eye. Eye (Lond) 2015;29:301-12.

101. Rosenthal P, Borsook D. Ocular neuropathic pain. Br J Ophthalmol 2016;100:128-34.

102. Dermer H, Lent-Schochet D, Theotoka D, et al. A Review of Management Strategies for Nociceptive and Neuropathic Ocular Surface Pain. Drugs 2020;80:547-571.

103. Vehof JH, C. J. Systemic Pain Conditions and Dry Eye Disease. In: Galor A, editor. Dry Eye Disease, 2022:135-145.

104. Papas EB, Ciolino JB, Jacobs D, et al. The TFOS International Workshop on Contact Lens Discomfort: report of the management and therapy subcommittee. Investigative ophthalmology & visual science 2013;54:Tfos183-203.

105. Bitton E, Aumond S. Demodex and eye disease: a review. Clin Exp Optom 2021;104:285-294.

106. Tariq MA, Amin H, Ahmed B, Ali U, Mohiuddin A. Association of dry eye disease with smoking: A systematic review and meta-analysis. Indian J Ophthalmol 2022;70:1892-1904.

107. Vehof J, Snieder H, Jansonius N, Hammond CJ. Prevalence and risk factors of dry eye in 79,866 participants of the population-based Lifelines cohort study in the Netherlands. The ocular surface 2021;19:83-93.

108. Karimi S, Nouri H, Mahmoudinejad-Azar S, Abtahi SH. Smoking and environmental tobacco smoke exposure: implications in ocular disorders. Cutan Ocul Toxicol 2023;42:1-7.

109. Abu-Ismail L, Abuawwad MT, Taha MJ, et al. Prevalence of Dry Eye Disease Among Medical Students and Its Association with Sleep Habits, Use of Electronic Devices and Caffeine Consumption: A Cross-Sectional Questionnaire. Clin Ophthalmol 2023;17:1013-1023.

110. Li YZ, Wang C, Peng X, et al. Caffeine's protective role in dry eye disease and meibomian gland Dysfunction: Insights from clinical and Experimental models. Int Immunopharmacol 2024;146:113863.

111. Magno MS, Utheim TP, Morthen MK, et al. The Relationship Between Caffeine Intake and Dry Eye Disease. Cornea 2023;42:186-193.

112. Garcia-Marques JV, Talens-Estarelles C, Garcia-Lazaro S, Wolffsohn JS, Cervino A. Systemic, environmental and lifestyle risk factors for dry eye disease in a mediterranean caucasian population. Cont Lens Anterior Eye 2022;45:101539.

113. Magno MS, Daniel T, Morthen MK, et al. The relationship between alcohol consumption and dry eye. Ocul Surf 2021;21:87-95.

114. You YS, Qu NB, Yu XN. Alcohol consumption and dry eye syndrome: a Meta-analysis. Int J Ophthalmol 2016;9:1487-1492.

115. Nguyen L, Magno MS, Utheim TP, Jansonius NM, Hammond CJ, Vehof J. The relationship between habitual water intake and dry eye disease. Acta Ophthalmol 2023;101:65-73.

116. Walsh NP, Fortes MB, Raymond-Barker P, et al. Is whole-body hydration an important consideration in dry eye? Invest Ophthalmol Vis Sci 2012;53:6622-7.

117. Wu Y, Xie Y, Yuan Y, et al. The Mediterranean Diet and Age-Related Eye Diseases: A Systematic Review. Nutrients 2023;15.

118. Galor A, Gardener H, Pouyeh B, Feuer W, Florez H. Effect of a Mediterranean dietary pattern and vitamin D levels on Dry Eye syndrome. Cornea 2014;33:437-41.

119. Ismail AMA, El-Azeim ASA, Saif H. Effect of aerobic exercise alone or combined with
Mediterranean diet on dry eye in obese hypertensive elderly. Ir J Med Sci 2023;192:3151-3161.
120. Magno MS, Moschowits E, Morthen MK, et al. Greater adherence to a mediterranean diet is
associated with lower C-reactive protein (CRP) levels, but not to lower odds of having dry eye
disease. Ocul Surf 2023;30:196-203.

121. Molina-Leyva I, Molina-Leyva A, Riquelme-Gallego B, Cano-Ibanez N, Garcia-Molina L, Bueno-Cavanillas A. Effectiveness of Mediterranean Diet Implementation in Dry Eye Parameters: A Study of PREDIMED-PLUS Trial. Nutrients 2020;12.

122. Sriprasert I, Warren DW, Mircheff AK, Stanczyk FZ. Dry eye in postmenopausal women: a hormonal disorder. Menopause 2016;23:343-51.

123. Sullivan DA, Rocha EM, Aragona P, et al. TFOS DEWS II Sex, Gender, and Hormones Report. The ocular surface 2017;15:284-333.

124. Chen SP, Massaro-Giordano G, Pistilli M, Schreiber CA, Bunya VY. Tear osmolarity and dry eye symptoms in women using oral contraception and contact lenses. Cornea 2013;32:423-8.

125. Moschos MM, Nitoda E. The impact of combined oral contraceptives on ocular tissues: a review of ocular effects. Int J Ophthalmol 2017;10:1604-1610.

126. Wolpert LE, Snieder H, Jansonius NM, Utheim TP, Hammond CJ, Vehof J. Medication use and dry eye symptoms: A large, hypothesis-free, population-based study in the Netherlands. The ocular surface 2021;22:1-12.

127. McMonnies CW, Ho A. Responses to a dry eye questionnaire from a normal population. J Am Optom Assoc 1987;58:588-91.

128. McMonnies CW. Key questions in a dry eye history. J Am Optom Assoc 1986;57:512-7.

129. Dumbleton KA, Guillon M, Theodoratos P, Patel T. Diurnal variation in comfort in contact lens and non-contact lens wearers. Optom Vis Sci 2016;93:820-827.

130. Chalmers RL, Begley CG. Dryness symptoms among an unselected clinical population with and without contact lens wear. Cont Lens Anterior Eye 2006;29:25-30.

131. Begley CG, Chalmers RL, Mitchell GL, et al. Characterization of ocular surface symptoms from optometric practices in North America. Cornea 2001;20:610-8.

132. González-Méijome JM, Parafita MA, Yebra-Pimentel E, Almeida JB. Symptoms in a population of contact lens and noncontact lens wearers under different environmental conditions. Optom Vis Sci 2007;84:296-302.

133. Guillon M, Shah S. Rationale for 24-hour management of dry eye disease: A review. Cont Lens Anterior Eye 2019;42:147-154.

134. Ayaki M, Negishi K. Seasonality of Tear Meniscus Volume and Dry Eye-Related Symptoms - A Cross-Sectional Retrospective Cohort Study. Clin Ophthalmol 2023;17:3809-3816.

135. Eidet JR, Chen X, Ræder S, Badian RA, Utheim TP. Seasonal variations in presenting symptoms and signs of dry eye disease in Norway. Sci Rep 2022;12:21046.

136. Wolffsohn JS. Chapter 1 - Questionnaire Design and Use to Assess Dry Eye Disease. In: Galor A, editor. Dry Eye Disease: Elsevier, 2023:1-14.

137. Okumura Y, Inomata T, Iwata N, et al. A Review of Dry Eye Questionnaires: Measuring Patient-Reported Outcomes and Health-Related Quality of Life. Diagnostics (Basel) 2020;10.

138. Chalmers RL, Begley CG, Caffery B. Validation of the 5-Item Dry Eye Questionnaire (DEQ-5): Discrimination across self-assessed severity and aqueous tear deficient dry eye diagnoses. Cont Lens Anterior Eye 2010;33:55-60.

139. Wang MTM, Xue AL, Craig JP. Comparative Evaluation of 5 Validated Symptom
Questionnaires as Screening Instruments for Dry Eye Disease. JAMA Ophthalmol 2019;137:228-229.
140. Akowuah PK, Adjei-Anang J, Nkansah EK, et al. Comparison of the performance of the dry eye questionnaire (DEQ-5) to the ocular surface disease index in a non-clinical population. CONTACT LENS & ANTERIOR EYE 2022;45.

141. Chatterjee S, Agrawal D, Chaturvedi P. Ocular Surface Disease Index© and the five-item dry eye questionnaire: A comparison in Indian patients with dry eye disease. INDIAN JOURNAL OF OPHTHALMOLOGY 2021;69:2396-2400.

142. Pult H, Wolffsohn JS. The development and evaluation of the new Ocular Surface Disease Index-6. The Ocular Surface 2019.

143. Chidi-Egboka NC, Golebiowski B, Lee SY, Vi M, Jalbert I. Dry eye symptoms in children: can we reliably measure them? Ophthalmic Physiol Opt 2021;41:105-115.

144. Pult H, Wolffsohn JS. The Development and Evaluation of the New Ocular Surface Disease Index-6. NCC 2018. Veldhoven, NL, 2018.

145. Qin G, Moutari S, Chen J, et al. Validity of OSDI-6 questionnaire in a Chinese adult population. Sci Rep 2024;14:14320.

146. Wolffsohn JS, Travé-Huarte S, Craig JP, Muntz A, Stapleton FJ. Appropriateness of Questionnaires for the Diagnosis and Monitoring Treatment of Dry Eye Disease. Journal of clinical medicine, 2024.

147. Pult H, Wolffsohn JS. The development and evaluation of the new Ocular Surface Disease Index-6. The ocular surface 2019;17:817-821.

148. Bartlett JD, Keith MS, Sudharshan L, Snedecor SJ. Associations between signs and symptoms of dry eye disease: a systematic review. Clinical ophthalmology 2015;9:1719-30.

149. Craig JP, Nichols KK, Akpek EK, et al. TFOS DEWS II Definition and Classification Report. The ocular surface 2017;15:276-283.

150. Vehof J, Sillevis Smitt-Kamminga N, Nibourg SA, Hammond CJ. Sex differences in clinical characteristics of dry eye disease. The ocular surface 2018;16:242-248.

151. Adatia FA, Michaeli-Cohen A, Naor J, Caffery B, Bookman A, Slomovic A. Correlation between corneal sensitivity, subjective dry eye symptoms and corneal staining in Sjogren's syndrome. Can J Ophthalmol 2004;39:767-71.

152. Bourcier T, Acosta MC, Borderie V, et al. Decreased corneal sensitivity in patients with dry eye. Investigative ophthalmology & visual science 2005;46:2341-5.

153. Chin JY, Liu C, Lee IXY, et al. Impact of Age on the Characteristics of Corneal Nerves and Corneal Epithelial Cells in Healthy Adults. Cornea 2024;43:409-418.

154. Galor A, Felix ER, Feuer W, et al. Dry eye symptoms align more closely to non-ocular conditions than to tear film parameters. The British journal of ophthalmology 2015;99:1126-9.

155. Viso E, Rodriguez-Ares MT, Abelenda D, Oubina B, Gude F. Prevalence of asymptomatic and symptomatic meibomian gland dysfunction in the general population of Spain. Investigative ophthalmology & visual science 2012;53:2601-6.

156. McCann P, Kruoch Z, Lopez S, Malli S, Qureshi R, Li T. Interventions for Dry Eye: An Overview of Systematic Reviews. JAMA Ophthalmol 2024;142:58-74.

157. Hynnekleiv L, Magno M, Vernhardsdottir RR, et al. Hyaluronic acid in the treatment of dry eye disease. Acta Ophthalmol 2022;100:844-860.

158. Downie LE, Ng SM, Lindsley KB, Akpek EK. Omega-3 and omega-6 polyunsaturated fatty acids for dry eye disease. Cochrane Database Syst Rev 2019;12:CD011016.

159. Baudouin C, de la Maza MS, Amrane M, et al. One-Year Efficacy and Safety of 0.1% Cyclosporine a Cationic Emulsion in the Treatment of Severe Dry Eye Disease. European journal of ophthalmology 2017;27:678-685.

160. Liu SH, Saldanha IJ, Abraham AG, et al. Topical corticosteroids for dry eye. Cochrane Database Syst Rev 2022;10:CD015070.

161. de Paiva CS, Pflugfelder SC, Ng SM, Akpek EK. Topical cyclosporine A therapy for dry eye syndrome. Cochrane Database Syst Rev 2019;9:CD010051.

162. Vernhardsdottir RR, Magno MS, Hynnekleiv L, et al. Antibiotic treatment for dry eye disease related to meibomian gland dysfunction and blepharitis - A review. The ocular surface 2022;26:211-221.

163. Craig JP, Muntz A, Wang MTM, et al. Developing evidence-based guidance for the treatment of dry eye disease with artificial tear supplements: A six-month multicentre, double-masked randomised controlled trial. The ocular surface 2021;20:62-69.

164. Wirta DL, Galor A, Aune CA, et al. Long-Term Safety and Efficacy of a Water-Free Cyclosporine 0.1% Ophthalmic Solution for Treatment of Dry Eye Disease: ESSENCE-2 OLE. Cornea 2024.

165. Vehof J, Sillevis Smitt-Kamminga N, Nibourg SA, Hammond CJ. Predictors of Discordance between Symptoms and Signs in Dry Eye Disease. Ophthalmology 2017;124:280-286.

166. Ong ES, Felix ER, Levitt RC, Feuer WJ, Sarantopoulos CD, Galor A. Epidemiology of discordance between symptoms and signs of dry eye. The British journal of ophthalmology 2018;102:674-679.

167. Lu PT, Lee CY, Sun CC. Sex Differences and Discordance Between Symptoms and Signs of Dry Eye Disease. American journal of ophthalmology 2024;260:14-20.

168. D'Souza S, Shetty R, Nair AP, et al. Corneal Confocal Microscopy Features and Tear Molecular Profile in Study Participants with Discordance between Ocular Surface Disease Clinical Signs and Discomfort. Journal of clinical medicine 2022;11.

169. Shtein RM, Harper DE, Pallazola V, et al. Discordant Dry Eye Disease (An American Ophthalmological Society Thesis). Trans Am Ophthalmol Soc 2016;114:T4.

170. Li B, Tian Y, Wang S. The correlation of cytokines and sensory hypersensitivity in mild dry eye patients characterized by symptoms outweighing signs. Molecular vision 2020;26:359-369.

171. Ayaki M, Negishi K, Kawashima M, Uchino M, Kaido M, Tsubota K. Age Is a Determining Factor of Dry Eye-Related Signs and Symptoms. Diagnostics (Basel) 2020;10.

172. Kim M, Chun YS, Kim KW. Different perception of dry eye symptoms between patients with and without primary Sjogren's syndrome. Sci Rep 2022;12:2172.

173. Ucar IC, Esen F, Turhan SA, Oguz H, Ulasoglu HC, Aykut V. Corneal neuropathic pain in irritable bowel syndrome: clinical findings and in vivo corneal confocal microscopy. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2021;259:3011-3017.

174. Vehof J, Sillevis Smitt-Kamminga N, Kozareva D, Nibourg SA, Hammond CJ. Clinical Characteristics of Dry Eye Patients With Chronic Pain Syndromes. American journal of ophthalmology 2016;162:59-65 e2.

175. Galor A, Covington D, Levitt AE, et al. Neuropathic Ocular Pain due to Dry Eye Is Associated With Multiple Comorbid Chronic Pain Syndromes. The journal of pain : official journal of the American Pain Society 2016;17:310-8.

176. Stapleton F, Velez FG, Lau C, Wolffsohn JS. Dry eye disease in the young: A narrative review. The ocular surface 2024;31:11-20.

177. Chidi-Egboka NC, Golebiowski B, Lee SY, Vi M, Jalbert I. Dry eye symptoms in children: can we reliably measure them? OPHTHALMIC AND PHYSIOLOGICAL OPTICS 2021;41:105-115.

178. Tsubota K, Yokoi N, Shimazaki J, et al. New Perspectives on Dry Eye Definition and Diagnosis: A Consensus Report by the Asia Dry Eye Society. The ocular surface 2017;15:65-76.

179. Tsubota K, Yokoi N, Watanabe H, et al. A New Perspective on Dry Eye Classification: Proposal by the Asia Dry Eye Society. Eye Contact Lens 2020;46 Suppl 1:S2-S13.

180. Shimazaki J. Definition and Diagnostic Criteria of Dry Eye Disease: Historical Overview and Future Directions. Investigative ophthalmology & visual science 2018;59:DES7-DES12.

181. Kim DH, Eom Y, Yoon CH, et al. New Korean Guidelines for the Diagnosis and Management of Dry Eye Disease. Korean J Ophthalmol 2024;38:156-163.

182. Papas EB. Diagnosing dry-eye: Which tests are most accurate? CONTACT LENS & ANTERIOR EYE 2023;46.

183. Papas EB. The global prevalence of dry eye disease: A Bayesian view. Ophthalmic Physiol Opt 2021;41:1254-1266.

184. Llewelyn HJB. Sensitivity and specificity are not appropriate for diagnostic reasoning. 2017;358.

185. Wolffsohn JS, Travé-Huarte S, Craig JP, Wang MTM, Jones L. Letter to Editor RE "Diagnosing dry-eye: Which tests are most accurate?". Cont Lens Anterior Eye 2023;46:102061.

186. Alves M, Reinach PS, Paula JS, et al. Comparison of diagnostic tests in distinct well-defined conditions related to dry eye disease. PLoS One 2014;9:e97921.

187. de Oliveira FR, Motta ACF, Módulo CM, et al. Clinical and laboratory evaluation of sicca complaints: distinctive aspects of primary, secondary and non-Sjogren syndrome. Advances in rheumatology (London, England) 2022;62:23.

188. Garcia DM, Reis de Oliveira F, Módulo CM, et al. Is Sjögren's syndrome dry eye similar to dry eye caused by other etiologies? Discriminating different diseases by dry eye tests. PLoS One 2018;13:e0208420.

189. Sullivan BD, Crews LA, Messmer EM, et al. Correlations between commonly used objective signs and symptoms for the diagnosis of dry eye disease: clinical implications. Acta Ophthalmol 2014;92:161-6.

190. Mizuno Y, Yamada M, Miyake Y, Dry Eye Survey Group of the National Hospital Organization of J. Association between clinical diagnostic tests and health-related quality of life surveys in patients with dry eye syndrome. Jpn J Ophthalmol 2010;54:259-65.

191. Sullivan BD, Smith GT, Gupta A, Harman F, Ansari E. Impact of Clinician Subjectivity on the Assessment of Dry Eye Disease Prevalence in a UK Public Health Care Patient Population. Clinical ophthalmology 2024;18:743-753.

Szalai E, Berta A, Szekanecz Z, Szucs G, Modis L, Jr. Evaluation of tear osmolarity in non-Sjogren and Sjogren syndrome dry eye patients with the TearLab system. Cornea 2012;31:867-71.
Zhang Q, Wu Y, Song Y, et al. Screening Evaporative Dry Eyes Severity Using an Infrared Image. J Ophthalmol 2021;2021:8396503.

