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Influence of operating parameters on surface properties of RF glow discharge oxygen plasma treated TiO₂/PET film for biomedical application

K.Navaneetha Pandiayaraj^{1*}, R.R. Deshmukh², Pi-G Su³, Emre Yassitepe⁴, Ismat Shah⁴, Stefano Perni⁵, Polina Prokopovich^{5,6}, Mallikarjuna N. Nadagouda⁷

¹Surface Engineering Laboratory, Department of Physics, Sri Shakthi Institute of Engineering and Technology, L&T by pass, Chinniyam Palayam (post), Coimbatore-641062. India

²Department of Physics, Institute of Chemical Technology, Matunga, Mumbai 400 019, India.

³Department of Chemistry, Chinese Culture University, Taipei 111, Taiwan.

⁴Department of Physics and Astronomy, Department of Materials Science and Engineering, University of Delaware, 208 Dupont Hall, Newark, United States.

⁵School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK.

⁶Institute of Medical Engineering & Medical Physics, School of Engineering, Cardiff University, UK

⁷The U.S. Environmental Protection Agency, ORD, NRMRL, WSWRD, 26 W. Martin Luther King Drive. Cincinnati, OH, 45268, USA

Abstract

In this paper, a thin transparent titania (TiO₂) film was coated on the surface of flexible poly(ethylene terephthalate) (PET) film using sol gel method. The surface properties of the obtained TiO₂/PET film were further improved by RF glow discharge oxygen plasma as a function of exposure time and discharge power. The changes in hydrophilicity of TiO₂/PET films were analyzed by contact angle measurements and surface energy. The influence of plasma on the surface of the TiO₂/PET films was analyzed by atomic force microscopy (AFM) as well as the change in chemical state and composition that were investigated by X-ray photo electron spectroscopy (XPS). The cytotoxicity of the TiO₂/PET films was analyzed using human osteoblast cells and the bacterial eradication behaviors of TiO₂/PET films were also evaluated by

against *Staphylococcus* bacteria. It was found that the surface roughness and incorporation of oxygen containing polar functional groups of the plasma treated TiO₂/PET films increased substantially as compared to untreated one. Moreover the increased concentration of Ti³⁺ on the surface of plasma treated TiO₂/PET films was due to the transformation of chemical states (Ti⁴⁺→Ti³⁺). These morphological and chemical changes are responsible for enhanced hydrophilicity of the TiO₂/PET films. Furthermore, the plasma treated TiO₂/PET film exhibited no cytotoxicity against osteoblast cells and antibacterial activity against *Staphylococcus* bacteria which can find application in manufacturing of biomedical devices.

Key words: RF glow discharge oxygen plasma, TiO₂/PET, surface properties, anti-bacterial adhesion, cell compatibility.

*Corresponding author

E-address: dr.knpr@gmail.com, Nadagouda.Mallikarjuna@epamail.epa.gov

Mobile no: +91-9003590874

1.0 Introduction

Currently, polymeric materials are employed in the bio-medical industry for artificial organs, medical devices and disposable clinical apparatus due to their versatility in producing devices with remarkably different mechanical and physical properties. However, the major complications of polymeric materials are the infection of bacteria, when the materials are implanted in the human body. The bacterial adhesion causes a high rate of morbidity and mortality thereby significantly increasing health care costs [1-4]. Hence polymeric materials need some additional treatment to improve their antibacterial and biological properties. As the surface properties of the materials play a vital role in determining their bioactivity and long term performance *in vivo* [5-6]. Several surface modification techniques such as corona discharge, glow discharge plasma treatment, chemical etching, incorporation of silver nano particles, surface-immobilized polyethylene oxide (PEO), surface thiocyanation, mechanical and thermal treatment have been employed to improve the surface and biological properties of the polymeric materials [7-13]. Among them, glow discharge plasma treatment has been found to be an extremely attractive way to modify the surface properties of polymeric materials through surface functionalization, cross linking, degradation or combination of these processes. Furthermore, the glow discharge plasma processes are precisely controllable, conformal and the surface chemistry can be tailored for the required end use. An interesting feature of glow discharge plasma treatment is that the changes are confined to a depth of a few nanometers at the surface because of the low level of penetration and also is a dry reaction in the gas-solid phase. Depending on the gas composition and treatment conditions, ions, electrons, fast neutrals, radicals and UV radiation contribute to the polymer treatment, resulting in enhancement of the surface properties of materials [10-16]. This kind of treatment saves water and energy and does not pollute the environment.

