

Assembling evidence for identifying reservoirs of infection

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Many pathogens persist in multihost systems, making the identification of infection reservoirs crucial for devising effective interventions. Here, we present a conceptual framework for classifying patterns of incidence and prevalence, and review recent scientific advances that allow us to study and manage reservoirs simultaneously. We argue that interventions can have a crucial role in enriching our mechanistic understanding of how reservoirs function and should be embedded as quasi-experimental studies in adaptive management frameworks. Single approaches to the study of reservoirs are unlikely to generate conclusive insights whereas the formal integration of data and methodologies, involving interventions, pathogen genetics, and contemporary surveillance techniques, promises to open up new opportunities to advance understanding of complex multihost systems.

Advancing our understanding of reservoirs

Most disease-causing organisms, including many important human, livestock, and wildlife pathogens, are capable of infecting multiple hosts [1–3]. Therefore, determining how hosts enable persistence [4] and which hosts are crucial for the persistence of these multihost pathogens [5] is essential for the design of effective control measures. Failure to establish this understanding can hamper policy formulation and lead to ineffective or counter-productive control measures with costly implications for socially, economically, or ecologically important populations.

Reservoirs of infection can be ecologically complicated structures comprising one or more interacting populations or species (Box 1 [5]). Although a range of developments has led to better theoretical conceptualisation of reservoirs [5–9], their empirical characterisation remains a challenge. In this article, we review methods currently used to characterise each of the components that comprise a reservoir according to the framework in Box 1. Specifically, we first present a conceptual approach for classifying

patterns of incidence and prevalence (see Glossary) that result from the connectivity between source and target populations (black arrows in Figure I in Box 1). We then review methods that allow us to identify maintenance or nonmaintenance populations (squares or circles in Figure I, Box 1), how they are connected (arrows in Figure I, Box 1), and the role that each of these populations has in maintaining the pathogen (i.e., reservoir capacity).

Long-term ecological data on multihost systems are sparse and challenging to collect [10–12]; this, combined with the inherent difficulty of identifying reservoirs of infection, means that each data set or approach in isolation is unlikely to result in a sufficient evidence base to inform control strategies. Here, we further discuss how to enrich

Glossary

Basic reproduction number (R_0): expected number of secondary cases caused by a single infectious individual in a fully susceptible population.

Critical community size (CCS): host population size below which a disease cannot persist in the long term.

Effective reproductive number: expected number of secondary cases caused by each infectious individual in a partially immune population.

Endemic: an infection is endemic in a population when it is maintained without the need for external introductions.

Force of infection: hazard rate of infection from a defined source to susceptible host individuals in a defined population.

Incidence: number of new cases in a particular time interval.

Maintenance community: any set of connected host (sub)populations that together can maintain a pathogen over the long term. A minimal maintenance community is a maintenance community of which all subsets are nonmaintenance. Trivially, a maintenance population is also a (minimal) maintenance community.

Maintenance population: single host population capable of maintaining a pathogen over the long term.

Metapopulation: set of populations that are connected by transmission. It can comprise structured populations of the same species (e.g., in space), populations of different species, or a combination thereof.

Patch value: measure of the contribution of individual populations to the reservoir capacity of a metapopulation.

Prevalence: proportion of positive cases in a population at a particular time point.

Reservoir capacity: measure of the potential of a host metapopulation to support long-term pathogen persistence in the absence of external imports.

Reservoir of infection: one or more epidemiologically connected populations or environments in which a pathogen can be permanently maintained and from which infection is transmitted to the target population.

Stuttering chain: pattern of cases in the form of short chains when transmission among hosts occurs but is too weak to support endemic or epidemic transmission.

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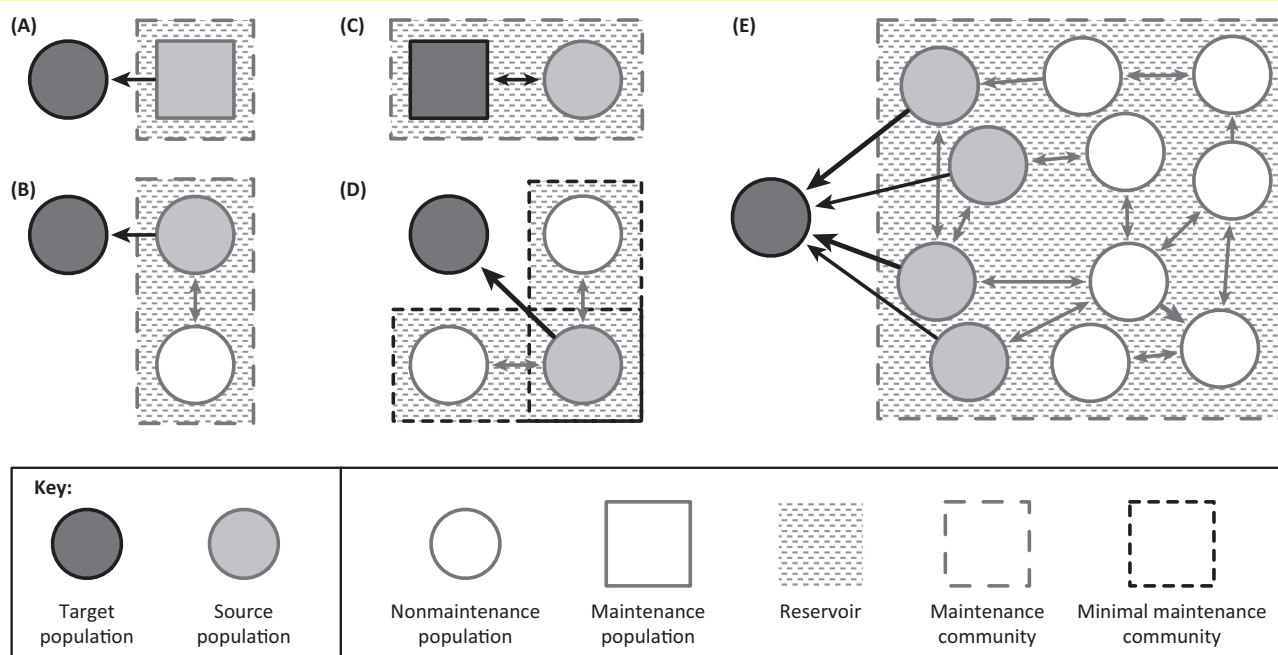
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Box 1. Disease reservoirs framework

Our study of epidemiology is usually motivated by the need to control disease in a particular host population or a subset of a population. Following Haydon *et al.* [5], we refer to this as the ‘target population’. Populations that are direct sources of infection for the target are termed ‘source populations’. A ‘reservoir of infection’ is defined with respect to a target population as ‘one or more epidemiologically connected populations or environments in which a pathogen can be permanently maintained and from which infection is transmitted to the target population’ [5]. Some reservoirs can be simple and comprise a single nontarget host population (Figure 1A). However, they can comprise a more structured set of connected host subpopulations termed ‘maintenance community’ (Figure 1B–D). Individually, some of these populations can maintain the pathogen (‘maintenance populations’), whereas others cannot (‘nonmaintenance populations’).

