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A Review of Stability Issues Associated with Vitamins in Parenteral Nutrition

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Abstract

Background & aims: There has been a move to increased emphasis on delivering parenteral nutrition to patients at home, which may improve patient care and reduce costs. However, safe provision of home, and indeed any, parenteral nutrition necessitates consideration of the physical and chemical stability of the parenteral nutrition and its components.

Methods: Medline and Embase were used to search for all English-language publications on vitamin stability. Identified publications were then analysed and summarised in the following review.

Results: Vitamins are one of the least stable components in PN and there are three main ways in which they have been shown to degrade: photodegradation, oxidation and through storage material interaction. Previous research on vitamins has demonstrated that significant losses can occur in the bag, which could have clinical consequences, particularly for long-term users of parenteral nutrition. These losses are most dramatic for vitamin C, which is rapidly degraded by oxygen, and vitamin A, which is rapidly degraded in the presence of sunlight.

Conclusions: There are a number of stability issues associated with vitamins in parenteral nutrition and further investigation is needed to assure their stability and compatibility with other parenteral nutrition constituents.

Key words

Parenteral Nutrition, Vitamins, Stability, Degradation

Introduction

Historically, Parenteral Nutrition (PN) was prepared at ward level in a multi-bottle dose system¹. This method was labour intensive and was readily susceptible to contamination. This called for the introduction of an all-in-one (AIO) bag, which would decrease the incidence of infection and be a convenient and cost-effective alternative to the multi-bottle system previously employed¹⁻³. The use of an AIO bag meant that all macronutrients, vitamins, trace elements and electrolytes were introduced to the same bag prior to administration, leading to a number of formulation and stability issues.

At present, there is a drive within the NHS to treat patients at home in an attempt to significantly reduce costs and improve clinical outcomes^{4,5}. An increased emphasis on delivering treatments at home has stimulated researchers to investigate the sources of instability within PN admixtures, thereby validating shelf lives and allowing more convenient administration at home.

One aspect of PN that has not been fully investigated is the addition of vitamins. Vitamins are highly reactive and their addition to PN admixtures can cause a number of pharmaceutical issues. Reactions involving vitamins are dependent on: relative concentrations of reactants, pH, temperature, time, container material and the presence of any other catalytically active components⁶. The reactive nature of vitamins mean they should be added shortly before administration to ensure that the integrity of the admixture is

maintained⁷. There are three main types of vitamin instability seen in PN: photodegradation, oxidation and interactions with storage material⁸. In every PN admixture administered a patient should receive a days supply of vitamins and trace elements⁹. Ensuring that patients are administered with sufficient amounts of vitamins is essential for normal bodily function and to prevent the manifestation of clinical symptoms of deficiency. Water-soluble vitamins in particular require regular dosing as they are not stored in significant amounts in the body, with the exception of vitamin B₁₂. Ensuring adequate dosing of fat-soluble vitamins is also important especially in patient groups such as infants, who only have small quantities of fat-soluble vitamins stored¹⁰.

Stability problems encountered with PN admixtures can occur in containers or administration sets. This is especially problematic for premature infants who receive their PN at slow infusion rates¹⁰. These stability issues can prevent patients from getting their intended doses or may harm the patient either through generation of potentially harmful by-products or interfere with the physical stability of the admixture⁸. A better grasp of how vitamins interact with the environment they are stored in and other PN components will ensure the safe use of them in the future.

The following review examines the current information available on the stability issues associated with vitamins.

Fat Soluble Vitamins

Vitamin A

Vitamin A, or retinol, is the most light-sensitive micronutrient found in PN admixtures. When subjected to light in unprotected bags or administration sets, it undergoes extensive photodegradation¹¹⁻¹³. The mechanism of this reaction is still not fully understood, but it is known that the wavelength and the intensity of the light interacting with the vitamin determines the rate of the photochemical reaction¹². A study by Allwood and Plane¹⁴ showed that retinol is more susceptible to photodegradation when exposed to wavelengths of less than 400 nm, with maximum degradation occurring between 330-350 nm. Such wavelengths are more commonly found in natural daylight, with artificial light emitting smaller amounts of wavelengths in the UV range¹². Nevertheless, a recent study by Ferguson et al.¹⁵ has found significant degradation (in excess of 10%) when retinol is exposed to artificial lighting that would be commonly found in hospitals and homes: cool white light and warm white light.

The inclusion of lipid in all-in-one (AIO) admixtures has resulted in the opacity of the admixture being increased significantly. There have been conflicting reports^{13,16}, but the addition of lipid does not seem to provide sufficient protection for light sensitive vitamins such as retinol¹². Therefore, to assure retinol photostability in PN light-protective covers should be used.