Titanium dioxide (TiO_2) is widely used in biomedical implants such as hip and knee prostheses, heart valves etc. TiO_2 is also used as coatings on eye-glasses, flat panel displays and different optical systems due to its superior mechanical properties, chemical stability, biocompatibility and lack of toxicity. The synthesis method to Produce TiO_2 thin films and the substrate on which it is deposited are known to play an important role for design efficacy of a biomaterial [17-18]. For example TiO_2 films coated on hard surfaces such as glass and stainless steel are generally not suitable for the production of artificial implants [14-19]. Soft matter substrates such as polypropylene (PP), polyethylene (PE), poly (ethylene terephthalate) (PET) and polycarbonate (PC) are increasingly being considered for tissue and organ replacements [20-23]. PET in particular is one of the most significant polymeric materials being used for the production of biomedical materials because of its excellent mechanical properties and moderate biocompatibility. Specially, it is widely adopted in cardiovascular implants, such as artificial heart valves and blood vessels.

In this work, transparent TiO_2 films were coated on the surface of PET polymers by simple sol-gel techniques. The surface properties of obtained TiO_2 /PET films were further improved by radio frequency (RF) glow discharge oxygen plasma as a function of discharge power and exposure time. The films were characterized using contact angle measurements, atomic force microscopy and X-ray photo electron spectroscopy for hydrophilicity, surface morphology and chemical analysis respectively. The influence of surface modification on antibacterial properties was assessed by bacterial eradication tests with *Staphylococcus* bacteria and its cytotoxicity determined on human osteoblast cells.

2.0 Materials and methods

2.1 Materials

Titanium isopropoxide ($\text{Ti}(\text{O}-i\text{-C}_3\text{H}_7)_4$) was purchased from Alfa Aesar, India and biaxial oriented PET films of 100- μm thickness were kindly supplied by Garware Polyester PVT Ltd., India. Other chemicals such as nitric acid, acetone and ethanol were obtained from MERCK, India. PET film was cut into 5 cm X 5 cm pieces and washed ultrasonically in acetone followed by distilled water for one hour and then dried at room temperature before using them for TiO_2 coating and plasma treatment.

2.2 Immobilization of Titania (TiO_2) on PET surface

Transparent TiO_2 films were immobilized on the surface of PET films through titanium isopropoxide sol by simple dip coating technique. Initially, one mole titanium isopropoxide dissolved in nine mole of ethanol was vigorously stirred for 2 h at 0°C and then a mixture of water (1mole) and HNO_3 (0.1 mole) was added drop wise to the above solution with a burette under stirring. The resultant alkoxide solution was kept standing at 45°C temperature for hydrolysis reaction for 36 h, resulting in the TiO_2 sol. PET films were used as the substrate for thin film deposition. Before TiO_2 immobilization, PET substrates were cleaned ultrasonically in distilled water and acetone for 45 min and dried at room temperature. After that the PET film was subjected to oxygen plasma for 15 min, the working pressure and discharge power was 0.2 mbar and 30W respectively for improvement of hydrophilicity of PET films which in turn enhances the adsorption strength of the TiO_2 on the surface of PET films. Finally the oxygen plasma treated PET films were dipped in TiO_2 precursor solution following dipping withdrawing cycles at 25°C and then dried in air at the temperature of 85°C for 24 h resulting TiO_2 was immobilized on the surface of PET films. Subsequently TiO_2 / PET films were washed with

distilled water for 20 min to remove the excess TiO₂ particles from the PET surfaces. The thickness of the obtained TiO₂/PET films was estimated using Mitutoyo SurfTest SJ-301 profilometer and was found to be 350nm. During the thickness measurement, not observed any irregularity and porous structure on the surface of TiO₂/PET films which clearly revealed the compactness and uniformity of same. The obtained TiO₂/PET films were further treated by RF glow discharge oxygen plasma. The plasma reactor consists of plasma chamber made of glass of length 45 cm and diameter of 7.5 cm. The system has two aluminum electrodes having the dimension of 22 cm X 3.5 cm X 0.1cm placed externally at 8.5 cm apart and were capacitively coupled to radio frequency power supply ($\nu = 13.56$ MHz). The sample was kept on the glass sample holder placed inside the chamber. Before plasma treatment, the chamber was initially evacuated to a pressure of $\sim 10^{-3}$ mbar using a rotary pump (EDWARDS) and the same was measured by Pirani gauge. Precursor gas (oxygen) was purged 3 times and the working pressure was adjusted to 0.15 mbar using needle valve. The RF power was applied between the two electrodes to generate stable glow discharge. Finally the samples were treated for different exposure time, discharge power at a fixed pressure level of 0.15 mbar and electrode separation of 8.5 cm.