Thus, infection reservoirs can be constituted in a variety of ways. Reservoirs can be wildlife species [e.g., possums (*Eichosurus vulpecula*) as a reservoir of bovine TB in cattle in New Zealand; or wildebeest (*Connochaetes taurinus*) as a reservoir of malignant catarrh fever for cattle in Tanzania]; domesticated species (e.g., dogs as a reservoir of rabies for humans in many developing countries; cattle as a reservoir of *Escherichia coli* 0157 for humans in the UK), or subsets of the same species (e.g., adults as a reservoir of respiratory syncytial virus for children, men as an element of the reservoir of human papillomavirus for women).

Other definitions of reservoirs have been proposed [7,55]. Although Ashford’s [7] definition is appealing for its generality, and Drexler *et al.*’s [55] for its evolutionary perspective, we use Haydon *et al.*’s [5] due to not only its acceptance within the epidemiological literature, but also its direct application for designing interventions.



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Figure 1. Illustrative examples of reservoir–target systems. Arrow thickness denotes rate of transmission. In (A), the reservoir comprises a single source maintenance population that transmits to a nonmaintenance target population. In (B), the reservoir comprises two connected nonmaintenance populations (of which one is the source) that together form a maintenance community. In (C), the target is a maintenance population and a source of infection and, thus, is part of the reservoir. In (D), the reservoir comprises three nonmaintenance populations, together forming two minimal maintenance communities each capable of maintaining the pathogen; together, these form a larger maintenance community. In (E), the reservoir comprises a maintenance community of multiple connected nonmaintenance populations, four of which are source populations. Modified from [5].

this evidence base. Almost inevitably, the need to intervene will precede adequate understanding of the dynamics of reservoir–target systems. Our central thesis is that interventions that are meticulously planned to optimise both the immediate short-term benefits to the target population and the longer-term understanding of how reservoirs function, applied together with a formal integration of data and methods [13], can provide powerful new opportunities for studying complex multihost systems (e.g., [14]).

Patterns of incidence and prevalence in the target

Data on patterns of incidence and prevalence provide indirect information on the connectivity between source and target populations (i.e., black arrows in Figure 1, Box 1). Building upon the ‘community–epidemiology continuum’ framework developed by Fenton and Pedersen [15], specific

patterns can be assigned to ‘zones’ (Figure 1 and Table 1) defined in relation to the relative magnitudes of the force of infection from one or more source(s) (x-axis in Figure 1; thickness of arrows in Figure 1, Box 1), and $R_{0,T}$, the basic reproduction number of the pathogen within the target.

If the target population is a ‘dead-end’ host from which transmission does not occur, then $R_{0,T} = 0$. For a sufficiently low force of infection, the interval between cases in the target host is longer than the infectious period of single cases (Figure 1, zone A) and cases are not directly linked. As the force of infection from alternative sources increases, we observe cases in the target population with increasing frequency. At higher values, cases can overlap in time and space but remain epidemiologically unlinked and, as long as variability in the pathogen is high enough, genetically distinct (Figure 1, zone B).

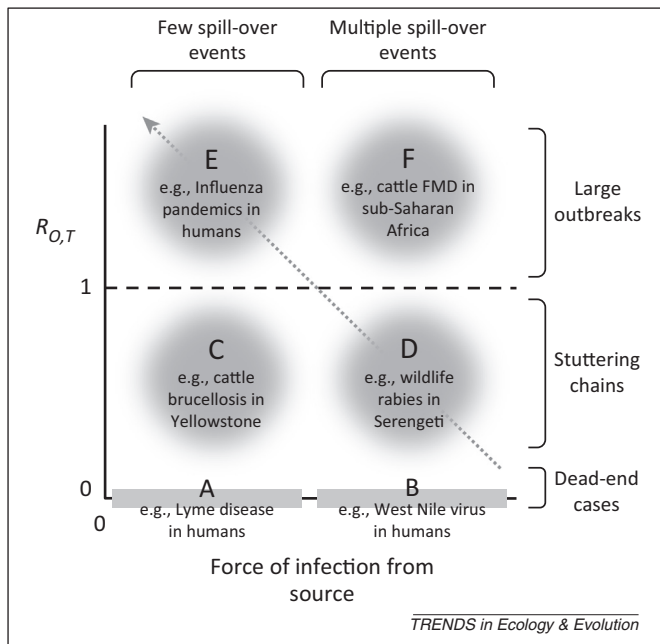


Figure 1. Illustration of disease zones characterised by low and high frequencies and/or rates of transmission from an external source of infection (force of infection, x-axis) and target-to-target transmission represented here by the basic reproduction number in the target population ($R_{0,T}$; y-axis). We note that the source of infection can be a reservoir, a maintenance population, or a nonmaintenance population. Further details of the dynamic and genetic signatures of each zone are provided in Table 1 (main text).

Target populations in which limited transmission can occur but $R_{0,T} < 1$ will, when the force of infection from the source is low, exhibit the classic ‘stuttering chain’ dynamics (Figure 1, zone C) in which outbreak sizes follow an overdispersed distribution [16]. As $R_{0,T} \rightarrow 1$, these outbreaks can become large. However, as the interval between introductions becomes shorter than the average duration of outbreaks, we observe a pseudo-endemic pattern in which the target population can appear to be a maintenance population even when it is not (Figure 1, zone D). Systems in which $R_{0,T}$ is close to 1 present particular threats because small changes in their epidemiology within the target population, through either pathogen evolution or changes in the target population structure, can cause $R_{0,T}$ to exceed 1 and lead to an endemic situation and/or epidemic behaviour [17].

If $R_{0,T} > 1$ then any spill-over events can give rise to substantial epidemics. Stochastic extinction will still occur frequently if $R_{0,T}$ is only slightly greater than 1 (Figure 1, zone E); however, if the outbreak ‘takes off’ or $R_{0,T} \gg 1$, then there are three broad possible outcomes: (i) the target population sustains a major epidemic after which the pathogen becomes extinct in the target population [e.g., distemper virus in wolves (*Canis lupus*) and harbour seals (*Phoca vitulina*)]; (ii) the target population sustains a major epidemic after which the pathogen proceeds towards an endemic state in the target population (e.g., HIV; the target population is then a square in Figure 1, Box 1); (iii) control measures within the target population reduce $R_{0,T}$ to below 1, so a major epidemic is averted and the pathogen becomes extinct in the target population (e.g., severe acute respiratory syndrome). If $R_{0,T} > 1$ and the force of infection

from the reservoir is large (Figure 1, zone F), fadeout is unlikely (e.g., Southern African Territories strains of foot-and-mouth disease in cattle in sub-Saharan Africa).

Dynamics ranging from pseudo-endemicity to true endemicity lie on an ascending diagonal from right to left (Figure 1, arrow), along which increasing $R_{0,T}$ compensates for a declining force of infection from the reservoir. These different situations are likely to be hard to distinguish using patterns of incidence and prevalence alone. However, higher resolution spatiotemporal data and pathogen genetic sequence data, together with sophisticated analytical techniques such as state-space modelling, can provide some of the necessary tools to examine these patterns (See ‘Connectivity within the reservoir’).