Since these studies on retinol degradation were performed there has been a change in lighting preference, with a number of aseptic units using energy-saving light bulbs. Similar changes have also occurred in homes in an attempt

to reduce costs. The impact of this change in lighting on retinol degradation has not yet been investigated fully. Until more is known about its influence on the degradation of retinol the use of light protection is essential¹⁷.

Besides photodegradation, sorption of retinol may occur with bags and administration set tubing, further reducing the amount of vitamin being administered to the patient⁸. This problem has been much reduced through the use of the less reactive palmitate ester, rather than the acetate ester¹⁰. The introduction of tubing containing polyolefine, which is free of PVC, plasticisers, adhesives or latex, has further reduced the absorption of vitamin A¹⁸.

Another source of degradation of vitamins are peroxides generated by lipid emulsions. Lipid emulsions containing polyunsaturated fatty acids (PUFAs) are at an increased risk of peroxidation¹⁹. Vitamin E acts as a major scavenger for free radicals and prevents lipid peroxy radicals from reacting with fatty acid side chains²⁰. Nevertheless, peroxidation still occurs to some degree. Guidetti et al.²¹ have studied the impact of different lipid emulsion compositions on vitamin degradation via peroxidation. Following 24 hours of light protected storage at room temperature Guidetti et al. found that retinol recovery was significantly increased in soybean-medium chain triacylglycerol oil-based emulsions when compared to soybean oil-based emulsions and olive/soybean oil-based emulsions. Further investigation is required to understand the relationship between the composition of lipid emulsions and the degradation of vitamins.

Vitamin D

As described by Allwood and Kearney⁸, there are very limited stability data available for vitamin D in PN. The only study on this vitamin has shown that following a 24-hour infusion period, 68% of the vitamin D concentration was recovered. Comparison of sample concentrations at various sites within the infusion set-up suggested that vitamin D may bind to plastic found in bags and administration sets²².

There has been a lot of recent interest in vitamin D, especially with the re-emergence of rickets in some urban areas. Consequently, there has been a call to increase its recommended daily allowance in countries such as the United States of America and Canada²³. This may, in turn, lead to an increase in the recommended amount of vitamin D being given to PN patients.

Vitamin E

Vitamin E is degraded by oxygen in a reaction catalysed by light. The intensity and wavelength of light as well as the amount of oxygen available influences the rate of degradation. Vitamin E is particularly sensitive to wavelengths between 285 nm and 305 nm²⁴.

Vitamin E seems to be relatively stable in admixtures especially when protected from light^{11-13,25}. A study by Allwood and Martin¹² investigated the

effect of light exposure on PN admixtures in multi-layered bags. They found that if oxygen was prevented from permeating into the bag, exposure to sunlight did not significantly reduce the concentration of vitamin E in the admixture. Additionally, in the presence of ascorbic acid, vitamin E oxidation is decreased as these two vitamins compete for oxygen¹².

Guidetti et al.²¹ also investigated the impact of lipid emulsion composition on tocopherol degradation. Like retinol, this study found that the recovery of both of the vitamin E isomers examined, α -tocopherol and γ -tocopherol, were significantly increased in AIO bags containing soybean-medium chain triacylglycerol oil-based emulsion when compared to soybean oil-based emulsions and olive/soybean oil-based emulsions.

It is important to note that vitamin E is commonly presented as a mixture of eight tocopherol isomers and the stability profiles have not been determined for each of the isomers. The introduction of increased levels of tocopherol in an attempt to protect fish oils found in some PN from oxidation, necessitates a thorough understanding of tocopherol isomer stability profiles²⁶.

Vitamin K₁

Phylloquinone (vitamin K₁) is a naturally occurring compound synthesized in plants. As it develops naturally in lipid emulsions some emulsions have higher concentrations than others²⁷, so patients may receive different amounts. However, reports of the impact of phylloquinone levels on neonates suggest

that symptoms associated with increased levels (e.g. constipation and pain) are non-serious and self limiting²⁸.

Phylloquinone is sensitive to sunlight but is considered stable in PN mixtures in the presence and absence of lipid emulsions, supporting the theory that lipid emulsions have little, if any, protective influence on light sensitive vitamins. It has been reported that the concentration of phylloquinone can decrease by 50% following 3 hours in strong sunlight¹³. Another study has shown degradation of 5.9-8.5% over 4.5 hours in artificial daylight²⁹.