2.3 Surface analysis

The surface hydrophilicity of the plasma treated TiO₂/PET films was analyzed using sessile drop contact angle measurements [16, 20-21], with three different liquids such as distilled water (W), formamide (F) and ethylene glycol (EG) of known surface tension components (**Table-1**) [15-16]. The error in the measurement was found to be $\pm 2^\circ$. The contact angle measurements were made under controlled room temperature and humidity conditions.

Further, the polar (γ_s^p) and disperse (γ_s^d) components of the TiO₂/PET films were estimated using Fowkes approximation as follows [16, 20-21, 24].

$$\left[\frac{1 + \cos \theta}{2} \right] \times \left[\frac{\gamma_l}{\sqrt{\gamma_l^d}} \right] = \sqrt{\gamma_s^p} \times \sqrt{\frac{\gamma_l^p}{\gamma_l^d}} + \sqrt{\gamma_s^d} \quad (1)$$

Equation (1) is in the form:

$$Y(\text{LHS}) = m.X(\text{RHS}) + C \quad (2)$$

where the value of LHS and RHS could be obtained by contact angle value (θ), surface energy of testing liquids (γ_l), polar (γ_l^p) and disperse (γ_l^d) components of liquid used from **Table 1**. The plot of LHS vs RHS gave a straight line with intercept on Y-axis (Figure-1). Slope and intercept obtained from the plot were squared and added up to give a total solid surface free energy (γ_s) of the TiO₂/PET films as given in equation (3)

$$\gamma_s = \gamma_s^p + \gamma_s^d \quad (\text{mJ/m}^2) \quad (3)$$

The surface morphology of the TiO₂/PET films was evaluated using a Seiko Instruments Scanning force microscopy (AFM, Ben-Yuan, CSPM 4000) operated in tapping mode with horizontal and vertical resolution of 0.26 and 0.10 nm respectively. Change in surface roughness of the plasma treated TiO₂ / PET films was expressed as difference in the root mean square (RMS) of the vertical Z-dimension values within the examined areas, which was calculated using following equation [25].

$$RMS_{xy} = \sqrt{\frac{\sum_{x,y=1}^N (Z_{x,y} - Z_{average})^2}{N}} \quad (4)$$

where $Z_{average}$ is the average Z value within the examined area, $Z_{x,y}$ is the local Z value and N indicates the number of points within the area.

The change in chemical state of the TiO₂/PET films induced by the oxygen plasma treatment was examined using X-ray photo electron spectroscopy (Omicron Surface Science Instruments with EAC2000-125 Energy Analyzer). Monochromatic AlK α X-rays (1486.7 eV) was employed. Typical operating conditions for the X-ray source were a 400- μ m nominal X-ray spot size (full width at half-maximum) at 15 kV, 8.9 mA, and 124 W for both survey and high resolution spectra. The C1s, Ti2p, and O1s envelopes were analyzed and peak-fitted using a combination of Gaussian and Lorentzian peak shapes obtained from XPSPEAK41 software package.

2.4 Bacterial adhesion analysis

The antibacterial activity of the TiO₂/PET films was analyzed using *Staphylococcus* bacteria. Prior to analysis, the PET, TiO₂/PET and plasma processed TiO₂/PET samples were washed with acetone and ethanol and then the samples were dried at room temperature. After this they were immersed in a bacterial suspension which was incubated at 37°C temperature for 24 h. After incubation, the samples were washed with 20 ml of 0.87 % NaCl solution containing polysorbate 80 at a pH of 7. Afterwards the growth of the bacteria on the TiO₂/PET surface was analyzed using Optical microscopy.

2.5 Cytotoxicity analysis

The cytotoxicity of untreated and plasma treated TiO₂/PET films was analyzed using osteoblast cells. Osteoblast cells were cultured in Dulbecco's Modified Eagle's medium supplemented with fetal bovine serum (10 % v/v) in an incubator at 37 °C in humidified atmosphere with 5 % CO₂. Cells were grown till confluence, washed twice with sterile PBS and detached with trypsin; osteoblast cells were counted (using trypan blue to differentiate between viable and no-viable cells) and a solution containing 10⁵ cells/ml was prepared diluting with

PBS. TiO₂/PET films samples (1 x 1 cm) were placed in 24 wells plates and 1 ml of solution containing osteoblast, prepared as described above, was added. Samples were incubated at 37 °C in humidified atmosphere with 5 % CO₂ for two days. After that the samples were removed from cultured medium and washed with PBS twice. The obtained samples were further placed in clean 24-well plate with 1 ml fresh medium. The MTT assay kit (Invitrogen, UK) was used to measure the cell viability of the samples. The MTT solution was prepared as per manufacturer guidance and added to the samples. The samples were incubated for 2 h and then the MTT solubilization solution was added; after dissolution of the crystals, the absorbance of each sample was recorded at 540 nm. The results are presented as the averages and standard deviations of three independent samples.