Analysis of serology data

Given the challenges of isolating pathogens from wildlife populations, patterns of incidence and prevalence are typically obtained from longitudinal seroprevalence surveys or age-seroprevalence curves. These have been used to investigate infection dynamics of various multihost systems, such as canine distemper virus (CDV) in carnivore communities of the Serengeti [18,19], Kenya [20], and Yellowstone [21], *Trypanosoma cruzi* in wildlife hosts in the USA [22] and hepatitis E in wild boars (*Sus scrofa*) in Europe [23]. However, their interpretation remains fraught with uncertainties mainly owing to cross-reactivity, declining antibody titres, cut-off thresholds used to distinguish positive and negative reactions, and difficulties with the detectability of antibodies because these depend on the relation between immunity and infection resistance (e.g., a detectable antibody does not always imply protection and the time of exposure remains unknown for pathogens that create life-long immunity in the host) [24]. New statistical approaches, such as latent class methods and site-occupancy modelling, have been suggested recently to improve estimates of prevalence from imperfect tests by allowing uncertainty in the detection of infection state [25]. Although still in early stages of development, advanced modelling techniques, such as Bayesian process models, can enable inferences of timing of exposure from age-seroprevalence data, accounting for non-stationary epidemiological dynamics [26], and/or detect cross-species transmission [27], to identify which host species is the most likely source of infection.

Identifying maintenance populations

Methods to identify plausible reservoirs typically focus on thresholds that define individual populations as maintenance or nonmaintenance (squares or circles in Figure 1, Box 1). Therefore, we discuss critical community size (CCS) as an intuitive measure of persistence that can be traced back to the reservoir framework proposed by Haydon *et al.* [5].

CCS can be loosely defined as the host population size below which a pathogen cannot persist [28,29]. Thus, a maintenance population can be defined as a host population in which a pathogen persists because the population size is greater than CCS, whereas a nonmaintenance population is one smaller than CCS [5]. However, there are several challenges to the study of CCS in practice.

Table 1. Description of the dynamics and genetic signature of each disease zone captured in Figure 1 (main text)

Zone	Process	Observation		Example
		Dynamics	Genetics	
A	Low frequency of spillover infection with no onward transmission in the target population. Low incidence with isolated, epidemiologically independent cases	Low incidence with long gaps between outbreaks that exceed the average combined incubation and infection periods	Genetic independence between cases	Lyme disease in humans Human rabies
B	Frequent, dead-end spillover leads to cases at a rate that could appear to indicate target-to-target transmission (but it is not)	Sometimes low incidence with frequent outbreaks (e.g., West Nile virus in humans). However, low frequency with high incidence can also occur (e.g., Rift Valley fever in humans)	Genetic independence between cases allows distinction from zones with similar incidence rates arising from target-to-target transmission (zones C/D)	West Nile virus in humans and horses Rift Valley fever in humans Wildebeest-associated malignant catarrhal fever in cattle Vampire bat rabies in humans and/or livestock
C	Limited target-to-target transmission causes isolated stuttering chains of transient nature and, thus, self-limiting outbreaks	Low-to-medium incidence with frequent small outbreaks	Genetics reveals that stuttering chains are unlinked based on cases having shared ancestry only in the distant past. Critical to distinguish from zones B/D	Monkeypox Cattle brucellosis in Yellowstone Early severe acute respiratory syndrome H5N1 avian influenza Food-borne <i>Escherichia coli</i>
D	Similar dynamics to zone C but chains initiated at high enough frequency to create a pseudo-endemic pattern (i.e., cases are always present in the target population)	Medium-to-high incidence with frequent small outbreaks. Reveals pseudo-endemicity	Genetics reveal that chains are separate and temporally superimposed (rather than linked), showing frequent transmission from source. Critical to distinguish from zone C	Wildlife CDV in the Serengeti Possibly TB in African lions Wildlife rinderpest (but see [78])
E	Rare introductions that result in large and usually sustained outbreaks due to $R_{0,T} > 1$. Size of target population is important because higher $R_{0,T}$ leads to a faster depletion of susceptibles, increasing the CCS required for persistence	High incidence with endemic circulation influenced by, for example, seasonal dynamics	Invasion can be traced to a single or a small number of spillover events	HIV Influenza in humans <i>Mycoplasma ovipneumonia</i> in bighorn sheep Bat rabies in skunks
F	Frequent introductions and large outbreaks associated with a high number of spillover events. Difficult to identify dynamically. Contribution from source unclear due to high $R_{0,T}$ in target population	High incidence	Genetics reveal multiple co-circulating lineages in the target population, with new lineages appearing through spillover events. Multiple spillovers from the source mean that it is more difficult to eliminate	Southern African Territories strains of cattle foot-and-mouth disease in sub-Saharan Africa Bovine TB in UK Jackal-dog rabies in southern Africa

The first challenge is to define the population in which persistence is to be measured. Given that persistence is sensitive to the complex relation between demographic and epidemiological factors, it is difficult to estimate in the presence of population structure [30,31]; therefore, CCS is most commonly discussed in the context of single well-mixed populations, although these are rare in natural systems. The second challenge is in defining persistence, particularly because any estimate of CCS is likely to be sensitive to the choice of persistence metric [32]. In his original work, Bartlett defined CCS as the size of a population in which extinction was as likely as not following a major outbreak [29]. However, persistence might also be measured from an initial condition corresponding to the endemic equilibrium [33,34]. Lloyd-Smith *et al.* [35] point out that the relation between persistence and population size is not well described as a step function, but instead increases in a gradual manner. CCS can also be thought of in relation to the probability of extinction within a given time or the time until a given proportion (usually 50%) of introductions (or simulations) have gone

extinct [36]. Once appropriate definitions are adopted, the final challenge is estimating CCS. The main approaches used are: (i) empirical observation, which consist of plotting incidence data against population size [28,37–39]; (ii) analytic expressions [34,40–42], although all approximations so far exclude many processes relevant to CCS, such as latency, spatial heterogeneity, seasonality, age structure, and non-exponential infectious periods [40,43]; and (iii) stochastic computer simulations, in which parameterised compartmental models are used to generate distributions of persistence times for populations of different sizes and from which CCS can be estimated [33,34]. However, these studies assume a linear relationship between population size and recruitment, which is unrealistic in natural systems [35]. Beyond studies of measles [44–46], little work has been done to estimate CCS.

Next-generation matrix (NGM) methods have also been used to identify reservoir communities from endemic prevalence data. This method estimates a threshold that is similar to R_0 (or the effective reproduction number in the endemic case) separately for individual host populations

within a multigroup population rather than averaging across all populations [47]. For example, using NGM, mallards and other dabbling ducks were reported to be part of the most likely reservoir community of influenza A in the global water bird population [8]. However, this method focuses strictly on whether persistence is possible (i.e., whether the appropriate reproductive number exceeds 1), and ignores the stochasticity and nonequilibrium dynamics that are central to classical thinking on CCS. Also, implementations of the method have relied on the strong assumption that infection prevalence is at endemic equilibrium in all host species.

Connectivity within the reservoir: tracing transmission

Identifying which populations constitute the reservoir requires understanding how the populations are epidemiologically connected to each other. Here, we discuss two approaches used to trace transmission within the reservoir and between the reservoir and a target population: simulations and modelling, and genetics. Evidence to test the hypothesis that a particular population is a source of infection can also be acquired through real-world interventions that either reduce prevalence of disease in the putative source or block source-to-target transmission while monitoring incidence in the target. These are discussed in a subsequent section.