Water Soluble Vitamins

Ascorbic acid

Vitamin C is one of the most reactive vitamins added to PN admixtures. In the body it is a strong antioxidant that quenches reactive oxygen and nitrogen species³⁰. When stored outside of the body, ascorbic acid acts in a similar way, reacting readily with oxygen. As shown in figure 1., ascorbic acid in the presence of oxygen is initially converted, by way of a reversible reaction, to an equally biologically active compound called dehydroascorbic acid. Hydrolysis of dehydroascorbic acid produces 2,3-diketo-gulonic acid, which is thought to be biologically inactive. Further oxidation of this intermediate produces threonic acid and oxalic acid. The degradation of ascorbic acid is directly linked to the amount of oxygen present in the medium³¹. Exclusion of free oxygen from the PN admixture limits the initial conversion of ascorbic acid to dehydroascorbic acid, thereby minimizing the resultant cascade³². Oxygen in

PN bags can originate from permeation of air through the bag wall during storage, residual headspace formed following filling and sealing and from dissolved air in injections of additives and infusions. The use of multilayered AIO bags has significantly reduced the amount of oxygen able to diffuse into the bags, thereby improving ascorbic acid stability⁸. However, this development has not eliminated the problem completely as oxygen transferred into the bag during filling cannot diffuse out of the bag and therefore remains in contact with the vitamin.

Additives to PN not only accommodate the transfer of oxygen to the medium, but also can directly influence ascorbic acid degradation. Copper, and to a lesser extent manganese, zinc and ferric ions, catalyse the oxidation of ascorbic acid to dehydroascorbic acid³². This theory has recently been supported by an extensive study conducted by Ferreyra et al.³³ who found significant degradation of ascorbic acid following the addition of nine trace elements to two-in-one (TIO) PN bags when compared to bags with no trace element additions. With copper, vitamin C is oxidized causing the concomitant reduction of copper from the cupric (II) to the cuprous (I) form. As a result of this, the cascade and the eventual production of threonic acid and oxalic acid speeds up. This reaction is enhanced by the introduction of such ions present as trace contaminants in PN components, resulting in higher concentration of copper available to catalyse the degradation of ascorbic acid³⁴. Allwood³⁵ investigated compatibility and stability in 3 Litre bags and found that the amino acid cysteine inhibits the catalytic effect of copper. Therefore, inclusion of

cysteine in amino acid solutions may be beneficial in slowing the degradation of ascorbic acid.

Physical conditions such as temperature and pH can also influence degradation of ascorbic acid in PN: at higher temperatures ascorbic acid degradation is increased^{36,37} and pH values above 4.0 make ascorbic acid more susceptible to oxidation³⁰.

The products of ascorbic acid degradation, oxalic acid and threonic acid, may compromise the stability of the emulsion by increasing the acidity. pHs below 5 can destabilize the PN emulsions³⁸. In addition, oxalic acid interacts with free calcium to produce calcium oxalate precipitate⁶. The impact of calcium oxalate formation in adults is unresolved but it is known to be hazardous to neonates³¹. Further investigation into the additional risks posed by oxalic acid and its precipitate is required.

Thiamine

In the past, thiamine in PN was degraded mainly by means of a reduction reaction. Sodium metabisulphite, a common antioxidant used in older generations of amino acid infusions, readily reacts with thiamine in solution⁸. This reaction involves the cleavage of thiamine molecules by sulphite into pyrimidine and thiazole. The stability of thiamine was directly linked to the concentration of sodium metabisulphite. Sodium metabisulphite is no longer

routinely used, with no amino acid solutions available in the UK containing it. Removal of this antioxidant has increased the stability of thiamine³⁹.

Riboflavin

Riboflavin has long been thought to degrade when it is exposed to daylight. In the presence of light and oxygen, riboflavin is irreversibly converted to lumino flavin and luminochromo, amongst other compounds⁶. A recent study by Ferguson et al.¹⁵ found significant degradation (in excess of 10%) of riboflavin when exposed to cool and warm white artificial light over a period of 24 hours. Significant degradation was also observed in a study by Mirkovic et al.⁴⁰ following 12 hours of exposure of riboflavin to daylight. In contrast, studies by Dahl et al.¹¹ and much more recently by Ribeiro et al.⁶ have shown no significant losses when stored at 25°C for 3 days and very little riboflavin loss when stored over a period of up to four days with and without light protection. However, the nature of the room illumination is not stated in any of these studies, so its influence is not quantifiable.

One of riboflavin's more undesirable properties is that it can act as a photochemical sensitizer⁴¹. As shown in figure 2., when riboflavin is in an excited state it can react directly with substrates or aid in the production of reactive oxygen species. Production of such reactive species may in turn, cause the oxidation of other PN constituents. Investigations into the effects of this process on various components of PN are ongoing⁴²⁻⁴⁴.