The cell viability is linked to the activity of the enzyme responsible for the reduction of MTT to purple formazan, the latter concentration is measured spectrophotometrically. The higher the reading, the higher the formazan produced, therefore, the higher the amount of enzyme presented and, consequently, the higher the number of cells.

3.0 Results and discussion

3.1 Hydrophilic analysis: Contact angle and surface energy results

The extent of hydrophilic modification of TiO₂/ PET films was measured by contact angle analysis using three different test liquids such as water, formamide, ethylene glycol. The variations in contact angle with the plasma treatment time at 30 and 45 W are shown in **Table-2**. It was observed that the contact angle value of the PET substrate was 89.6° for distilled water, 80.5° for formamide and 75.2° for ethylene glycol and it decreased considerably after TiO₂ coating (UT sample). The contact angle of TiO₂/PET films with respect to all the three liquids decreased considerably at an exposure time of 5 min; it further decreased with increasing the

exposure time, for a given power level reaching the lowest contact angle value of 22.7°, 17.3° and 14.7° for W, F and EG respectively after 15min of plasma treatment. The lowest contact angle value was observed for the TiO₂/PET film treated under higher power level of 45W compared with 30W of discharge power (**Table-2**). The decrease in the contact angle is an indication of the formation of high magnitude of oxygen containing polar groups onto the surface of TiO₂/PET film due to plasma treatment and also an increase in surface roughness of the same. The PET film used in the present investigation is of 100 micron. The surface modification achieved during plasma treatment is just ~ 10 nm as reported in the literature [23, 26]. It means that in our case one part in ten thousand is modified. Also the power used in the present experiment is very less (30 and 45 W). The electrodes are kept outside the tubular plasma reactor with 8.5 cm distance between them, which is quite a large distance. Therefore, bulk property - the mechanical strength remains unchanged. In the past, we have tested gaseous plasma treated samples randomly for their mechanical properties and have found no adverse effect. Therefore, in the present experiment we have not measured this parameter. The factors, surface roughness and functional changes facilitate the surface of TiO₂/PET film to become highly hydrophilic in nature related to untreated one [16, 21, 26-29].

Changes in polar and dispersive components of the total surface energy of untreated and plasma treated TiO₂/PET films were estimated using Fowkes approximation from contact angle measurements data. The surface energy of the PET film was found to be 26.84 mJ/m², whereas its polar and disperse components were 5.89 mJ/m² and 20.95 mJ/m² respectively and they increased considerably after TiO₂ coating (untreated sample). The surface energy of unmodified TiO₂/PET film was found to be 36.29 mJ/m². The surface energy value increased with increase in plasma treatment time at a given power and reached the maximum value of 66.49 mJ/m² for

the sample (TiO₂/PET) treated at the higher power level of 45 W (**Table-2**). The increase in total surface energy was essentially due to the incorporation of polar components (γ_s^p), whereas, there was no appreciable change in the dispersive component (γ_s^d) of the surface. The above changes may be due to the increase in power level leading to an increase in the degree of ionization of plasma that resulted in an enhancement of the active species concentration. The strong interaction between active species and TiO₂/ PET produces active sites on the materials surface leading to an increase in the formation of the oxygen containing polar groups onto the TiO₂/ PET surfaces. The properties such as wettability, biocompatibility, cell compatibility and anti bacterial strongly depend upon the surface energy of TiO₂/PET films.

3.2 Surface topographical analysis: AFM results

The surface topography of the materials could play a vital role in improving the biocompatibility of an implant. **Figure 2a-e** shows AFM images of TiO₂/PET films as a function of exposure time at an operating power of 30 W. The surface of TiO₂/PET untreated film was relatively smoother and moderate roughness which exhibits the uniform distribution TiO₂ layer on the surface PET film i.e compactness of the TiO₂/PET film as shown in **Figure 2a**. Even a short duration (2 min) of plasma treatment results in rougher morphology. These topographical changes were further increased with increasing exposure times, as can be seen in **Figure 2b-e**. Similar trend was observed when the samples were treated at higher power level of 45 W. However, the samples exhibited enhanced rougher morphology when treated at 45 W which may be due to the magnitude of bombardment of more energetic plasma species on the surface leads to increase in surface roughness of the TiO₂/PET films at the higher discharge power level (**Figure 3b-e**). Moreover RMS value of surface roughness of the untreated TiO₂/PET film was 13.3 nm and gradually increased with increasing the exposure time. However, the maximum

value of RMS surface roughness was obtained at the discharge power of 45 W for all the treatment times compared with the samples treated at the power level of 30 W. This phenomenon is related with the etching process by plasma particles. The results indicated an apparent increase in the surface roughness and effective surface area for contact, causing improvement in the hydrophilicity of the materials.