Tracing transmission using simulations and modelling

Statistical modelling is increasingly used to identify plausible sources of infection. One of the advantages of modelling is that they can be used for partially observed processes [e.g., approximate Bayesian computation [48,49], state-space models (SSM; [50]), and Markov models [50–52]]. For example, SSMs make an explicit distinction between data that can be observed (e.g., infected individuals detected by surveillance) and the underlying process itself, which might be largely unobserved (e.g., all infection events). Beyer *et al.* [50] constructed an SSM of rabies persistence in the Serengeti District in Tanzania that used records of humans reporting to hospital with dog bite injuries. Using a state-space implementation of a metapopulation process describing the unobserved process of dog-to-dog transmission between villages, they were able to estimate parameters capturing the effects of intervillage distance and the size of dog populations on rabies dynamics. Based on these, they inferred that it was more likely that dog rabies infections were being imported from unvaccinated domestic dogs in outlying districts, or from wild peri-urban carnivores in the Serengeti district itself, rather than from wildlife residing within the National Park. Despite their advantages, inferences rely on valid assumptions being made about the biological processes embodied in model structure.

Given a possible set of transmission parameters, plausible reservoirs of infection can also be identified using simulation models [43,53,54]. Transmission parameters are typically obtained from epidemiological, demographic, or genetic data, and can be manipulated to explore the sensitivity of the reservoir dynamics to these parameters. For example, Cross *et al.* [54] used an age-structured model of two interacting elk populations (free-ranging and those receiving supplemental feeding) to investigate the extent to which dispersal from feeding grounds could explain

changes in brucellosis seroprevalence in elk around the Yellowstone ecosystem. They found that R_0 in the free-ranging elk population (the target population) had increased to above 1 over the past 20 years, probably due to changes in elk aggregations that led to enhanced elk-to-elk transmission (i.e., moved from zone C to E in Figure 1).

Tracing transmission using genetics

Genetic inference of cross-species transmission has so far tended to borrow analytical approaches from population genetics and phylogeography [55–57] (Box 2). Genealogy-based methods have particular appeal because, for many pathogens, the accumulation of mutations takes place on approximately the same timescale as transmission. If transmission chains are genetically distinguishable, they can provide complementary information to incidence and prevalence data. For example, given sufficient pathogen genetic variability in the reservoir, genetic data should readily distinguish rare spillover and subsequent transmission in the target from scenarios involving the same incidence due only to a high

Box 2. Using pathogen genetics to untangle reservoir–target dynamics

Methods based on discrete ancestral state inference offer an appealing statistical way to approach the problem of multihost transmission by fitting probabilistic models to pathogen sequence data [79]. In these methods, genealogies are constructed from the data, and host associations (states) observed at the tips of the trees are used to estimate the conditional probability of being affiliated with a particular host population along all interior branches. Transmission events between host populations and, thus, the net contribution of the reservoir to dynamics in the target, can be enumerated through Markov jump counts [56,80]. A more formal population genetic framework, centred on joint estimation of population sizes and migration rates across all patches [81,82], can similarly be adapted to deal with pathogen gene flow [83].

Although novel approaches based on genetic data open up intriguing opportunities, their resolution has defined limits. As introductions into the target become more frequent, it is increasingly difficult to distinguish contributions of the reservoir from continuous transmission within the target. Increasing genetic resolution by using longer sequences can compensate for this, but only to the point of sequencing entire pathogen genomes. The ability of genetic markers to resolve cross-species transmission processes will also be reduced by potential pathogen flow from the target population back into the reservoir [84].

Genetic inference of reservoir–target dynamics has so far also received little formal testing. There is strong reason to suspect that biased sampling can have a profound influence on the inferences generated. For example, genetically unsampled sources of infection will not only remain undetectable, but their contribution will also be wrongly attributed to populations included in the sample (a problem akin to that caused by ‘ghost populations’ in population genetics [85]). Moreover, inferred transmission dynamics can depend on the relative spatial and temporal density of sampling among host populations. Finally, stuttering chains within the target population can boost the frequency of cases (compare zones A and C in Figure 1, main text) and, hence, the likelihood of detection under sparse sampling; if unaccounted for, this can lead to overestimation of cross-species transmission rates. These complexities can generate significant challenges for the investigation of reservoir–target systems because balanced, representative sampling, proportional to the incidence in each host, is rarely achievable. Therefore, developing robust ways to deal with problems related to sampling in the context of genealogical inference and the reconstruction of transmission histories remains an important focal area for future research.

force of infection from outside the target population (Figure 1, zones B and C, and zones D and E) [57,58].

Reservoir capacity

A reservoir can comprise multiple connected populations of the same or different species (see Figure 1E in Box 1), and, thus, can be represented as a metapopulation. To assess whether this metapopulation is capable of supporting pathogen persistence, we can draw a parallel with ecological theory. Representing the reservoir as a metapopulation, we can extend the notion of metapopulation capacity [59] to that of reservoir capacity (Box 3). Reservoir capacity

ty, λ'_M , is a measure of the potential of a structured host population to support pathogen persistence in the absence of external imports and, thus, can be used to assess whether a population constitutes a reservoir. A useful benefit is that associated patch values V_i (i.e., the relative contribution of each population to overall persistence; Box 3) can be used to prioritise populations when designing interventions. The modelling framework encapsulates three processes (within-population processes, transmission between populations, and community-level persistence) and is normally used to investigate one of these processes when it is possible to parameterise the other two.

Box 3. Reservoir capacity of a metapopulation

The notion of 'metapopulation capacity' [59] captures in a single number λ'_M the capacity of a fragmented landscape, comprising patches, to support the long-term persistence of a species in the absence of external imports. By analogy, we define 'reservoir capacity' as the capacity of a metapopulation to support the long-term persistence of a pathogen. It can be regarded as a measure of effective host abundance, weighted to take into account structural factors, such as local population sizes and connectivity, that influence fadeout rates within populations and transmission between them.

The dynamics of a general metapopulation are governed by Equation I (Figure I), in which invasion and fadeout rates are functions of infection status of other populations, as well as factors such as population sizes and transmission rates. Persistence in the metapopulation is controlled by the ratio of population invasion events to disease fadeouts, and these are balanced at equilibrium. Reservoir capacity for this general model is defined in Equation II (Figure I). Reservoir capacity also suggests a persistence threshold. In this deterministic model, a pathogen persists in a given landscape if and only if $\lambda'_M > 1$, corresponding to the threshold above which Equation I has a stable nontrivial equilibrium. Although parameterising this kind of metapopulation model is challenging, methodology shared between ecologists and epidemiologists [86] can allow assessments of likely persistence.