Pyridoxine

Pyridoxine is known to be light sensitive, although, there is limited information available on its stability in admixtures. It has been reported to be stable in PN admixtures for up to 96 hours at 2-8°C in darkness¹¹. Chen et al.⁴⁵ reported an 86% loss of pyridoxine occurring in 8 hours of direct sunlight. A more recent study by Ribeiro et al.⁶, found that pyridoxine was stable for 3 days when stored between 4°C and 25°C with and without photo-protection. Again, the illumination of the room in which the samples were stored is not stated therefore it is difficult to ascertain the extent of its influence on degradation.

Juhasz et al.⁴⁶ examined the thermal decomposition of pyridoxine. In this experiment, they calculated the amount of time it takes to reach 90% pyridoxine recovery at 25°C to be 1.7×10^{-2} years (approximately 6.2 days). This experiment was conducted using a pure sample of pyridoxine and may not correspond to pyridoxine degradation in parenteral nutrition.

Folic acid

There are very few studies investigating the degradation of folic acid. The main source of instability arises from changes in pH. Folic acid injections are usually formulated at a pH in excess of 8.0 because the vitamin is prone to precipitation at lower pHs⁴⁷. One investigation examined the effect of pH on folic acid precipitation and found that if the pH remains above 5.0, folic acid

remains in solution⁴⁸. As PN usually has a pH of between 5.0-6.0, folic acid should not precipitate.

There has been a suggestion by Lee et al.⁴⁹ that adsorption of folic acid onto polyvinylchloride (PVC) infusion bags may occur and was responsible for a 33% loss seen after 42 days of storage. However, later studies have found the vitamin to be compatible with PN bags^{11,48}.

Nicotinamide, Pantothenic acid, Biotin and Cyanocobalamin

There is very limited information available on the stability of cyanocobalamin, pantothenic acid, biotin and nicotinamide in PN. Dahl et al.¹¹ report that all four are stable in PN admixtures when stored for 96 hours. However, as no further studies have been reported, the stability of these water-soluble vitamins requires further investigation.

Conclusion

The inclusion of vitamins in PN provides a number of formulation issues. Ascorbic acid is the most unstable vitamin added to PN and is degraded by oxygen into a number of different products including oxalic acid. There is some concern over the formation of oxalic acid, which may form a calcium oxalate precipitate. Further investigation into its impact is required.

Photodegradation is a problem encountered with vitamins such as retinol and can cause significant losses. In addition, the impact of light on degradation in giving sets is an important consideration as the surface area of PN in contact with light is vastly increased.

The role of riboflavin as a photochemical sensitizer is being explored.

Its ability to form reactive oxygen species may have a significant impact on the stability of PN as a whole.

Little is known about the stability of vitamins such as nicotinamide, cyanocobalamin, biotin and pantothenic acid in PN. Clearly this needs some further investigation.

Trace elements can also cause a number of problems during compounding and administration. A recent paper by Hardy et al.³⁴, is an excellent review of the current literature available on trace elements.

Many of the degradation processes of vitamins in PN admixtures can be reduced or prevented by controlling physical conditions. Light exposure can cause degradation of a number of water and fat-soluble vitamins, most notably riboflavin and retinol. Use of light protection on bags and

administration sets can reduce vitamin loss significantly. Degradation can be further reduced in administration sets through shortening the lines to the patient. This would decrease degradation caused by light sources and also help to decrease vitamin sorption onto the tubing¹⁰. The light protective effects of lipids on vitamins is a contentious issue that needs further investigation, nevertheless, the use of lipid may provide added protection for light sensitive vitamins. Oxygen is another problematic physical condition that is the cause of substantial vitamin degradation, with ascorbic acid being the worst affected. Use of multi-layered bags and removal of excess air after filling reduces degradation rates even in the presence of trace elements. The use of cysteine may also help inactivate copper catalysis of ascorbic acid oxidation to its degradation product dehydroascorbic acid.

This review illustrates the importance of vitamin stability in parenteral nutrition. Maintaining vitamin stability will ensure that patients receive the correct doses and prevent the production of potentially harmful degradation products. Furthermore, it shows that there is still plenty of research to be done on vitamin stability in parenteral nutrition admixtures.

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TF prepared the first draft of the review. TF, SE, RPD and AGC contributed to the final version of the review. All authors have made substantial contributions to and approved the final version of the review.