3.3 Surface composition analysis: XPS results

The elemental composition of the untreated and plasma treated TiO₂/PET films is presented in **Table-3** which mainly contain C1s, O1s and Ti2p contents. It is clear from **Table-3** that carbon content decreases where as there is increase in oxygen and Ti2p content with increase in plasma treatment time. The higher concentration of O1s and Ti2p was obtained for the sample exposed at the higher power level of 45W. The above change revealed that the incorporation of significant amount O1s onto the surface of the TiO₂/PET films when treated by oxygen plasma. **Table-3** also shows the ratio of the O1s/C1s and C/Ti of the untreated and plasma treated TiO₂/PET films. It is clearly seen that the O/C ratio has increased with respect to increase in exposure time which may be attributed to the increase in the newly formed oxygen functional groups by oxygen plasma treatment [30]. Furthermore, the ratio of C/Ti in TiO₂/PET film was decreased steadily with increase in exposure time. The changes may be attributed to removal of the carbon atom from the surface of the plasma treated TiO₂/PET film surface, while increment of oxygen content on the surface. The introduction of oxygen containing functional groups onto the TiO₂/PET surface may be the main reason for improvement of hydrophilic property and biological response.

Information on how the chemical states of TiO₂/PET films were modified as a consequence of oxygen plasma treatment can be obtained from deconvolution of XPS signals using Gaussian-Lorentzian fit. **Figure-4** shows the Ti2p core level of the untreated and plasma treated TiO₂/PET film for different treatment time at 30 and 45 W. The peaks may be identified for untreated TiO₂/PET to be at 454.9 eV due to Ti³⁺2p_{3/2}, 455.8 eV due to Ti⁴⁺2p_{3/2}, at 460.4 eV due to Ti³⁺2p_{1/2} and at 462.4 eV due to Ti⁴⁺2p_{1/2}. The Ti2p XPS spectrum of plasma treated TiO₂/PET were also fitted with four peaks at 455.0, 456.6, 460.2 and 462.2 eV due to Ti³⁺2p_{3/2}, Ti⁴⁺2p_{3/2}, Ti³⁺2p_{1/2} and Ti⁴⁺2p_{1/2} respectively [31-34]. However, the intensity of the peaks due to Ti³⁺2p_{3/2} and Ti³⁺2p_{1/2} was increased by increase in the exposure time whereas there is decrease in the chemical state of Ti⁴⁺ (Ti⁴⁺2p_{3/2} and Ti⁴⁺2p_{1/2}). The percentage contribution of Ti2p (Ti⁴⁺ and Ti³⁺) components of the untreated and plasma treated TiO₂/PET films for different exposure time at 30W power level, as calculated from Ti2p core level spectra, is shown in **Figure 4c**. It can be seen that the surface of the untreated TiO₂/PET films exhibited an higher concentration of Ti⁴⁺ (Ti⁴⁺2p_{3/2} and Ti⁴⁺2p_{1/2}). After plasma treatment at 30W, the concentration of Ti⁴⁺ decreased with increasing exposure time, whilst the concentration of Ti³⁺ (Ti³⁺2p_{3/2} and Ti³⁺2p_{1/2}) on the surface of the TiO₂/PET films (**Figure-4c**) increased. The higher concentration of Ti³⁺ in TiO₂/PET film was achieved when the samples were exposed under higher discharge power of 45W (**Figure-4d**). The above changes may be attributed to desorption of water molecules from the surface of TiO₂/PET films by ions, electrons, neutral species and UV radiation in plasma which can be described an one electron from OH⁻ can transfer to the Ti⁴⁺ state and H⁺ can take one electron from O²⁻ in TiO₂. Hence, the plasma treated surface mainly exhibited Ti³⁺, O⁻ and highly reactive radicals such as H and OH as shown in **Scheme-1**. The increase in Ti³⁺, O⁻ and highly

reactive radicals facilitated the improvement in biocompatibility and antibacterial properties of TiO₂/PET films [25, 27, 31].