194. Kamao T, Yamaguchi M, Kawasaki S, Mizoue S, Shiraishi A, Ohashi Y. Screening for dry eye with newly developed ocular surface thermographer. American journal of ophthalmology 2011;151:782-791 e1.

195. Su TY, Hwa CK, Liu PH, et al. Noncontact detection of dry eye using a custom designed infrared thermal image system. Journal of biomedical optics 2011;16:046009.

196. Kikukawa Y, Tanaka S, Kosugi T, Pflugfelder SC. Non-invasive and objective tear film breakup detection on interference color images using convolutional neural networks. PLoS One 2023;18:e0282973.

197. Wang MTM, Power B, Xue AL, Craig JP. Diagnostic performance and optimal cut-off values for tear film lipid layer grading and thickness in dry eye disease: An investigator-masked, randomised crossover study. The ocular surface 2024;34:415-417.

198. Monfared N, Murphy PJ. Features and influences on the normal tear evaporation rate. Contact Lens and Anterior Eye 2023;46:101809.

199. Fineide F, Storås AM, Chen X, et al. Predicting an unstable tear film through artificial intelligence. Sci Rep 2022;12:21416.

200. Lemp MA, Holly FJ, Iwata S, Dohlman CH. The precorneal tear film. I. Factors in spreading and maintaining a continuous tear film over the corneal surface. Arch Ophthalmol 1970;83:89-94.
201. Norn M. Desiccation of the precorneal tear film I. Corneal wetting time. Acta Ophthalmologica 1969;47:865 - 880.

202. Peterson RC, Wolffsohn JS, Fowler CW. Optimization of anterior eye fluorescein viewing. American journal of ophthalmology 2006;142:572-5.

203. Acet Y, Dağ Y. Changes caused by fluorescein in tear film by hybrid break-up time test- part one; on quantitative values. Photodiagnosis and photodynamic therapy 2022;40:103137.

204. Itokawa T, Suzuki T, Koh S, Hori Y. Evaluating the Differences Between Fluorescein Tear Break-up Time and Noninvasive Measurement Techniques. Eye Contact Lens 2023;49:104-109.

205. Wang MTM, Craig JP. Comparative Evaluation of Clinical Methods of Tear Film Stability Assessment: A Randomized Crossover Trial. JAMA Ophthalmol 2018;136:291-294.

206. Mengher LSB, A. J.; Tonge, S. R.; Gilbert, D. J. Effect of fluorescein instillation on the precorneal tear film stability. Current eye research 1985;4:9-12.

207. Mooi JK, Wang MT, Lim J, Muller A, Craig JP. Minimising instilled volume reduces the impact of fluorescein on clinical measurements of tear film stability. Cont Lens Anterior Eye 2017.

208. Pflugfelder SC, Kikukawa Y, Tanaka S, Kosugi T. The utility of software-detected non-invasive tear break-up in comparison to fluorescein tear break-up measurements. Front Med (Lausanne) 2024;11:1351013.

209. Ramos L, Barreira N, Mosquera A, Penedo MG, Yebra-Pimentel E, García-Resúa C. Analysis of parameters for the automatic computation of the tear film break-up time test based on CCLRU standards. Comput Methods Programs Biomed 2014;113:715-724.

210. Downie LE. Automated Tear Film Surface Quality Breakup Time as a Novel Clinical Marker for Tear Hyperosmolarity in Dry Eye Disease. Investigative ophthalmology & visual science 2015;56:7260-8.

211. Alonso-Caneiro D, Iskander DR, Collins MJ. Tear film surface quality with soft contact lenses using dynamic-area high-speed videokeratoscopy. Eye Contact Lens 2009;35:227-31.

212. Szczesna DH, Alonso-Caneiro D, Robert Iskander D, Read SA, Collins MJ. Predicting dry eye using noninvasive techniques of tear film surface assessment. Invest Ophthalmol Vis Sci 2011;52:751-756.

213. Zeri F, Rizzo GC, Ponzini E, Tavazzi S. Comparing automated and manual assessments of tear break-up time using different non-invasive devices and a fluorescein procedure. Sci Rep 2024;14:2516.

214. Bandlitz S, Peter B, Pflugi T, et al. Agreement and repeatability of four different devices to measure non-invasive tear breakup time (NIBUT). Cont Lens Anterior Eye 2020;43:507-511.

215. Lee R, Yeo S, Aung HT, Tong L. Agreement of noninvasive tear break-up time measurement between Tomey RT-7000 Auto Refractor-Keratometer and Oculus Keratograph 5M. Clinical ophthalmology 2016;10:1785-1790.

216. Szczesna-Iskander DH, Llorens-Quintana C. Agreement between invasive and noninvasive measurement of tear film breakup time. Sci Rep 2024;14:3852.

217. Lim J, Wang MTM, Craig JP. Evaluating the diagnostic ability of two automated non-invasive tear film stability measurement techniques. Cont Lens Anterior Eye 2021;44:101362.

218. Johnson ME, Murphy PJ. Measurement of ocular surface irritation on a linear interval scale with the ocular comfort index. Investigative ophthalmology & visual science 2007;48:4451-8.

219. Yazdani M, Chen X, Tashbayev B, et al. Evaluation of the Ocular Surface Disease Index Questionnaire as a Discriminative Test for Clinical Findings in Dry Eye Disease Patients. Current eye research 2019;44:941-947.

220. Liu H, Begley C, Chen M, et al. A link between tear instability and hyperosmolarity in dry eye. Investigative ophthalmology & visual science 2009;50:3671-9.

221. Yi HC, Lee YP, Shin YJ. Influence of Nasal Tear Osmolarity on Ocular Symptoms Related to Dry Eye Disease. American journal of ophthalmology 2018;189:71-76.

222. Lemp MA, Bron AJ, Baudouin C, et al. Tear os molarity in the diagnosis and management of dry eye disease. American journal of ophthalmology 2011;151:792-798 e1.

223. Park J, Choi Y, Han G, et al. Evaluation of tear osmolarity measured by I-Pen osmolarity system in patients with dry eye. Sci Rep 2021;11:7726.

224. Tavakoli A, Markoulli M, Flanagan J, Papas E. The validity of point of care tear film osmometers in the diagnosis of dry eye. Ophthalmic Physiol Opt 2022;42:140-148.

225. Nolfi J, Caffery B. Randomized comparison of in vivo performance of two point-of-care tear film osmometers. Clinical ophthalmology 2017;11:945-950.

226. Greiner JV, Ying GS, Pistilli M, Maguire MG, Asbell PA. Association of Tear Osmolarity With Signs and Symptoms of Dry Eye Disease in the Dry Eye Assessment and Management (DREAM) Study. Investigative ophthalmology & visual science 2023;64:5.

227. Tashbayev B, Utheim TP, Utheim Ø A, et al. Utility of Tear Osmolarity Measurement in Diagnosis of Dry Eye Disease. Sci Rep 2020;10:5542.

228. Choi M, Park YM, Ko BY. Comparative Evaluation of Matrix Metalloproteinase-9 Immunoassay and Tear Osmolarity Measurement for Diagnosing Severity of Dry Eye Disease. Korean J Ophthalmol 2023;37:409-416.

229. van Setten GB. Osmokinetics: A new dynamic concept in dry eye disease. J Fr Ophtalmol 2019;42:221-225.

230. Kursite A, Laganovska G. Effect of tear osmolarity on postoperative refractive error after cataract surgery. Journal of Ophthalmology (Ukraine) 2023:11-15.

231. Vidal-Rohr M, Craig JP, Davies LN, Wolffsohn JS. The epidemiology of dry eye disease in the UK: The Aston dry eye study. Contact Lens Anterior Eye 2023;46:101837.

232. Wolffsohn JS, Travé-Huarte S, Stapleton F, et al. Investigating the relative importance of tear homeostatic signs for the diagnosis of dry eye disease. The Ocular Surface 2025;36:151-155.

233. Fong PY, Shih KC, Lam PY, Chan TCY, Jhanji V, Tong L. Role of tear film biomarkers in the diagnosis and management of dry eye disease. Taiwan journal of ophthalmology 2019;9:150-159.

234. Li Y, Smith RG, Steel JC, Layton CJ. Attitudes of eye health practitioners towards diagnostic tests and therapies for dry eye disease in Australasia. Cont Lens Anterior Eye 2022;45:101724.
235. Casemore RK, Wolffsohn JS, Dutta D. Dry eye clinical practice patterns of UK optometrists. Cont Lens Anterior Eye 2023;46:101889.

236. Arita R, Morishige N, Koh S, et al. Increased Tear Fluid Production as a Compensatory Response to Meibomian Gland Loss: A Multicenter Cross-sectional Study. Ophthalmology 2015;122:925-33.

237. Tawfik A, Pistilli M, Maguire MG, et al. Association of Dry Eye Symptoms and Signs in Patients with Dry Eye Disease. Ophthalmic Epidemiol 2024;31:274-282.

238. Bron AJ, Argueso P, Irkec M, Bright FV. Clinical staining of the ocular surface: mechanisms and interpretations. Progress in retinal and eye research 2015;44:36-61.

239. Woods J, Hutchings N, Srinivasan S, Jones L. Geographic distribution of corneal staining in symptomatic dry eye. The ocular surface 2020;18:258-266.

240. Feenstra RP, Tseng SC. Comparison of fluorescein and rose bengal staining. Ophthalmology 1992;99:605-17.

241. Korb DR, Herman JP, Finnemore VM, Exford JM, Blackie CA. An evaluation of the efficacy of fluorescein, rose bengal, lissamine green, and a new dye mixture for ocular surface staining. Eye Contact Lens 2008;34:61-64.

242. Kim J, Foulks GN. Evaluation of the effect of lissamine green and rose bengal on human corneal epithelial cells. Cornea 1999;18:328-32.

243. Chodosh J, Dix RD, Howell RC, Stroop WG, Tseng SC. Staining characteristics and antiviral activity of sulforhodamine B and lissamine green B. Investigative ophthalmology & visual science 1994;35:1046-58.

244. Duan H, Yang T, Zhou Y, et al. Comparison of mucin levels at the ocular surface of visual display terminal users with and without dry eye disease. BMC Ophthalmol 2023;23:189.

245. Liu S, Dong H, Huang XH, Tang SH. Analysis of factors leading to lid wiper epitheliopathy. European review for medical and pharmacological sciences 2020;24:1593-1601.

246. Kato H, Yokoi N, Watanabe A, et al. Clinical Factors for Determining the Severity of Lid Wiper Epitheliopathy in Dry Eye. Cornea 2022;41:545-551.

247. Li W, Yeh TN, Leung T, Yuen T, Lerma M, Lin MC. The Relationship of Lid Wiper Epitheliopathy to Ocular Surface Signs and Symptoms. Investigative ophthalmology & visual science 2018;59:1878-1887.

248. Wang MTM, Dean SJ, Xue AL, Craig JP. Comparative performance of lid wiper epitheliopathy and corneal staining in detecting dry eye disease. Clin Experiment Ophthalmol 2019;47:546-548.

249. Wang MTM, Muntz A, Lim J, et al. Ageing and the natural history of dry eye disease: A prospective registry-based cross-sectional study. The ocular surface 2020;18:736-741.

250. Wang MTM, Power B, Xue AL, Craig JP. Discriminative performance of ocular surface staining and lid wiper epitheliopathy in dry eye disease: An investigator-masked, prospective registry-based, diagnostic accuracy study. The ocular surface 2024;34:165-172.

251. Blackie CA, Korb DR. A novel lid seal evaluation: the Korb-Blackie light test. Eye Contact Lens 2015;41:98-100.

252. Mejía-Salgado G, Rojas-Carabali W, Cifuentes-González C, et al. Real-world performance of the inflammadry test in dry eye diagnosis: an analysis of 1,515 patients. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2025.

253. Levy A, Georgeon C, Knoeri J, et al. Corneal Epithelial Thickness Mapping in the Diagnosis of Ocular Surface Disorders Involving the Corneal Epithelium: A Comparative Study. Cornea 2022;41:1353-1361.

254. Cui X, Hong J, Wang F, et al. Assessment of corneal epithelial thickness in dry eye patients. Optom Vis Sci 2014;91:1446-54.

255. Buffault J, Zéboulon P, Liang H, et al. Assessment of corneal epithelial thickness mapping in epithelial basement membrane dystrophy. PLoS One 2020;15:e0239124.

256. Rattan SA, Anwar DS. Comparison of corneal epithelial thickness profile in dry eye patients, keratoconus suspect, and healthy eyes. European journal of ophthalmology 2020;30:1506-1511.

257. Shrestha E, Shrestha JK, Shayami G, Chaudhary M. The conjunctival impression cytology between the diagnosed cases of dry eye and normal individuals. Nepalese journal of ophthalmology : a biannual peer-reviewed academic journal of the Nepal Ophthalmic Society : NEPJOPH 2011;3:39-44.

258. Baudouin C, Haouat N, Brignole F, Bayle J, Gastaud P. Immunopathological findings in conjunctival cells using immunofluorescence staining of impression cytology specimens. The British journal of ophthalmology 1992;76:545-9.

259. Brignole F, Pisella PJ, De Saint Jean M, Goldschild M, Goguel A, Baudouin C. Flow cytometric analysis of inflammatory markers in KCS: 6-month treatment with topical cyclosporin A. Investigative ophthalmology & visual science 2001;42:90-5.

260. Reddy M, Reddy PR, Reddy SC. Conjunctival impression cytology in dry eye states. Indian J Ophthalmol 1991;39:22-4.

261. Laver NMV. Ocular cytology: Diagnostic features and ongoing practices. Cancer cytopathology 2021;129:419-431.

262. Foulks GN. Challenges and pitfalls in clinical trials of treatments for dry eye. The ocular surface 2003;1:20-30.

263. Pena-Verdeal H, Garcia-Queiruga J, Sabucedo-Villamarin B, Giraldez MJ, Garcia-Resua C, Yebra-Pimentel E. Capability of the Inter-Eye Differences in Osmolarity, Break-Up Time and Corneal Staining on the Diagnostic of Dry Eye. Ocular immunology and inflammation 2023:1-8.

264. Nilsen C, Graae Jensen P, Gundersen M, et al. The Significance of Inter-Eye Osmolarity Difference in Dry Eye Diagnostics. Clinical ophthalmo ogy 2023;17:829-835.

265. Wang MTM, Ormonde SE, Muntz A, Craig JP. Diagnostic profile of tear osmolarity and interocular variability for dry eye disease. Clin Exp Ophthalmol 2020;48:255-257.

266. Korb DR, Herman JP, Greiner JV, et al. Lid wiper epitheliopathy and dry eye symptoms. Eye Contact Lens 2005;31:2-8.

267. Begley CG, Caffery B, Nelson JD, Situ P. The effect of time on grading corneal fluorescein and conjunctival lissamine green staining. The ocular surface 2022;25:65-70.

268. Delaveris A, Stahl U, Madigan M, Jalbert I. Comparative performance of lissamine green stains. Cont Lens Anterior Eye 2018;41:23-27.

269. Ghorbani-Mojarrad N, Wolffsohn JS, Craig JP, et al. A comparison of physical characteristics in different brands and staining techniques in a brand of lissamine green strips. Journal of Clinical Medicine 2025;14:2022.

270. Ghorbani-Mojarrad N, Vianya-Estopa M, Martin E, et al. Optimizing the methodology for the assessment of bulbar conjunctival lissamine green staining. Optom Vis Sci 2024;101:589-594.

271. Lievens CW, Norgett Y, Briggs N, Allen PM, Vianya-Estopa M. Optimal methodology for lid wiper epitheliopathy identification. Cont Lens Anterior Eye 2021;44:101332.

272. Hamrah P, Alipour F, Jiang S, Sohn JH, Foulks GN. Optimizing evaluation of Lissamine Green parameters for ocular surface staining. Eye (Lond) 2011;25:1429-34.

273. Shaw A, Collins M, Huang J, et al. Lid wiper epitheliopathy: The influence of multiple lid eversions and exposure time. Cont Lens Anterior Eye 2019;42:304-310.

274. Kunnen CME, Wolffsohn JS, Ritchey ER. Comparison of subjective grading of lid wiper epitheliopathy with a semi-objective method. Cont Lens Anterior Eye 2018;41:28-33.

275. Wolffsohn JS, Recchioni A, Hunt OA, et al. Optimising subjective grading of corneal staining in Sjögren's syndrome dry eye disease. Ocul Surf 2024;32:166-172.

276. Bağbaba A, Şen B, Delen D, Uysal BS. An Automated Grading and Diagnosis System for Evaluation of Dry Eye Syndrome. Journal of medical systems 2018;42:227.

277. Arvind A, Nanjappa R, Gupta KK, Macedo-de-Araújo RJ, Fadel D. Lid wiper epitheliopathy: Topical review of current identification strategies and future perspectives. Cont Lens Anterior Eye 2024;47:102312.

278. Nagino K, Okumura Y, Yamaguchi M, et al. Diagnostic Ability of a Smartphone App for Dry Eye Disease: Protocol for a Multicenter, Open-Label, Prospective, and Cross-sectional Study. JMIR Res Protoc 2023;12:e45218.

279. Oganov A, Yazdanpanah G, Jabbehdari S, Belamkar A, Pflugfelder S. Dry eye disease and blinking behaviors: A narrative review of methodologies for measuring blink dynamics and inducing blink response. The ocular surface 2023;29:166-174.

280. Palakuru JR, Wang J, Aquavella JV. Effect of blinking on tear dynamics. Investigative ophthalmology & visual science 2007;48:3032-7.

281. Seo J, Byun WY, Alisafaei F, et al. Multiscale reverse engineering of the human ocular surface. Nature medicine 2019;25:1310-1318.

282. Wang MTM, Tien L, Han A, et al. Impact of blinking on ocular surface and tear film parameters. The ocular surface 2018;16:424-429.

283. Johnston PR, Rodriguez J, Lane KJ, Ousler G, Abelson MB. The interblink interval in normal and dry eye subjects. Clinical ophthalmology 2013;7:253-9.

284. Tsubota K, Hata S, Okusawa Y, Egami F, Ohtsuki T, Nakamori K. Quantitative videographic analysis of blinking in normal subjects and patients with dry eye. Arch Ophthalmol 1996;114:715-20.
285. Pena-Verdeal H, García-Resúa C, Ramos L, Yebra-Pimentel E, Giráldez MJ. Diurnal variations in tear film break-up time determined in healthy subjects by software-assisted interpretation of tear film video recordings. Clin Exp Optom 2016;99:142-8.

286. Nakamori K, Odawara M, Nakajima T, Mizutani T, Tsubota K. Blinking is controlled primarily by ocular surface conditions. Am J Ophthalmol 1997;124:24-30.