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Conflict of interest

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References

1. Barnett MI, Pertkiewicz M, Cosslett AG, Mühlebach S. Basics in clinical nutrition: Parenteral nutrition admixtures, how to prepare parenteral nutrition (PN) admixtures. *e-SPEN. Eur e-J Clin Nutr Metab.* 2009;4(3):e114–e116.
2. Allwood MC. Pharmaceutical aspects of parenteral nutrition: from now to the future. *Nutrition.* 2000;16(7-8):615–8.
3. Menne R, Adolph M, Brock E, Schneider H, Senkal M. Cost analysis of parenteral nutrition regimens in the intensive care unit: three-compartment bag system vs multibottle system. *JPEN. J Parenter Enteral Nutr.* 2008;32(6):606–12.
4. NHS 2010 – 2015: from good to great. preventative, people-centred, productive. London: Department of Health, 2009 (Cm 7775). (Accessed 13/12/13, at http://webarchive.nationalarchives.gov.uk/20130107105354/http://www.dh.gov.uk/prod_consum_dh/groups/dh_digitalassets/@dh/@en/@ps/@sta/@perf/documents/digitalasset/dh_109887.pdf)
5. Scanzano C, Iacone R, Santarpia L, Alfonsi L, Pastore E, D'Isanto A, Pagano MC, Galeotanza MR, Contaldo F. PP182-Sun Cost Saving of Home Artificial Nutrition. *Clin Nutr.* 2013;32:S91.
6. Ribeiro DO, Pinto DC, Lima LMTR, Volpato NM, Cabral LM, de Sousa VP. Chemical stability study of vitamins thiamine, riboflavin, pyridoxine and ascorbic acid in parenteral nutrition for neonatal use. *Nutr J.* 2011;10:47–56.
7. Wormleighton CV, Catling TB. Stability issues in home parenteral nutrition. *Clin Nutr.* 1998;17(5):199–203.
8. Allwood MC, Kearney MCJ. Compatibility and stability of additives in parenteral nutrition admixtures. *Nutrition.* 1998;14(9):697–706.
9. Singer P, Berger MM, Van den Berghe G, Biolo G, Calder P, Forbes A, Griffiths R, Kreyman G, Lerverve X, Pichard C. ESPEN Guidelines on Parenteral Nutrition: intensive care. *Clin Nutr.* 2009;28(4):387–400.
10. Koletzko B, Goulet O, Hunt J, Krohn K, Shamir R. 8. Vitamins. *J Pediatr Gastroenterol Nutr.* 2005;41:S47–S53.
11. Dahl GB, Jeppsson RI, Tengborn HJ. Vitamin stability in a TPN mixture stored in an EVA plastic bag. *J Clin Hosp Pharm.* 1986;11:271–9.
12. Allwood MC, Martin HJ. The photodegradation of vitamins A and E in parenteral nutrition mixtures during infusion. *Clin Nutr.* 2000;19:339–42.

13. Billion-Rey F, Guillaumont M, Frederich A, Aulagner G. Stability of fat-soluble vitamins A (Retinol Palmitate), E (Tocopherol Acetate) and K1 (Phylloquinone) in total parenteral nutrition at home. *JPEN. J Parenter Enter Nutr.* 1993;17:56–60.
14. Allwood MC, Plane JH. The wavelength-dependent degradation of vitamin A exposed to ultraviolet radiation. *Int J Pharm.* 1986;31(1-2):1–7.
15. Ferguson TI, Price-Davies R, Cosslett A. PP259-Mon Vitamins – an Unknown Quantity. *Clin Nutr.* 2013;32:S219.
16. Haas C, Genzel-Boroviczény O, Koletzko B. Losses of vitamin A and E in parenteral nutrition suitable for premature infants. *Eur J Clin Nutr.* 2002;56(9):906–12.
17. Allwood MC, Ball PA, Driscoll DF, Sizer T. Light protection during parenteral nutrition Infusion: Is It Really Necessary? *Nutrition.* 2000;16:234–5.
18. Henton DH, Merritt RJ. Vitamin A Sorption to Polyvinyl and Polyolefin Intravenous Tubing. *JPEN. J Parenter Enter Nutr.* 1990;14:79–81.
19. Mühlebach S, Steger PJ. Lipid peroxidation of intravenous fat emulsions: a pharmaceutical issue with clinical impact? *Nutrition.* 1998;14(9):720–1.
20. Skouroliakou M, Matthaiou C, Chiou A, Panagiotakos D, Gounaris A, Nunn T, Andrikopoulos N. Physicochemical stability of parenteral nutrition supplied as all-in-one for neonates. *JPEN. J Parenter Enter Nutr.* 2008;32(2):201–9.
21. Guidetti M, Sforzini A, Bersani G, Corsini C, Zolezzi C, Fasano C, Pironi L. Vitamin A and Vitamin E Isoforms Stability and Peroxidation Potential of All-In-One Admixtures for Parenteral Nutrition. *Int J Vitam Nutr Res.* 2008;78(3):156–66.
22. Gillis J, Jones G, Pencharz P. Delivery of vitamins A, D and E in total parenteral nutrition solutions. *JPEN. J Parenter Enter Nutr.* 1983;7:11–4.
23. DeLuca HF. Evolution of our understanding of vitamin D. *Nutr Rev.* 2008;66(10 Suppl 2):S73–87.
24. Drott P, Meurling S, Meurling L. Clinical adsorption and photodegradation of the fat soluble vitamins A and E. *Clin Nutr.* 1991;10:348–51.
25. Lavoie J, Chessex P, Rouleau T, Tsopmo A, Friel J. Shielding parenteral multivitamins from light increases vitamin A and E