In order to investigate the different oxygen states on the surface of TiO₂/PET induced by oxygen plasma treatment, a high-resolution XPS analysis of O1s peaks were performed. **Figure-5a-b** show O1s spectra of the untreated sample; deconvoluted into four peaks at 526.9, 527.9, 529.3 and 525 eV which corresponds to lattice O= in [TiO₂], -O- in [Ti₂O₃], and H₂O/O(C/H)Ti in [TiO(C/H)Ti] and O²⁻ ions in (-Ti-O-Ti) respectively; the spectra of plasma treated films also showed peaks for TiO₂, Ti₂O₃, TiO (C/H) Ti [29-31]. However, the fourth peak due to -Ti-O-Ti almost disappeared when the sample was treated by oxygen plasma. The concentration of each chemical component with O1s can be calculated by deconvolution using Gaussian-Lorentzian fit; the results are shown in **Figure 5c-d**. It is clearly seen that the concentration of the component TiO₂ (Ti⁴⁺ state) and TiO (C/H) Ti decreased with increasing the exposure time at a given power level, whereas Ti₂O₃ (Ti³⁺ state) found to be increased (**Figure-5c**). The higher concentration of Ti₂O₃ is obtained at a power of 45 W as compared to the power level of 30W (**Figure 5d**). Hence, it was expected that higher power level of the plasma treatment has increased TiO₂ active states at the surface. The process of conversion of Ti⁴⁺ state to Ti³⁺ is induced by the plasma treatment that can generate oxygen moieties at the surface and excess electrons in Ti, this causes the change in the electronic structure of TiO₂.

The C1s high-resolution XPS spectra of untreated and plasma treated TiO₂/PET films for different treatment time at 30 W are shown in **Figure-6**. The spectra of untreated TiO₂/PET film indicated the presence of five peaks with binding energy of 283.31 for C-C/C-H, 285.3 for C-C(=O)OX, 286.2 for C-OX, 287.6 for C=O and 288.1eV for C(=O)OX [34] (**Figure 6a**). The high resolution spectra of plasma treated films also showed the peaks at 283.31, 285.3, 286.2,

287.6 and 288.1 eV due to C-C/C-H, C-C(=O)OX, C-OX, C=O and C(=O)OX respectively. It revealed that the intensity of the C-C/C-H component of plasma treated films decreased while the intensity of oxygen containing functional groups such as C-C(=O)OX, C-OX, C=O and C(=O)OX increased with increasing the plasma treatment time. **Figure 6c and d** show the percentage contribution of the C1s components of the plasma treated film. It can be seen that, after the plasma treatment, the C-C groups found to be decreased, while C-C(=O)OX, C-OX, C=O and C(=O)OX were found to be increased with plasma exposure time. This effect was pronounced for the samples exposed under higher discharge power of 45W. These increased oxygen containing functional groups are useful to enhance the biocompatibility and antibacterial properties of the material.

3.4 Bacterial adhesion and Cell compatibility analysis

Figure-7a shows the antibacterial activity of TiO₂/PET films as a function of exposure time at discharge power level of 45W which were analyzed against *staphylococcus* bacteria by the inhibition zone method. It was observed that the higher concentration of staphylococcus bacteria's were adhered and proliferated on the surface of virgin PET film as shown in **Figure 7a**. Comparatively less adhesion and proliferation of bacteria was found on the TiO₂ immobilized PET film surface as shown in **Figure-7b**. Nonetheless, *staphylococcus* bacteria exhibit significantly lower adherence and proliferation on the surface of the oxygen plasma treated TiO₂/PET films (**Figure 7 c-e**) in comparison to the untreated TiO₂/PET and PET and thus 15 min treated TiO₂/PET film surface suppressed growth and adhesion of staphylococcus bacteria dramatically. The inhibition of bacteria is mainly attributed to change in chemical state (Ti⁴⁺ → Ti³⁺), incorporation of highly reactive species such as OH[°], H[°], O⁻ and hydrophilic polar functional groups onto the surface of TiO₂/PET films by oxygen plasma treatment. The above

results are clearly demonstrated that the oxygen plasma treated TiO₂/PET films resist the growth and adhesion of *staphylococcus* bacteria.

The cell compatibility is one of the most significant factors to an initial screening on materials toxicity effects against human cells. Cell viability of the plasma treated and untreated TiO₂/PET films were analyzed by using human osteoblast cells. As indicated in **Figure-8**, the OD₅₄₀ (Absorbance) value of the control TiO₂/PET film is around 0.08 and it increased considerably at the short exposure (2 min) of oxygen plasma. Interestingly, the values do not change significantly with longer exposure times for the power level of 30W. However, maximum OD₅₄₀ value was observed for the TiO₂/PET films samples treated at the higher exposure time and discharge power of 45W, indicating that the plasma treated (at higher power) possessed good biocompatibility compared with that of untreated one (control). Since absorbance values (OD₅₄₀) is proportional to proliferation cells (cell density) on the surfaces. Therefore, Osteoblast cells showed a higher proliferation rate and cell density on plasma treated TiO₂/PET films as compared to untreated one. In other word, cell viability on oxygen plasma treated TiO₂/PET was superior to that on untreated TiO₂/PET films due to formation of high density of functional groups and Ti³⁺ chemical states by oxygen plasma treatment [34-36].