287. Inomata T, Nakamura M, Sung J, et al. Smartphone-based digital phenotyping for dry eye toward P4 medicine: a crowdsourced cross-sectional study. NPJ Digit Med 2021;4:171.

288. Wolffsohn JS, Craig JP, Vidal-Rohr M, Huarte ST, Ah Kit L, Wang M. Blink Test enhances ability to screen for dry eye disease. Cont Lens Anterior Eye 2018;41:421-425.

289. Inomata T, Iwagami M, Hiratsuka Y, et al. Maximum blink interval is associated with tear film breakup time: A new simple, screening test for dry eye disease. Sci Rep 2018;8:13443.

290. Inomata T, Nakamura M, Iwagami M, et al. Risk Factors for Severe Dry Eye Disease: Crowdsourced Research Using DryEyeRhythm. Ophthalmology 2019;126:766-768.

291. Hirosawa K, Inomata T, Sung J, et al. Diagnostic ability of maximum blink interval together with Japanese version of Ocular Surface Disease Index score for dry eye disease. Sci Rep 2020;10:18106.

292. Nagino K, Okumura Y, Akasaki Y, et al. Smartphone App-Based and Paper-Based Patient-Reported Outcomes Using a Disease-Specific Questionnaire for Dry Eye Disease: Randomized Crossover Equivalence Study. J Med Internet Res 2023;25:e42638.

Fujio K, Nagino K, Huang T, et al. Clinical utility of maximum blink interval measured by smartphone application DryEyeRhythm to support dry eye disease diagnosis. Sci Rep 2023;13:13583.
Okumura Y, Inomata T, Midorikawa-Inomata A, et al. DryEyeRhythm: A reliable and valid smartphone application for the diagnosis assistance of dry eye. The ocular surface 2022;25:19-25.
Su Y, Liang Q, Su G, Wang N, Baudouin C, Labbé A. Spontaneous Eye Blink Patterns in Dry Eye: Clinical Correlations. Investigative ophthalmology & visual science 2018;59:5149-5156.

296. Zheng Q, Wang L, Wen H, et al. Impact of Incomplete Blinking Analyzed Using a Deep Learning Model With the Keratograph 5M in Dry Eye Disease. Translational vision science & technology 2022;11:38.

297. Boyd C, Weiss C, Halter J, Han K, Ershler W, Fried L. Framework for Evaluating Disease Severity Measures in Older Adults With Comorbidity. The journals of gerontology Series A, Biological sciences and medical sciences 2007;62:286-95. 298. Ferris FL, Davis MD, Clemons TE, et al. A simplified severity scale for age-related macular degeneration: AREDS Report No. 18. Arch Ophthalmol 2005;123:1570-4.

299. Alio JL, Shabayek MH. Corneal higher order aberrations: a method to grade keratoconus. J Refract Surg 2006;22:539-45.

300. Belin MW, Duncan JK. Keratoconus: The ABCD Grading System. Klinische Monatsblatter fur Augenheilkunde 2016;233:701-7.

301. Issarti I, Consejo A, Jimenez-Garcia M, Kreps EO, Koppen C, Rozema JJ. Logistic index for keratoconus detection and severity scoring (Logik). Comput Biol Med 2020;122:103809.

302. Age-Related Eye Disease Study Research G. The Age-Related Eye Disease Study (AREDS): design implications. AREDS report no. 1. Controlled clinical trials 1999;20:573-600.

303. Koh S, Inoue R, Ambrosio R, Jr., Maeda N, Miki A, Nishida K. Correlation Between Corneal Biomechanical Indices and the Severity of Keratoconus. Cornea 2020;39:215-221.

304. Mrugacz M, Ostrowska L, Bryl A, Szulc A, Zelazowska-Rutkowska B, Mrugacz G. Proinflammatory cytokines associated with clinical severity of dry eye disease of patients with depression. Adv Med Sci 2017;62:338-344.

305. Kim S, Park D, Shin Y, et al. Deep learning-based fully automated grading system for dry eye disease severity. PLoS One 2024;19:e0299776.

306. Yu J, Asche CV, Fairchild CJ. The economic burden of dry eye disease in the United States: a decision tree analysis. Cornea 2011;30:379-87.

307. Baudouin C, Aragona P, Van Setten G, et al. Diagnosing the severity of dry eye: a clear and practical algorithm. British Journal of Ophthalmology 2014;98:1168-76.

308. Sullivan BD, Whitmer D, Nichols KK, et al. An objective approach to dry eye disease severity. Investigative ophthalmology & visual science 2010;51:6125-30.

309. Ebrahimi F, Ayatollahi H, Aghaei H. A clinical decision support system for diagnosing and determining severity of dry eye disease. Eye (Lond) 2023;37:1619-1624.

310. Guo M, Huang B, Jia X, Liu B. Utilizing corneal fluorescein and conjunctival lissamine green staining combined with in vivo confocal microscopy for grading the severity of dry eye disease. Clinical ophthalmology 2025;19:807-817.

311. Chester T, Garg SS, Johnston J, Ayers B, Gupta P. How Can We Best Diagnose Severity Levels of Dry Eye Disease: Current Perspectives. Clinical ophthalmology 2023;17:1587-1604.

312. Methodologies to diagnose and monitor dry eye disease: report of the Diagnostic Methodology Subcommittee of the International Dry Eye WorkShop (2007). The ocular surface 2007;5:108-52.

313. Behrens A, Doyle JJ, Stern L, et al. Dysfunctional tear syndrome: a Delphi approach to treatment recommendations. Cornea 2006;25:900-7.

314. Amescua G, Ahmad S, Cheung AY, et al. American Academy of Ophthalmology preferred Practice pattern guidance - Dry Eye Syndrome. Preferred Practice Guidines, 2023.

315. Chiang JCB, Wolffsohn JS. Perspectives of dry eye patients in the United Kingdom on risk factors and desired treatment outcomes Cont Lens Anterior Eye 2024:In press.

316. Calvo F, Karras BT, Phillips R, Kimball AM, Wolf F. Diagnoses, syndromes, and diseases: a knowledge representation problem. AMIA Annual Symposium proceedings AMIA Symposium 2003;2003:802.

317. Albietz JM. Prevalence of dry eye subtypes in clinical optometry practice. Optom Vis Sci 2000;77:357-63.

318. Rege A, Kulkarni V, Puthran N, Khandgave T. A Clinical Study of Subtype-based Prevalence of Dry Eye. Journal of clinical and diagnostic research : JCDR 2013;7:2207-10.

319. Asiedu K, Dzasimatu SK, Kyei S. Clinical subtypes of dry eye in youthful clinical sample in Ghana. Cont Lens Anterior Eye 2019;42:206-211.

320. Vidal-Rohr M, Craig JP, Davies LN, Wolffsohn JS. Classification of dry eye disease subtypes Cont Lens Anterior Eye 2024:In Press.

321. Wang MTM, Craig JP, Power B, Wolffsohn JS. Prospective registry-based external validation cohort analysis of dry eye disease subtype classification criteria. Cont Lens Anterior Eye 2024;47:102290.

322. Craig JP, Nichols KK, Akpek EK, et al. TFOS DEWS II Definition and Classification Report. The ocular surface 2017;15:276-283.

323. Labetoulle M, Bourcier T, Doan S, group D. Classifying signs and symptoms of dry eye disease according to underlying mechanism via the Delphi method: the DIDACTIC study. The British journal of ophthalmology 2019;103:1475-1480.

Lemp MA, Crews LA, Bron AJ, Foulks GN, Sullivan BD. Distribution of aqueous-deficient and evaporative dry eye in a clinic-based patient cohort: a retrospective study. Cornea 2012;31:472-8.
Misra S, Wagner R, Ozkan B, et al. Precision subclassification of type 2 diabetes: a systematic review. Communications Medicine 2023;3:138.

326. Jones L, Downie LE, Korb D, et al. TFOS DEWS II Management and Therapy Report. The ocular surface 2017;15:575-628.

327. Yokoi N, Georgiev GA. Tear-film-oriented diagnosis for dry eye. Jpn J Ophthalmol 2019;63:127-136.

328. Yokoi N, Georgiev GA. Tear Film-Oriented Diagnosis and Tear Film-Oriented Therapy for Dry Eye Based on Tear Film Dynamics. Investigative ophthalmology & visual science 2018;59:DES13-DES22.

329. Yokoi N, Georgiev GA, Kato H, et al. Classification of Fluorescein Breakup Patterns: A Novel Method of Differential Diagnosis for Dry Eye. American journal of ophthalmology 2017;180:72-85.

330. Han SB, Liu YC, Mohamed-Noriega K, Tong L, Mehta JS. Objective Imaging Diagnostics for Dry Eye Disease. J Ophthalmol 2020;2020:3509064.

331. Geerling G, Baudouin C, Aragona P, et al. Emerging strategies for the diagnosis and treatment of meibomian gland dysfunction: Proceedings of the OCEAN group meeting. The ocular surface 2017;15:179-192.

332. Singh S, Donthineni PR, Srivastav S, Jacobi C, Basu S, Paulsen F. Lacrimal and meibomian gland evaluation in dry eye disease: A mini-review. Indian J Ophthalmol 2023;71:1090-1098.

333. Yoon DY, Hee Eun J, Hyon JY. Correlation between tear film lipid layer thickness and transepidermal water loss from the ocular area in patients with dry eye disease and in healthy controls. PLoS One 2022;17:e0270810.

334. McGinnigle S, Naroo SA, Eperjesi F. Evaluation of dry eye. Surv Ophthalmol 2012;57:293-316.
335. Craig JP, Tomlinson A. Importance of the lipid layer in human tear film stability and evaporation. Optom Vis Sci 1997;74:8-13.

336. Rinert J, Branger G, Bachmann LM, et al. Accuracy of a New Noninvasive Automatic Ocular Surface Analyzer for the Diagnosis of Dry Eye Disease-Two-Gate Design Using Healthy Controls. Cornea 2023;42:416-422.

337. Sanchez-Gonzalez MC, Capote-Puente R, Garcia-Romera MC, et al. Dry eye disease and tear film assessment through a novel non-invasive ocular surface analyzer: The OSA protocol. Front Med (Lausanne) 2022;9:938484.

338. Toth N, Szalai E, Rak T, Lillik V, Nagy A, Csutak A. Reliability and clinical applicability of a novel tear film imaging tool. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2021;259:1935-1943.

339. Ward CD, Murchison CE, Petroll WM, Robertson DM. Evaluation of the Repeatability of the LacryDiag Ocular Surface Analyzer for Assessment of the Meibomian Glands and Tear Film. Translational vision science & technology 2021;10:1.

340. Garcia-Marques JV, Talens-Estarelles C, Garcia-Lazaro S, Cervino A. Validation of a new objective method to assess lipid layer thickness without the need of an interferometer. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2022;260:655-676.

341. Finis DP, N.; Schrader, S.; Geerling, G. Evaluation of lipid layer thickness measurement of the tear film as a diagnostic tool for meibomian gland dysfunction. Cornea 2013; 32: 1549–1553.
342. Lee Y, Hyon JY, Jeon HS. Characteristics of dry eye patients with thick tear film lipid layers evaluated by a LipiView II interferometer. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2021;259:1235-1241.

Arita R, Morishige N, Fujii T, et al. Tear Interferometric Patterns Reflect Clinical Tear
Dynamics in Dry Eye Patients. Investigative ophthalmology & visual science 2016;57:3928-34.
Rolando M, Valente C, Barabino S. New test to quantify lipid layer behavior in healthy

subjects and patients with keratoconjunctivitis sicca. Cornea 2008;27:866-70.

345. Arita R, Fukuoka S, Morishige N. New Insights Into the Lipid Layer of the Tear Film and Meibomian Glands. Eye Contact Lens 2017;43:335-339.

346. Zhao Y, Tan CL, Tong L. Intra-observer and inter-observer repeatability of ocular surface interferometer in measuring lipid layer thickness. BMC Ophthalmol 2015;15:53.

347. Kim WJ, Ahn YJ, Kim MH, Kim HS, Kim MS, Kim EC. Lipid layer thickness decrease due to meibomian gland dysfunction leads to tear film instability and reflex tear secretion. Ann Med 2022;54:893-899.

348. Cohen Y, Epshtein S, Harris A, Gefen R, Kagemann L, Arieli Y. Tear film imager for dynamic mapping of the human tear film. Applied optics 2019;58:7987-7995.

349. Segev F, Geffen N, Galor A, et al. Dynamic assessment of the tear film muco-aqueous and lipid layers using a novel tear film imager (TFI). The British journal of ophthalmology 2020;104:136-141.

350. Mochizuki H, Yamada M, Hatou S, Tsubota K. Turnover rate of tear-film lipid layer determined by fluorophotometry. The British journal of ophthalmology 2009;93:1535-8.

351. Napoli PE, Coronella F, Satta GM, Fossarello M. A novel technique of contrast-enhanced optical coherence tomography imaging in evaluation of clearance of lipids in human tears. PLoS One 2014;9:e109843.

352. Bai Y, Ngo W, Nichols JJ. Characterization of the thickness of the tear film lipid layer using high resolution microscopy. The ocular surface 2019;17:356-359.

353. Bai Y, Ngo W, Khanal S, Nichols KK, Nichols JJ. Human precorneal tear film and lipid layer dynamics in meibomian gland dysfunction. The ocular surface 2021;21:250-256.

354. Stegmann HDS, V.A.; Schmidl, D.; Garhöfer, G.; Fard, A.; Bagherinia, H.; Schmetterer, L.; Werkmeister R.M. Classification of tear film lipid layer en face maps obtained using optical coherence tomography and their correlation with clinical parameters. Cornea 2023;42:490–497.

355. Wong S, Murphy PJ, Jones L. Tear evaporation rates: What does the literature tell us? Cont Lens Anterior Eye 2018;41:297-306.

356. Mathers WD, Lane JA, Zimmerman MB. Tear film changes associated with normal aging. Cornea 1996;15:229-34.

357. Kim YH, Graham AD, Li W, et al. Tear-film evaporation flux and its relationship to tear properties in symptomatic and asymptomatic soft-contact-lens wearers. Cont Lens Anterior Eye 2023;46:101850.

358. Goto E, Endo K, Suzuki A, Fujikura Y, Matsumoto Y, Tsubota K. Tear evaporation dynamics in normal subjects and subjects with obstructive meibomian gland dysfunction. Investigative ophthalmology & visual science 2003;44:533-9.

359. Petznick A, Tan JH, Boo SK, Lee SY, Acharya UR, Tong L. Repeatability of a New Method for Measuring Tear Evaporation Rates. Optom Vis Sci 2013;90:366-371.

360. Yeo S, Tan JH, Acharya UR, Sudarshan VK, Tong L. Longitudinal Changes in Tear Evaporation Rates After Eyelid Warming Therapies in Meibomian Gland Dysfunction. Investigative ophthalmology & visual science 2016;57:1974-81.

361. Monfared N, Murphy PJ. Features and influences on the normal tear evaporation rate. Cont Lens Anterior Eye 2023;46:101809.

362. Kimball SH, King-Smith PE, Nichols JJ. Evidence for the major contribution of evaporation to tear film thinning between blinks. Investigative ophthalmology & visual science 2010;51:6294-7.
363. Persiya J, Sasithradevi A. Thermal mapping the eye: A critical review of advances in infrared

imaging for disease detection. J Therm Biol 2024;121:103867.
364. Tan LL, Sanjay S, Morgan PB. Repeatability of infrared ocular thermography in assessing healthy and dry eyes. Cont Lens Anterior Eye 2016;39:284-92.

365. Moghadas M, Nosch DS, Käser E, et al. Ocular thermography and clinical measurements in symptomatic and asymptomatic soft contact lens wearers. Optom Vis Sci 2024;101:595-602.

366. Shah AM, Galor A. Impact of Ocular Surface Temperature on Tear Characteristics: Current Insights. Clin Optom (Auckl) 2021;13:51-62.

367. Matteoli S, Favuzza E, Mazzantini L, et al. Ocular surface temperature in patients with evaporative and aqueous-deficient dry eyes: a thermographic approach. Physiol Meas 2017;38:1503-1512.

368. Yamaguchi M, Sakane Y, Kamao T, et al. Noninvasive Dry Eye Assessment Using High-Technology Ophthalmic Examination Devices. Cornea 2016;35 Suppl 1:S38-S48.

369. Craig JP, Singh I, Tomlinson A, Morgan PB, Efron N. The role of tear physiology in ocular surface temperature. Eye (Lond) 2000;14 (Pt 4):635-41.

370. Su TY, Ho WT, Chiang SC, Lu CY, Chiang HK, Chang SW. Infrared thermography in the evaluation of meibomian gland dysfunction. J Formos Med Assoc 2017;116:554-559.

371. Korb DR, Blackie CA. Meibomian gland diagnostic expressibility: correlation with dry eye symptoms and gland location. Cornea 2008;27:1142-7.

372. Bilkhu P, Vidal-Rohr M, Trave-Huarte S, Wolffsonn JS. Effect of meibomian gland morphology on functionality with applied treatment. Cont Lens Anterior Eye 2022;45:101402.

373. Li Y, Lu J, Zhou Q, et al. Analysis of Clinical and Regional Distribution Characteristics of Obstructive Meibomian Gland Dysfunction in China: A Multicenter Study. Current eye research 2020;45:1373-1379.

374. Sang X, Li Y, Yang L, et al. Lipid layer thickness and tear meniscus height measurements for the differential diagnosis of evaporative dry eye subtypes. International journal of ophthalmology 2018;11:1496-1502.

375. Yang B, Wen K, Li J, et al. Quantitative evaluation of lipid layer thickness and blinking in children with allergic conjunctivitis. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2021;259:2795-2805.

376. Yokoi N, Komuro A. Non-invasive methods of assessing the tear film. Experimental eye research 2004;78:399-407.

377. Nakayama N, Kawashima M, Kaido M, Arita R, Tsubota K. Analysis of Meibum Before and After Intraductal Meibomian Gland Probing in Eyes With Obstructive Meibomian Gland Dysfunction. Cornea 2015;34:1206-8.

378. Li Q, Jiao SJ, Wang YQ, Xie HT, Zhang MC. Development of a Novel Tear Lipid Test Strip. Cornea 2022;41:232-237.

379. Niedernolte B, Trunk L, Wolffsohn JS, Pult H, Bandlitz S. Evaluation of tear meniscus height using different clinical methods. Clin Exp Optom 2021;104:583-588.

380. Sabucedo-Villamarin B, Pena-Verdeal H, Garcia-Queiruga J, Giraldez MJ, Garcia-Resua C, Yebra-Pimentel E. Categorization of the Aqueous Deficient Dry Eye by a Cut-Off Criterion of TMH Measured with Tearscope. Life (Basel) 2022;12.