An attractive feature of this approach is the ability to estimate patch values, V_i , which are measures of the contribution of individual populations to the persistence threshold that is used to guide interventions. For example, Figure II shows the relative contribution

of 75 villages in the Serengeti District, Tanzania, to local rabies reservoir capacity, estimated from the Beyer *et al.* [50,87] model discussed in the main text. Although the village shaded in red is the biggest both in terms of size and patch value, for villages with approximately 400 dogs, the patch values range from 0.00 to 0.04 depending on their spatial location relative to other villages (Figure II(B)).

$$\frac{dp_i}{dt} = C_i(\mathbf{p})(1 - p_i) - p_i E_i(\mathbf{p}) \quad \text{[I]}$$

Rate of change in p_i , the probability that infection is present in population i

Invasion rate C_i of population i by infection from other populations $\mathbf{p}$ when infection absent

Fadeout rate E_i of pathogen in population i when infection present

$$\lambda'_M := \sup_{\mathbf{p} \in \Omega} \left(\min_i \left[\frac{(1 - p_i) C_i(\mathbf{p})}{p_i E_i(\mathbf{p})} \right] \right) \quad \text{[II]}$$

Persistence is possible if and only if $\lambda'_M > 1$, i.e., there exists a vector $\mathbf{p}$ (in the set of all such vectors Ω) such that the minimum expected ratio of invasions to fadeouts across populations > 1

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Figure I. Metapopulation model and definition of reservoir capacity.

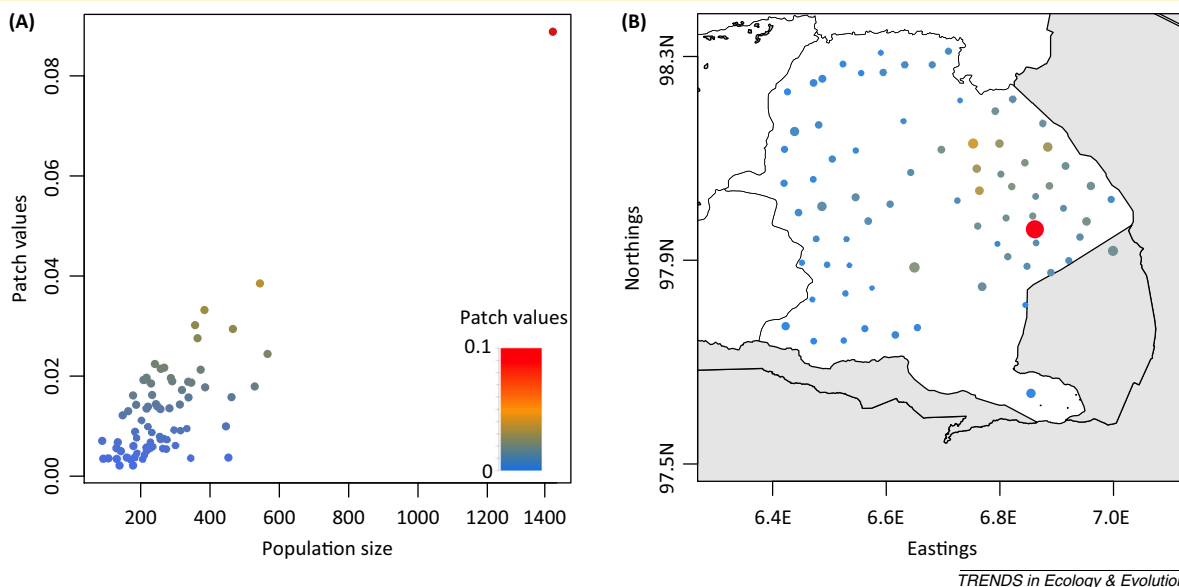


Figure II. Dog population size against relative patch values (A) estimated for rabies in 75 villages of the Serengeti District, Tanzania, and their geographic location (B). Colour gradient represents patch values V_i and circle sizes (B) are proportional to dog population size.

Enriching the evidence base

No one line of evidence is likely to support unambiguous inferences about the structure and functioning of a reservoir system. However, two general strategies are worth emphasising: interventions embedded into adaptive management and data integration.

Interventions embedded into adaptive management programs

Interventions are normally designed to maximise the benefits of disease control [60]. However, they must often be devised with an incomplete understanding of the overall disease dynamics. We argue that using interventions as quasi-experiments can provide valuable opportunities to learn more about the functioning of a reservoir (see examples in Table 2). Through adaptive management [61,62], disease control objectives can be met while generating and enriching the evidence base to improve future control policies and resource allocation.

Interventions that generate substantial (and, thus, more easily measurable) changes to the system are the most likely to provide useful information. Such interventions can be akin to ‘press’ (sustained action; e.g., long-term vaccination)

or ‘pulse’ (one-off action; e.g., single culling or vaccination campaign) phenomena that are familiar to ecologists [63], or ‘block’ perturbations (where potential transmission between reservoir and target is impeded; e.g., fences). Power can be assessed from predictions based on classical sensitivity or elasticity analysis [64]. Interventions that induce no changes can also be informative if, for example, they allow us to rule out a particular transmission route or source population. What can be learned from such interventions is dependent on whether they enable relevant hypotheses to be tested. It is also important to note that these interventions can alter the target-reservoir transmission dynamics, leading to difficulties in distinguishing causes and effects of the intervention. For example, the Randomised Badger Culling Trial conducted over 3000 km² and a 10-year time period [65] generated a wealth of data and analysis that should be instrumental in determining the circumstances in which badger culling might usefully contribute to the effective control of bovine TB in the UK. However, it might not be simple to determine whether particular changes in the reservoir–target disease transmission dynamics were a direct or indirect (due to dispersal and behaviour change) consequence of the culling.

Table 2. Intervention studies shedding light on maintenance host status

Pathogen	Focal (other) species	Location	Type of intervention: intervention	Outcome	Is focal species a maintenance host?	Refs
<i>Mycobacterium bovis</i>	Wild boar (red deer)	Spain	Block: isolated and/or fenced from livestock hosts for over 20 years	Pathogen persisted	Yes	[88]
	Red deer (possum)	New Zealand	Pulse: possum density reduced by poisoning	Incidence in deer dropped to approximately zero in treatment plots	No	[89]
	Possum (pigs, deer, and ferrets)	New Zealand	Press: reduced possums by poisoning to 22% of former population density, followed by maintenance through trapping and shooting	Major reduction in TB prevalence in possums and TB incidence in cattle	Yes	[90]
	Badgers (cattle)	England	Pulse: randomised controlled trial of badger culling	Mixed results, but evidence for temporary decrease in incidence	Insufficient evidence	[91]
<i>Brucella</i> spp.	Red deer (cattle, sheep, and goats)	Spain	Press: controlled in livestock through a yearly ‘test-and-slaughter’ program; no control in deer	Pathogen eliminated in deer	No	[92]
Louping ill virus	Hares (grouse)	Scotland	Press: hare density reduced by shooting and snaring; control plot without intervention	Huge drop in tick burden and viral prevalence in grouse on treatment plot	Yes	[93]
<i>Leishmania chagasi</i>	Dogs (humans)	Brazil	Pulse: seropositive dogs eliminated in two valleys, no treatment in two other valleys	No difference in human incidence across treatments	No (although see [69])	[94]
Rabies virus	Red foxes (skunks, dogs, cats, and bats)	Ontario	Press: oral vaccination of foxes	Elimination from foxes, followed by elimination from skunks	Yes	[95]
	Red foxes (carnivores and dogs)	Europe	Press: oral vaccination of foxes	Elimination	Yes	[96]
	Dogs (wild carnivores, livestock, and humans)	Serengeti, Tanzania	Press: long-term mass vaccination of dogs	Elimination in parts of the ecosystem	Yes	[97]
Rinderpest virus	Cattle (wildlife)	Africa	Press: coordinated large-scale vaccination of cattle with sudden interruption to identify remaining affected areas	Eradication in cattle and wildlife	Yes	[78]