4.0 Conclusion

The transparent TiO₂/PET films were prepared effectively by simple sol gel technique. Oxygen glow discharge RF plasma has been used to improve the surface properties of the obtained TiO₂/PET films. The oxygen plasma treatment incorporated oxygen containing polar functional groups onto the surface of the TiO₂/PET films caused decrease in contact angle and increase in surface energy. The increase in surface energy is mainly due to the incorporation of oxygen containing polar components onto the plasma treated TiO₂/PET film surfaces. The results

from the AFM showed a gradual increase in the surface roughness of plasma treated surfaces with exposure time. The XPS results clearly exhibited that the decrease in carbon content on the plasma treated film surfaces and significant increase in oxygen and Ti2p on the surface of the TiO₂/PET films. Furthermore the XPS analysis identified oxygen containing polar groups on the TiO₂/PET film surfaces and also the increase in the proportion of Ti³⁺ in Ti2p whereas decrease in Ti⁴⁺ state. The increase in surface roughness and concentration of polar functional groups and Ti³⁺ states induced by plasma treatment was observed more for the samples treated at higher power level (45W) and exposure time (15 min). These chemical and morphological changes contributed to enhanced / improved biocompatibility and anti-adhesion bacterial properties of TiO₂/PET films. This plasma based surface engineering technology has great potential for wide applications in biomedical industry.

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Notice

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References

- [1] R.Langer, N.A. Peppas, *AIChE J*, 49(2003)2990.
- [2] A.E.Madkour, G.N. Tew, *Polym. Int.* 1(2008)6.
- [3] W.Zhang, P.K. Chu, J.H. Ji, Y.H. Zhang, X.Y. Liu, R.K.Y. Fu, P.C.T. Ha, Q. Yan, *Biomaterials*, 27(2006) 44.
- [4] W-S.Wang, X. Wang, X. Fu, J. Weng, Wenyue, *Sur. Coat. Techno*, 205(2010) 465.
- [5] D.Sakthi Kumar, M.Fujioka, K.Asano, A.Shoji, A. Jayakrishnan, Y. Yoshida, *J. Mater. Sci. Mater. Med*, 18(2007)1831.
- [6] T. Matsunaga, Y. Ikada, "Modification of Polymers", C. E. Carraher, M. Tsuda, eds., *ACS Symposium Series*, 121(1980) 391.
- [7] C.Yao, X.S. Li, K.G. Neoh, Z.L Shi, E.T. Kang, *J. Membr. Sci*, 320(2008)259.
- [8] Alenka Vesel, Miran Mozetic, *Vacuum*, 86(2012) 634.
- [9] J. Behnisch, A. Hollander, H. Zimmermann, *J Appl Polym Sci*, 49(1993)117.
- [10] S.Bhowmik, P.K. Ghosh, S. Ray, S.K. Barthwal, *J Adhes Sci Technol*, 12(1998)1181.
- [11] M.R. Yang, K.S. Chen, J.C. Tsai, C.C. Tseng, S.F. Lin, *Mater Sci Eng C*, 20(2002)167.
- [12] P.Favia, R.D'Agostino, *Surf Coat Technol*, 98(1998)1102.
- [13] D.J.Balazs, K. Triandafillu, P. Wood, Y. Chevolut, C.V. Delden, H. Harms, C. Hollenstein, H.J Mathieu, *Biomaterials*, 25(2004)2139.