381. Guinel M, Bitton E. Does the palpebral morphology influence the tear meniscus height between Caucasian and Asian eyes? Optom Vis Sci 2024;101:537-541.

382. Wolffsohn JS, Ayaz M, Bandlitz S, von der Höh F, Ebner A, Craig JP. Optimising the methodology for assessing tear meniscus height when using a digital camera. Contact Lens and Anterior Eye 2025:In press.

383. Yokoi N, Bron A, Tiffany J, Brown N, Hsuan J, Fowler C. Reflective meniscometry: a noninvasive method to measure tear meniscus curvature. The British journal of ophthalmology 1999;83:92-7.

384. Yokoi N, Bron AJ, Tiffany JM, Kinoshita S. Reflective meniscometry: a new field of dry eye assessment. Cornea 2000;19:S37-43.

385. Fukuda R, Usui T, Miyai T, Yamagami S, Amano S. Tear meniscus evaluation by anterior segment swept-source optical coherence tomography. American journal of ophthalmology 2013;155:620-624, 624 e1-2.

386. Ibrahim OM, Dogru M, Takano Y, et al. Application of visante optical coherence tomography tear meniscus height measurement in the diagnosis of dry eye disease. Ophthalmology 2010;117:1923-9.

387. Qiu X, Gong L, Lu Y, Jin H, Robitaille M. The diagnostic significance of Fourier-domain optical coherence tomography in Sjögren syndrome, aqueous tear deficiency and lipid tear deficiency patients. Acta Ophthalmol 2012;90:e359-66.

388. Qiu X, Gong L, Sun X, Jin H. Age-related variations of human tear meniscus and diagnosis of dry eye with Fourier-domain anterior segment optical coherence tomography. Cornea 2011;30:543-9.

389. Akiyama R, Usui T, Yamagami S. Diagnosis of Dry Eye by Tear Meniscus Measurements Using
Anterior Segment Swept Source Optical Coherence Tomography. Cornea 2015;34 Suppl 11:S115-20.
390. Elhusseiny AM, Khalil AA, El Sheikh RH, Bakr MA, Eissa MG, El Sayed YM. New approaches for
diagnosis of dry eye disease. International journal of ophthalmology 2019;12:1618-1628.

391. Singh S, Srivastav S, Modiwala Z, Ali MH, Basu S. Repeatability, reproducibility and agreement between three different diagnostic imaging platforms for tear film evaluation of normal and dry eye disease. Eye (Lond) 2023;37:2042-2047.

392. Pult H, Riede-Pult BH. Impact of Conjunctival Folds on Central Tear Meniscus Height. Investigative ophthalmology & visual science 2015;56:1459-66.

393. Wang Y, Xu Z, Gong Q, et al. The role of different tear volume detection methods in the evaluation and diagnosis of mild dry eye disease. Translational vision science & technology 2022;11:15.

394. Tomlinson A, Blades KJ, Pearce EI. What does the phenol red thread test actually measure? Optom Vis Sci 2001;78:142-6.

395. Hao Y, Jin T, Zhu L, et al. Validation of the phenol red thread test in a Chinese population. BMC Ophthalmol 2023;23:498.

396. Moore JE, Graham JE, Goodall EA, et al. Concordance between common dry eye diagnostic tests. The British journal of ophthalmology 2009;93:66-72.

397. Vashisht S, Singh S. Evaluation of Phenol Red Thread test versus Schirmer test in dry eyes: A comparative study. Int J Appl Basic Med Res 2011;1:40-2.

398. Masmali A, Alqahtani TA, Alharbi A, El-Hiti GA. Comparative study of repeatability of phenol red thread test versus Schirmer test in normal adults in Saudi Arabia. Eye Contact Lens 2014;40:127-31.

399. Patel S, Farrell J, Blades KJ, Grierson DJ. The value of a phenol red impregnated thread for differentiating between the aqueous and non-aqueous deficient dry eye. Ophthalmic Physiol Opt 1998;18:471-6.

400. Senchyna M, Wax MB. Quantitative assessment of tear production: A review of methods and utility in dry eye drug discovery. J Ocul Biol Dis Infor 2008;1:1-6.

401. Puentes B, Hisey EA, Ferneding M, et al. Development and validation of a method to generate phenol red thread tests. The ocular surface 2024;34:262-266.

402. Li N, Deng XG, He MF. Comparison of the Schirmer I test with and without topical anesthesia for diagnosing dry eye. International journal of ophthalmology 2012;5:478-81.

403. Wang X, Fan X, Wu Y, Mou Y, Min J, Jin X. Rear 4-min Schirmer test, a modified indicator of Schirmer test in diagnosing dry eye. Sci Rep 2022;12:6272.

## Journal Pre-proof

404. Miyasaka K, Ayaki M, Negishi K. Tear Strip Meniscometry and Its Clinical Application: Analysis of More Than 2000 Cases. Translational vision science & technology 2022;11:3.

405. Dogru M, Ishida K, Matsumoto Y, et al. Strip meniscometry: a new and simple method of tear meniscus evaluation. Investigative ophthalmology & visual science 2006;47:1895-901.

406. Alshammeri S, Madden L, Hagan S, Pearce El. Strip meniscometry tube: a rapid method for assessing aqueous deficient dry eye. Clin Exp Optom 2020;103:469-473.

407. Hao Y, Tian L, Cao K, Jie Y. Repeatability and Reproducibility of SMTube Measurement in Dry Eye Disease Patients. J Ophthalmol 2021;2021:1589378.

408. Negishi K, Ayaki M, Uchino M, Takei K, Tsubota K. Strip Meniscometry Correlates With Ocular Surface Tests and Symptoms. Translational vision science & technology 2020;9:31.

409. Garaszczuk IK, Iskander DR. Qualitative assessment of tear dynamics with fluorescein profilometry. Cont Lens Anterior Eye 2017;40:208-212.

410. Lim P, Han TA, Tong L. Short-Term Changes in Tear Lipid Layer Thickness After Instillation of Lipid Containing Eye Drops. Translational vision science & technology 2020;9:29.

411. Alotaibi WM, Abusharha A, Pearce El, Althomali M, Afsar T, Razak S. Assessment of Tamarind Seed Polysaccharide (TSP) and Hyaluronic Acid (HA) Containing Ophthalmic Solution to Maintain Tear Osmolarity, Ocular Surface Temperature (OST) and Tear Production. Clinical ophthalmology 2024;18:3503-3513.

412. Abusharha A, Pearce EI, Afsar T, Razak S. Evaluation of therapeutic efficacy of Emustil drops for ocular discomfort and tear film osmolarity using different treatment management modes under dry environmental conditions. BMC Ophthalmol 2024;24:129.

413. Abusharha A, Pearce EI, Alsaqar A, Fagehi R, Afsar T, Razak S. Efficacy of hydroxypropyl-guar drops in improving tear film index and ocular surface dynamics using two treatment methods under a controlled desiccating environment. International ophthalmology 2024;44:175.

414. Garaszczuk IK, Mousavi M, Cervino Exposito A, Bartuzel MM, Montes-Mico R, Iskander DR. Evaluating tear clearance rate with optical coherence tomography. Cont Lens Anterior Eye 2018;41:54-59.

415. Erhamamci S, Karalezli A, Yilmaz S, Aktas A. The clinical value and histopathological correlation of lacrimal scintigraphy in patients with primary Sjogren's syndrome. Nucl Med Commun 2012;33:689-94.

416. Gencoglu EA, Dursun D, Akova YA, Cengiz F, Yalcin H, Koyuncu A. Tear clearance measurement in patients with dry eye syndrome using quantitative lacrimal scintigraphy. Ann Nucl Med 2005;19:581-7.

417. Singh S, Shanbhag SS, Basu S. Tear secretion from the lacrimal gland: variations in normal versus dry eyes. The British journal of ophthalmology 2022;106:772-776.

418. Merino ML, Belmonte J, Rosas J, Acosta MC, Gallar J, Belmonte C. Maximal tear secretion evoked by controlled stimulation of corneal sensory nerves in healthy individuals and dry eye subjects. The ocular surface 2023;27:80-88.

419. Chang AY, Purt B. Biochemistry, Tear Film. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Boonkit Purt declares no relevant financial relationships with ineligible companies.: StatPearls Publishing LLC., 2024.

420. Versura P, Giannaccare G, Vukatana G, Mule R, Malavolta N, Campos EC. Predictive role of tear protein expression in the early diagnosis of Sjogren's syndrome. Ann Clin Biochem 2018;55:561-570.

421. Caffery B, Joyce E, Boone A, et al. Tear lipocalin and lysozyme in Sjogren and non-Sjogren dry eye. Optom Vis Sci 2008;85:661-7.

422. Sonobe H, Ogawa Y, Yamada K, et al. A novel and innovative paper-based analytical device for assessing tear lactoferrin of dry eye patients. The ocular surface 2019;17:160-166.

423. Lin Y, Zhang Y, Shi K, Wu H, Ou S. Advances in clinical examination of lacrimal gland. Front Med (Lausanne) 2023;10:1257209.

424. Ballow M, Donshik PC, Rapacz P, Samartino L. Tear lactoferrin levels in patients with external inflammatory ocular disease. Investigative ophthalmology & visual science 1987;28:543-5.

425. Kiratli H, Irkeç M, Orhan M. Tear lactoferrin levels in chronic meibomitis associated with acne rosacea. European journal of ophthalmology 2000;10:11-4.

426. Rapacz P, Tedesco J, Donshik PC, Ballow M. Tear lysozyme and lactoferrin levels in giant papillary conjunctivitis and vernal conjunctivitis. Clao j 1988;14:207-9.

427. Velamakanni GS, Sharma A, Batra HS, Velamakanni SM, Khan M, Gupta S. Evaluation of Tear Film Urea Level as a Diagnostic Marker of Dry Eye Disease: A Pilot Study. J Lab Physicians 2022;14:43-46.

428. Hori Y. Secreted Mucins on the Ocular Surface. Investigative ophthalmology & visual science 2018;59:DES151-DES156.

429. Argüeso P, Balaram M, Spurr-Michaud S, Keutmann HT, Dana MR, Gipson IK. Decreased levels of the goblet cell mucin MUC5AC in tears of patients with Sjögren syndrome. Investigative ophthalmology & visual science 2002;43:1004-11.

430. Mantelli F, Tiberi E, Micera A, Lambiase A, Visintini F, Bonini S. MUC5AC overexpression in tear film of neonates. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2007;245:1377-81.

431. You Y, Chen J, Chen H, et al. Investigation of Conjunctival Goblet Cell and Tear MUC5AC Protein in Patients With Graves' Ophthalmopathy. Translational vision science & technology 2023;12:19.

432. Danjo Y, Watanabe H, Tisdale AS, et al. Alteration of mucin in human conjunctival epithelia in dry eye. Investigative ophthalmology & visual science 1998;39:2602-9.

433. Argüeso P, Spurr-Michaud S, Russo CL, Tisdale A, Gipson IK. MUC16 mucin is expressed by the human ocular surface epithelia and carries the H185 carbohydrate epitope. Investigative ophthalmology & visual science 2003;44:2487-95.

434. Choi M, Tichenor AA. Regional Conjunctival Differences in Glycocalyx Mucin Expression in Dry Eye and Normal Subjects. Investigative ophthalmology & visual science 2024;65:20.

435. Inatomi T, Spurr-Michaud S, Tisdale AS, Gipson IK. Human corneal and conjunctival epithelia express MUC1 mucin. Investigative ophthalmology & visual science 1995;36:1818-27.

436. Nakamura Y, Yokoi N, Tokushige H, Kinoshita S. Sialic Acid in human tear fluid decreases in dry eye. Jpn J Ophthalmol 2004;48:519-23.

437. Uchino Y, Mauris J, Woodward AM, et al. Alteration of galectin-3 in tears of patients with dry eye disease. American journal of ophthalmology 2015;159:1027-1035.e3.

438. Hata-Mizuno M, Uchino Y, Uchino M, et al. Analysis of the Association between Galectin-3 Concentration in Tears and the Severity of Dry Eye Disease: A Case-Control Study. Journal of clinical medicine 2021;11.

439. Argueso P, Tisdale A, Spur-Michaud S, Sumiyoshi M, Gipson IK. Mucin characteristics of human corneal-limbal epithelial cells that exclude the rose bengal anionic dye. Investigative ophthalmology & visual science 2006;47:113-119.

440. Srinivas SP, Rao SK. Ocular surface staining: Current concepts and techniques. Indian J Ophthalmol 2023;71:1080-1089.

441. Vujkovic V, Mikac G, Kozomara R. Distribution and density of conjunctival goblet cells. Med Pregl 2002;55:195-200.

442. Liu Y, Duan Z, Yuan J, Xiao P. Imaging assessment of conjunctival goblet cells in dry eye disease. Clin Exp Ophthalmol 2024.

443. Matsumoto Y, Ibrahim OMA, Kojima T, Dogru M, Shimazaki J, Tsubota K. Corneal In Vivo Laser-Scanning Confocal Microscopy Findings in Dry Eye Patients with Sjögren's Syndrome. Diagnostics (Basel) 2020;10.

444. Mastropasqua R, Fasanella V, Brescia L, et al. In Vivo Confocal Imaging of the Conjunctiva as a Predictive Tool for the Glaucoma Filtration Surgery Outcome. Investigative ophthalmology & visual science 2017;58:Bio114-bio120.

445. Simsek C, Dogru M, Shinzawa M, et al. The Efficacy of 2% Topical Rebamipide on Conjunctival Squamous Metaplasia and Goblet Cell Density in Dry Eye Disease. Journal of ocular pharmacology and therapeutics : the official journal of the Association for Ocular Pharmacology and Therapeutics 2019;35:350-358.

446. Masmali AM, Purslow C, Murphy PJ. The tear ferning test: a simple clinical technique to evaluate the ocular tear film. Clin Exp Optom 2014;97:399-406.

447. Horwath J, Ettinger K, Bachernegg M, Bodner E, Schmut O. Ocular Ferning test - effect of temperature and humidity on tear Ferning patterns. Ophthalmologica 2001;215:102-7.

448. Rolando M. Tear mucus ferning test in normal and keratoconjunctivitis sicca eyes. . Chibret Int J Ophthalmol 1984;2:33-41.

449. Pearce EI, Tomlinson A. Spatial location studies on the chemical composition of human tear ferns. Ophthalmic and Physiological Optics 2000;20:306-313.

450. Masmali AM. Improvement of ferning patterns of lubricant eye drops mixed with various electrolytes and carboxymethylcellulose. Cont Lens Anterior Eye 2019;42:633-639.

451. Dutta D, Kim J, Sarkes M, Nath S, Markoulli M. The repeatability of subjective and objective tear ferning assessment and its association with lipid layer thickness, non-invasive tear break-up time and comfort. Cont Lens Anterior Eye 2019;42:420-427.

452. Sharanjeet K, Ho CY, Mutalib HA, Ghazali AR. The Relationship Between Tear Ferning
Patterns and Non-invasive Tear Break-up Time in Normal Asian Population. J Optom 2016;9:175-81.
453. Puderbach S, Stolze HH. Tear ferning and other lacrimal tests in normal persons of different ages. International ophthalmology 1991;15:391-5.

454. Mantelli F, Argueso P. Functions of ocular surface mucins in health and disease. Current opinion in allergy and clinical immunology 2008;8:477-83.

455. Bron AJ, Tiffany JM, Gouveia SM, Yokoi N, Voon LW. Functional aspects of the tear film lipid layer. Experimental eye research 2004;78:347-60.

456. Tsubota K. Tear dynamics and dry eye. Progress in retinal and eye research 1998;17:565-96. 457. Tong J, Lopez MJ, Patel BC. Anatomy, Head and Neck: Orbicularis Oculi Muscle. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Michael Lopez declares no relevant financial relationships with ineligible companies. Disclosure: Bhupendra Patel declares no relevant financial relationships with ineligible companies.: StatPearls Publishing

Copyright © 2024, StatPearls Publishing LLC., 2024.

458. Knight B, Lopez MJ, Patel BC. Anatomy, Head and Neck: Levator Palpebrae Superioris Muscle. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Michael Lopez declares no relevant financial relationships with ineligible companies. Disclosure: Bhupendra Patel declares no relevant financial relationships with ineligible companies.: StatPearls Publishing

Copyright © 2024, StatPearls Publishing LLC., 2024.

459. Ogino MH, Tadi P. Neuroanatomy, Trigeminal Reflexes. StatPearls. Treasure Island (FL) ineligible companies. Disclosure: Prasanna Tadi declares no relevant financial relationships with ineligible companies.: StatPearls Publishing

Copyright © 2024, StatPearls Publishing LLC., 2024.

460. Cruz AA, Garcia DM, Pinto CT, Cechetti SP. Spontaneous eyeblink activity. The ocular surface 2011;9:29-41.

461. Godfrey KJ, Wilsen C, Satterfield K, Korn BS, Kikkawa DO. Analysis of Spontaneous Eyelid Blink Dynamics Using a 240 Frames per Second Smartphone Camera. Ophthalmic Plast Reconstr Surg 2019;35:503-505.

462. Mitchell T, Murri M, Pflugfelder SC. Video Viewing Blink Rate in Normal and Dry Eyes. Eye Contact Lens 2021;47:442-444.

463. Wang MTM, Power B, Xue AL, Craig JP. Blink completeness and rate in dry eye disease: an investigator-masked, prospective registry-based, cross-sectional, prognostic study. Cont Lens Anterior Eye 2025:In press.

464. Jie Y, Sella R, Feng J, Gomez ML, Afshari NA. Evaluation of incomplete blinking as a measurement of dry eye disease. The ocular surface 2019;17:440-446.

465. Li Y, Li S, Zhou J, Liu C, Xu M. Relationship between lipid layer thickness, incomplete blinking rate and tear film instability in patients with different myopia degrees after small-incision lenticule extraction. PLoS One 2020;15:e0230119.

466. Jeon YJ, Song MY, Kim KY, Hwang KY, Kwon YA, Koh K. Relationship between the partial blink rate and ocular surface parameters. International ophthalmology 2021;41:2601-2608.

467. Shew W, Muntz A, Dean SJ, Pult H, Wang MTM, Craig JP. Blinking and upper eyelid morphology. Cont Lens Anterior Eye 2022;45:101702.

468. Nelson JD, Shimazaki J, Benitez-del-Castillo JM, et al. The international workshop on meibomian gland dysfunction: report of the definition and classification subcommittee. Investigative ophthalmology & visual science 2011;52:1930-7.

469. Knop E, Knop N, Millar T, Obata H, Sullivan DA. The international workshop on meibomian gland dysfunction: report of the subcommittee on anatomy, physiology, and pathophysiology of the meibomian gland. Investigative ophthalmology & visual science 2011;52:1938-78.