The main challenge in using adaptive management lies in balancing knowledge gain that enables improved future control with achieving the best short-term outcome based on current knowledge [66]. It might be that an intervention that optimises the short-term outcome (e.g., one resulting in the greatest reduction of disease prevalence in the target) is also the one that provides the greatest statistical power to test the hypothesis of interest. In other situations, there can be a trade-off between the short-term goals of rapid disease control and the longer-term goals of learning about the system to optimise future control. This is particularly true given the cost of allocating resources to monitoring, the need to include experimental control areas [67], and the challenge of defining efficient experimental designs (e.g., stepped wedged trial; [68]) for the hypothesis being tested [62,69]. For example, faced with limited knowledge about the dynamics of chronic wasting disease (CWD) in Wisconsin (US), the US Department of Natural Resources established an adaptive management programme to eradicate CWD from the area [70]. The intervention was based on random deer culling, but a key component of the program was the collection of lymph nodes and brain tissue from the culled and harvested deer to assess the spatial distribution of CWD and provide further insights into its epidemiology.

Integration of data and findings

In most cases, understanding reservoir dynamics requires the use of multiple data sources. Integration of findings can occur at the analysis or study design stage [71] or later, using techniques such as meta-analysis or mathematical modelling (e.g., [69]). Triangulation of multiple sources should improve understanding of the validity and generalisation of inferences [72]. By synthesising several lines of evidence, Lembo *et al.* [14] found support for the hypothesis that domestic dogs, rather than wildlife, constitute the maintenance population for canine rabies in northern Tanzania. Their analyses included post-hoc integration of long-term case monitoring data (suggesting that rabies can persist in high-density domestic dog populations), genetic data (showing that a single rabies virus variant circulates among a range of species), and analysis of incidence patterns (indicating that spillover from domestic dog populations initiated only short-lived chains of transmission in other carnivores, consistent with zone C in Figure 1).

Ultimately, we seek a formal statistical integration of different sources of evidence that can be used to characterise reservoir systems. Such integrative approaches are rare, but increasingly powerful methods are being developed. For example, genetic, spatial, and epidemiological data can now be combined to enable detailed reconstruction of transmission within and between host populations (e.g., [73,74]) and time-calibrated phylogenies can be layered with geographical and epidemiological data in a joint framework that enables estimation of the frequency and directionality of interspecies transmission (e.g., [56,75]). For example, based on viral gene sequences and epidemiological data, Faria *et al.* [75] reconstructed the cross-species transmission history of rabies virus between North American bats and identified ecological and evolutionary

Box 4. Outstanding questions

Although there are increasingly powerful tools to characterise different components of reservoir–target systems, there are two central challenges that remain to be overcome: (i) determining persistence thresholds for different host populations; and (ii) estimating the rates of cross-species transmission. Instead, we typically make qualitative inferences (as shown in Table 2, main text), which inevitably results in less effective control policies. Advances in the use of genetic data and widely available serology data sets, including their integration with relevant ecological theory such as reservoir capacity, offer promising new ways to approach these challenges.

The use of interventions as quasi-experiments can provide robust empirical ‘top-down’ approaches to characterising reservoirs, particularly if they are designed as ‘crucial experiments’ [98] that test among multiple hypotheses to eliminate host populations progressively as contributing to a reservoir. However, this approach raises two other challenges: (iii) how to design interventions that allow us to test the quantitative predictions of the level of control needed to eliminate infection in one population; and (iv) how to coordinate the close engagement of the research community with managers charged with improving veterinary and public health.

constraints on transmission patterns. Latent variable models that explicitly parameterise both process and observation models are also well suited to combining data types, particularly when observations are sparse [26]. Developing statistically rigorous analytical methods that integrate multiple data layers is a challenging but exciting area, and key to future progress in infectious disease ecology [76,77]. Box 4 summarises outstanding questions in the study of reservoirs of infection.

Concluding remarks

Each of the approaches discussed here provides important threads of evidence on its own. However, these threads are part of a more extensive tapestry and, when viewed in isolation, they convey only a fragmentary understanding of how reservoirs work. Appropriately designed interventions can simultaneously provide direct tests of disease control methodology, deliver health benefits within the target population, and create important research opportunities that can advance understanding of reservoir dynamics. However, to realise these benefits fully, we must form broad-based interdisciplinary teams, engage with their full range of expertise from the earliest planning stages, and support them throughout the lifetime of the intervention. Understanding reservoir structure and function requires not only an integration of approaches to data collection and analysis, but also a step-change in the way that research communities integrate their activities with animal and human health practitioners.