- concentration in lung of newborn guinea pigs. *Clin Nutr*. 2007;26(3):341–7.
26. Waitzberg DL, Torrinhas RS, Jacintho TM. New Parenteral Lipid Emulsions for Clinical Use. *JPEN. J Parenter Enter Nutr*. 2006;30(4):351–67.
 27. Shearer MJ. Vitamin K in parenteral nutrition. *Gastroenterology*. 2009;137(5 Suppl):S105–18.
 28. Martindale. Vitamin K Substances [Internet]. Martindale: The complete drug reference. 2012 [Accessed 05/04/2012]. Available from: http://www.medicinescomplete.com/mc/martindale/current/ms-7902-c.htm?q=Phylloquinone&t=search&ss=text&p=1#_hit
 29. Schmutz CW, Martinelli E, Mühlebach S. Stability of Vitamin K1 assessed by HPLC in total parenteral nutrition (TPN) admixtures. *Clin Nutr*. 1992;12(169):110–1.
 30. Berger MM. Vitamin C requirements in parenteral nutrition. *Gastroenterology*. 2009;137(5 Suppl):S70–8.
 31. Allwood MC. Oxalogenesis in parenteral nutrition mixtures. *Nutrition*. 1999;15(1):70.
 32. Rockwell GF, Campfield T, Nelson BC, Uden PC. Oxalogenesis in parenteral nutrition solution components. *Nutrition*. 1998;14(11-12):836–9.
 33. Ferreyra ME, Ocaña MC, Vega R. PP260-Sun Trace Elements Influence on Ascorbic Acid Availability. *Clin Nutr*. 2013;32:S121.
 34. Hardy G, Menendez AM, Manzanares W. Trace element supplementation in parenteral nutrition: pharmacy, posology, and monitoring guidance. *Nutrition*. 2009;25(11-12):1073–84.
 35. Allwood MC. Factors Influencing the stability of Ascorbic Acid in total parenteral nutrition infusions. *J Clin Hosp Pharm*. 1984;9:75–85.
 36. Gibbons E, Allwood MC, Neal T, Hardy G. Degradation of dehydroascorbic acid in parenteral nutrition mixtures. *J Pharm Biomed Anal*. 2001;25(3-4):605–11.
 37. Proot P, De Pourcq L, Raymakers AA. Stability of ascorbic acid in a standard total parenteral nutrition mixture. *Clin Nutr*. 1994;13(5):273–9.
 38. Lobo BW, Veiga VF, Cabral LM, Michel RC, Volpato NM. Influence of the relative composition of trace elements and vitamins in physicochemical stability of total parenteral nutrition formulations for neonatal use. *Nutr J*. 2012;11(26):1–12.

39. Gallitelli L. Trace element and vitamin requirements in patients receiving parenteral nutrition. *Clin Nutr.* 1995;14 Suppl 1:70–4.
40. Mirkovic DC, Basic Z, Roganovic B. PP185-Sun Influence of Type of Bag, Sort of Admixtures and Exposure To Daylight Upon Stability of Vitamins C and B2 in Parenteral Nutrition Admixtures. *Clin Nutr Suppl.* 2011;6(1):93–4.
41. Silva E, Ugarte R, Andrade A, Edwards AM. Riboflavin-sensitized photoprocesses of tryptophan. *J Photochem Photobiol B.* 1994;23(1):43–8.
42. Natera J, Massad WA, García NA. Vitamin B1 as a scavenger of reactive oxygen species photogenerated by vitamin B2. *Photochem Photobiol.* 2011;87(2):317–23.
43. Huvaere K, Cardoso DR, Homem-de-Mello P, Westermann S, Skibsted LH. Light-induced oxidation of unsaturated lipids as sensitized by flavins. *J Phys Chem B.* 2010;114(16):5583–93.
44. Shen L, Ji H-F. How α -tocopherol quenches triplet state riboflavin? Insights from theory. *J Photochem Photobiol A Chem.* 2008;199(1):119–21.
45. Chen MF, Worth Boyce H, Triplett L. Stability of B vitamins mixed in parenteral nutrition solution. *JPEN. J Parenter Enter Nutr.* 1983;7:462–4.
46. Juhász M, Takahashi S, Kitahara Y, Fujii T. Thermal decomposition of pyridoxine: an evolved gas analysis-ion attachment mass spectrometry study. *Rapid Commun Mass Spectrom.* 2012;26(7):759–64.
47. Allwood MC. Compatibility and stability of TPN mixtures in big bags. *J Clin Hosp Pharm.* 1984;9(3):181–98.
48. Barker A, Hebron BS, Beck PR, Ellis B. Folic Acid and total parenteral nutrition. *JPEN. J Parenter Enter Nutr.* 1984;8:3–8.
49. Lee DR, Ware I, Winsley BE. Survival of folic acid in TPN solutions. *Br J Intraven Ther.* 1980;1(2):13–6.