470. Rynerson JM, Perry HD. DEBS - a unification theory for dry eye and blepharitis. Clinical ophthalmology 2016;10:2455-2467.

471. Guillon M, Maissa C, Wong S. Symptomatic relief associated with eyelid hygiene in anterior blepharitis and MGD. Eye Contact Lens 2012;38:306-12.

472. Assayag E, Abulafia A, Teren D, Gelman E, Givoni H, Zadok D. Efficacy of 1% Povidone-Iodine in the Treatment of Anterior Blepharitis-Randomized Single-Center Controlled Trial. Journal of clinical medicine 2024;13.

473. Siddireddy JS, Tan J, Vijay AK, Willcox MDP. The Effect of Microblepharon Exfoliation on Clinical Correlates of Contact Lens Discomfort. Optom Vis Sci 2019;96:187-199.

474. Siddireddy JS, Vijay AK, Tan J, Willcox M. Effect of Eyelid Treatments on Bacterial Load and Lipase Activity in Relation to Contact Lens Discomfort. Eye Contact Lens 2020;46:245-253.

475. Kim AD, Muntz A, Lee J, Wang MTM, Craig JP. Therapeutic benefits of blinking exercises in dry eye disease. Cont Lens Anterior Eye 2021;44:101329.

476. de Paula A, Oliva G, Barraquer RI, de la Paz MF. Prevalence and antibiotic susceptibility of bacteria isolated in patients affected with blepharitis in a tertiary eye centre in Spain. European journal of ophthalmology 2020;30:991-997.

477. Rocha KM, Farid M, Raju L, et al. EYELID MARGIN DISEASE (BLEPHARITIS AND MEIBOMIAN GLAND DYSFUNCTION): CLINICAL REVIEW OF EVIDENCE-BASED AND EMERGING TREATMENTS. J Cataract Refract Surg 2024.

478. Teweldemedhin M, Saravanan M, Gebreyesus A, Gebreegziabiher D. Ocular bacterial infections at Quiha Ophthalmic Hospital, Northern Ethiopia: an evaluation according to the risk factors and the antimicrobial susceptibility of bacterial isolates. BMC Infect Dis 2017;17:207.

479. Marasini S, Swift S, Dean SJ, Ormonde SE, Craig JP. Spectrum and Sensitivity of Bacterial Keratitis Isolates in Auckland. J Ophthalmol 2016;2016:3769341.

480. Sakimoto T, Sugiura T. Clinical features of anterior blepharitis after cataract surgery. Sci Rep 2023;13:6615.

481. Rhee MK, Yeu E, Barnett M, et al. Demodex Blepharitis: A Comprehensive Review of the Disease, Current Management, and Emerging Therapies. Eye Contact Lens 2023;49:311-318.

482. Rabensteiner DF, Aminfar H, Boldin I, et al. Demodex Mite Infestation and its Associations with Tear Film and Ocular Surface Parameters in Patients with Ocular Discomfort. American journal of ophthalmology 2019;204:7-12.

483. Luo X, Li J, Chen C, Tseng S, Liang L. Ocular Demodicosis as a Potential Cause of Ocular Surface Inflammation. Cornea 2017;36 Suppl 1:S9-S14.

484. Norn MS. Incidence of Demodex Folliculorum on skin of lids and nose. Acta Ophthalmologica 1982;60:575-583.

485. Sharma N, Martin E, Pearce EI, Hagan S. A Delphi approach to establishing consensus on best practice for the diagnosis and treatment of Demodex blepharitis. Cont Lens Anterior Eye 2024;47:102080.

486. Ayres BD, Donnenfeld E, Farid M, et al. Clinical diagnosis and management of Demodex blepharitis: the Demodex Expert Panel on Treatment and Eyelid Health (DEPTH). Eye (Lond) 2023;37:3249-3255.

487. Man Peles I, Zahavi A, Chemodanova E, Vardizer Y. Novel In-Office Technique for Visual Confirmation of Demodex Infestation in Blepharitic Patients. Cornea 2020;39:858-861.

488. Shah PP, Stein RL, Perry HD. Update on the Management of Demodex Blepharitis. Cornea 2022;41:934-939.

489. Mastrota KM. Method to identify Demodex in the eyelash follicle without epilation. Optom Vis Sci 2013;90:e172-4.

490. Bitton E, Wittich W. Double-Blind Comparison of Eyelash Epilation Techniques for the Purpose of Demodex folliculorum Mite Retrieval. Eye Contact Lens 2024;50:152-157.

491. Muntz A, Purslow C, Wolffsohn JS, Craig JP. Improved Demodex diagnosis in the clinical setting using a novel in situ technique. Cont Lens Anterior Eye 2020;43:345-349.

492. Zhang AC, Muntz A, Wang MTM, Craig JP, Downie LE. Ocular Demodex: a systematic review of the clinical literature. Ophthalmic Physiol Opt 2020;40:389-432.

493. Gupta PK, Karpecki P. Comprehensive Assessment of the Meibomian Glands by Meibography: Why the Upper Eyelids Matter. Cornea 2025;44.

494. Amano S, Shimazaki J, Yokoi N, Hori Y, Arita R, Guidelines CfMGDCP. Meibomian Gland Dysfunction Clinical Practice Guidelines. Jpn J Ophthalmol 2023;67:448-539.

495. Feng J, Wang J, Wu B, et al. Association of meibomian gland morphology with orifice plugging and lid margin thickening in meibomian gland dysfunction patients. International ophthalmology 2023;43:3207-3218.

496. Crespo-Treviño RR, Salinas-Sánchez AK, Amparo F, Garza-Leon M. Comparative of meibomian gland morphology in patients with evaporative dry eye disease versus non-dry eye disease. Sci Rep 2021;11:20729.

497. Arita R, Minoura I, Morishige N, et al. Development of Definitive and Reliable Grading Scales for Meibomian Gland Dysfunction. American journal of ophthalmology 2016;169:125-137.

498. Daniel E, Maguire MG, Pistilli M, et al. Grading and baseline characteristics of meibomian glands in meibography images and their clinical associations in the Dry Eye Assessment and Management (DREAM) study. The ocular surface 2019;17:491-501.

499. Pult H, Riede-Pult B. Comparison of subjective grading and objective assessment in meibography. Cont Lens Anterior Eye 2013;36:22-7.

500. Fu J, Chou Y, Hao R, Jiang X, Liu Y, Li X. Evaluation of ocular surface impairment in meibomian gland dysfunction of varying severity using a comprehensive grading scale. Medicine 2019;98:e16547.

501. Opitz DL, Tyler KF. Efficacy of azithromycin 1% ophthalmic solution for treatment of ocular surface disease from posterior blepharitis. Clin Exp Optom 2011;94:200-6.

502. Hwang HS, Parfitt GJ, Brown DJ, Jester JV. Meibocyte differentiation and renewal: Insights into novel mechanisms of meibomian gland dysfunction (MGD). Experimental eye research 2017;163:37-45.

503. Adil MY, Xiao J, Olafsson J, et al. Meibomian Gland Morphology Is a Sensitive Early Indicator of Meibomian Gland Dysfunction. American journal of ophthalmology 2019;200:16-25.

504. Arita R, Itoh K, Inoue K, Amano S. Noncontact infrared meibography to document agerelated changes of the meibomian glands in a normal population, 2008/05/03 ed. Ophthalmology, 2008:911-5.

505. Arita R, Itoh K, Maeda S, et al. Proposed diagnostic criteria for obstructive meibomian gland dysfunction. Ophthalmology 2009;116:2058-63 e1.

506. Fineide F, Arita R, Utheim TP. The role of meibography in ocular surface diagnostics: A review. The ocular surface 2021;19:133-144.

507. Vunnava KP, Shetty R, Prabhu SM, Tiwari P, Kummelil MK. Automatic segmentation and quantified analysis of meibomian glands from infrared images. Indian J Ophthalmol 2023;71:1426-1431.

508. Lin X, Fu Y, Li L, et al. A Novel Quantitative Index of Meibomian Gland Dysfunction, the Meibomian Gland Tortuosity. Translational vision science & technology 2020;9:34.

509. Wang MTM, Power B, Xue AL, Kim JS, Craig JP. Diagnostic performance of qualitative and quantitative methods of meibomian gland dropout evaluation in dry eye disease: An investigator-masked, randomised crossover study. Cont Lens Anterior Eye 2024:102324.

510. Arita R. Meibography: A Japanese Perspective. Investigative ophthalmology & visual science 2018;59:Des48-des55.

511. Wang MTM, Wolffsohn JS, Kim JS, et al. Meibomian gland tortuosity, truncation, and dilation in paediatric dry eye disease: A multi-centre, investigator-masked, cross-sectional study. The ocular surface 2024;34:441-443.

512. von Ahrentschildt A, Hanenberg L, Robich ML, et al. Morphological characteristics of Meibomian Glands and their Influence on Dry Eye disease in contact lens wearers. The ocular surface 2022;24:93-99.

513. Ravichandran S, Pucker AD. Comparing meibomian gland visibility on optical coherence tomography and Keratograph 5M images using objective and subjective grading methods. Cont Lens Anterior Eye 2024;47:102162.

514. Randon M, Aragno V, Abbas R, Liang H, Labbé A, Baudouin C. In vivo confocal microscopy classification in the diagnosis of meibomian gland dysfunction. Eye (Lond) 2019;33:754-760.

515. Zhou S, Robertson DM. Wide-Field In Vivo Confocal Microscopy of Meibomian Gland Acini and Rete Ridges in the Eyelid Margin. Investigative ophthalmology & visual science 2018;59:4249-4257.

516. Li M, Yu Y, Yuan Y, et al. Rete Ridges in Eyelid Margin and Inflammatory Cytokines in Meibomian Gland Dysfunction Associated with Dry Eye Symptom. Current eye research 2021;46:202-209.

517. Muntz A, Subbaraman LN, Craig JP, Jones L. Cytomorphological assessment of the lid margin in relation to symptoms, contact lens wear and lid wiper epitheliopathy. The ocular surface 2020;18:214-220.

518. Knop E, Knop N, Zhivov A, et al. The lid wiper and muco-cutaneous junction anatomy of the human eyelid margins: an in vivo confocal and histological study. J Anat 2011;218:449-61.

519. Shanbhag SS, Singh S, Koshy PG, Donthineni PR, Basu S. A beginner's guide to mucous membrane grafting for lid margin keratinization: Review of indications, surgical technique and clinical outcomes. Indian J Ophthalmol 2021;69:794-805.

520. Ahn H, Ji YW, Jun I, Kim TI, Lee HK, Seo KY. Effects of meibomian gland dysfunction and aqueous deficiency on friction-related disease. The ocular surface 2022;26:295-299.

521. Koduri MA, Pingali T, Prasad D, et al. Neutrophil-driven and interleukin-36γ-associated ocular surface inflammation in chronic Stevens-Johnson syndrome. Allergy 2024;79:2128-2143.

522. D'Souza S, Nair AP, Ashok N, et al. Elevated neutrophils and reduced NK cells are associated with altered tear molecular signatures and clinical sequelae of chronic ocular Stevens-Johnson syndrome. The ocular surface 2024;34:9-21.

523. Cermak JM, Krenzer KL, Sullivan RM, Dana MR, Sullivan DA. Is complete androgen insensitivity syndrome associated with alterations in the meibomian gland and ocular surface? Cornea 2003;22:516-21.

524. Chen H, Gao H, Xie HT, Liu ST, Huang YK, Zhang MC. Hyperkeratinization and Proinflammatory Cytokine Expression in Meibomian Glands Induced by Staphylococcus aureus. Investigative ophthalmology & visual science 2021;62:11. 525. Schröder A, Abrar DB, Hampel U, Schicht M, Paulsen F, Garreis F. In vitro effects of sex hormones in human meibomian gland epithelial cells. Experimental eye research 2016;151:190-202.
526. Ji C, Guo Y, Liu Y, et al. Inhibition of ceramide de novo synthesis ameliorates meibomian gland dysfunction induced by SCD1 deficiency. The ocular surface 2021;22:230-241.

527. Bu J, Wu Y, Cai X, et al. Hyperlipidemia induces meibomian gland dysfunction. The ocular surface 2019;17:777-786.

528. Ibrahim MAA, Elwan WM. Role of topical dehydroepiandrosterone in ameliorating isotretinoin-induced Meibomian gland dysfunction in adult male albino rat. Annals of anatomy = Anatomischer Anzeiger : official organ of the Anatomische Gesellschaft 2017;211:78-87.

529. Redd TK, Seitzman GD. Ocular rosacea. Current opinion in ophthalmology 2020;31:503-507.
530. Gallo RL, Granstein RD, Kang S, et al. Standard classification and pathophysiology of rosacea: The 2017 update by the National Rosacea Society Expert Committee. Journal of the American Academy of Dermatology 2018;78:148-155.

531. Sobolewska B, Schaller M, Zierhut M. Rosacea and Dry Eye Disease. Ocular immunology and inflammation 2022;30:570-579.

532. Ye F, Zhou F, Xia Y, Zhu X, Wu Y, Huang Z. Evaluation of meibomian gland and tear film changes in patients with pterygium. Indian Journal of Ophthalmology 2017;65.

533. Wanzeler ACV, Barbosa IAF, Duarte B, Barbosa EB, Borges DA, Alves M. Impact of pterygium on the ocular surface and meibomian glands. PLoS One 2019;14:e0213956.

534. Dong N, Li W, Lin H, et al. Abnormal epithelial differentiation and tear film alteration in pinguecula. Investigative ophthalmology & visual science 2009;50:2710-5.

535. Ballesteros-Sánchez A, Sánchez-González J-M, Borrone MA, Borroni D, Rocha-de-Lossada C. The Influence of Lid-Parallel Conjunctival Folds and Conjunctivochalasis on Dry Eye Symptoms with and Without Contact Lens Wear: A Review of the Literature. Ophthalmology and Therapy 2024;13:651-670.

536. Linaburg T, Choi D, Bunya VY, Massaro-Giordano M, Briceno CA. Systematic Review: Effects of Pterygium and Pingueculum on the Ocular Surface and Efficacy of Surgical Excision. Cornea 2021;40:258-267.

537. Pult H, Tosatti SGP, Spencer ND, Asfour J-M, Ebenhoch M, Murphy PJ. Spontaneous Blinking from a Tribological Viewpoint. The ocular surface 2015;13:236-49.

538. Gumus K, Pflugfelder SC. Increasing prevalence and severity of conjunctivochalasis with aging detected by anterior segment optical coherence tomography. American journal of ophthalmology 2013;155:238-242.e2.

539. Höh H, Schirra F, Kienecker C, Ruprecht KW. Lid-parallel conjunctival folds (LIPCOF): A definite diagnostic sign of dry eye. Ophthalmologe 1995;92:802-808.

540. Pult H, Purslow C, Murphy PJ. The relationship between clinical signs and dry eye symptoms. Eye (Lond) 2011;25:502-10.

541. Pult H, Purslow C, Berry M, Murphy PJ. Clinical tests for successful contact lens wear: relationship and predictive potential. Optom Vis Sci 2008;85:E924-9.

542. Pult H, Bandlitz S. Lid-Parallel Conjunctival Folds and Their Ability to Predict Dry Eye. Eye Contact Lens 2018;44 Suppl 2:S113-s119.

543. Németh J, Fodor E, Lang Z, et al. Lid-parallel conjunctival folds (LIPCOF) and dry eye: A multicentre study. British Journal of Ophthalmology 2012.

544. Miller WL, Narayanan S, Jackson J, Bergmanson J. The association of bulbar conjunctival folds with other clinical findings in normal and moderate dry eye subjects. Optometry (St Louis, Mo) 2003;74:576-82.

545. Veres A, Tapaszto B, Kosina-Hagyo K, Somfai GM, Nemeth J. Imaging lid-parallel conjunctival folds with OCT and comparing its grading with the slit lamp classification in dry eye patients and normal subjects. Investigative ophthalmology & visual science 2011;52:2945-51.

546. Bandlitz S, Purslow C, Murphy PJ, Pult H. Lid-parallel conjunctival fold (LIPCOF) morphology imaged by optical coherence tomography and its relationship to LIPCOF grade. Cont Lens Anterior Eye 2019;42:299-303.

547. Barbosa Ribeiro B, Marques JH, Baptista PM, et al. Corneal Epithelial Thickness Correlation with Dry Eye Symptom Severity: A Cross-Sectional Study. Clinical ophthalmology 2024;18:3313-3320.
548. Watson SL, Le DT-M. Corneal neuropathic pain: a review to inform clinical practice. Eye 2024;38:2350-2358.

549. Kaido M, Kawashima M, Ishida R, Tsubota K. Relationship of Corneal Pain Sensitivity With Dry Eye Symptoms in Dry Eye With Short Tear Break-Up Time. Investigative ophthalmology & visual science 2016;57:914-919.

550. Wang MTM, Meyer JJ, Xue AL, Power B, Craig JP. Predictive performance of corneal and lid margin sensitivity for dry eye disease: An investigator-masked, prospective, prognostic accuracy study. Ocul Surf 2024;33:11-15.

551. Al-Aqaba MA, Dhillon VK, Mohammed I, Said DG, Dua HS. Corneal nerves in health and disease. Progress in retinal and eye research 2019;73:100762.

552. Chiang JCB, Tran V, Wolffsohn JS. The impact of dry eye disease on corneal nerve parameters: A systematic review and meta-analysis. Ophthalmic Physiol Opt 2023;43:1079-1091.
553. Matsumoto Y, Ibrahim OMA. Application of In Vivo Confocal Microscopy in Dry Eye Disease. Investigative ophthalmology & visual science 2018;59:DES41-DES47.

554. Talens-Estarelles C, Golebiowski B, Ehrmann K, García-Lázaro S, Cerviño A, Stapleton F. Corneal hypersensitivity to cold stimuli in symptomatic computer users. Optometry Vision Sci 2024;101.

555. Kalteniece A, Ferdousi M, Adam S, et al. Corneal confocal microscopy is a rapid reproducible ophthalmic technique for quantifying corneal nerve abnormalities. PLoS One 2017;12:e0183040.
556. Cardigos J, Barcelos F, Carvalho H, et al. Tear Meniscus and Corneal Sub-basal Nerve Plexus

Assessment in Primary Sjogren Syndrome and Sicca Syndrome Patients. Cornea 2019;38:221-228. 557. Tepelus TC, Chiu GB, Huang J, et al. Correlation between corneal innervation and inflammation evaluated with confocal microscopy and symptomatology in patients with dry eye

syndromes: a preliminary study. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2017;255:1771-1778. 558. Kheirkhah A, Satitpitakul V, Hamrah P, Dana R. Patients With Dry Eye Disease and Low Subbasal Nerve Density Are at High Risk for Accelerated Corneal Endothelial Cell Loss. Cornea 2017;36:196-201.