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References

- Cleaveland, S. *et al.* (2001) Diseases of humans and their domestic mammals: pathogen characteristics, host range and the risk of emergence. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.* 356, 991–999
- Taylor, L.H. *et al.* (2001) Risk factors for human disease emergence. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.* 356, 983–989
- Woolhouse, M.E. *et al.* (2001) Population biology of multihost pathogens. *Science* 292, 1109–1112
- Streicker, D.G. *et al.* (2013) Differential sources of host species heterogeneity influence the transmission and control of multihost parasites. *Ecol. Lett.* 16, 975–984
- Haydon, D.T. *et al.* (2002) Identifying reservoirs of infection: a conceptual and practical challenge. *Emerg. Infect. Dis.* 8, 1468–1473
- Ashford, R.W. (1997) What it takes to be a reservoir host. *Belg. J. Zool.* 127 (Suppl.), 85–90
- Ashford, R.W. (2003) When is a reservoir not a reservoir? *Emerg. Infect. Dis.* 9, 1495–1496
- Nishiura, H. *et al.* (2009) How to find natural reservoir hosts from endemic prevalence in a multi-host population: a case study of influenza in waterfowl. *Epidemics* 1, 118–128
- Dobson, A.P. (2004) Population dynamics of pathogens with multiple host species. *Am. Nat.* 164, S64–S78
- Begon, M. *et al.* (1999) Transmission dynamics of a zoonotic pathogen within and between wildlife host species. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.* 266, 1939–1945
- Carslake, D. *et al.* (2006) Inference of cowpox virus transmission rates between wild rodent host classes using space–time interaction. *Proc. Biol. Sci.* 273, 775–782
- Kilpatrick, A.M. *et al.* (2006) Host heterogeneity dominates West Nile virus transmission. *Proc. Biol. Sci.* 273, 2327–2333
- O’Cathain, A. *et al.* (2010) Three techniques for integrating data in mixed methods studies. *BMJ* 341, c4587
- Lembo, T. *et al.* (2008) Exploring reservoir dynamics: a case study of rabies in the Serengeti ecosystem. *J. Appl. Ecol.* 45, 1246–1257
- Fenton, A. and Pedersen, A.B. (2005) Community epidemiology framework for classifying disease threats. *Emerg. Infect. Dis.* 11, 1815–1821
- Blumberg, S. and Lloyd-Smith, J.O. (2013) Inference of $R(0)$ and transmission heterogeneity from the size distribution of stuttering chains. *PLoS Comput. Biol.* 9, e1002993
- Antia, R. *et al.* (2003) The role of evolution in the emergence of infectious diseases. *Nature* 426, 658–661
- Cleaveland, S. *et al.* (2000) Serological and demographic evidence for domestic dogs as a source of canine distemper virus infection for Serengeti wildlife. *Vet. Microbiol.* 72, 217–227
- Packer, C. *et al.* (1999) Viruses of the Serengeti: patterns of infection and mortality in African lions. *J. Anim. Ecol.* 68, 1161–1178
- Prager, K.C. *et al.* (2012) Rabies virus and canine distemper virus in wild and domestic carnivores in northern Kenya: are domestic dogs the reservoir? *Ecohealth* 9, 483–498
- Almberg, E.S. *et al.* (2009) A serological survey of infectious disease in Yellowstone National Park’s canid community. *PLoS ONE* 4, e7042
- Brown, E.L. *et al.* (2010) Seroprevalence of *Trypanosoma cruzi* among eleven potential reservoir species from six states across the southern United States. *Vector Borne Zoonotic Dis.* 10, 757–763
- Carpentier, A. *et al.* (2012) High hepatitis E virus seroprevalence in forestry workers and in wild boars in France. *J. Clin. Microbiol.* 50, 2888–2893
- Gilbert, A. *et al.* (2013) Deciphering serology to understand the ecology of infectious diseases in wildlife. *Ecohealth* 3, 298–313
- Lachish, S. *et al.* (2011) Infection dynamics of endemic malaria in a wild bird population: parasite species-dependent drivers of spatial and temporal variation in transmission rates. *J. Anim. Ecol.* 80, 1207–1216
- Heisey, D.M. *et al.* (2010) Linking process to pattern: estimating spatiotemporal dynamics of a wildlife epidemic from cross-sectional data. *Ecol. Monogr.* 80, 221–240
- Blackwood, J. *et al.* (2013) Resolving the roles of immunity, pathogenesis and immigration for rabies persistence in vampire bats. *Proc. Natl. Acad. Sci. U.S.A.* <http://dx.doi.org/10.1073/pnas.13088171110>
- Bartlett, M.S. (1957) Measles periodicity and community size. *J. R. Stat. Soc. Ser. A* 120, 48–70
- Bartlett, M.S. (1956) Deterministic and stochastic models for recurrent epidemics. In *Proceedings of the Third Berkeley Symposium on Mathematical Statistics and Probability*, pp. 81–109 University of California Press (<http://projecteuclid.org/euclid.bsm/1200502549>)
- Ferrari, M.J. *et al.* (2008) The dynamics of measles in sub-Saharan Africa. *Nature* 451, 679–684
- Swinton, J. *et al.* (1998) Persistence thresholds for phocine distemper virus infection in harbour seal *Phoca vitulina* metapopulations. *J. Anim. Ecol.* 67, 54–68
- Conlan, A.J.K. *et al.* (2010) Resolving the impact of waiting time distributions on the persistence of measles. *J. R. Soc. Interface* 7, 623–640
- Lloyd, A.L. (2001) Realistic distributions of infectious periods in epidemic models: changing patterns of persistence and dynamics. *Theor. Popul. Biol.* 60, 59–71
- Nåsell, I. (1999) On the time to extinction in recurrent epidemics. *J. R. Stat. Soc. Ser. B* 61, 309–330
- Lloyd-Smith, J.O. *et al.* (2005) Should we expect population thresholds for wildlife disease? *Trends Ecol. Evol.* 20, 511–519
- Farrington, C.P. and Grant, A.D. (1999) The distribution of time to extinction in subcritical branching processes: applications to outbreaks of infectious disease. *J. Appl. Probab.* 36, 621–950
- Bartlett, M.S. (1960) The critical community size for measles in the United States. *J. R. Stat. Soc. Ser. A* 123, 37–44
- Choisy, M. and Rohani, P. (2012) Changing spatial epidemiology of pertussis in continental USA. *Proc. Biol. Sci.* 279, 4574–4581
- Gunning, C.E. and Wearing, H.J. (2013) Probabilistic measures of persistence and extinction in measles (meta)populations. *Ecol. Lett.* 16, 985–994
- Nåsell, I. (2005) A new look at the critical community size for childhood infections. *Theor. Popul. Biol.* 67, 203–216
- Schenzle, D. and Dietz, K. (1987) Critical population sizes for endemic virus transmission. In *Raumliche Persistenz und Diffusion von Krankheiten* (Fricke, W. and Hinz, E., eds), Heidelberg Geographisches Arbeiten. 83, 31–42
- Diekmann, O. and Heesterbeek, J.A.P. (1999) *Mathematical Epidemiology of Infectious Diseases: Model Building, Analysis and Interpretation*, Wiley
- Almberg, E.S. *et al.* (2010) Persistence of canine distemper virus in the Greater Yellowstone ecosystem’s carnivore community. *Ecol. Appl.* 20, 2058–2074
- Grenfell, B. and Harwood, J. (1997) (Meta)population dynamics of infectious diseases. *Trends Ecol. Evol.* 12, 395–399
- Grenfell, B.T. *et al.* (2002) Dynamics of measles epidemics: scaling noise, determinism, and predictability with the TSIR model. *Ecol. Monogr.* 72, 185–202
- Keeling, M.J. and Grenfell, B.T. (1997) Disease extinction and community size: modeling the persistence of measles. *Science* 275, 65–67
- Roberts, M.G. and Heesterbeek, J.A. (2003) A new method for estimating the effort required to control an infectious disease. *Proc. Biol. Sci.* 270, 1359–1364
- Toni, T. *et al.* (2009) Approximate Bayesian computation scheme for parameter inference and model selection in dynamical systems. *J. R. Soc. Interface* 6, 187–202
- Beaumont, M.A. (2010) Approximate bayesian in evolution and ecology. *Annu. Rev. Ecol. Evol. Syst.* 41, 379–406
- Beyer, H.L. *et al.* (2011) Metapopulation dynamics of rabies and the efficacy of vaccination. *Proc. Biol. Sci.* 278, 2182–2190
- He, D. *et al.* (2010) Plug-and-play inference for disease dynamics: measles in large and small populations as a case study. *J. R. Soc. Interface* 7, 271–283
- Ionides, E.L. *et al.* (2006) Inference for nonlinear dynamical systems. *Proc. Natl. Acad. Sci. U.S.A.* 103, 18438–18443
- Craft, M.E. *et al.* (2010) Disease transmission in territorial populations: the small-world network of Serengeti lions. *J. R. Soc. Interface* 8, 776–786
- Cross, P.C. *et al.* (2010) Probable causes of increasing brucellosis in free-ranging elk of the Greater Yellowstone Ecosystem. *Ecol. Appl.* 20, 278–288
- Drexler, J.F. *et al.* (2012) Bats host major mammalian paramyxoviruses. *Nat. Commun.* 3, 796