Figure 1. The ascorbic acid degradation cascade adapted from Allwood and Kearney⁴.

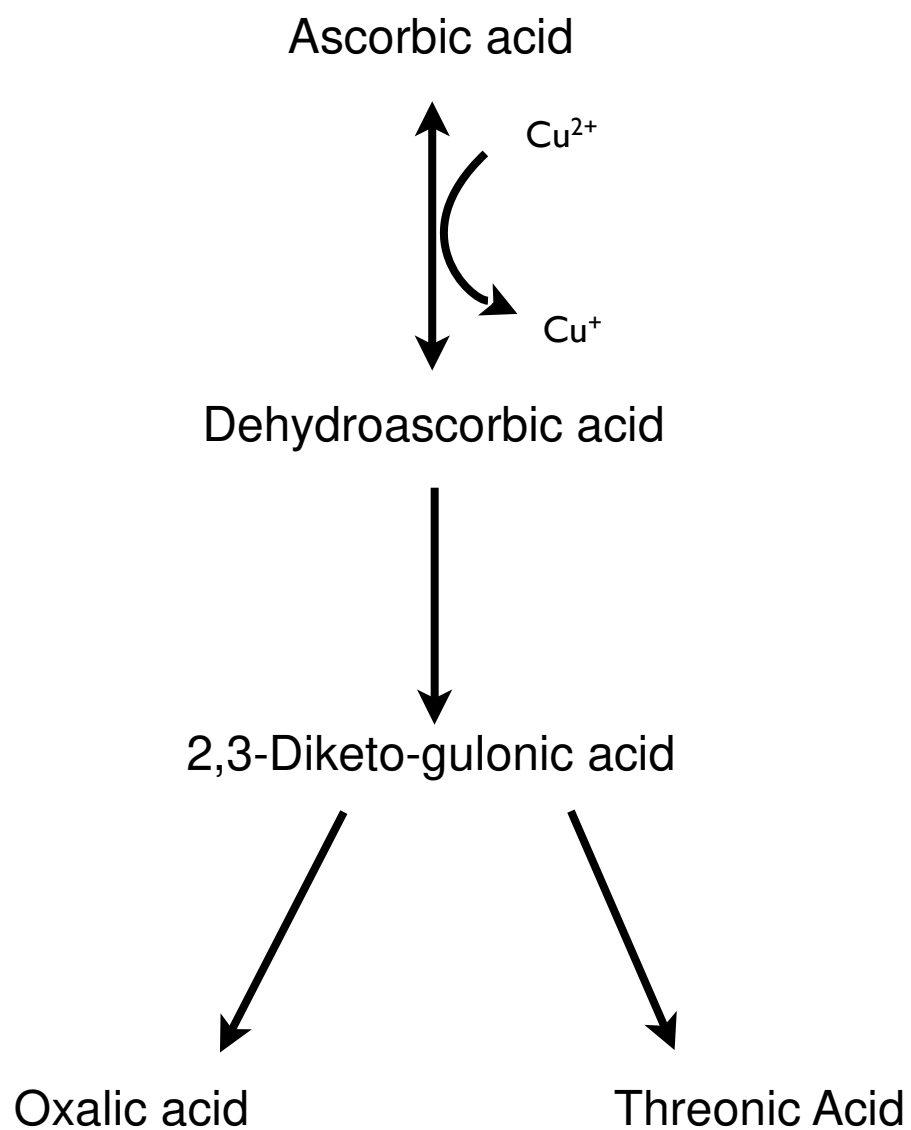


Figure 2. A flow diagram illustrating riboflavin as a photosensitizer.