559. Levy O, Labbe A, Borderie V, et al. Increased corneal sub-basal nerve density in patients with Sjogren syndrome treated with topical cyclosporine A. Clin Exp Ophthalmol 2017;45:455-463.

560. Machalińska A, Kuligowska A, Ziontkowska-Wrzałek A, et al. The Severity of Diabetic Retinopathy Corresponds with Corneal Nerve Alterations and Ocular Discomfort of the Patient. Int J Mol Sci 2024;25.

561. Gong Q, Huang K, Li K, et al. Structural and functional changes of binocular corneal innervation and ocular surface function after unilateral SMILE and tPRK. The British journal of ophthalmology 2024;108:1492-1499.

562. Luzu J, Labbé A, Réaux-Le Goazigo A, et al. In vivo confocal microscopic study of corneal innervation in Sjögren's Syndrome with or without small fiber neuropathy. The ocular surface 2022;25:155-162.

563. Cañadas P, Lantigua Y, Enríquez-de-Salamanca A, et al. Ocular Surface Pathology in Patients Suffering from Mercury Intoxication. Diagnostics (Basel) 2021;11.

564. Colak D, Kocabeyoglu S, Karakaya J, Irkec M. Association of ocular surface and meibomian gland alterations with silicone hydrogel contact lens wear. Cont Lens Anterior Eye 2024;47:102093.
565. Rahman EZ, Lam PK, Chu CK, Moore Q, Pflugfelder SC. Corneal Sensitivity in Tear Dysfunction and its Correlation With Clinical Parameters and Blink Rate. American journal of ophthalmology 2015;160:858-866.e5.

566. Ross AR, Al-Aqaba MA, Almaazmi A, et al. Clinical and in vivo confocal microscopic features of neuropathic corneal pain. The British journal of ophthalmology 2020;104:768-775.

567. Moein HR, Akhlaq A, Dieckmann G, et al. Visualization of microneuromas by using in vivo confocal microscopy: An objective biomarker for the diagnosis of neuropathic corneal pain? The ocular surface 2020;18:651-656.

568. Maity M, Allay MB, Ali MH, Deshmukh R, Basu S, Singh S. Association of tear osmolarity and corneal nerves structure in dry eye disease: an in vivo study. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2024.

569. Kawashima M, Sano K, Takechi S, Tsubota K. Impact of lifestyle intervention on dry eye disease in office workers: a randomized controlled trial. J Occup Health 2018;60:281-288.

570. Giannaccare G, Pellegrini M, Sebastiani S, et al. Efficacy of Omega-3 Fatty Acid Supplementation for Treatment of Dry Eye Disease: A Meta-Analysis of Randomized Clinical Trials. Cornea 2019;38:565-573.

571. Molina-Leyva I, Molina-Leyva A, Bueno-Cavanillas A. Efficacy of nutritional supplementation with omega-3 and omega-6 fatty acids in dry eye syndrome: a systematic review of randomized clinical trials. Acta Ophthalmologica 2017;95.

572. So WZ, Qi Wong NS, Tan HC, et al. Diabetic corneal neuropathy as a surrogate marker for diabetic peripheral neuropathy. Neural Regen Res 2022;17:2172-2178.

573. Lagali NS, Allgeier S, Guimaraes P, et al. Reduced Corneal Nerve Fiber Density in Type 2 Diabetes by Wide-Area Mosaic Analysis. Investigative ophthalmology & visual science 2017;58:6318-6327.

574. Cozzini T, Piona C, Marchini G, et al. In vivo confocal microscopy study of corneal nerve alterations in children and youths with Type 1 diabetes. Pediatr Diabetes 2021;22:780-786.

575. Wu L-Q, Mou P, Chen Z-Y, et al. Altered Corneal Nerves in Chinese Thyroid-Associated Ophthalmopathy Patients Observed by In Vivo Confocal Microscopy. Medical Science Monitor 2019;25:1024-1031.

576. Fenner BJ, Tong L. Corneal staining characteristics in limited zones compared with whole cornea documentation for the detection of dry eye subtypes. Invest Ophthalmol Vis Sci 2013;54:8013-8019.

577. Efron N, Brennan NA, Morgan PB, Wilson T. Lid wiper epitheliopathy. Progress in retinal and eye research 2016;53:140-74.

578. Begley C, Caffery B, Chalmers R, Situ P, Simpson T, Nelson JD. Review and analysis of grading scales for ocular surface staining. The ocular surface 2019;17:208-220.

579. van Bijsterveld OP. Diagnostic tests in the Sicca syndrome. Arch Ophthalmol 1969;82:10-4.
580. Barr JT, Schechtman KB, Fink BA, et al. Corneal scarring in the Collaborative Longitudinal
Evaluation of Keratoconus (CLEK) Study: baseline prevalence and repeatability of detection. Cornea
1999;18:34-46.

581. Bron AJ, Evans VE, Smith JA. Grading of corneal and conjunctival staining in the context of other dry eye tests. Cornea 2003;22:640-50.

582. Sall K, Foulks GN, Pucker AD, Ice KL, Zink RC, Magrath G. Validation of a Modified National Eye Institute Grading Scale for Corneal Fluorescein Staining. Clinical ophthalmology 2023;17:757-767.

583. Miyata K, Amano S, Sawa M, Nishida T. A novel grading method for superficial punctate keratopathy magnitude and its correlation with corneal epithelial permeability. Arch Ophthalmol 2003;121:1537-9.

584. Whitcher JP, Shiboski CH, Shiboski SC, et al. A simplified quantitative method for assessing keratoconjunctivitis sicca from the Sjogren's Syndrome International Registry. American journal of ophthalmology 2010;149:405-15.

585. Yeniad B, Beginoglu M, Bilgin LK. Lid-wiper epitheliopathy in contact lens users and patients with dry eye. Eye Contact Lens 2010;36:140-143.

586. Wang MTM, Craig JP. Natural history of dry eye disease: Perspectives from inter-ethnic comparison studies. Ocul Surf 2019;17:424-433.

587. Nichols KK, Mitchell GL, Zadnik K. The repeatability of clinical measurements of dry eye. Cornea 2004;23:272-85.

588. Rose-Nussbaumer J, Lietman TM, Shiboski CH, et al. Inter-grader Agreement of the Ocular Staining Score in the Sjogren's International Clinical Collaborative Alliance (SICCA) Registry. American journal of ophthalmology 2015;160:1150-1153 e3.

589. Matossian C, Crowley M, Periman L, Sorkin S. Personalized Management of Dry Eye Disease: Beyond Artificial Tears. Clin Ophthalmol 2022;16:3911-3918.

590. Barabino S, Benitez-Del-Castillo JM. Dry eye disease pathogenesis and clinical signs: searching for correspondence in the clinical practice. Eur Rev Med Pharmacol Sci 2024;28:1881-1890.

591. Baudouin C. The pathology of dry eye. Surv Ophthalmol 2001;45 Suppl 2:S211-20.

592. Kojima T, Dogru M, Kawashima M, Nakamura S, Tsubota K. Advances in the diagnosis and treatment of dry eye. Progress in retinal and eye research 2020:100842.

593. Kemeny-Beke A, Szodoray P. Ocular manifestations of rheumatic diseases. Int Ophthalmol 2020;40:503-510.

594. Wang YH, Li W, McDermott M, et al. IFN-γ-producing T(H)1 cells and dysfunctional regulatory T cells contribute to the pathogenesis of Sjögren's disease. Sci Transl Med 2024;16:eado4856.

595. Hassan AS, Clouthier SG, Ferrara JL, et al. Lacrimal gland involvement in graft-versus-host disease: a murine model. Investigative ophthalmology & visual science 2005;46:2692-7.

596. Chen X, Aqrawi LA, Utheim TP, et al. Elevated cytokine levels in tears and saliva of patients with primary Sjogren's syndrome correlate with clinical ocular and oral manifestations. Sci Rep 2019;9:7319.

597. Park JY, Kim BG, Kim JS, Hwang JH. Matrix Metalloproteinase 9 Point-of-Care Immunoassay Result Predicts Response to Topical Cyclosporine Treatment in Dry Eye Disease. Transl Vis Sci Technol 2018;7:31.

598. Daull P, Baudouin C, Liang H, Feraille L, Barabino S, Garrigue JS. Review of Preclinical Outcomes of a Topical Cationic Emulsion of Cyclosporine A for the Treatment of Ocular Surface Diseases. Ocular immunology and inflammation 2022;30:1945-1955.

599. Aragona P, Aguennouz M, Rania L, et al. Matrix metalloproteinase 9 and transglutaminase 2 expression at the ocular surface in patients with different forms of dry eye disease. Ophthalmology 2015;122:62-71.

600. Tauber J. A 6-Week, Prospective, Randomized, Single-Masked Study of Lifitegrast Ophthalmic Solution 5% Versus Thermal Pulsation Procedure for Treatment of Inflammatory Meibomian Gland Dysfunction. Cornea 2020;39:403-407.

601. Valdes-Arias D, Locatelli EVT, Sepulveda-Beltran PA, Mangwani-Mordani S, Navia JC, Galor A. Recent United States Developments in the Pharmacological Treatment of Dry Eye Disease. Drugs 2024.

602. Zhuang D, Misra SL, Mugisho OO, Rupenthal ID, Craig JP. NLRP3 Inflammasome as a Potential Therapeutic Target in Dry Eye Disease. Int J Mol Sci 2023;24.

603. Messmer EM, von Lindenfels V, Garbe A, Kampik A. Matrix Metalloproteinase 9 Testing in Dry Eye Disease Using a Commercially Available Point-of-Care Immunoassay. Ophthalmology 2016;123:2300-2308.

604. Shon C, Muntz A, Xue A, Zhuang D, Craig JP. Optimising diagnosis of clinically significant inflammation in dry eye disease. American Academy of Optometry. San Diego, 2022.

605. Peterson RC, Wolffsohn JS. Sensitivity and reliability of objective image analysis compared to subjective grading of bulbar hyperaemia. The British journal of ophthalmology 2007;91:1464-6.

606. Amparo F, Wang H, Emami-Naeini P, Karimian P, Dana R. The Ocular Redness Index: a novel automated method for measuring ocular injection. Investigative ophthalmology & visual science 2013;54:4821-6.

607. Rodriguez JD, Hamm A, Bensinger E, et al. Statistical Evaluation of Smartphone-Based Automated Grading System for Ocular Redness Associated with Dry Eye Disease and Implications for Clinical Trials. Clinical ophthalmology 2025;19:907-914.

608. Altman J, Jones G, Ahmed S, Sharma S, Sharma A. Tear Film MicroRNAs as Potential Biomarkers: A Review. Int J Mol Sci 2023;24.

609. Winiarczyk M, Biela K, Michalak K, Winiarczyk D, Mackiewicz J. Changes in Tear Proteomic Profile in Ocular Diseases. Int J Environ Res Public Health 2022;19.

610. Tong L, Petznick A. Correlation between tear matrix metalloproteinases and the Schirmer's test. Investigative ophthalmology & visual science 2012;53:1592.

611. Papas EB. Key factors in the subjective and objective assessment of conjunctival erythema. Investigative ophthalmology & visual science 2000;41:687-91.

612. Fieguth P, Simpson T. Automated measurement of bulbar redness. Investigative ophthalmology & visual science 2002;43:340-7.

613. Bron AJ, Mengher LS, Davey CC. The normal conjunctiva and its responses to inflammation. Trans Ophthalmol Soc U K (1962) 1985;104 (Pt 4):424-35.

614. Roy FH. The red eye. Ann Ophthalmol (Skokie) 2006;38:35-8.

615. Mandell RB. Slit lamp classification system. J Am Optom Assoc 1987;58:198-201.

616. McMonnies CW, Chapman-Davies A. Assessment of conjunctival hyperemia in contact lens wearers. Part I. Am J Optom Physiol Opt 1987;64:246-50.

617. Efron N, Morgan PB, Katsara SS. Validation of grading scales for contact lens complications. Ophthalmic Physiol Opt 2001;21:17-29.

618. Schulze MM, Jones DA, Simpson TL. The development of validated bulbar redness grading scales. Optom Vis Sci 2007;84:976-83.

619. Schulze MM, Hutchings N, Simpson TL. Grading bulbar redness using cross-calibrated clinical grading scales. Invest Ophthalmol Vis Sci 2011;52:5812-7.

620. Villumsen J, Ringquist J, Alm A. Image analysis of conjunctival hyperemia. A personal computer based system. Acta Ophthalmol (Copenh) 1991;69:536-9.

621. Willingham FF, Cohen KL, Coggins JM, Tripoli NK, Ogle JW, Goldstein GM. Automatic quantitative measurement of ocular hyperemia. Curr Eye Res 1995;14:1101-8.

622. Wolffsohn JS, Purslow C. Clinical monitoring of ocular physiology using digital image analysis. Cont Lens Anterior Eye 2003;26:27-35.

623. Yoneda T, Sumi T, Takahashi A, Hoshikawa Y, Kobayashi M, Fukushima A. Automated hyperemia analysis software: reliability and reproducibility in healthy subjects. Jpn J Ophthalmol 2012;56:1-7.

Rodriguez JD, Johnston PR, Ousler GW, 3rd, Smith LM, Abelson MB. Automated grading
system for evaluation of ocular redness associated with dry eye. Clin Ophthalmol 2013;7:1197-1204.
Wu S, Hong J, Tian L, Cui X, Sun X, Xu J. Assessment of Bulbar Redness with a Newly

Developed Keratograph. Optom Vis Sci 2015;92:892-9.

626. Huntjens B, Basi M, Nagra M. Evaluating a new objective grading software for conjunctival hyperaemia. Cont Lens Anterior Eye 2020;43:137-143.

627. Peterson RC, Wolffsohn JS. Objective grading of the anterior eye. Optom Vis Sci 2009;86:273-8.

628. Shetty R, Dua HS, Tong L, et al. Role of in vivo confocal microscopy in dry eye disease and eye pain. Indian J Ophthalmol 2023;71:1099-1104.

629. Yu M, Liu C, Mehta JS, Liu YC. A review of the application of in-vivo confocal microscopy on conjunctival diseases. Eye and vision (London, England) 2024;11:43.

630. Kasikci M, Erogul O, Polat O. Evaluation of aqueous-deficient and evaporative dry eye cases with confocal microscopy. J Fr Ophtalmol 2023;46:1161-1168.

631. Azizi S, Uçak T, Yaşar I, Karakurt Y, Erdogan E, Salman I. Evaluation of the Corneal Layers in Meibomian-Gland-Dysfunction-Related Dry Eye by In Vivo Slit-Scanning Confocal Microscopy. Semin Ophthalmol 2017;32:377-383.

632. Yang T, Ma B, Xie J, et al. Evaluation of Ocular Surface Characteristics in Dry Eye Disease With and Without Soft Contact Lens Wear: A Comparative Study. Eye Contact Lens 2022;48:377-383.

633. He J, Ogawa Y, Mukai S, et al. In Vivo Confocal Microscopy Evaluation of Ocular Surface with Graft-Versus-Host Disease-Related Dry Eye Disease. Sci Rep 2017;7:10720.

634. Setu MAK, Schmidt S, Musial G, Stern ME, Steven P. Segmentation and Evaluation of Corneal Nerves and Dendritic Cells From In Vivo Confocal Microscopy Images Using Deep Learning. Translational vision science & technology 2022;11:24.

635. Hwang J, Dermer H, Galor A. Can in vivo confocal microscopy differentiate between subtypes of dry eye disease? A review. Clin Exp Ophthalmol 2021;49:373-387.

636. Villani E, Garoli E, Termine V, Pichi F, Ratiglia R, Nucci P. Corneal Confocal Microscopy in Dry Eye Treated with Corticosteroids. Optom Vis Sci 2015;92:e290-5.

637. Kheirkhah A, Dohlman TH, Amparo F, et al. Effects of corneal nerve density on the response to treatment in dry eye disease. Ophthalmology 2015;122:662-8.

638. Iaccheri B, Torroni G, Cagini C, et al. Corneal confocal scanning laser microscopy in patients with dry eye disease treated with topical cyclosporine. Eye (Lond) 2017;31:788-794.

639. Manicone AM, McGuire JK. Matrix metalloproteinases as modulators of inflammation. Semin Cell Dev Biol 2008;19:34-41.

640. Caban M, Owczarek K, Lewandowska U. The Role of Metalloproteinases and Their Tissue Inhibitors on Ocular Diseases: Focusing on Potential Mechanisms. Int J Mol Sci 2022;23.

641. Kapoor C, Vaidya S, Wadhwan V, Hitesh, Kaur G, Pathak A. Seesaw of matrix metalloproteinases (MMPs). J Cancer Res Ther 2016;12:28-35.

642. Acera A, Rocha G, Vecino E, Lema I, Duran JA. Inflammatory markers in the tears of patients with ocular surface disease. Ophthalmic Res 2008;40:315-21.

643. Chotikavanich S, de Paiva CS, Li de Q, et al. Production and activity of matrix metalloproteinase-9 on the ocular surface increase in dysfunctional tear syndrome. Investigative ophthalmology & visual science 2009;50:3203-9.

644. Knight BE, Kozlowski N, Havelin J, et al. TIMP-1 Attenuates the Development of Inflammatory Pain Through MMP-Dependent and Receptor-Mediated Cell Signaling Mechanisms. Front Mol Neurosci 2019;12:220.

645. Bang SP, Son MJ, Kim H, Lee YH, Jun JH. In vitro validation of the tear matrix metalloproteinase 9 in-situ immunoassay. Sci Rep 2020;10:15126.

646. Periman LM, Perez VL, Saban DR, Lin MC, Neri P. The Immunological Basis of Dry Eye Disease and Current Topical Treatment Options. Journal of ocular pharmacology and therapeutics : the official journal of the Association for Ocular Pharmacology and Therapeutics 2020;36:137-146.

647. Pflugfelder SC, De Paiva CS, Moore QL, et al. Aqueous Tear Deficiency Increases Conjunctival Interferon-γ (IFN-γ) Expression and Goblet Cell Loss. Investigative ophthalmology & visual science 2015;56:7545-50.

648. Zhao L, Yu JJ, Liu Y, et al. An enhanced model for environmental dry eye: Exploring pathological features and underlying factors. Experimental eye research 2024;239:109744.

649. Kumar NR, Praveen M, Narasimhan R, et al. Tear biomarkers in dry eye disease: Progress in the last decade. Indian J Ophthalmol 2023;71:1190-1202.

650. Jackson DC, Zeng W, Wong CY, et al. Tear Interferon-Gamma as a Biomarker for Evaporative Dry Eye Disease. Investigative ophthalmology & visual science 2016;57:4824-4830.