- 56 Mather, A.E. *et al.* (2013) Distinguishable epidemics of multidrug-resistant *Salmonella* typhimurium DT104 in different hosts. *Science* 341, 1514–1517
- 57 Streicker, D.G. *et al.* (2010) Host phylogeny constrains cross-species emergence and establishment of rabies virus in bats. *Science* 329, 676–679
- 58 Kuzmin, I.V. *et al.* (2012) Molecular inferences suggest multiple host shifts of rabies viruses from bats to mesocarnivores in Arizona during 2001–2009. *PLoS Pathog.* 8, e1002786
- 59 Hanski, I. and Ovaskainen, O. (2000) The metapopulation capacity of a fragmented landscape. *Nature* 404, 755–758
- 60 Wobeser, G.A. (2002) Disease management strategies for wildlife. *Rev. Sci. Tech.* 21, 159–178
- 61 Holling, C.S. (1978) *Adaptive Environmental Assessment and Management*, Wiley
- 62 Stankey, G.H. *et al.* (2005) *Adaptive Management of Natural Resources: Theory, Concepts, and Management Institutions*, US Department of Agriculture
- 63 Case, T.J. and Bender, E.A. (1981) Testing for higher order interactions. *Am. Nat.* 118, 920–929
- 64 Caswell, H. (2001) *Matrix Population Models: Construction, Analysis and Interpretation*. (2nd edn), Sinauer Associates
- 65 Donnelly, C.A. *et al.* (2005) Positive and negative effects of widespread badger culling on tuberculosis in cattle. *Nature* 439, 843–846
- 66 Allen, C. and Stankey, G.H. (2009) *Adaptive Environmental Management: A Practitioner's Guide*, Springer
- 67 McDonald-Madden, E. *et al.* (2010) Monitoring does not always count. *Trends Ecol. Evol.* 25, 547–550
- 68 Brown, C.A. and Lilford, R.J. (2006) The stepped wedge trial design: a systematic review. *BMC Med. Res. Method.* 6, 54
- 69 Quinnell, R.J. and Courtenay, O. (2009) Transmission, reservoir hosts and control of zoonotic visceral leishmaniasis. *Parasitology* 136, 1915–1934
- 70 Bartelt, G. *et al.* (2003) *Environmental Impact Statement: On Rules to Eradicate Chronic Wasting Disease from Wisconsin's Free-Ranging White-Tailed Deer Herd*, Wisconsin Department of Natural Resources
- 71 Moran-Ellis, J. (2006) Triangulation and integration: processes, claims and implications. *Qual. Res.* 6, 45–59
- 72 Farmer, T. *et al.* (2006) Developing and implementing a triangulation protocol for qualitative health research. *Qual. Health Res.* 16, 377–394
- 73 Brunker, K. *et al.* (2012) Integrating the landscape epidemiology and genetics of RNA viruses: rabies in domestic dogs as a model. *Parasitology* 139, 1899–1913
- 74 Morelli, G. *et al.* (2010) *Yersinia pestis* genome sequencing identifies patterns of global phylogenetic diversity. *Nat. Genet.* 42, 1140–1143
- 75 Faria, N.R. *et al.* (2013) Simultaneously reconstructing viral cross-species transmission history and identifying the underlying constraints. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.* 368, 20120196
- 76 Jombart, T. *et al.* (2014) Bayesian reconstruction of disease outbreaks by combining epidemiologic and genomic data. *PLoS Comput. Biol.* 10, e1003457
- 77 Mollentze, N. *et al.* (2014) A Bayesian approach for inferring the dynamics of partially observed endemic infectious diseases from space-time-genetic data. *Proc. Biol. Sci.* 281, 20133251
- 78 Roeder, P. *et al.* (2013) Rinderpest: the veterinary perspective on eradication. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.* 368, 20120139
- 79 Lemey, P. *et al.* (2010) Phylogeography takes a relaxed random walk in continuous space and time. *Mol. Biol. Evol.* 27, 1877–1885
- 80 Minin, V.N. and Suchard, M.A. (2008) Counting labeled transitions in continuous-time Markov models of evolution. *J. Math. Biol.* 56, 391–412
- 81 Beerli, P. (2006) Comparison of Bayesian and maximum-likelihood inference of population genetic parameters. *Bioinformatics* 22, 341–345
- 82 Beerli, P. and Felsenstein, J. (2001) Maximum likelihood estimation of a migration matrix and effective population sizes in *n* subpopulations by using a coalescent approach. *Proc. Natl. Acad. Sci. U.S.A.* 98, 4563–4568
- 83 Bedford, T. *et al.* (2010) Global migration dynamics underlie evolution and persistence of human influenza A (H3N2). *PLoS Pathog.* 6, e1000918
- 84 Biek, R. *et al.* (2012) Whole genome sequencing reveals local transmission patterns of *Mycobacterium bovis* in sympatric cattle and badger populations. *PLoS Pathog.* 8, e1003008
- 85 Beerli, P. (2004) Effect of unsampled populations on the estimation of population sizes and migration rates between sampled populations. *Mol. Ecol.* 13, 827–836
- 86 Harrison, P.J. *et al.* (2011) Bayesian state-space modeling of metapopulation dynamics in the Glanville fritillary butterfly. *Ecol. Monogr.* 81, 581–598
- 87 Beyer, H.L. *et al.* (2012) The implications of metapopulation dynamics on the design of vaccination campaigns. *Vaccine* 30, 1014–1022
- 88 Gortazar, C. *et al.* (2005) Molecular characterization of *Mycobacterium tuberculosis* complex isolates from wild ungulates in south-central Spain. *Vet. Res.* 36, 43–52
- 89 Nugent, G. (2005) *The Role of Wild Deer in the Epidemiology and Management of Bovine Tuberculosis in New Zealand*, Lincoln University PhD thesis
- 90 Caley, P. *et al.* (1999) Effects of sustained control of brushtail possums on levels of *Mycobacterium bovis* infection in cattle and brushtail possum populations from Hohotaka, New Zealand. *N. Z. Vet. J.* 47, 133–142
- 91 Godfray, H.C.J. *et al.* (2013) A restatement of the natural science evidence base relevant to the control of bovine tuberculosis in Great Britain. *Proc. Biol. Sci.* 280, 20131634
- 92 Serrano, E. *et al.* (2011) Decreasing prevalence of brucellosis in red deer through efforts to control disease in livestock. *Epidemiol. Infect.* 139, 1626–1630
- 93 Laurenson, M.K. *et al.* (2003) Identifying disease reservoirs in complex systems: mountain hares as reservoirs of ticks and louping-ill virus, pathogens of red grouse. *J. Anim. Ecol.* 72, 177–185
- 94 Dietze, R. *et al.* (1997) Effect of eliminating seropositive canines on the transmission of visceral leishmaniasis in Brazil. *Clin. Infect. Dis.* 25, 1240–1242
- 95 MacInnes, C.D. *et al.* (2001) Elimination of rabies from red foxes in eastern Ontario. *J. Wildl. Dis.* 37, 119–132
- 96 Vitasek, J. (2004) A review of rabies elimination in Europe. *Vet. Med.* 49, 171–185
- 97 Lembo, T. *et al.* (2010) The feasibility of canine rabies elimination in Africa: dispelling doubts with data. *PLoS Negl. Trop. Dis.* 4, e626
- 98 Platt, J.R. (1964) Strong inference. *Science* 146, 347–353