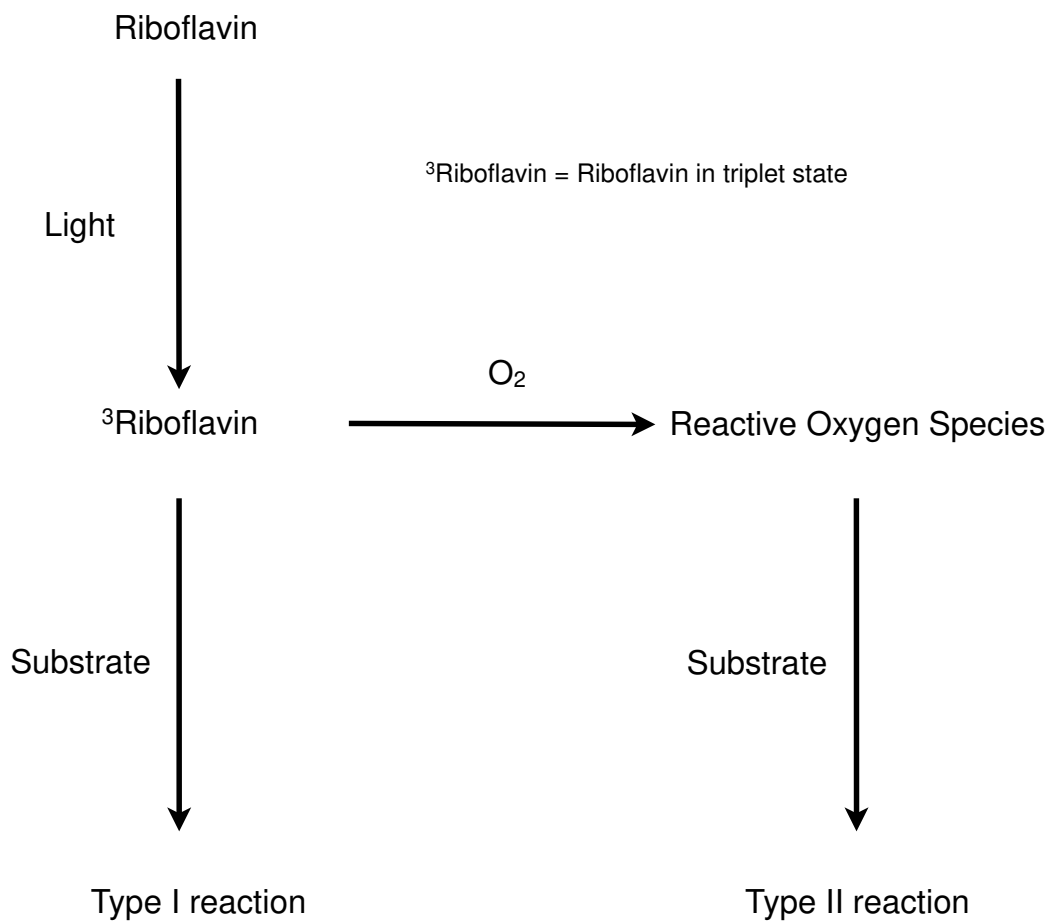


Table 1. Vitamin Reference Nutrient Intake (RNI) values for healthy males and females aged between 19-50 years adapted from Department of Health reference values for Food Energy and Nutrients for the United Kingdom (41):

| Vitamin | Males 19-50 years | Females 19-50 years |
|-------------------------|--------------------------|----------------------------|
| Vitamin A | 700 µg/day | 600 µg/day |
| Vitamin D | - | - |
| Vitamin E | Above 4 mg/day | Above 3 mg/day |
| Vitamin K | 1 µg/kg/day | 1 µg/kg/day |
| Ascorbic Acid | 40 mg/day | 40 mg/day |
| Thiamine | 1 mg/day | 0.8 mg/day |
| Riboflavin | 1.3 mg/day | 1.1 mg/day |
| Niacin | 17 mg/day | 13 mg/day |
| Pantothenic acid | 3-7 mg/day | 3-7 mg/day |
| Pyridoxine | 1.4 mg/day | 1.2 mg/day |
| Biotin | 10-200 µg/day | 10-200 µg/day |
| Folate | 200 µg/day | 200 µg/day |
| Cyanocobalamin | 1.5 µg/day | 1.5 µg/day |

N.B. Niacin describes the total amount of nicotinic acid and nicotinamide in the diet.

Table 2. Quick reference of known physical vitamin sensitivities:

| Vitamin | Light | Oxygen | pH | Temperature |
|---------------------|-------|--------|----|-------------|
| Vitamin A | ☐ | | | |
| Vitamin D | | | | |
| Vitamin E | ☐ | ☐ | | |
| Vitamin K | ☐ | | | |
| Ascorbic acid | | ☐ | ☐ | ☐ |
| Thiamine | | | | |
| Riboflavin | ☐ | | | |
| Nicotinamide | | | | |
| Pantothenic Acid | | | | |
| Pyridoxine | ☐ | | | |
| Biotin | | | | |
| Folic Acid | | | ☐ | |
| Cyanocobalamin | | | | |