651. Chen Y, Dana R. Autoimmunity in dry eye disease - An updated review of evidence on effector and memory Th17 cells in disease pathogenicity. Autoimmunity reviews 2021;20:102933. 652. Kuklinski EJ, Yu Y, Ying GS, Asbell PA. Association of Ocular Surface Immune Cells With Dry Eye Signs and Symptoms in the Dry Eye Assessment and Management (DREAM) Study. Investigative ophthalmology & visual science 2023;64:7.

653. Roy NS, Yu Y, Ying GS, Maguire MG, Asbell PA. Effect of Omega-3 on HLA-DR Expression by Conjunctival Cells and Tear Cytokine Concentrations in the Dry Eye Assessment and Management Study. Eye Contact Lens 2022;48:384-390.

654. Willems B, Tong L, Minh TDT, Pham ND, Nguyen XH, Zumbansen M. Novel Cytokine Multiplex Assay for Tear Fluid Analysis in Sjogren's Syndrome. Ocular immunology and inflammation 2021;29:1639-1644.

655. Roda M, Corazza I, Bacchi Reggiani ML, et al. Dry Eye Disease and Tear Cytokine Levels-A Meta-Analysis. Int J Mol Sci 2020;21.

656. Perez VL, Stern ME, Pflugfelder SC. Inflammatory basis for dry eye disease flares. Experimental eye research 2020;201:108294.

657. VanDerMeid KR, Su SP, Ward KW, Zhang JZ. Correlation of tear inflammatory cytokines and matrix metalloproteinases with four dry eye diagnostic tests. Investigative ophthalmology & visual science 2012;53:1512-8.

658. Enriquez-de-Salamanca A, Castellanos E, Stern ME, et al. Tear cytokine and chemokine analysis and clinical correlations in evaporative-type dry eye disease. Molecular vision 2010;16:862-73.

659. Lee JH, Min K, Kim SK, Kim EK, Kim TI. Inflammatory cytokine and osmolarity changes in the tears of dry eye patients treated with topical 1% methylprednisolone. Yonsei medical journal 2014;55:203-8.

660. González-García MJ, Murillo GM, Pinto-Fraga J, et al. Clinical and tear cytokine profiles after advanced surface ablation refractive surgery: A six-month follow-up. Experimental eye research 2020;193:107976.

661. López-Miguel A, Tesón M, Martín-Montañez V, et al. Clinical and Molecular Inflammatory Response in Sjögren Syndrome-Associated Dry Eye Patients Under Desiccating Stress. American journal of ophthalmology 2016;161:133-41.e1-2.

662. Gad A, Vingrys AJ, Wong CY, Jackson DC, Downie LE. Tear film inflammatory cytokine upregulation in contact lens discomfort. The ocular surface 2019;17:89-97.

663. Chhadva P, Lee T, Sarantopoulos CD, et al. Human Tear Serotonin Levels Correlate with Symptoms and Signs of Dry Eye. Ophthalmology 2015;122:1675-80.

664. Zhang X, Yin Y, Yue L, Gong L. Selective Serotonin Reuptake Inhibitors Aggravate Depression-Associated Dry Eye Via Activating the NF-κB Pathway. Investigative ophthalmology & visual science 2019;60:407-419.

665. Dartt DA. Regulation of lacrimal gland secretion by neurotransmitters and the EGF family of growth factors. Experimental eye research 2001;73:741-52.

666. Lambiase A, Micera A, Sacchetti M, Cortes M, Mantelli F, Bonini S. Alterations of tear neuromediators in dry eye disease. Arch Ophthalmol 2011;129:981-6.

667. Yang LWY, Mehta JS, Liu YC. Corneal neuromediator profiles following laser refractive surgery. Neural Regen Res 2021;16:2177-2183.

668. Liu C, Lin MT, Lee IXY, et al. Neuropathic Corneal Pain: Tear Proteomic and Neuromediator Profiles, Imaging Features, and Clinical Manifestations. American journal of ophthalmology 2024;265:6-20.

669. Hernández-Molina G, Avila-Casado C, Nuñez-Alvarez C, et al. Utility of the American-European Consensus Group and American College of Rheumatology Classification Criteria for Sjögren's syndrome in patients with systemic autoimmune diseases in the clinical setting. Rheumatology (Oxford, England) 2015;54:441-8.

670. Loiseau P, Lepage V, Djelal F, et al. HLA class I and class II are both associated with the genetic predisposition to primary Sjögren syndrome. Human immunology 2001;62:725-31.

671. Kessal K, Liang H, Rabut G, et al. Conjunctival Inflammatory Gene Expression Profiling in Dry Eye Disease: Correlations With HLA-DRA and HLA-DRB1. Frontiers in immunology 2018;9:2271.
672. Roy NS, Wei Y, Yu Y, et al. Conjunctival HLA-DR Expression and Its Association With

Symptoms and Signs in the DREAM Study. Translational vision science & technology 2019;8:31.

673. Baudouin C, Liang H, Riancho L, et al. Correlation between the inflammatory marker HLA DR and signs and symptoms in moderate to severe dry eye disease. Investigative ophthalmology & visual science 2015;56:298-CO183.

674. Nair AP, D'Souza S, Khamar P, Nuijts R, Sethu S, Shetty R. Ocular surface immune cell diversity in dry eye disease. Indian J Ophthalmol 2023;71:1237-1247.

675. de Zoete MR, Palm NW, Zhu S, Flavell RA. Inflammasomes. Cold Spring Harb Perspect Biol 2014;6:a016287.

676. Yerramothu P, Vijay AK, Willcox MDP. Inflammasomes, the eye and anti-inflammasome therapy. Eye (Lond) 2018;32:491-505.

677. Zheng Q, Ren Y, Reinach PS, et al. Reactive oxygen species activated NLRP3 inflammasomes initiate inflammation in hyperosmolarity stressed human corneal epithelial cells and environment-induced dry eye patients. Experimental eye research 2015;134:133-40.

678. Niu L, Zhang S, Wu J, Chen L, Wang Y. Upregulation of NLRP3 Inflammasome in the Tears and Ocular Surface of Dry Eye Patients. PLoS One 2015;10:e0126277.

679. Zheng Q, Ren Y, Reinach PS, et al. Reactive oxygen species activated NLRP3 inflammasomes prime environment-induced murine dry eye. Experimental eye research 2014;125:1-8.

680. Tovar A, Gomez A, Serrano A, et al. Role of Caspase-1 as a Biomarker of Ocular Surface Damage. Am J Ophthalmol 2022;239:74-83.

681. Zhang J, Dai Y, Yang Y, Xu J. Calcitriol Alleviates Hyperosmotic Stress-Induced Corneal Epithelial Cell Damage via Inhibiting the NLRP3-ASC-Caspase-1-GSDMD Pyroptosis Pathway in Dry Eye Disease. J Inflamm Res 2021;14:2955-2962.

682. Hammond SM. An overview of microRNAs. Adv Drug Deliv Rev 2015;87:3-14.

683. Kim YJ, Yeon Y, Lee WJ, et al. Comparison of MicroRNA Expression in Tears of Normal Subjects and Sjogren Syndrome Patients. Invest Ophthalmol Vis Sci 2019;60:4889-4895.

684. Wang Q, Xie X, Li H, Hao S. Discovery of microRNA expression profiles involved in regulating TGF-beta2 expression in the tears of dry eye patients. Ann Clin Biochem 2020;57:420-428.

685. Pucker AD, Ngo W, Postnikoff CK, Fortinberry H, Nichols JJ. Tear Film miRNAs and Their Association With Human Dry Eye Disease. Curr Eye Res 2022;47:1479-1487.

686. Ren Y, Feng J, Lin Y, et al. MiR-223 inhibits hyperosmolarity-induced inflammation through downregulating NLRP3 activation in human corneal epithelial cells and dry eye patients. Exp Eye Res 2022;220:109096.

687. Chan HW, Yang B, Wong W, et al. A Pilot Study on MicroRNA Profile in Tear Fluid to Predict Response to Anti-VEGF Treatments for Diabetic Macular Edema. J Clin Med 2020;9.

688. Bu J, Liu Y, Zhang R, et al. Potential New Target for Dry Eye Disease-Oxidative Stress. Antioxidants (Basel) 2024;13.

689. Li B, Sheng M, Li J, et al. Tear proteomic analysis of Sjogren syndrome patients with dry eye syndrome by two-dimensional-nano-liquid chromatography coupled with tandem mass spectrometry. Sci Rep 2014;4:5772.

690. Kruk J, Kubasik-Kladna K, Aboul-Enein HY. The Role Oxidative Stress in the Pathogenesis of Eye Diseases: Current Status and a Dual Role of Physical Activity. Mini Rev Med Chem 2015;16:241-57.

691. Gaschler MM, Stockwell BR. Lipid peroxidation in cell death. Biochem Biophys Res Commun 2017;482:419-425.

692. Wakamatsu TH, Dogru M, Matsumoto Y, et al. Evaluation of lipid oxidative stress status in Sjogren syndrome patients. Invest Ophthalmol Vis Sci 2013;54:201-10.

693. Navel V, Sapin V, Henrioux F, et al. Oxidative and antioxidative stress markers in dry eye disease: A systematic review and meta-analysis. Acta Ophthalmol 2022;100:45-57.

694. Sedlak L, Swierczynska M, Borymska W, Zych M, Wygledowska-Promienska D. Impact of dorzolamide, benzalkonium-preserved dorzolamide and benzalkonium-preserved brinzolamide on selected biomarkers of oxidative stress in the tear film. BMC Ophthalmol 2021;21:319.

695. Choi JH, Li Y, Kim SH, et al. The influences of smartphone use on the status of the tear film and ocular surface. PLoS One 2018;13:e0206541.

696. Macri A, Scanarotti C, Bassi AM, et al. Evaluation of oxidative stress levels in the conjunctival epithelium of patients with or without dry eye, and dry eye patients treated with preservative-free hyaluronic acid 0.15 % and vitamin B12 eye drops. Graefe's archive for clinical and experimental ophthalmology = Albrecht von Graefes Archiv fur klinische und experimentelle Ophthalmologie 2015;253:425-30.

697. Choi W, Lian C, Ying L, et al. Expression of Lipid Peroxidation Markers in the Tear Film and Ocular Surface of Patients with Non-Sjogren Syndrome: Potential Biomarkers for Dry Eye Disease. Curr Eye Res 2016;41:1143-9.

698. Jung JH, Ji YW, Hwang HS, et al. Proteomic analysis of human lacrimal and tear fluid in dry eye disease. Sci Rep 2017;7:13363.

699. Xiao W, Huang P. Effects of the preoperative use of artificial tears combined with recombinant bovine basic fibroblast growth factor on cataract patients complicated with dry eyes. Arq Bras Oftalmol 2022;87:0539.

700. Jee D, Park M, Lee HJ, Kim MS, Kim EC. Comparison of treatment with preservative-free versus preserved sodium hyaluronate 0.1% and fluorometholone 0.1% eyedrops after cataract surgery in patients with preexisting dry-eye syndrome. J Cataract Refract Surg 2015;41:756-63.

701. Chen A, Chen HT, Hwang YH, Chen YT, Hsiao CH, Chen HC. Severity of dry eye syndrome is related to anti-dsDNA autoantibody in systemic lupus erythematosus patients without secondary Sjogren syndrome: A cross-sectional analysis. Medicine (Baltimore) 2016;95:e4218.

702. Ma W, Wang G, Li X, et al. Study of Factors Influencing Dry Eye in Rheumatoid Arthritis. J Ophthalmol 2020;2020:5768679.

703. Psianou K, Panagoulias I, Papanastasiou AD, et al. Clinical and immunological parameters of Sjogren's syndrome. Autoimmun Rev 2018;17:1053-1064.

704. D'Souza S, James E, Jois R, Mahendradas P, Koul A, Pradhan A. Systemic investigations in dry eye disease. Indian J Ophthalmol 2023;71:1326-1331.

705. Osei-Bimpong A, Meek JH, Lewis SM. ESR or CRP? A comparison of their clinical utility. Hematology 2007;12:353-7.

706. Crane AM, Feuer W, Galor A. C-reactive protein levels and tear function parameters. Ophthalmology 2013;120:1099 e1.

707. Hashemi MH, Ambrus JL, Jr., Shukla AA, et al. Association of Systemic Markers of Inflammation with Signs and Symptoms of Dry Eye Disease and Sjogren's Syndrome in the Dry Eye Assessment and Management (DREAM(c)) Study. Curr Eye Res 2024;49:574-581.

708. Da Cunha E, Mariette X, Desmoulins F, et al. Associations between ocular and extra-ocular assessment in primary Sjögren's syndrome. Joint Bone Spine 2022;89:105426.

709. Lim SA, Nam S, Kwok SK, Park SH, Chung SH. Serologic Markers Are Associated With Ocular Staining Score in Primary Sjogren Syndrome. Cornea 2015;34:1466-70.

710. Tsubota K, Fujihara T, Takeuchi T. Soluble interleukin-2 receptors and serum autoantibodies in dry eye patients: correlation with lacrimal gland function. Cornea 1997;16:339-44.

711. Ruiz de Morales JMG, Puig L, Dauden E, et al. Critical role of interleukin (IL)-17 in inflammatory and immune disorders: An updated review of the evidence focusing in controversies. Autoimmun Rev 2020;19:102429.

712. Oh JY, Kim MK, Choi HJ, et al. Investigating the relationship between serum interleukin-17 levels and systemic immune-mediated disease in patients with dry eye syndrome. Korean J Ophthalmol 2011;25:73-6.

713. Panpruk R, Puangsricharern V, Klaewsongkram J, et al. Clinical parameters and biological markers associated with acute severe ocular complications in Stevens-Johnson syndrome and toxic epidermal necrolysis. Scientific Reports 2021;11:20275.

714. Vehof J, Hysi PG, Hammond CJ. A Metabolome-Wide Study of Dry Eye Disease Reveals Serum Androgens as Biomarkers. Ophthalmology 2017;124:505-511.

715. Wang MTM, Dean SJ, Muntz A, Craig JP. Evaluating the diagnostic utility of evaporative dry eye disease markers. Clin Exp Ophthalmol 2020;48:267-270.

716. Ruiz-Lozano RE, Quiroga-Garza ME, Ramos-Dávila EM, et al. Comparative Evaluation of the Corneal Sensitivity Thresholds between the Novel Non-Contact and Cochet-Bonnet Esthesiometers. American journal of ophthalmology 2025;271:407-416.

717. Tavakoli M, Ferdousi M, Petropoulos IN, et al. Normative values for corneal nerve morphology assessed using corneal confocal microscopy: a multinational normative data set. Diabetes Care 2015;38:838-43.

718. Pflugfelder SC, de Paiva CS. The Pathophysiology of Dry Eye Disease: What We Know and Future Directions for Research. Ophthalmology 2017;124:S4-s13.

719. Chen Y, Chauhan SK, Lee HS, et al. Effect of desiccating environmental stress versus systemic muscarinic AChR blockade on dry eye immunopathogenesis. Investigative ophthalmology & visual science 2013;54:2457-64.

720. Usuba FS, Saad CGS, Aikawa NE, et al. Improvement of conjunctival cytological grade and tear production in Ankylosing Spondylitis patients under TNF inhibitors: a long-term follow-up. Sci Rep 2020;10:334.

721. Khokhar M, Dey S, Tomo S, Jaremko M, Emwas AH, Pandey RK. Unveiling Novel Drug Targets and Emerging Therapies for Rheumatoid Arthritis: A Comprehensive Review. ACS pharmacology & translational science 2024;7:1664-1693.

722. Kelagere Y, Scholand KK, DeJong EN, et al. TNF is a critical cytokine in age-related dry eye disease. The ocular surface 2023;30:119-128.

723. Mavragani CP, Moutsopoulos HM. Sjögren's syndrome: Old and new therapeutic targets. Journal of autoimmunity 2020;110:102364.

724. Shen L, Suresh L, Lindemann M, et al. Novel autoantibodies in Sjogren's syndrome. Clinical immunology (Orlando, Fla) 2012;145:251-5.

725. de Paiva CS, Trujillo-Vargas CM, Schaefer L, Yu Z, Britton RA, Pflugfelder SC. Differentially Expressed Gene Pathways in the Conjunctiva of Sjögren Syndrome Keratoconjunctivitis Sicca. Frontiers in immunology 2021;12:702755.

726. Pflugfelder SC, Bian F, Gumus K, Farley W, Stern ME, De Paiva CS. Severity of Sjögren's Syndrome Keratoconjunctivitis Sicca Increases with Increased Percentage of Conjunctival Antigen-Presenting Cells. Int J Mol Sci 2018;19.

727. Schaumberg DA, Buring JE, Sullivan DA, Dana MR. Hormone replacement therapy and dry eye syndrome. Jama 2001;286:2114-9.

728. Britten-Jones AC, Craig JP, Anderson AJ, Downie LE. Association between systemic omega-3 polyunsaturated fatty acid levels, and corneal nerve structure and function. Eye (Basingstoke) 2023;37.

729. Sağdık HM, Ugurbas SH, Can M, et al. Tear film osmolarity in patients with diabetes mellitus. Ophthalmic Res 2013;50:1-5.

730. Darenskaya MA, Kolesnikova LI, Kolesnikov SI. Oxidative Stress: Pathogenetic Role in Diabetes Mellitus and Its Complications and Therapeutic Approaches to Correction. Bulletin of experimental biology and medicine 2021;171:179-189.

731. Suwajanakorn O, Puangsricharern V, Kittipibul T, Chatsuwan T. Ocular surface microbiome in diabetes mellitus. Sci Rep 2022;12:21527.

732. Ma A, Mak MS, Shih KC, et al. Association of long-term glycaemic control on tear break-up times and dry eye symptoms in Chinese patients with type 2 diabetes. Clin Exp Ophthalmol 2018;46:608-615.

733. Pereira MV, Glória AL. Lagophthalmos. Semin Ophthalmol 2010;25:72-8.

734. Craig JP, Muntz A, Wang MTM, et al. Developing evidence-based guidance for the treatment of dry eye disease with artificial tear supplements: A six-month multicentre, double-masked randomised controlled trial. The ocular surface 2021;20:62-69.

735. Donthineni PR, Kammari P, Shanbhag SS, Singh V, Das AV, Basu S. Incidence, demographics, types and risk factors of dry eye disease in India: Electronic medical records driven big data analytics report I. The ocular surface 2019;17:250-256.

736. Watson SL, Chidi-Egboka NC, Khoo P, et al. Efficient capture of dry eye data from the real world: The Save Sight Dry Eye Registry. AJO International 2024;1:100065.

737. Zhang S, Echegoyen J. Design and Usability Study of a Point of Care mHealth App for Early Dry Eye Screening and Detection. Journal of clinical medicine 2023;12.

738. Shi Y, Wang L, Hu Y, et al. Contact lens sensor for ocular inflammation monitoring. Biosensors & bioelectronics 2024;249:116003.