- [14] S.Bhowmik, T.K. Chaki, S. Ray, F. Hoffman, L. Dorn, Metall. Mater. Trans. A, 35 (2004) 865.
- [15] K.N. Pandiyaraj, V. Selvarajan, R.R. Deshmukh, C. Gao, Appl Surf Sci, 255 (2009) 3965.
- [16] R.R. Deshmukh, N.V. Bhat, Mat Res Innovat, 7(2003)283.
- [17] R.Pareek, A.S. Joshi, P.D. Gupta, P.K. Biswas, S. Das, Opt. Laser Technol, 37 (2005) 369.
- [18] J.Szczyrbowski, G. Brauer, G.Teschner, A. Zmelty, Surf. Coat. Technol, 98 (1998) 1460.
- [19] A.Fujishima, T.N. Rao, D.A. Tryk, J.Photochem C, Photobiol, Photochemistry Reviews, 1(2000)1.
- [20] F.M. Fowkes, J Phys Chem, 67 (1963) 2538.
- [21] R.R. Deshmukh, A.R. Shetty, J. Appl Polym Sci, 107(2008) 3707.
- [22] K. Navaneetha Pandiyaraj, J.Heeg, C.Mewes, M. Wienecke, T.Barfels, V. Uthayakumar, Pi Guey Su, Key Eng Mater, 52 (2012)191
- [23] K. Navaneetha Pandiyaraj, V. Selvarajan, R.R. Deshmukh Changyou Gao, Vacuum, 83 (2009) 332
- [24] N.V.Bhat, D.J. Upadhyay, R.R. Deshmukh, S.K Gupta, J Phys Chem, 107(2003) 4550.
- [25] E.Selli, G. Mazzone, C.Oliva, F. Martin, C. Riccardi, R.Barni, B. Marcandalli, M.R. Massafra, J Mater Chem, 11(2001)1985.
- [26] T.Oktem, N. Seventekin, H. Ayhan, E. Piskin, Indian J Fibre . Text. Res, 27(2002) 161
- [27] B.Gupta, J. Hilborn, Ch. Hollenstein, C.J.G. Plummer, R.Houriet, N. Xanthopoulos, J.Appl Polym Sci, 78(2000)1083.
- [28] R. N. Wenzel, Ind. Eng. Chem, 28 (1936) 988.
- [29] Mittal, K. L., Ed. Contact Angle, Wettability and Adhesion; VSP: The Netherlands, 2003.
- [30] M. Modic, I.Junkar, Alenka Vesel, Miran Mozetic, Surf.Coat.Technol, 213 (2012) 98

- [31] J.Jun, M.Dhayal, Joong-Hyeok Shin, Jo-Chun Kim, Nikola Getoff, Radiation Phy . Chem, 75(2006)583.
- [32] J.Jun, J-H.Shin, Marshal Dhayal, Appl Surf Sci, 252(2006)3871.
- [33] P.Zhang, B.K. Tay, Y.B. Zhang, S.P. Lau, K.P. Yung, Surf. Coat. Technol, 198 (2005)184.
- [34] G.Beamson, D.Briggs, High Resolution XPS of Organic Polymers, The Scienta ESCA300 Database, Wiley, Chichester, 1992.
- [35] C. Ohtsuki, H. Iida, S. Nakamura, A. Osaka, J. Biomed. Mater. Res, 35 (1997) 39.
- [36] C.Brunot, L.Ponsonnet, Christelle Lagneau, Pierre Farge, Catherine Picart, Brigitte Grosogeat , Biomaterials, 28 (2007) 632.

Figure Captions

Figure-1 Plot of Fowkes equation to estimate solid surface free energy

Figure-2 AFM images of the O₂ plasma treated TiO₂/PET films (a) Untreated, (b) 2 (c) 5 (d) 10 and (e) 15min at the power level of 30 W

Figure-3 AFM images of the O₂ plasma treated TiO₂/PET films (a) Untreated, (b) 2min (c) 5min (d) 10min and (e) 15min at the power level of 45 W

Figure-4 XPS Ti2p spectra and contribution of Ti2p components of O₂ plasma-treated TiO₂/PET film surfaces for different power level of 30 W (a&c) and 45W(b&d)

Figure-5 XPS O1s spectra and contribution of O1s components of O₂ plasma-treated TiO₂/PET film surfaces for different power level of 30 W (a&c) and 45W(b&d)

Figure-6 XPS C1s spectra and contribution of C1s components of O₂ plasma-treated TiO₂/PET film surfaces for different power level of 30 W (a&c) and 45W(b&d)

Figure-7 Optical photograph showing adhesion and proliferation of staphylococcus on plasma treated TiO₂/PET film (a) Control PET (b) Untreated, (c) 5 min (c) 10min and (d) 15 min at the power level of 45W

Figure-8 Osteoblast cell adhesion on plasma treated TiO₂ /PET films as a function of exposure time

Table Captions

Table-1 Surface energy of the testing liquids

Table-2 Variation of contact angle and surface energy with time for plasma treated TiO₂/PET films at different power levels.

Table-3 The surface elemental composition and ratio of the plasma modified TiO₂/ PET film for different exposure times

Scheme

Scheme-1 Mechanism of plasma treatment on the surface of TiO₂/PET films