739. Woods CA DK, Jones L and Fonn D. . Patient use of smartphones to communicate subjective data in clinical trials. Optom Vis Sci 2011;88:290-4.

740. Yang HK, Che SA, Hyon JY, Han SB. Integration of Artificial Intelligence into the Approach for Diagnosis and Monitoring of Dry Eye Disease. Diagnostics (Basel) 2022;12.

741. Neurotrophic Keratopathy Study Group. Neurotrophic keratopathy: An updated understanding. The ocular surface 2023;30:129-138.

742. Tamimi A, Sheikhzadeh F, Ezabadi SG, et al. Post-LASIK dry eye disease: A comprehensive review of management and current treatment options. Front Med (Lausanne) 2023;10:1057685.

743. Gupta PK, Drinkwater OJ, VanDusen KW, Brissette AR, Starr CE. Prevalence of ocular surface dysfunction in patients presenting for cataract surgery evaluation. J Cataract Refract Surg 2018;44:1090-1096.

744. Trattler WB, Majmudar PA, Donnenfeld ED, McDonald MB, Stonecipher KG, Goldberg DF. The Prospective Health Assessment of Cataract Patients' Ocular Surface (PHACO) study: the effect of dry eye. Clinical ophthalmology 2017;11:1423-1430.

745. Nosch DS, Käser E, Bracher T, Joos RE. Age-Related Changes in Corneal Sensitivity. Cornea 2023;42:1257-1262.

746. Zhao PF, Zhou YH, Hu YB, et al. Evaluation of preoperative dry eye in people undergoing corneal refractive surgery to correct myopia. International journal of ophthalmology 2021;14:1047-1051.

747. Woodward MA, Randleman JB, Stulting RD. Dissatisfaction after multifocal intraocular lens implantation. J Cataract Refract Surg 2009;35:992-7.

748. Yang F, Yang L, Ning X, Liu J, Wang J. Effect of dry eye on the reliability of keratometry for cataract surgery planning. J Fr Ophtalmol 2024;47:103999.

749. Nilsen C, Gundersen M, Jensen PG, et al. Effect of Artificial Tears on Preoperative Keratometry and Refractive Precision in Cataract Surgery. Clinical ophthalmology 2024;18:1503-1514.

750. Chuang J, Shih KC, Chan TC, Wan KH, Jhanji V, Tong L. Preoperative optimization of ocular surface disease before cataract surgery. J Cataract Refract Surg 2017;43:1596-1607.

751. Nibandhe AS, Donthineni PR. Understanding and Optimizing Ocular Biometry for Cataract Surgery in Dry Eye Disease: A Review. Semin Ophthalmol 2023;38:24-30.

752. Starr CE, Gupta PK, Farid M, et al. An algorithm for the preoperative diagnosis and treatment of ocular surface disorders. J Cataract Refract Surg 2019;45:669-684.

753. Castillo-Macías A, Arreola-Martínez JE, Loya-García D, Valdez-García JE. Prevalence and clinical characteristics of neurotrophic keratopathy in hispanic population in northeastern Mexico. International ophthalmology 2023;43:3257-3262.

754. Sambhi RS, Sambhi GDS, Mather R, Malvankar-Mehta MS. Dry eye after refractive surgery: a meta-analysis. Can J Ophthalmol 2020;55:99-106.

755. Shen Z, Zhu Y, Song X, Yan J, Yao K. Dry Eye after Small Incision Lenticule Extraction (SMILE) versus Femtosecond Laser-Assisted in Situ Keratomileusis (FS-LASIK) for Myopia: A Meta-Analysis. PLoS One 2016;11:e0168081.

756. Kobashi H, Kamiya K, Shimizu K. Dry Eye After Small Incision Lenticule Extraction and Femtosecond Laser-Assisted LASIK: Meta-Analysis. Cornea 2017;36:85-91.

757. Nair S, Kaur M, Sharma N, Titiyal JS. Refractive surgery and dry eye - An update. Indian J Ophthalmol 2023;71:1105-1114.

758. Miura M, Inomata T, Nakamura M, et al. Prevalence and Characteristics of Dry Eye Disease After Cataract Surgery: A Systematic Review and Meta-Analysis. Ophthalmol Ther 2022;11:1309-1332.

759. Lu Q, Lu Y, Zhu X. Dry Eye and Phacoemulsification Cataract Surgery: A Systematic Review and Meta-Analysis. Front Med (Lausanne) 2021;8:649030.

760. Qian L, Wei W. Identified risk factors for dry eye syndrome: A systematic review and metaanalysis. PLoS One 2022;17:e0271267.

761. Naderi K, Gormley J, O'Brart D. Cataract surgery and dry eye disease: A review. European journal of ophthalmology 2020;30:840-855.

762. Bonini S, Rama P, Olzi D, Lambiase A. Neurotrophic keratitis. Eye (Lond) 2003;17:989-95.

763. Dua HS, Said DG, Messmer EM, et al. Neurotrophic keratopathy. Progress in retinal and eye research 2018;66:107-131.

764. Mackie IA. Neuroparalytic keratitis. . In: Fraunfelder F, Roy FH, Meyer SM, editors. Current ocular therapy. Philadelphia, PA: Saunders, 1995.

765. Zieske JD, Gipson IK. Protein synthesis during corneal epithelial wound healing. Investigative ophthalmology & visual science 1986;27:1-7.

766. Cardona G, Seres C, Quevedo L, Auge M. Knowledge and use of tear film evaluation tests by spanish practitioners. Optom Vis Sci 2011;88:1106-11.

767. Belmonte C, Acosta MC, Schmelz M, Gallar J. Measurement of corneal sensitivity to mechanical and chemical stimulation with a CO2 esthesiometer. Investigative ophthalmology & visual science 1999;40:513-9.

768. Merayo-Lloves J, Gómez Martín C, Lozano-Sanroma J, Renedo Laguna C. Assessment and safety of the new esthesiometer BRILL: Comparison with the Cochet-Bonnet Esthesiometer. European journal of ophthalmology 2024;34:1036-1045.

769. Villalba M, Sabates V, Orgul S, Perez VL, Swaminathan SS, Sabater AL. Detection of Subclinical Neurotrophic Keratopathy by Noncontact Esthesiometry. Ophthalmol Ther 2024;13:2393-2404.

770. Swanevelder SK, Misra SL, Tyler EF, McGhee CN. Precision, agreement and utility of a contemporary non-contact corneal aesthesiometer. Clin Exp Optom 2020;103:798-803.

771. Dana R, Farid M, Gupta PK, et al. Expert consensus on the identification, diagnosis, and treatment of neurotrophic keratopathy. BMC Ophthalmol 2021;21:327.

772. Giovannetti F, Sacchetti M, Marenco M, et al. New disposable esthesiometer (KeraSense®) to improve diagnosis and management of neurotrophic keratitis. The ocular surface 2024;32:192-197.

773. Merayo-Lloves J, Gómez Martín C, Lozano-Sanroma J, Renedo Laguna C. Assessment and safety of the new esthesiometer BRILL: Comparison with the Cochet-Bonnet Esthesiometer. European journal of ophthalmology 2023:11206721231210754.

774. Crabtree JR, Tannir S, Tran K, Boente CS, Ali A, Borschel GH. Corneal Nerve Assessment by Aesthesiometry: History, Advancements, and Future Directions. Vision (Basel, Switzerland) 2024;8.

775. Rosenberg ME, Tervo TM, Immonen IJ, Müller LJ, Grönhagen-Riska C, Vesaluoma MH. Corneal structure and sensitivity in type 1 diabetes mellitus. Investigative ophthalmology & visual science 2000;41:2915-21.

776. Martone G, Alegente M, Balestrazzi A, et al. In vivo confocal microscopy in bilateral herpetic keratitis: a case report. European journal of ophthalmology 2008;18:994-7.

777. Patel DV, McGhee CN. Laser scanning in vivo confocal microscopy demonstrating significant alteration of human corneal nerves following herpes zoster ophthalmicus. Archives of neurology 2010;67:640-1.

778. Hamrah P, Sahin A, Dastjerdi MH, et al. Cellular changes of the corneal epithelium and stroma in herpes simplex keratitis: an in vivo confocal microscopy study. Ophthalmology 2012;119:1791-7.

779. Cruzat A, Qazi Y, Hamrah P. In Vivo Confocal Microscopy of Corneal Nerves in Health and Disease. The ocular surface 2017;15:15-47.

780. Hamrah P, Cruzat A, Dastjerdi MH, et al. Unilateral herpes zoster ophthalmicus results in bilateral corneal nerve alteration: an in vivo confocal microscopy study. Ophthalmology 2013;120:40-7.

781. Hamrah P, Cruzat A, Dastjerdi MH, et al. Corneal sensation and subbasal nerve alterations in patients with herpes simplex keratitis: an in vivo confocal microscopy study. Ophthalmology 2010;117:1930-6.

782. Cruzat A, Witkin D, Baniasadi N, et al. Inflammation and the nervous system: the connection in the cornea in patients with infectious keratitis. Investigative ophthalmology & visual science 2011;52:5136-43.

783. Guerrero-Moreno A, Baudouin C, Melik Parsadaniantz S, Réaux-Le Goazigo A. Morphological and Functional Changes of Corneal Nerves and Their Contribution to Peripheral and Central Sensory Abnormalities. Frontiers in cellular neuroscience 2020;14:610342.

784. McNally TW, Figueiredo FC. Corneal Neuropathic Pain: A Patient and Physician Perspective. Ophthalmol Ther 2024;13:1041-1050.

785. Galor A, Batawi H, Felix ER, et al. Incomplete response to artificial tears is associated with features of neuropathic ocular pain. The British journal of ophthalmology 2016;100:745-9.

786. Fernandez CA, Galor A, Arheart KL, et al. Dry eye syndrome, posttraumatic stress disorder, and depression in an older male veteran population. Investigative ophthalmology & visual science 2013;54:3666-72.

787. Galor A, Feuer W, Lee DJ, et al. Depression, post-traumatic stress disorder, and dry eye syndrome: a study utilizing the national United States Veterans Affairs administrative database. American journal of ophthalmology 2012;154:340-346 e2.

788. Segal BM, Pogatchnik B, Henn L, Rudser K, Sivils KM. Pain severity and neuropathic pain symptoms in primary Sjögren's syndrome: a comparison study of seropositive and seronegative Sjögren's syndrome patients. Arthritis Care Res (Hoboken) 2013;65:1291-8.

789. Dieckmann G, Goyal S, Hamrah P. Neuropathic Corneal Pain: Approaches for Management. Ophthalmology 2017;124:S34-s47.

790. Woltsche JN, Horwath-Winter J, Dorn C, et al. Neuropathic Corneal Pain as Debilitating Manifestation of LONG-COVID. Ocular immunology and inflammation 2023;31:1216-1218.

791. Costa Í F, Bonifácio LP, Bellissimo-Rodrigues F, et al. Ocular findings among patients surviving COVID-19. Sci Rep 2021;11:11085.

792. Yavuz Saricay L, Bayraktutar BN, Kenyon BM, Hamrah P. Concurrent ocular pain in patients with neurotrophic keratopathy. The ocular surface 2021;22:143-151.

793. Qazi Y, Hurwitz S, Khan S, Jurkunas UV, Dana R, Hamrah P. Validity and Reliability of a Novel Ocular Pain Assessment Survey (OPAS) in Quantifying and Monitoring Corneal and Ocular Surface Pain. Ophthalmology 2016;123:1458-68.

794. Farhangi M, Feuer W, Galor A, et al. Modification of the Neuropathic Pain Symptom Inventory for use in eye pain (NPSI-Eye). Pain 2019;160:1541-1550.

795. Goyal S, Hamrah P. Understanding Neuropathic Corneal Pain--Gaps and Current Therapeutic Approaches. Semin Ophthalmol 2016;31:59-70.

796. Crane AM, Feuer W, Felix ER, et al. Evidence of central sensitisation in those with dry eye symptoms and neuropathic-like ocular pain complaints: incomplete response to topical anaesthesia and generalised heightened sensitivity to evoked pain. The British journal of ophthalmology 2017;101:1238-1243.

797. Patel S, Mittal R, Sarantopoulos KD, Galor A. Neuropathic ocular surface pain: Emerging drug targets and therapeutic implications. Expert opinion on therapeutic targets 2022;26:681-695.

798. Ozmen MC, Dieckmann G, Cox SM, et al. Efficacy and tolerability of nortriptyline in the management of neuropathic corneal pain. The ocular surface 2020;18:814-820.

799. Aykut V, Elbay A, Çigdem Uçar I, et al. Corneal sensitivity and subjective complaints of ocular pain in patients with fibromyalgia. Eye (Lond) 2018;32:763-767.

800. Spierer O, Felix ER, McClellan AL, et al. Corneal Mechanical Thresholds Negatively Associate With Dry Eye and Ocular Pain Symptoms. Investigative ophthalmology & visual science 2016;57:617-25.

801. Aggarwal S, Kheirkhah A, Cavalcanti BM, et al. Autologous Serum Tears for Treatment of Photoallodynia in Patients with Corneal Neuropathy: Efficacy and Evaluation with In Vivo Confocal Microscopy. The ocular surface 2015;13:250-62.

802. Storås AM, Strümke I, Riegler MA, et al. Artificial intelligence in dry eye disease. The ocular surface 2022;23:74-86.

803. Shimizu E, Ishikawa T, Tanji M, et al. Artificial intelligence to estimate the tear film breakup time and diagnose dry eye disease. Scientific Reports 2023;13:5822.

804. Wang Y, Shi F, Wei S, Li X. A deep learning model for evaluating meibomian glands morphology from meibography. Journal of clinical medicine 2023;12

805. Wang J, Yeh TN, Chakraborty R, Yu SX, Lin MC. A Deep Learning Approach for Meibomian Gland Atrophy Evaluation in Meibography Images. Translational vision science & technology 2019;8:37.

806. Setu MAK, Horstmann J, Schmidt S, Stern ME, Steven P. Deep learning-based automatic meibomian gland segmentation and morphology assessment in infrared meibography. Sci Rep 2021;11:7649.

807. Yu Y, Zhou Y, Tian M, et al. Automatic identification of meibomian gland dysfunction with meibography images using deep learning. International ophthalmology 2022;42:3275-3284.

808. Saha RK, Chowdhury AMM, Na KS, et al. Automated quantification of meibomian gland dropout in infrared meibography using deep learning. The ocular surface 2022;26:283-294.

809. Swiderska K, Blackie CA, Maldonado-Codina C, Morgan PB, Read ML, Fergie M. A Deep Learning Approach for Meibomian Gland Appearance Evaluation. Ophthalmology science 2023;3:100334.

810. Wang S, He J, He X, et al. AES-CSFS: an automatic evaluation system for corneal sodium fluorescein staining based on deep learning. Therapeutic advances in chronic disease 2023;14:20406223221148266.

811. Nair PP, Keskar M, Borghare PT, Methwani DA, Nasre Y, Chaudhary M. Artificial Intelligence in Dry Eye Disease: A Narrative Review. Cureus 2024;16:e70056.

812. Huet E, Vallée B, Delbé J, et al. EMMPRIN modulates epithelial barrier function through a MMP-mediated occludin cleavage: implications in dry eye disease. The American journal of pathology 2011;179:1278-86.

813. Jamerson EC, Elhusseiny AM, ElSheikh RH, Eleiwa TK, El Sayed YM. Role of Matrix Metalloproteinase 9 in Ocular Surface Disorders. Eye Contact Lens 2020;46 Suppl 2:S57-s63.

814. Sivak JM, Fini ME. MMPs in the eye: emerging roles for matrix metalloproteinases in ocular physiology. Progress in retinal and eye research 2002;21:1-14.

815. Murphy G, Stanton H, Cowell S, et al. Mechanisms for pro matrix metalloproteinase
activation. APMIS : acta pathologica, microbiologica, et immunologica Scandinavica 1999;107:38-44.
816. Lu Z, Liu T, Zhou X, et al. Rapid and quantitative detection of tear MMP-9 for dry eye patients
using a novel silicon nanowire-based biosensor. Biosensors & bioelectronics 2022;214:114498.

817. McDevitt H, Munson S, Ettinger R, Wu A. Multiple roles for tumor necrosis factor-alpha and lymphotoxin alpha/beta in immunity and autoimmunity. Arthritis research 2002;4 Suppl 3:S141-52.

818. Körner H, Sedgwick JD. Tumour necrosis factor and lymphotoxin: molecular aspects and role in tissue-specific autoimmunity. Immunology and cell biology 1996;74:465-72.

819. Ruddle NH. Lymphoid neo-organogenesis: lymphotoxin's role in inflammation and development. Immunologic research 1999;19:119-25.

820. Howard Tripp N, Tarn J, Natasari A, et al. Fatigue in primary Sjögren's syndrome is associated with lower levels of proinflammatory cytokines. RMD open 2016;2:e000282.

821. Davies K, Mirza K, Tarn J, et al. Fatigue in primary Sjögren's syndrome (pSS) is associated with lower levels of proinflammatory cytokines: a validation study. Rheumatology international 2019;39:1867-1873.

822. Chen H, Chen H, Liang L, et al. Evaluation of Tear Protein Markers in Dry Eye Disease with Different Lymphotoxin-Alpha Expression Levels. American journal of ophthalmology 2020;217:198-211.

823. March de Ribot F, Benitez Del Castillo JM, Geerling G, Messmer EM, Baudouin C, Alves M. Dry eye disease sustainability. The ocular surface 2023;30:104-106.

824. Nasa P, Jain R, Juneja D. Delphi methodology in healthcare research: How to decide its appropriateness. World J Methodol 2021;11:116-129.

825. Graham B, Regehr G, Wright JG. Delphi as a method to establish consensus for diagnostic criteria. J Clin Epidemiol 2003;56:1150-6.

826. Mathewson PA, Williams GP, Watson SL, Hodson J, Bron AJ, Rauz S. Defining Ocular Surface Disease Activity and Damage Indices by an International Delphi Consultation. The ocular surface 2017;15:97-111.

827. Baudouin C, Aragona P, Van Setten G, et al. Diagnosing the severity of dry eye: a clear and practical algorithm. The British journal of ophthalmology 2014;98:1168-76.

828. Barabino S, Aragona P, di Zazzo A, Rolando M, with the contribution of selected ocular surface experts from the Societa Italiana di Dacriologia e Superficie O. Updated definition and classification of dry eye disease: Renewed proposals using the nominal group and Delphi techniques. European journal of ophthalmology 2021;31:42-48.

829. Farid M, Ayres BD, Donnenfeld E, et al. Delphi Panel Consensus Regarding Current Clinical Practice Management Options for Demodex blepharitis. Clinical ophthalmology 2023;17:667-679.

**Conflict of Interest** 

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.